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(54) **TISSUE TARGETED COMPLEMENT MODULATORS**

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(58) **Field of Classification Search**
None
See application file for complete search history.

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(57) **ABSTRACT**

Systemic suppression of the complement system has been shown to be effective to treat inflammatory disease, yet at the potential cost of compromising host defense and immune homeostasis. Herein disclosed are methods for antigen-specific targeting of complement inhibitors that show that complement inhibitors targeted to the proximal tubular epithelium protect against tubulointerstitial injury and renal dysfunction in a rat model of nephrosis. It is shown that appropriate targeting of a systemically administered complement inhibitor to a site of disease markedly enhances efficacy and obviates the need to systemically inhibit complement. Additionally, it is shown by specifically inhibiting the terminal pathway of complement, that the membrane attack complex (MAC) plays a key role in proteinuria-induced tubulointerstitial injury, thus establishing the MAC as a valid target for pharmacological intervention in proteinuric disorders. The disclosed compositions can be used in methods of treating pathogenic diseases and inflammatory conditions by modulating the complement system.

5 Claims, 6 Drawing Sheets

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Fig. 1

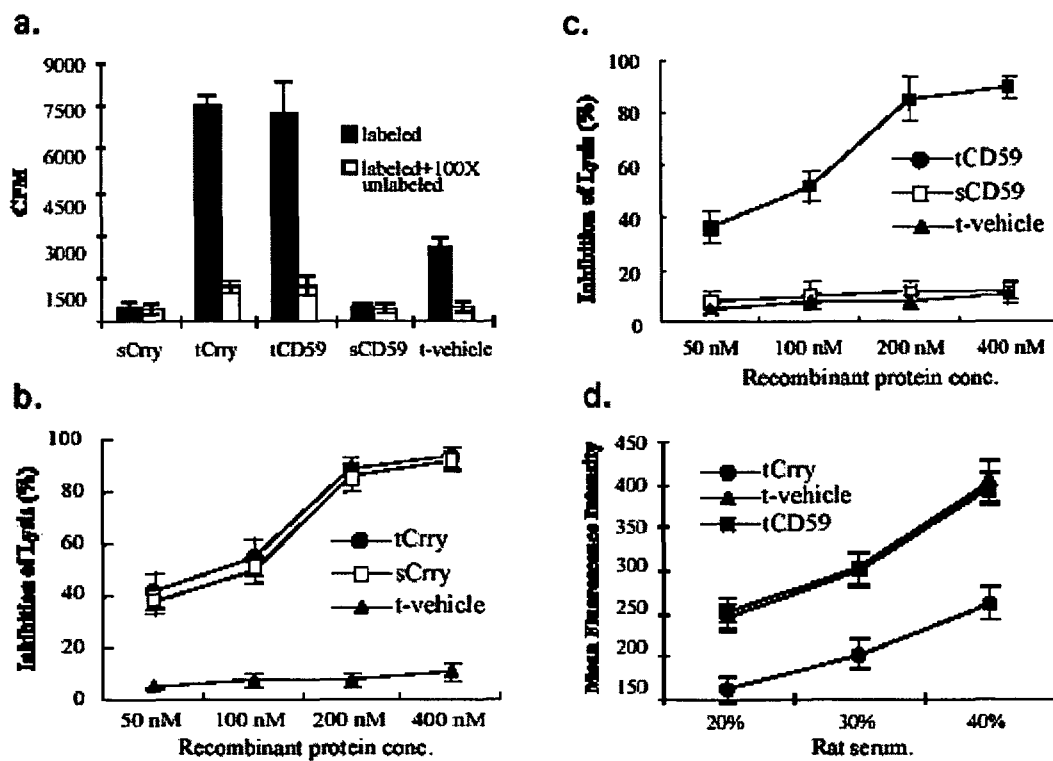


Fig. 2

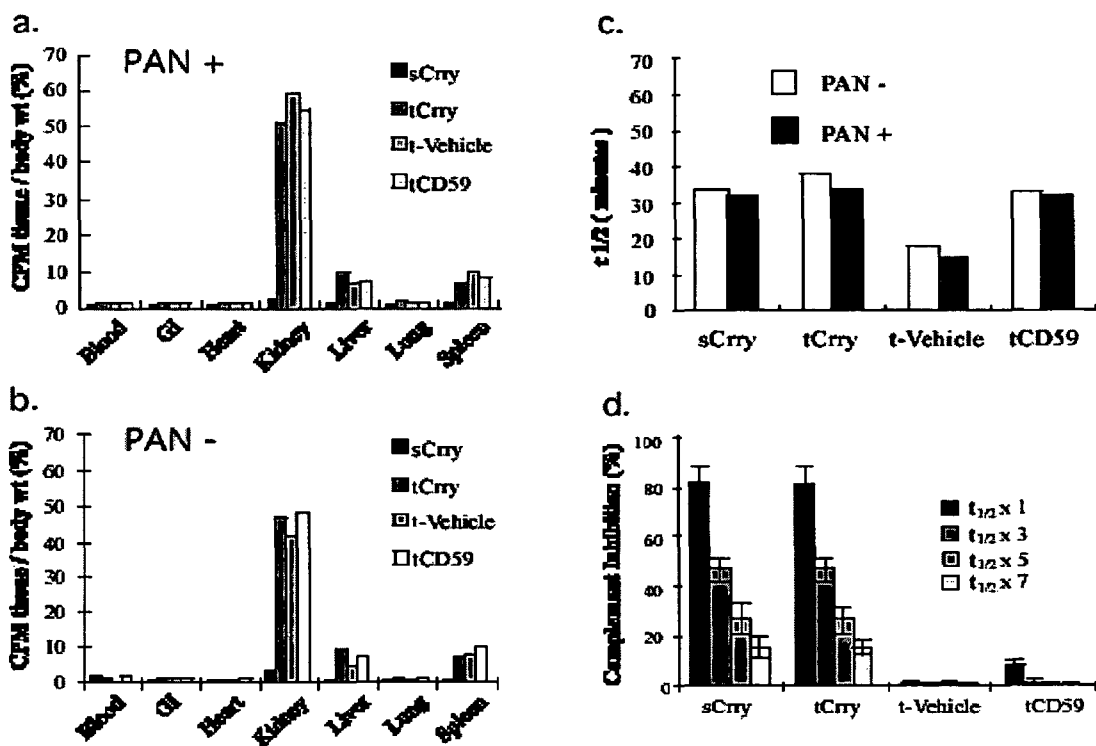
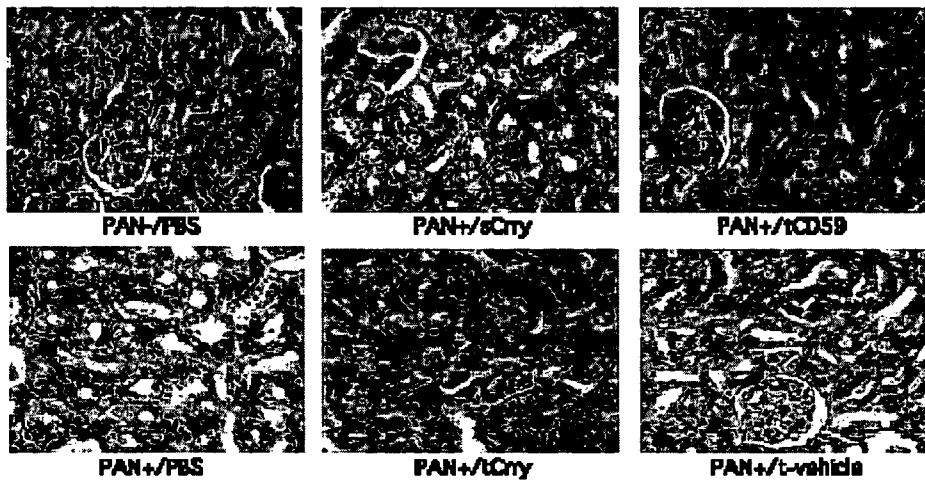
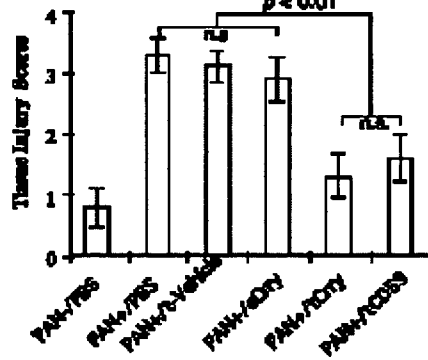


Fig. 3

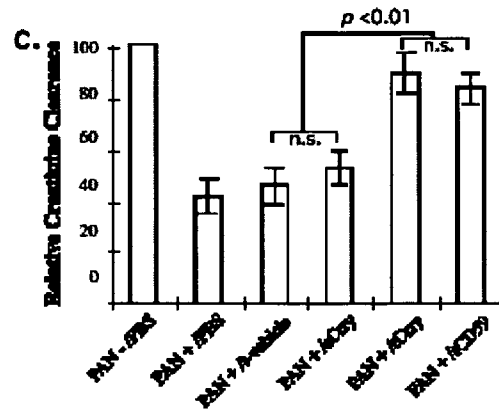
a.



b.



c.



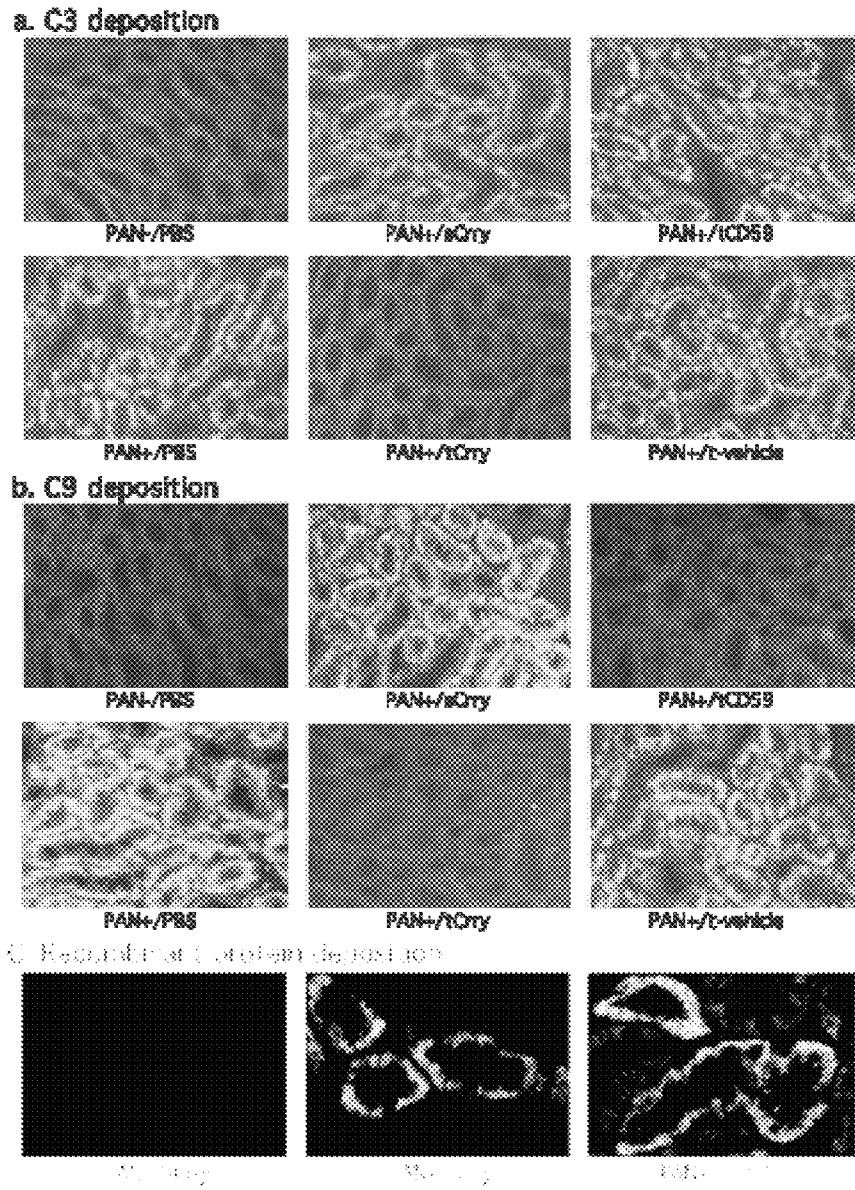


Fig. 4

Fig. 5

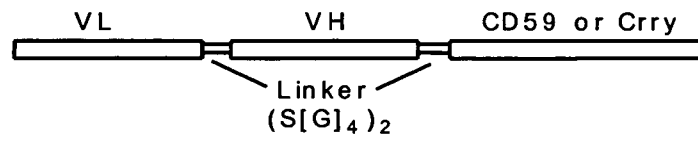
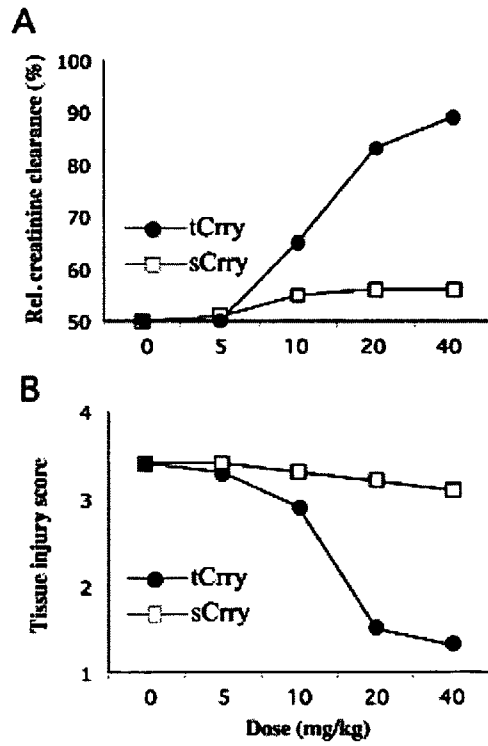


Fig. 6



TISSUE TARGETED COMPLEMENT MODULATORS

This application is a continuation-in-part of U.S. application Ser. No. 10/534,772, which is a national stage application of International Application No. PCT/US03/36459, filed Nov. 13, 2003, which is incorporated by reference herein in its entirety. This application also claims benefit of U.S. Provisional Application No. 60/565,907, filed Apr. 28, 2004, which is incorporated herein by reference in its entirety.

This invention was made with government support under National Institutes of Health Grants AI34451 and DK41873. The government has certain rights in the invention.

I. BACKGROUND OF THE INVENTION

Complement is the collective term for a series of blood proteins and is a major effector mechanism of the immune system. Complement activation and its deposition on target structures can lead to direct complement-mediated cell lysis, or can lead indirectly to cell or tissue destruction due to the generation of powerful modulators of inflammation and the recruitment and activation of immune effector cells. Complement activation products that mediate tissue injury are generated at various points in the complement pathway. Inappropriate complement activation on host tissue plays an important role in the pathology of many autoimmune and inflammatory diseases, and is also responsible for many disease states associated with bioincompatibility, e.g. post-cardiopulmonary inflammation and transplant rejection. Complement inhibition represents a potential therapeutic modality for the treatment of such immune-mediated diseases and disease states. Complement inhibitory proteins that systemically inhibit complement have been shown to be effective in various animal models of disease (and in a few clinical trials), but complement inhibitors that target a site of disease and complement activation offer significant potential advantages with regard to safety and efficacy.

In healthy individuals, complement deposition on host cell membranes is prevented by complement inhibitory proteins expressed at the cell surface. These complement inhibitory proteins are also expressed on the surface of tumor cells, often at increased levels, and are considered to be an important contributing factor to the resistance of tumor cells to monoclonal antibody-mediated immunotherapy (monoclonal antibodies that target to tumor cells and activate complement).

The complement system comprises a collection of about 30 proteins and is one of the major effector mechanisms of the immune system. The complement cascade is activated principally via either the classical (usually antibody-dependent) or alternative (usually antibody-independent) pathways. Activation via either pathway leads to the generation of C3 convertase, which is the central enzymatic complex of the cascade. C3 convertase cleaves serum C3 into C3a and C3b, the latter of which binds covalently to the site of activation and leads to the further generation of C3 convertase (amplification loop). The activation product C3b (and also C4b generated only via the classical pathway) and its breakdown products are important opsonins and are involved in promoting cell-mediated lysis of target cells (by phagocytes and NK cells) as well as immune complex transport and solubilization. C3/C4 activation products and their receptors on various cells of the immune system are also important in modulating the cellular immune response. C3 convertases participate in the formation of C5 convertase, a complex that cleaves C5 to yield C5a and C5b. C5a has powerful proinflammatory and chemotactic properties and can recruit and activate immune

effector cells. Formation of C5b initiates the terminal complement pathway resulting in the sequential assembly of complement proteins C6, C7, C8 and (C9)_n to form the membrane attack complex (MAC or C5b-9). Formation of MAC in a target cell membrane can result in direct cell lysis, but can also cause cell activation and the expression/release of various inflammatory modulators.

There are two broad classes of membrane complement inhibitor; inhibitors of the complement activation pathway (inhibit C3 convertase formation), and inhibitors of the terminal complement pathway (inhibit MAC formation). Membrane inhibitors of complement activation include complement receptor 1 (CR1), decay-accelerating factor (DAF) and membrane cofactor protein (MCP). They all have a protein structure that consists of varying numbers of repeating units of about 60-70 amino acids termed short consensus repeats (SCR) that are a common feature of C3/C4 binding proteins. Rodent homologues of human complement activation inhibitors have been identified. The rodent protein Crry is a widely distributed inhibitor of complement activation that functions similar to both DAF and MCP. Rodents also express DAF and MCP, although Crry appears to be functionally the most important regulator of complement activation in rodents. Although there is no homolog of Crry found in humans, the study of Crry and its use in animal models is clinically relevant.

Control of the terminal complement pathway and MAC formation in host cell membranes occurs principally through the activity of CD59, a widely distributed 20 kD glycoprotein attached to plasma membranes by a glucosylphosphatidylinositol (GPI) anchor. CD59 binds to C8 and C9 in the assembling MAC and prevents membrane insertion.

Various types of complement inhibitory proteins, including several that are based on soluble forms of membrane complement inhibitors, are currently under investigation for therapy of inflammatory and ischemic disease and disease states associated with bioincompatibility (reviewed in refs (9-11)). Almost all previous therapeutic studies (in animal models and in the clinic) have been performed with systemic complement inhibitors, even though it is recognized that systemic suppression of the complement system is likely to compromise host defense and immune homeostasis (9, 12, 13) Systemic inhibition at the C3 step is particularly undesirable due to important physiological functions of C3 and C5 activation products. In conditions where C3 and C5 activation products are also involved in disease pathogenesis, we hypothesize that appropriate targeting of a complement inhibitor that functions early in the pathway (eg. Crry) will minimize systemic inhibition while maintaining an effective local concentration. On the other hand, if the MAC plays a critical role in pathogenesis, complement inhibition late in the pathway (eg. by CD59) will be advantageous since the generation of C3 and C5 activation products will not be altered. However, soluble CD59 is not an effective inhibitor and specific inhibition of the MAC following systemic administration has not been accomplished *in vivo*.

II. SUMMARY OF THE INVENTION

In accordance with the purposes of this invention, as embodied and broadly described herein, this invention, in one aspect, relates to tubule targeted modulators of complement activity.

Additional advantages of the invention will be set forth in part in the description which follows, and in part will be obvious from the description, or can be learned by practice of the invention. The advantages of the invention will be realized

and attained by means of the elements and combinations particularly pointed out in the appended claims. It is to be understood that both the foregoing general description and the following detailed description are exemplary and explanatory only and are not restrictive of the invention, as claimed.

III. BRIEF DESCRIPTION OF THE DRAWINGS

The accompanying drawings, which are incorporated in and constitute a part of this specification, illustrate several embodiments of the invention and together with the description, serve to explain the principles of the invention.

FIG. 1 shows the *in vitro* characterization of targeted complement inhibitors. (a) Binding of iodinated recombinant proteins to rat kidney proximal tubular epithelial cells (PTEC) in the absence and presence of 100-fold excess of unlabeled recombinant protein (mean+/-SD, n=3). (b) and (c) Inhibition of complement mediated cell lysis by recombinant complement inhibitors, Crry and CD59. Antibody sensitized rat PTEC were preincubated with recombinant proteins followed by the addition of 10% normal rat serum. Cell lysis was determined after 40 min at 37° C. Percent inhibitory activity relative to 100% lysis control group shown (mean+/-SD, n=4). (d) Effect of recombinant proteins on C3 deposition on rat PTEC. Antibody sensitized rat PTEC were incubated with the indicated protein, the cells washed and then incubated with C6-deficient rat serum. C3 deposition was determined by flow cytometry (mean+/-SD, n=3).

FIG. 2 shows the *in vivo* characterization of targeted complement inhibitors. (a) and (b) Biodistribution of iodinated recombinant proteins in rats treated with (a) or without (b) puromycin aminonucleoside (PAN). Biodistribution was determined 48 h after tail vein injection. Average of two determinations shown. (c) Blood clearance of the recombinant proteins in rats. Iodinated proteins were injected *i.v.* into rats treated with or without PAN and blood samples taken periodically from the tail vein for radioactive counting and half life determination ($t_{1/2}$). (d) Complement inhibitory activity in rat serum after administration of recombinant proteins. Rat serum samples were obtained periodically following injection of 40 mg/kg recombinant protein and complement inhibitory activity of serum determined using sensitized sheep erythrocytes as targets. Percent inhibitory activity relative to serum from PBS-treated rats is shown (mean+/-SD, n=3).

FIG. 3 shows the histological and functional assessment of tubulointerstitial injury. (a) PAS staining of kidney sections from different treatment groups. Representative images are shown. (b) semiquantitative analysis of tubulointerstitial injury as assessed by tubular dilation and degeneration. (c) Creatinine clearance in proteinuric rats treated with recombinant complement inhibitors. Percent clearance relative to clearance in healthy rats (PAN-/PBS) is shown (mean+/-SD, n=4).

FIG. 4 shows the immunofluorescence microscopy showing C3 deposition (a) C9 deposition (b) and complement inhibitor binding to tubular cells (c). Sections were prepared from kidneys isolated from rats treated with recombinant complement inhibitors. Representative images are shown.

FIG. 5 shows the single chain antibody CD59 or Crry construct. The figure shows the construct comprises a variable light chain (VL) and a variable heavy chain (VH) from K9/9 mAb. The construct was prepared in the yeast expression vector pPICZalpha (Invitrogen).

FIG. 6 shows the dose response of targeted and untargeted Crry in proteinuric rats. Effect of tCrry and sCrry on creati-

nine clearance (A), relative to clearance in healthy rats, and tubulointerstitial injury (B) as assessed by tubular dilation and degeneration. Injury was defined using a scale of 0-4 as defined in *Materials and Methods*. Shown is the average of two determinations.

IV. DETAILED DESCRIPTION

The present invention can be understood more readily by reference to the following detailed description of preferred embodiments of the invention and the Examples included therein and to the Figures and their previous and following description.

Before the present compounds, compositions, articles, devices, and/or methods are disclosed and described, it is to be understood that this invention is not limited to specific synthetic methods, specific recombinant biotechnology methods unless otherwise specified, or to particular reagents unless otherwise specified, as such can, of course, vary. It is also to be understood that the terminology used herein is for the purpose of describing particular embodiments only and is not intended to be limiting.

A. Definitions,

As used in the specification and the appended claims, the singular forms "a," "an" and "the" include plural referents unless the context clearly dictates otherwise. Thus, for example, reference to "a pharmaceutical carrier" includes mixtures of two or more such carriers, and the like.

Ranges can be expressed herein as from "about" one particular value, and/or to "about" another particular value. When such a range is expressed, another embodiment includes from the one particular value and/or to the other particular value. Similarly, when values are expressed as approximations, by use of the antecedent "about," it will be understood that the particular value forms another embodiment. It will be further understood that the endpoints of each of the ranges are significant both in relation to the other endpoint, and independently of the other endpoint.

In this specification and in the claims which follow, reference will be made to a number of terms which shall be defined to have the following meanings:

"Optional" or "optionally" means that the subsequently described event or circumstance may or may not occur, and that the description includes instances where said event or circumstance occurs and instances where it does not.

"Treatment" or "treating" means to administer a composition to a subject with a condition, wherein the condition can be any pathogenic disease, autoimmune disease, cancer or inflammatory condition. The effect of the administration of the composition to the subject can have the effect of but is not limited to reducing the symptoms of the condition, a reduction in the severity of the condition, or the complete ablation of the condition.

Herein, "inhibition" or "inhibits" means to reduce activity. It is understood that inhibition can mean a slight reduction in activity to the complete ablation of all activity. An "inhibitor" can be anything that reduces activity.

Herein, "activation" or "activates" means to increase activity. It is understood that activation can mean an increase in existing activity as well as the induction of new activity. An "activator" can be anything that increases activity.

B. Complement Inhibiting and Activating Constructs

The present disclosure establishes the use of targeted complement inhibitors *in vivo* and demonstrate important benefits of targeted versus untargeted systemic complement inhibition in a model of disease. Disclosed are compositions

comprising a construct, wherein the construct comprises renal tubule targeted antibody and a modulator of complement activity.

K9/9 is a single chain anti-proximal tubule antibody. The antibody is specific for dipeptidylpeptidase IV (DPPIV) that is expressed on rat glomerular epithelial cells and PTEC.

It is understood and herein contemplated that other proximal tubule targeting vehicles can be used in the methods and compositions disclosed to target complement modulators. Although a K9/9 targeting vehicle is used in these studies, the same principles apply for other targeting vehicles. Examples of other targeting moieties include but are not limited to antibodies to the Human homologue of K9/9 antigen, dipeptidylpeptidase IV (DPPIV) (SEQ ID NO: 13), Lrp2 (megalin), Cubn (cubilin), Abcc2 (ATP binding cassette, sub-family C, member 2), Abcc4 (ATP binding cassette, sub-family C, member 4), Abcb1b (ATP binding cassette, sub-family B, member 1; P-glycoprotein), Slc1a1 (excitatory amino acid carrier 1), Slc3a1 (cystine, dibasic and neutral amino acid transporters), Slc5a1 (sodium/glucose cotransporter 1), Slc5a2 (sodium/glucose cotransporter 2), Slc9a3 (sodium/hydrogen exchanger 3), Slc10a2 (sodium/taurocholate cotransporting polypeptide), Slc13a2 (sodium dependent dicarboxylate cotransporter), Slc15a1 (oligopeptide transporter 1), Slc15a2 (oligopeptide transporter 2), Slc17a1 (sodium phosphate transporter 1), Slc17a2 (sodium phosphate transporter 3), Slc17a3 (sodium phosphate transporter 4), Slc1a1 (organic anion transporter protein 1), Slc22a4 (organic cation transporter OCTN1), Slc22a5 (organic cation transporter OCTN2), Slc22a11 (organic anion transporter 4), Slc34a1 (sodium phosphate transporter IIa), megalin (low density lipoprotein receptor-related protein 2, LRP2), neutral endopeptidase (NEP), CD10, mucin 20 (or other mucins) (SEQ ID NO: 14), kidney-injury molecule 1 (KIM-1) (SEQ ID NO: 15) or hepatitis A virus cellular receptor 1 and megalin. Additionally one of skill in the art understands that certain advantages may exist that will aid in determining a preference for a particular targeting moiety. For example, the last four targeting moieties have high expression in the apical membrane of the PTEC, their protein structure is known, and, in the case of the latter two, are highly expressed in states in which the PTEC is injured. Thus, for example, disclosed and herein contemplated are compositions of the invention comprising anti-human DPPIV antibody or fragment thereof linked to CD59.

It is understood that species and strain variations exist for the disclosed peptides, polypeptides, proteins, protein fragments and compositions. Specifically disclosed are all species and strain variations for the disclosed peptides, polypeptides, proteins, protein fragments and compositions.

Also disclosed are compositions, wherein the construct is a fusion protein.

Herein a "fusion protein" means two or more components comprising peptides, polypeptides, or proteins operably linked. K9/9 can be linked to complement inhibitors or activators by an amino acid linking sequence. Examples of linkers are well known in the art. Examples of linkers can include but are not limited to (Gly₄Ser)₂, (Gly₄Ser)₃ (G4S), (Gly₃Ser)₄ (G3S), SerGly₄, and SerGly₄SerGly₄. Linking sequences can also consist of "natural" linking sequences found between SCR units within human (or mouse) proteins, for example VSVFPLE, the linking sequence between SCR 2 and 3 of human CR2, can be used to link the inhibitors of the invention with K9/9. Fusion proteins can also be constructed without linking sequences. Fusion proteins are also referred to as "recombinant proteins" herein.

Also disclosed are compositions of the invention, wherein the fusion protein inhibits complement.

Also disclosed are compositions of the invention, wherein the modulator of complement activity comprises a complement inhibitor.

Also disclosed are compositions of the invention wherein; for example, the complement inhibitor is decay accelerating factor (DAF), for example SEQ ID NO: 2 (amino acid) encoded by SEQ ID NO: 1 (nucleotide). For example, the DAF can be soluble human DAF comprising the four SCR domains without glycoposphatidyl anchor and serine-threonine rich region. The DAF can also be soluble human DAF comprising the four SCR domains and the serine-threonine rich region but without glycoposphatidyl anchor.

The DAF extracellular region consists of 4 SCR units at N-terminus followed by serine/threonine rich region. Amino acids 1-34 comprise the leader peptide, amino acids 35-95 comprise SCR1, amino acids 97-159 comprise SCR2, amino acids 162-221 comprise SCR3, amino acids 224-284 comprise SCR4, and amino acids 287-356 comprise the S/T region. Thus specifically disclosed herein are compositions of the invention wherein the composition comprises all 4 SCR units. Also disclosed herein are compositions of the invention, wherein the composition comprises for example, SCR2-4 of DAF.

Disclosed are compositions of the invention, wherein the complement inhibitor comprises a fusion protein between CD59 and another complement inhibitor selected from the group consisting of DAF, MCP, Crry, and CR1. Also disclosed are compositions of the invention, wherein the complement inhibitor is a fusion protein of two or more complement inhibitors.

Also disclosed are compositions of the invention, wherein the fusion protein comprises K9/9-DAF. Also disclosed are compositions of the invention wherein the fusion protein is encoded by a nucleic acid comprising K9/9-DAF encoding sequences.

Also disclosed are compositions of the invention, wherein the complement inhibitor is human CD59 (SEQ ID NO: 3 (nucleotide) and SEQ ID NO: 4 (amino acid)). The human CD59 can be soluble human CD59 comprising the mature protein without glycoposphatidyl anchor.

Also disclosed are compositions of the invention, wherein the fusion protein comprises K9/9-CD59 (SEQ ID NO: 12). Also disclosed are compositions of the invention wherein the fusion protein is encoded by a nucleotide.

Also disclosed are compositions of the invention, wherein the complement inhibitor is CR1 (SEQ ID NO: 5 (nucleotide) and SEQ ID NO: 6 (amino acid)). The extracellular region of CR1 can comprise 30 SCR units. It is an embodiment of the invention that the composition can comprise the entire extracellular region of CR1. In another embodiment of the invention, the composition comprises [the] one active site[s] of CR1. The active sites of CR1 are amino acids 1-46 which comprise the leader peptide, amino acids 47-300 which comprise SCR1-4 (C4b binding site, lower affinity for C3b), amino acids 497-750 which comprise SCR8-11 (C3b binding site, lower affinity for C4b), amino acids 947-1200 which comprise SCR15-18 (C3b binding site, lower affinity for C4b), and amino acids 1400-1851 which comprise the C1q binding site. In an additional embodiment of the invention, the composition of the invention can comprise any [one or] combination or all of the active sites of CR1.

Also disclosed are compositions of the invention, wherein the complement inhibitor comprises the active sites of CR1, and wherein [the] one active site[s] further comprise a leader peptide comprising amino acids 6-46, amino acids 47-300

which comprise SCR1-4 (C4b binding site, lower affinity for C3b), amino acids 497-750 which comprise SCR8-11 (C3b binding site, lower affinity for C4b), amino acids 947-1200 which comprise SCR15-18 (C3b binding site, lower affinity for C4b), and amino acids 1400-1851 which comprise the C1q binding site. In an additional embodiment of the invention, the composition of the invention can comprise any [one or] combination or all of the active sites of CR1.

Also disclosed are compositions of the invention, wherein the complement inhibitor is MCP (SEQ ID NO: 7 (nucleotide) and SEQ ID NO: 8 (amino acid)). The extracellular region consists of 4 SCR units followed by ser/thr region. Amino acids 1-34 comprise the leader peptide, amino acids 35-95 comprise SCR1, amino acids 96-158 comprise SCR2, amino acids, 159-224 comprise SCR3, amino acids 225-285 comprise SCR4, and amino acids 286-314 comprise the S/T region

Also disclosed are compositions of the invention, wherein the complement inhibitor is Crry (SEQ ID NO: 9). The Crry can be soluble mouse Crry comprising the 5 N-terminal SCR domains without transmembrane region.

Also disclosed are compositions of the invention, wherein the fusion protein comprises K9/9-Crry (SEQ ID NO: 11). Also disclosed are compositions of the invention wherein the fusion protein is encoded by a nucleotide.

Also disclosed are compositions of the invention, wherein the complement inhibitor is murine CD59. The murine CD59 can be soluble murine CD59 comprising the mature protein without glycoposphatidyl anchor.

It is understood and herein disclosed that instances can exist where the administration of a composition comprising a single fusion protein or immunoconjugate may be improved upon through the administration of multiple fusion proteins or immunoconjugates. Therefore, disclosed are compositions of the invention comprising more than one fusion protein or immunoconjugate of the invention. The compositions with more than one fusion protein can include fusion proteins chosen from the list of fusion proteins described herein, for example in Example 5. As a specific example, a composition comprising K9/9-CD59 fusion protein and K9/9-Crry fusion protein is provided. It is understood that compositions comprising more than one fusion protein or immunoconjugate can be administered such that all fusion proteins or immunoconjugates are administered concurrently. This would include but not be limited to, for example, the administration of a single composition comprising more than one fusion construct, and the administration of two separate fusion constructs concurrently. Also disclosed are methods of the invention comprising administering the compositions of the invention sequentially. For example, a K9/9-Crry fusion is administered followed by a K9/9-CD59 fusion. Alternatively, a K9/9-CD59 fusion is administered first, followed by a K9/9-Crry fusion.

Disclosed are compositions of the invention, wherein the construct is in a vector.

Disclosed are cells comprising the vector of the invention.

Also disclosed are compositions, wherein the construct is an immunoconjugate. Herein "immunoconjugate" means two or more components comprising peptides, polypeptides, or proteins, one of which is an immunoreactive component, operably linked by a chemical cross-linker. Linking of the components of the immunoconjugate can occur on reactive groups located on the component. Reactive groups that can be targeted using a cross-linker include primary amines, sulfhydryls, carbonyls, carbohydrates and carboxylic acids, or active groups can be added to proteins. Examples of chemical linkers are well known in the art and can include but are not

limited to bismaleimidohexane, m-maleimidobenzoyl-N-hydroxysuccinimide ester, NHS-Esters-Maleimide Crosslinkers such as MBS, Sulfo-MBS, SMPB, Sulfo-SMPB, GMBS, Sulfo-GMBS, EMCS, Sulfo-EMCS; Imidoester Crosslinkers such as DMA, DMP, DMS, DTBP; EDC [1-Ethyl-3-(3-Dimethylaminopropyl)carbodiimide Hydrochloride], [2-(4-Hydroxyphenyl)ethyl]-4-N-maleimidomethyl)-cyclohexane-1-carboxamide, DTME: Dithio-bismaleimidoethane, DMA (Dimethyl adipimidate•2 HCl), DMP (Dimethyl pimelimidate•2 HCl), DMS (Dimethyl suberimidate•2 HCl), DTBP (Dimethyl 3,3'-dithiobispropionimidate•2HCl), MBS, (m-Maleimidobenzoyl-N-hydroxysuccinimide ester), Sulfo-MBS (m-Maleimidobenzoyl-N-hydroxysuccinimide ester), Sulfo-SMPB (Sulfosuccinimidyl 4-[p-maleimidophenyl]butyrate), GMBS (N-[•-maleimidobutyryloxy]succinimide ester), EMCS(N-[•-maleimidocaproyloxy]succinimide ester), and Sulfo-EMCS(N-[•-maleimidocaproyloxy]sulfosuccinimide ester).

C. Methods of Using the Compositions

Various types of complement inhibitory proteins are currently under investigation for therapy of inflammatory disease and disease states associated with bioincompatibility.

Disclosed are methods of treating a condition affected by complement in a subject comprising administering to the subject the composition of the invention. It is understood that administration of the composition to the subject can have the effect of, but is not limited to, reducing the symptoms of the condition, a reduction in the severity of the condition, or the complete ablation of the condition.

1. Methods of Using the Compositions to Inhibit Complement

Disclosed are methods of treating a condition affected by complement in a subject comprising administering to the subject the composition of the invention, wherein the composition will inhibit complement activity. It is understood that the effect of the administration of the composition to the subject can be, but is not limited to, reducing the symptoms of the condition, a reduction in the severity of the condition, or the complete ablation of the condition.

Disclosed are methods of reducing complement-mediated damage comprising administering to a subject the composition of the invention, which inhibits complement.

Disclosed are methods of the invention, wherein the condition treated is an inflammatory condition, an ischemic condition or a condition relating to bioincompatibility. Also disclosed are methods of the invention, wherein the inflammatory condition can be selected from the group consisting of nephritis, Proteinuria, Diabetic nephropathy, Focal and segmental glomerulosclerosis, Membranous nephropathy, IgA nephropathy, Lupus nephritis, Minimal change disease, Amyloidosis, Membranoproliferative glomerulonephritis, Essential mixed cryoglobulinemia (includes secondary to hepatitis C), Light chain deposition disease, Vasculitis (includes Wegener's granulomatosis, microscopic polyangiitis and renal limited vasculitis), Congenital nephrotic syndrome, Fibrillary glomerulonephritis, Mesangial proliferative, glomerulonephritis, Postinfectious glomerulonephritis, Drug-induced nephrotic syndrome, Chronic allograft rejection, Preeclampsia/eclampsia, Hypertensive nephrosclerosis, and Immunotactoid glomerulonephritis.

Disclosed are methods of the invention, wherein complement inhibitor can enhance the outcome of apoptosis-base therapy (e.g., gene therapy with adenovirus expressing Fas ligand).

Apoptosis occurring during normal development is non inflammatory and is involved in induction of immunological tolerance. Although apoptotic cell death can be inflammatory

depending on how it is activated and in what cell types (for example, therapeutic agents that ligate Fas are able to induce inflammation), necrotic cell death results in a sustained and powerful inflammatory response mediated by released cell contents and by proinflammatory cytokines released by stimulated phagocytes. Apoptotic cells and vesicles are normally cleared by phagocytes, thus preventing the pro-inflammatory consequences of cell lysis. In this context, it has been shown that apoptotic cells and apoptotic bodies directly fix complement, and that complement can sustain an anti-inflammatory response due to opsonization and enhanced phagocytosis of apoptotic cells.

Inflammation is involved in non specific recruitment of immune cells that can influence innate and adaptive immune responses. Modulating complement activation during apoptosis-based tumor therapy to inhibit phagocytic uptake of apoptotic cells/bodies enhances the inflammatory/innate immune response within the tumor environment. In addition, apoptotic cells can be a source of immunogenic self antigens and uncleared apoptotic bodies can result in autoimmunization. In addition to creating an enhanced immuno-stimulatory environment, modulating complement at a site in which tumor cells have been induced to undergo apoptosis further augments or triggers specific immunity against a tumor to which the host is normally tolerant.

In the methods of the invention, the subject can be a mammal. For example, the mammal can be a human, nonhuman primate, mouse, rat, pig, dog, cat, monkey, cow, or horse.

D. Compositions

Disclosed are the components to be used to prepare the disclosed compositions as well as the compositions themselves to be used in the methods disclosed herein. These and other materials are disclosed herein, and it is understood that when combinations, subsets, interactions, groups, etc. of these materials are disclosed that while specific reference for each of the various individual and collective combinations and permutation of these compounds may not be explicitly made, each is specifically contemplated and described herein. For example, if a particular K9/9, DAF, CD59, CR1, MCP, Crry is described, and/or a specific combination thereof is disclosed and discussed and/or a number of modifications that can be made to a number of molecules including the K9/9, DAF, CD59, CR1, MCP, Crry, and/or combination thereof are discussed, specifically contemplated is each and every combination and permutation of K9/9, DAF, CD59, CR1, MCP, Crry, or combination thereof and the modifications that are possible, unless specifically indicated to the contrary. Thus, if a class of molecules A, B, and C are disclosed as well as a class of molecules D, E, and F and an example of a combination molecule, A-D is disclosed, then even if each is not individually recited each is individually and collectively contemplated meaning combinations, A-E, A-F, B-D, B-E, B-F, C-D, C-E, and C-F are considered disclosed. Likewise, any subset or combination of these is also disclosed. Thus, for example, the sub-group of A-E, B-F, and C-E would be considered disclosed. This concept applies to all aspects of this application including, but not limited to, steps in methods of making and using the disclosed compositions. Thus, if there are a variety of additional steps that can be performed it is understood that each of these additional steps can be performed with any specific embodiment or combination of embodiments of the disclosed methods.

1. Sequence Similarities

It is understood that as discussed herein the use of the terms homology and identity mean the same thing as similarity. Thus, for example, if the use of the word homology is used between two non-natural sequences it is understood that this

is not necessarily indicating an evolutionary relationship between these two sequences, but rather is looking at the similarity or relatedness between their nucleic acid sequences. Many of the methods for determining homology between two evolutionarily related molecules are routinely applied to any two or more nucleic acids or proteins for the purpose of measuring sequence similarity regardless of whether they are evolutionarily related or not.

In general, it is understood that one way to define any known variants and derivatives or those that might arise, of the genes and proteins disclosed herein, is through defining the variants and derivatives in terms of homology to specific known sequences. This identity of particular sequences disclosed herein is also discussed elsewhere herein. Specifically disclosed are variants of these and other genes and proteins herein disclosed which have at least, 70, 71, 72, 73, 74, 75, 76, 77, 78, 79, 80, 81, 82, 83, 84, 85, 86, 87, 88, 89, 90, 91, 92, 93, 94, 95, 96, 97, 98, 99 percent homology to the stated sequence. Those of skill in the art readily understand how to determine the homology of two proteins or nucleic acids, such as genes. For example, the homology can be calculated after aligning the two sequences so that the homology is at its highest level.

Another way of calculating homology can be performed by published algorithms. Optimal alignment of sequences for comparison can be conducted by the local homology algorithm of Smith and Waterman *Adv. Appl. Math.* 2: 482 (1981), by the homology alignment algorithm of Needleman and Wunsch, *J. Mol. Biol.* 48: 443 (1970), by the search for similarity method of Pearson and Lipman, *Proc. Natl. Acad. Sci. U.S.A.* 85: 2444 (1988), by computerized implementations of these algorithms (GAP, BESTFIT, FASTA, and TFASTA in the Wisconsin Genetics Software Package, Genetics Computer Group, 575 Science Dr., Madison, Wis.), or by inspection.

The same types of homology can be obtained for nucleic acids by for example the algorithms disclosed in Zuker, *M. Science* 244:48-52, 1989, Jaeger et al. *Proc. Natl. Acad. Sci. USA* 86:7706-7710, 1989, Jaeger et al. *Methods Enzymol.* 183:281-306, 1989 which are herein incorporated by reference for at least material related to nucleic acid alignment. It is understood that any of the methods typically can be used and that in certain instances the results of these various methods can differ, but the skilled artisan understands if identity is found with at least one of these methods, the sequences would be said to have the stated identity, and be disclosed herein.

For example, as used herein, a sequence recited as having a particular percent homology to another sequence refers to sequences that have the recited homology as calculated by any one or more of the calculation methods described above. For example, a first sequence has 80 percent homology, as defined herein, to a second sequence if the first sequence is calculated to have 80 percent homology to the second sequence using the Zuker calculation method even if the first sequence does not have 80 percent homology to the second sequence as calculated by any of the other calculation methods. As another example, a first sequence has 80 percent homology, as defined herein, to a second sequence if the first sequence is calculated to have 80 percent homology to the second sequence using both the Zuker calculation method and the Pearson and Lipman calculation method even if the first sequence does not have 80 percent homology to the second sequence as calculated by the Smith and Waterman calculation method, the Needleman and Wunsch calculation method, the Jaeger calculation methods, or any of the other calculation methods. As yet another example, a first sequence has 80 percent homology, as defined herein, to a second

sequence if the first sequence is calculated to have 80 percent homology to the second sequence using each of calculation methods (although, in practice, the different calculation methods will often result in different calculated homology percentages).

2. Nucleic Acids

There are a variety of molecules disclosed herein that are nucleic acid based, including for example the nucleic acids that encode, for example K9/9, DAF, CD59, CR1, MCP, Crry, K9/9-DAF, K9/9-CD59, K9/9-CR1, K9/9-MCP, K9/9-CD59 (human), or K9/9-Crry as well as various functional nucleic acids. The disclosed nucleic acids are made up of for example, nucleotides, nucleotide analogs, or nucleotide substitutes. Non-limiting examples of these and other molecules are discussed herein. It is understood that for example, when a vector is expressed in a cell, the expressed mRNA will typically be made up of A, C, G, and U. Likewise, it is understood that if, for example, an antisense molecule is introduced into a cell or cell environment through, for example exogenous delivery, it is advantageous that the antisense molecule be made up of nucleotide analogs that reduce the degradation of the antisense molecule in the cellular environment.

a) Nucleotides and Related Molecules

A nucleotide is a molecule that contains a base moiety, a sugar moiety and a phosphate moiety. Nucleotides can be linked together through their phosphate moieties and sugar moieties creating an internucleoside linkage. The base moiety of a nucleotide can be adenin-9-yl (A), cytosin-1-yl (C), guanin-9-yl (G), uracil-1-yl (U), and thymine-1-yl (T). The sugar moiety of a nucleotide is a ribose or a deoxyribose. The phosphate moiety of a nucleotide is pentavalent phosphate. A non-limiting example of a nucleotide would be 3'-AMP (3'-adenosine monophosphate) or 5'-GMP (5'-guanosine monophosphate).

A nucleotide analog is a nucleotide which contains some type of modification to any of the base, sugar, or phosphate moieties. Modifications to the base moiety would include natural and synthetic modifications of A, C, G, and T/U as well as different purine or pyrimidine bases, such as uracil-5-yl (*psi.*), hypoxanthin-9-yl (I), and 2-aminoadenin-9-yl. A modified base includes but is not limited to 5-methylcytosine (5-me-C), 5-hydroxymethyl cytosine, xanthine, hypoxanthine, 2-aminoadenine, 6-methyl and other alkyl derivatives of adenine and guanine, 2-propyl and other alkyl derivatives of adenine and guanine, 2-thiouracil, 2-thiothymine and 2-thiocytosine, 5-halouracil and cytosine, 5-propynyl uracil and cytosine, 6-azo uracil, cytosine and thymine, 5-uracil (pseudouracil), 4-thiouracil, 8-halo, 8-amino, 8-thiol, 8-thioalkyl, 8-hydroxyl and other 8-substituted adenines and guanines, 5-halo particularly 5-bromo, 5-trifluoromethyl and other 5-substituted uracils and cytosines, 7-methylguanine and 7-methyladenine, 8-azaguanine and 8-azaadenine, 7-deazaguanine and 7-deazaadenine and 3-deazaguanine and 3-deazaadenine. Additional base modifications can be found for example in U.S. Pat. No. 3,687,808, Englisch et al., *Angewandte Chemie, International Edition*, 1991, 30, 613, and Sanghvi, Y. S., Chapter 15, *Antisense Research and Applications*, pages 289-302, Crooke, S. T. and Lebleu, B. ed., CRC Press, 1993. Certain nucleotide analogs, such as 5-substituted pyrimidines, 6-azapyrimidines and N-2, N-6 and O-6 substituted purines, including 2-aminopropyladenine, 5-propynyluracil and 5-propynylcytosine. 5-methylcytosine can increase the stability of duplex formation. Often time base modifications can be combined with for example a

sugar modification, such as 2'-O-methoxyethyl, to achieve unique properties such as increased duplex stability. There are numerous United States patents such as U.S. Pat. Nos. 4,845,205; 5,130,302; 5,134,066; 5,175,273; 5,367,066; 5,432,272; 5,457,187; 5,459,255; 5,484,908; 5,502,177; 5,525,711; 5,552,540; 5,587,469; 5,594,121; 5,596,091; 5,614,617; and 5,681,941, which detail and describe a range of base modifications. Each of these patents is herein incorporated by reference.

Nucleotide analogs can also include modifications of the sugar moiety. Modifications to the sugar moiety would include natural modifications of the ribose and deoxy ribose as well as synthetic modifications. Sugar modifications include but are not limited to the following modifications at the 2' position: OH; F; O-, S-, or N-alkyl; O-, S-, or N-alkenyl; O-, S- or N-alkynyl; or O-alkyl-O-alkyl, wherein the alkyl, alkenyl and alkynyl can be substituted or unsubstituted C₁ to C₁₀, alkyl or C₂ to C₁₀ alkenyl and alkynyl. 2' sugar modifications also include but are not limited to —O[(CH₂)_nO]_m, CH₃, —O(CH₂)_nOCH₃, —O(CH₂)_nNH₂, —O(CH₂)_nCH₃, —O(CH₂)_n—ONH₂, and —O(CH₂)_nON[(CH₂)_mCH₃]₂, where n and m are from 1 to about 10.

Other modifications at the 2' position include but are not limited to: C, to C₁₀ lower alkyl, substituted lower alkyl, alkaryl, aralkyl, O-alkaryl or O-aralkyl, SH, SCH₃, OCN, Cl, Br, CN, CF₃, OCF₃, SOCH₃, SO₂ CH₃, ONO₂, NO₂, N₃, NH₂, heterocycloalkyl, heterocycloalkaryl, aminoalkylamino, polyalkylamino, substituted silyl, an RNA cleaving group, a reporter group, an intercalator, a group for improving the pharmacokinetic properties of an oligonucleotide, or a group for improving the pharmacodynamic properties of an oligonucleotide, and other substituents having similar properties. Similar modifications can also be made at other positions on the sugar, particularly the 3' position of the sugar on the 3' terminal nucleotide or in 2'-5' linked oligonucleotides and the 5' position of 5' terminal nucleotide. Modified sugars would also include those that contain modifications at the bridging ring oxygen, such as CH₂ and S. Nucleotide sugar analogs can also have sugar mimetics such as cyclobutyl moieties in place of the pentofuranosyl sugar. There are numerous United States patents that teach the preparation of such modified sugar structures such as U.S. Pat. Nos. 4,981,957; 5,118,800; 5,319,080; 5,359,044; 5,393,878; 5,446,137; 5,466,786; 5,514,785; 5,519,134; 5,567,811; 5,576,427; 5,591,722; 5,597,909; 5,610,300; 5,627,053; 5,639,873; 5,646,265; 5,658,873; 5,670,633; and 5,700,920, each of which is herein incorporated by reference in its entirety.

Nucleotide analogs can also be modified at the phosphate moiety. Modified phosphate moieties include but are not limited to those that can be modified so that the linkage between two nucleotides contains a phosphorothioate, chiral phosphorothioate, phosphorodithioate, phosphotriester, aminoalkylphosphotriester, methyl and other alkyl phosphonates including 3'-alkylene phosphonate and chiral phosphonates, phosphinates, phosphoramidates including 3'-amino phosphoramidate and aminoalkylphosphoramidates, thionophosphoramidates, thionoalkylphosphonates, thionoalkylphosphotriesters, and boranophosphates. It is understood that these phosphate or modified phosphate linkage between two nucleotides can be through a 3'-5' linkage or a 2'-5' linkage, and the linkage can contain inverted polarity such as 3'-5' to 5'-3' or 2'-5' to 5'-2'. Various salts, mixed salts and free acid forms are also included. Numerous United States patents teach how to make and use nucleotides containing modified phosphates and include but are not limited to, U.S. Pat. Nos. 3,687,808; 4,469,863; 4,476,301; 5,023,243; 5,177,196; 5,188,897; 5,264,423; 5,276,019; 5,278,302; 5,286,717;

5,321,131; 5,399,676; 5,405,939; 5,453,496; 5,455,233; 5,466,677; 5,476,925; 5,519,126; 5,536,821; 5,541,306; 5,550,111; 5,563,253; 5,571,799; 5,587,361; and 5,625,050, each of which is herein incorporated by reference.

It is understood that nucleotide analogs need only contain a single modification, but can also contain multiple modifications within one of the moieties or between different moieties.

Nucleotide substitutes are molecules having similar functional properties to nucleotides, but which do not contain a phosphate moiety, such as peptide nucleic acid (PNA). Nucleotide substitutes are molecules that will recognize nucleic acids in a Watson-Crick or Hoogsteen manner, but which are linked together through a moiety other than a phosphate moiety. Nucleotide substitutes are able to conform to a double helix type structure when interacting with the appropriate target nucleic acid.

Nucleotide substitutes are nucleotides or nucleotide analogs that have had the phosphate moiety and/or sugar moieties replaced. Nucleotide substitutes do not contain a standard phosphorus atom. Substitutes for the phosphate can be for example, short chain alkyl or cycloalkyl internucleoside linkages, mixed heteroatom and alkyl or cycloalkyl internucleoside linkages, or one or more short chain heteroatomic or heterocyclic internucleoside linkages. These include those having morpholino linkages (formed in part from the sugar portion of a nucleoside); siloxane backbones; sulfide, sulfoxide and sulfone backbones; formacetyl and thioformacetyl backbones; methylene formacetyl and thioformacetyl backbones; alkene containing backbones; sulfamate backbones; methyleneimino and methylenehydrazino backbones; sulfonate and sulfonamide backbones; amide backbones; and others having mixed N, O, S and CH₂ component parts. Numerous United States patents disclose how to make and use these types of phosphate replacements and include but are not limited to U.S. Pat. Nos. 5,034,506; 5,166,315; 5,185,444; 5,214,134; 5,216,141; 5,235,033; 5,264,562; 5,264,564; 5,405,938; 5,434,257; 5,466,677; 5,470,967; 5,489,677; 5,541,307; 5,561,225; 5,596,086; 5,602,240; 5,610,289; 5,602,240; 5,608,046; 5,610,289; 5,618,704; 5,623,070; 5,663,312; 5,633,360; 5,677,437; and 5,677,439, each of which is herein incorporated by reference.

It is also understood in a nucleotide substitute that both the sugar and the phosphate moieties of the nucleotide can be replaced, by for example an amide type linkage (aminoethylglycine) (PNA). U.S. Pat. Nos. 5,539,082; 5,714,331; and 5,719,262 teach how to make and use PNA molecules, each of which is herein incorporated by reference. (See also Nielsen et al., *Science*, 1991, 254, 1497-1500).

It is also possible to link other types of molecules (conjugates) to nucleotides or nucleotide analogs to enhance for example, cellular uptake. Conjugates can be chemically linked to the nucleotide or nucleotide analogs. Such conjugates include but are not limited to lipid moieties such as a cholesterol moiety (Letsinger et al., *Proc. Natl. Acad. Sci. USA*, 1989, 86, 6553-6556), cholic acid (Manoharan et al., *Bioorg. Med. Chem. Lett.*, 1994, 4, 1053-1060), a thioether, e.g., hexyl-S-tritylthiol (Manoharan et al., *Ann. N.Y. Acad. Sci.*, 1992, 660, 306-309; Manoharan et al., *Bioorg. Med. Chem. Lett.*, 1993, 3, 2765-2770), a thiocholesterol (Oberhauser et al., *Nucl. Acids Res.*, 1992, 20, 533-538), an aliphatic chain, e.g., dodecandiol or undecyl residues (Saison-Behmoaras et al., *EMBO J.*, 1991, 10, 1111-1118; Kabanov et al., *FEBS Lett.*, 1990, 259, 327-330; Svinarchuk et al., *Biochimie*, 1993, 75, 49-54), a phospholipid, e.g., di-hexadecyl-

Let., 1995, 36, 3651-3654; Shea et al., *Nucl. Acids Res.*, 1990, 18, 3777-3783), a polyamine or a polyethylene glycol chain (Manoharan et al., *Nucleosides & Nucleotides*, 1995, 14, 969-973), or adamantane acetic acid (Manoharan et al., *Tetrahedron Lett.*, 1995, 36, 3651-3654), a palmitoyl moiety (Mishra et al., *Biochim. Biophys. Acta*, 1995, 1264, 229-237), or an octadecylamine or hexylamino-carbonyl-oxysterol moiety (Crooke et al., *J. Pharmacol. Exp. Ther.*, 1996, 277, 923-937. Numerous United States patents teach the preparation of such conjugates and include, but are not limited to U.S. Pat. Nos. 4,828,979; 4,948,882; 5,218,105; 5,525,465; 5,541,313; 5,545,730; 5,552,538; 5,578,717; 5,580,731; 5,580,731; 5,591,584; 5,109,124; 5,118,802; 5,138,045; 5,414,077; 5,486,603; 5,512,439; 5,578,718; 5,608,046; 4,587,044; 4,605,735; 4,667,025; 4,762,779; 4,789,737; 4,824,941; 4,835,263; 4,876,335; 4,904,582; 4,958,013; 5,082,830; 5,112,963; 5,214,136; 5,082,830; 5,112,963; 5,214,136; 5,245,022; 5,254,469; 5,258,506; 5,262,536; 5,272,250; 5,292,873; 5,317,098; 5,371,241; 5,391,723; 5,416,203; 5,451,463; 5,510,475; 5,512,667; 5,514,785; 5,565,552; 5,567,810; 5,574,142; 5,585,481; 5,587,371; 5,595,726; 5,597,696; 5,599,923; 5,599,928 and 5,688,941, each of which is herein incorporated by reference.

A Watson-Crick interaction is at least one interaction with the Watson-Crick face of a nucleotide, nucleotide analog, or nucleotide substitute. The Watson-Crick face of a nucleotide, nucleotide analog, or nucleotide substitute includes the C2, N1, and C6 positions of a purine based nucleotide, nucleotide analog, or nucleotide substitute and the C2, N3, C4 positions of a pyrimidine based nucleotide, nucleotide analog, or nucleotide substitute.

A Hoogsteen interaction is the interaction that takes place on the Hoogsteen face of a nucleotide or nucleotide analog, which is exposed in the major groove of duplex DNA. The Hoogsteen face includes the N7 position and reactive groups (NH₂ or O) at the C6 position of purine nucleotides.

b) Sequences

There are a variety of sequences related to the K9/9, DAF, CD59, CR1, MCP, Crry, K9/9-DAF, K9/9-CR1, K9/9-MCP, K9/9-CD59, K9/9-CD59 (human), or K9/9-Crry genes having, for example, the sequences as disclosed herein or sequences available in the literature. These sequences and others are herein incorporated by reference in their entireties as well as for individual subsequences contained therein.

One particular sequence set forth in SEQ ID NO: 11 used herein, as an example, to exemplify the disclosed compositions and methods. It is understood that the description related to this sequence is applicable to any sequence related to K9/9, K9/9-DAF, K9/9-CD59, K9/9-CD59 (human), K9/9-CR1, K9/9-MCP, or K9/9-Crry, unless specifically indicated otherwise. Those of skill in the art understand how to resolve sequence discrepancies and differences and to adjust the compositions and methods relating to a particular sequence to other related sequences (i.e. sequences of K9/9, DAF, CD59, CR1, MCP, Crry, K9/9-DAF, K9/9-CR1, K9/9-MCP, K9/9-CD59, K9/9-CD59 (human), or K9/9-Crry. Primers and/or probes can be designed for any K9/9, DAF, CD59, CR1, MCP, Crry, K9/9-DAF, K9/9-CD59, K9/9-CD59 (human), K9/9-CR1, K9/9-MCP, and K9/9-Crry sequence given the information disclosed herein and known in the art.

3. Delivery of the Compositions to Cells

There are a number of compositions and methods which can be used to deliver the present fusion protein compositions, immunoconjugate compositions, and nucleic acid compositions to cells, either in vitro or in vivo. Compositions of

the invention are preferably administered to a subject in a pharmaceutically acceptable carrier. Suitable carriers and their formulations are described in *Remington: The Science and Practice of Pharmacy* (19th ed.) ed. A. R. Gennaro, Mack Publishing Company, Easton, Pa. 1995. Typically, an appropriate amount of a pharmaceutically-acceptable salt is used in the formulation to render the formulation isotonic. Examples of the pharmaceutically-acceptable carriers include, but are not limited to, saline, water:oil emulsions, oil:water emulsions, water:oil:water emulsions, and Ringer's solution and dextrose solution. The pH of the solution is preferably from about 5 to about 8, and more preferably from about 7 to about 7.5. Further carriers include sustained release preparations such as semipermeable matrices of solid hydrophobic polymers containing the antibody, which matrices are in the form of shaped articles, e.g., films, liposomes or microparticles. It will be apparent to those persons skilled in the art that certain carriers may be more preferable depending upon, for instance, the route of administration and concentration of antibody being administered.

The compositions of the invention can be administered to the subject, patient, or cell by injection (e.g., intravenous, intraperitoneal, subcutaneous, intramuscular), or by other methods such as infusion that ensure its delivery to the bloodstream in an effective form. Local or intravenous injection is preferred.

Effective dosages and schedules for administering the compositions of the invention can be determined empirically, and making such determinations is within the skill in the art. Those skilled in the art will understand that the dosage of the compositions of the invention that must be administered will vary depending on, for example, the subject that will receive the composition, the route of administration, the particular type of composition used and other drugs being administered. A typical daily dosage of the compositions of the invention used alone might range from about 1 µg/kg to up to 100 mg/kg of body weight or more per day, depending on the factors mentioned above.

a) Nucleic Acid Based Delivery Systems

There are a number of compositions and methods which can be used to deliver nucleic acids to cells, either in vitro or in vivo. These methods and compositions can largely be broken down into two classes: viral based delivery systems and non-viral based delivery systems. For example, the nucleic acids can be delivered through a number of direct delivery systems such as, electroporation, lipofection, calcium phosphate precipitation, plasmids, viral vectors, viral nucleic acids, phage nucleic acids, phages, cosmids, or via transfer of genetic material in cells or carriers such as cationic liposomes. Appropriate means for transfection, including viral vectors, chemical transfectants, or physico-mechanical methods such as electroporation and direct diffusion of DNA, are described by, for example, Wolff, J. A., et al., *Science*, 247, 1465-1468, (1990); and Wolff, J. A. *Nature*, 352, 815-818, (1991) Such methods are well known in the art and readily adaptable for use with the compositions and methods described herein. In certain cases, the methods will be modified to specifically function with large DNA molecules. Further, these methods can be used to target certain diseases and cell populations by using the targeting characteristics of the carrier.

Transfer vectors can be any nucleotide construction used to deliver genes into cells (e.g., a plasmid), or as part of a general strategy to deliver genes, e.g., as part of recombinant retrovirus or adenovirus (Ram et al. *Cancer Res.* 53:83-88, (1993)).

As used herein, plasmid or viral vectors are agents that transport the disclosed nucleic acids, into the cell without degradation and include a promoter yielding expression of the gene in the cells into which it is delivered. In some embodiments the K9/9, DAF, CD59, CR1, MCP, Crry, K9/9-DAF, K9/9-CR1, K9/9-MCP, K9/9-CD59, K9/9-CD59 (human), or K9/9-Crry s are derived from either a virus or a retrovirus. Viral vectors are, for example, Adenovirus, Adeno-associated virus, Herpes virus, Vaccinia virus, Polio virus, AIDS virus, neuronal trophic virus, Sindbis and other RNA viruses, including these viruses with the HIV backbone. Also preferred are any viral families which share the properties of these viruses which make them suitable for use as vectors. Retroviruses include Murine Maloney Leukemia virus, MMLV, and retroviruses that express the desirable properties of MMLV as a vector. Retroviral vectors are able to carry a larger genetic payload, i.e., a transgene or marker gene, than other viral vectors, and for this reason are a commonly used vector. However, they are not as useful in non-proliferating cells. Adenovirus vectors are relatively stable and easy to work with, have high titers, and can be delivered in aerosol formulation, and can transfect non-dividing cells. Pox viral vectors are large and have several sites for inserting genes. They are thermostable and can be stored at room temperature. A preferred embodiment is a viral vector which has been engineered so as to suppress the immune response of the host organism, elicited by the viral antigens. Preferred vectors of this type will carry coding regions for Interleukin 8 or 10.

Viral vectors can have higher transaction (ability to introduce genes) abilities than chemical or physical methods to introduce genes into cells. Typically, viral vectors contain, nonstructural early genes, structural late genes, an RNA polymerase III transcript, inverted terminal repeats necessary for replication and encapsidation, and promoters to control the transcription and replication of the viral genome. When engineered as vectors, viruses typically have one or more of the early genes removed and a gene or gene/promotor cassette is inserted into the viral genome in place of the removed viral DNA. Constructs of this type can carry up to about 8 kb of foreign genetic material. The necessary functions of the removed early genes are typically supplied by cell lines which have been engineered to express the gene products of the early genes in trans.

(1) Retroviral Vectors

A retrovirus is an animal virus belonging to the virus family of Retroviridae, including any types, subfamilies, genus, or tropisms. Retroviral vectors, in general, are described by Verma, I. M., *Retroviral vectors for gene transfer*. In *Microbiology-1985*, American Society for Microbiology, pp. 229-232, Washington, (1985), which is incorporated by reference herein. Examples of methods for using retroviral vectors for gene therapy are described in U.S. Pat. Nos. 4,868,116 and 4,980,286; PCT applications WO 90/02806 and WO 89/07136; and Mulligan, (*Science* 260:926-932 (1993)); the teachings of which are incorporated herein by reference.

A retrovirus is essentially a package which has packed into it nucleic acid cargo. The nucleic acid cargo carries with it a packaging signal, which ensures that the replicated daughter molecules will be efficiently packaged within the package coat. In addition to the package signal, there are a number of molecules which are needed in cis, for the replication, and packaging of the replicated virus. Typically a retroviral genome, contains the gag, pol, and env genes which are involved in the making of the protein coat. It is the gag, pol, and env genes which are typically replaced by the foreign

DNA that it is to be transferred to the target cell. Retrovirus vectors typically contain a packaging signal for incorporation into the package coat, a sequence which signals the start of the gag transcription unit, elements necessary for reverse transcription, including a primer binding site to bind the tRNA primer of reverse transcription, terminal repeat sequences that guide the switch of RNA strands during DNA synthesis, a purine rich sequence 5' to the 3' LTR that serve as the priming site for the synthesis of the second strand of DNA synthesis, and specific sequences near the ends of the LTRs that enable the insertion of the DNA state of the retrovirus to insert into the host genome. The removal of the gag, pol, and env genes allows for about 8 kb of foreign sequence to be inserted into the viral genome, become reverse transcribed, and upon replication be packaged into a new retroviral particle. This amount of nucleic acid is sufficient for the delivery of a one to many genes depending on the size of each transcript. It is preferable to include either positive or negative selectable markers along with other genes in the insert.

Since the replication machinery and packaging proteins in most retroviral vectors have been removed (gag, pol, and env), the vectors are typically generated by placing them into a packaging cell line. A packaging cell line is a cell line which has been transfected or transformed with a retrovirus that contains the replication and packaging machinery, but lacks any packaging signal. When the vector carrying the DNA of choice is transfected into these cell lines, the vector containing the gene of interest is replicated and packaged into new retroviral particles, by the machinery provided in cis by the helper cell. The genomes for the machinery are not packaged because they lack the necessary signals.

(2) Adenoviral Vectors

The construction of replication-defective adenoviruses has been described (Berkner et al., *J. Virology* 61:1213-1220 (1987); Massie et al., *Mol. Cell. Biol.* 6:2872-2883 (1986); Haj-Ahmad et al., *J. Virology* 57:267-274 (1986); Davidson et al., *J. Virology* 61:1226-1239 (1987); Zhang "Generation and identification of recombinant adenovirus by liposome-mediated transfection and PCR analysis" *BioTechniques* 15:868-872 (1993)). The benefit of the use of these viruses as vectors is that they are limited in the extent to which they can spread to other cell types, since they can replicate within an initial infected cell, but are unable to form new infectious viral particles. Recombinant adenoviruses have been shown to achieve high efficiency gene transfer after direct, in vivo delivery to airway epithelium, hepatocytes, vascular endothelium, CNS parenchyma and a number of other tissue sites (Morsy, *J. Clin. Invest.* 92:1580-1586 (1993); Kirshenbaum, *J. Clin. Invest.* 92:381-387 (1993); Roessler, *J. Clin. Invest.* 92:1085-1092 (1993); Moullier, *Nature Genetics* 4:154-159 (1993); La Salle, *Science* 259:988-990 (1993); Gomez-Foix, *J. Biol. Chem.* 267:25129-25134 (1992); Rich, *Human Gene Therapy* 4:461-476 (1993); Zabner, *Nature Genetics* 6:75-83 (1994); Guzman, *Circulation Research* 73:1201-1207 (1993); Bout, *Human Gene Therapy* 5:3-10 (1994); Zabner, *Cell* 75:207-216 (1993); Caillaud, *Eur. J. Neuroscience* 5:1287-1291 (1993); and Ragot, *J. Gen. Virology* 74:501-507 (1993)). Recombinant adenoviruses achieve gene transduction by binding to specific cell surface receptors, after which the virus is internalized by receptor-mediated endocytosis, in the same manner as wild type or replication-defective adenovirus (Chardonnet and Dales, *Virology* 40:462-477 (1970); Brown and Burlingham, *J. Virology* 12:386-396 (1973); Svensson and Persson, *J. Virology* 55:442-449 (1985); Seth, et al., *J. Virol.* 51:650-655 (1984); Seth, et al., *Mol. Cell. Biol.*

4:1528-1533 (1984); Varga et al., *J. Virology* 65:6061-6070 (1991); Wickham et al., *Cell* 73:309-319 (1993)).

A viral vector can be one based on an adenovirus which has had the E1 gene removed and these virions are generated in a cell line such as the human 293 cell line. In another preferred embodiment both the E1 and E3 genes are removed from the adenovirus genome.

(3) Adeno-Associated Viral Vectors

Another type of viral vector is based on an adeno-associated virus (AAV). This defective parvovirus is a preferred vector because it can infect many cell types and is nonpathogenic to humans. AAV type vectors can transport about 4 to 5 kb and wild type AAV is known to stably insert into chromosome 19. Vectors which contain this site specific integration property are preferred. An especially preferred embodiment of this type of vector is the P4.1 C vector produced by Avigen, San Francisco, Calif., which can contain the herpes simplex virus thymidine kinase gene, HSV-tk, and/or a marker gene, such as the gene encoding the green fluorescent protein, GFP.

In another type of AAV virus, the AAV contains a pair of inverted terminal repeats (ITRs) which flank at least one cassette containing a promoter which directs cell-specific expression operably linked to a heterologous gene. Heterologous in this context refers to any nucleotide sequence or gene which is not native to the AAV or B19 parvovirus.

Typically the AAV and B19 coding regions have been deleted, resulting in a safe, noncytotoxic vector. The AAV ITRs, or modifications thereof, confer infectivity and site-specific integration, but not cytotoxicity, and the promoter directs cell-specific expression. U.S. Pat. No. 6,261,834 is herein incorporated by reference for material related to the AAV vector.

The vectors of the present invention thus provide DNA molecules which are capable of integration into a mammalian chromosome without substantial toxicity.

The inserted genes in viral and retroviral usually contain promoters, and/or enhancers to help control the expression of the desired gene product. A promoter is generally a sequence or sequences of DNA that function when in a relatively fixed location in regard to the transcription start site. A promoter contains core elements required for basic interaction of RNA polymerase and transcription factors, and can contain upstream elements and response elements.

(4) Large Payload Viral Vectors

Molecular genetic experiments with large human herpesviruses have provided a means whereby large heterologous DNA fragments can be cloned, propagated and established in cells permissive for infection with herpesviruses (Sun et al., *Nature genetics* 8: 33-41, 1994; Cotter and Robertson, *Curr Opin Mol Ther* 5: 633-644, 1999). These large DNA viruses (herpes simplex virus (HSV) and Epstein-Barr virus (EBV)), have the potential to deliver fragments of human heterologous DNA >150 kb to specific cells. EBV recombinants can maintain large pieces of DNA in the infected B-cells as episomal DNA. Individual clones carried human genomic inserts up to 330 kb appeared genetically stable. The maintenance of these episomes requires a specific EBV nuclear protein, EBNA1, constitutively expressed during infection with EBV. Additionally, these vectors can be used for transfection, where large amounts of protein can be generated transiently in vitro. Herpesvirus amplicon systems are also being used to package pieces of DNA >220 kb and to infect cells that can stably maintain DNA as episomes.

Other useful systems include, for example, replicating and host-restricted non-replicating vaccinia virus vectors.

b) Non-Nucleic Acid Based Systems

The disclosed compositions can be delivered to the target cells in a variety of ways. For example, the compositions can be delivered through electroporation, or through lipofection, or through calcium phosphate precipitation. The delivery mechanism chosen will depend in part on the type of cell targeted and whether the delivery is occurring for example *in vivo* or *in vitro*.

Thus, the compositions can comprise, in addition to the disclosed K9/9, DAF, CD59, CR1, MCP, Crry, K9/9-DAF, K9/9-CR1, K9/9-MCP, K9/9-CD59, K9/9-CD59 (human), or K9/9-Crry or vectors for example, lipids such as liposomes, such as cationic liposomes (e.g., DOTMA, DOPE, DC-cholesterol) or anionic liposomes. Liposomes can further comprise proteins to facilitate targeting a particular cell, if desired. Administration of a composition comprising a compound and a cationic liposome can be administered to the blood afferent to a target organ or inhaled into the respiratory tract to target cells of the respiratory tract. Regarding liposomes, see, e.g., Brigham et al. *Am. J. Resp. Cell. Mol. Biol.* 1:95-100 (1989); Felgner et al. *Proc. Natl. Acad. Sci USA* 84:7413-7417 (1987); U.S. Pat. No. 4,897,355. Furthermore, the compound can be administered as a component of a microcapsule that can be targeted to specific cell types, such as macrophages, or where the diffusion of the compound or delivery of the compound from the microcapsule is designed for a specific rate or dosage.

In the methods described above which include the administration and uptake of exogenous DNA into the cells of a subject (i.e., gene transduction or transfection), delivery of the compositions to cells can be via a variety of mechanisms. As one example, delivery can be via a liposome, using commercially available liposome preparations such as LIPOFECTIN, LIPOFECTAMINE (GIBCO-BRL, Inc., Gaithersburg, Md.), SUPERFECT (Qiagen, Inc. Hilden, Germany) and TRANSFECTAM (Promega Biotec, Inc., Madison, Wis.), as well as other liposomes developed according to procedures standard in the art. In addition, the nucleic acid or vector of this invention can be delivered *in vivo* by electroporation, the technology for which is available from Genetronics, Inc. (San Diego, Calif.) as well as by means of a SONOPORATION machine (ImaRx Pharmaceutical Corp., Tucson, Ariz.).

The materials can be in solution, suspension (for example, incorporated into microparticles, liposomes, or cells). These can be targeted to a particular cell type via antibodies, receptors, or receptor ligands. The following references are examples of the use of this technology to target specific proteins to tumor tissue (Senter, et al., *Bioconjugate Chem.*, 2:447-451, (1991); Bagshawe, K. D., *Br. J. Cancer*, 60:275-281, (1989); Bagshawe, et al., *Br. J. Cancer*, 58:700-703, (1988); Senter, et al., *Bioconjugate Chem.*, 4:3-9, (1993); Battelli, et al., *Cancer Immunol. Immunother.*, 35:421-425, (1992); Pietersz and McKenzie, *Immunolog. Reviews*, 129:57-80, (1992); and Roffler, et al., *Biochem. Pharmacol.* 42:2062-2065, (1991)). These techniques can be used for a variety of other specific cell types. Vehicles such as "stealth" and other antibody conjugated liposomes (including lipid mediated drug targeting to colonic carcinoma), receptor mediated targeting of DNA through cell specific ligands, lymphocyte directed tumor targeting, and highly specific therapeutic retroviral targeting of murine glioma cells *in vivo*. The following references are examples of the use of this technology to target specific proteins to tumor tissue (Hughes

et al., *Cancer Research*, 49:6214-6220, (1989); and Litzinger and Huang, *Biochimica et Biophysica Acta*, 1104:179-187, (1992)). In general, receptors are involved in pathways of endocytosis, either constitutive or ligand induced. These receptors cluster in clathrin-coated pits, enter the cell via clathrin-coated vesicles, pass through an acidified endosome in which the receptors are sorted, and then either recycle to the cell surface, become stored intracellularly, or are degraded in lysosomes. The internalization pathways serve a variety of functions, such as nutrient uptake, removal of activated proteins, clearance of macromolecules, opportunistic entry of viruses and toxins, dissociation and degradation of ligand, and receptor-level regulation. Many receptors follow more than one intracellular pathway, depending on the cell type, receptor concentration, type of ligand, ligand valency, and ligand concentration. Molecular and cellular mechanisms of receptor-mediated endocytosis has been reviewed (Brown and Greene, *DNA and Cell Biology* 10:6, 399-409 (1991)).

Nucleic acids that are delivered to cells which are to be integrated into the host cell genome, typically contain integration sequences. These sequences are often viral related sequences, particularly when viral based systems are used. These viral intergration systems can also be incorporated into nucleic acids which are to be delivered using a non-nucleic acid based system of deliver, such as a liposome, so that the nucleic acid contained in the delivery system can be come integrated into the host genome.

Other general techniques for integration into the host genome include, for example, systems designed to promote homologous recombination with the host genome. These systems typically rely on sequence flanking the nucleic acid to be expressed that has enough homology with a target sequence within the host cell genome that recombination between the vector nucleic acid and the target nucleic acid takes place, causing the delivered nucleic acid to be integrated into the host genome. These systems and the methods necessary to promote homologous recombination are known to those of skill in the art.

c) In Vivo/Ex Vivo

As described above, the compositions can be administered in a pharmaceutically acceptable carrier and can be delivered to the subject's cells *in vivo* and/or *ex vivo* by a variety of mechanisms well known in the art (e.g., uptake of naked DNA, liposome fusion, intramuscular injection of DNA via a gene gun, endocytosis and the like).

If *ex vivo* methods are employed, cells or tissues can be removed and maintained outside the body according to standard protocols well known in the art. The compositions can be introduced into the cells via any gene transfer mechanism, such as, for example, calcium phosphate mediated gene delivery, electroporation, microinjection or proteoliposomes. The transduced cells can then be infused (e.g., in a pharmaceutically acceptable carrier) or homotopically transplanted back into the subject per standard methods for the cell or tissue type. Standard methods are known for transplantation or infusion of various cells into a subject.

4. Expression Systems

The nucleic acids that are delivered to cells typically contain expression controlling systems. For example, the inserted genes in viral and retroviral systems usually contain promoters, and/or enhancers to help control the expression of the desired gene product. A promoter is generally a sequence or sequences of DNA that function when in a relatively fixed

location in regard to the transcription start site. A promoter contains core elements required for basic interaction of RNA polymerase and transcription factors, and can contain upstream elements and response elements.

a) Viral Promoters and Enhancers

Preferred promoters controlling transcription from vectors in mammalian host cells can be obtained from various sources, for example, the genomes of viruses such as: polyoma, Simian Virus 40 (SV40), adenovirus, retroviruses, hepatitis-B virus and most preferably cytomegalovirus, or from heterologous mammalian promoters, e.g. beta actin promoter. The early and late promoters of the SV40 virus are conveniently obtained as an SV40 restriction fragment which also contains the SV40 viral origin of replication (Fiers et al., *Nature*, 273: 113 (1978)). The immediate early promoter of the human cytomegalovirus is conveniently obtained as a HindIII E restriction fragment (Greenway, P. J. et al., *Gene* 18: 355-360 (1982)). Of course, promoters from the host cell or related species also are useful herein.

Enhancer generally refers to a sequence of DNA that functions at no fixed distance from the transcription start site and can be either 5' (Laimins, L. et al., *Proc. Natl. Acad. Sci.* 78: 993 (1981)) or 3' (Lusky, M. L., et al., *Mol. Cell Bio.* 3: 1108 (1983)) to the transcription unit. Furthermore, enhancers can be within an intron (Banerji, J. L. et al., *Cell* 33: 729 (1983)) as well as within the coding sequence itself (Osborne, T. F., et al., *Mol. Cell Bio.* 4: 1293 (1984)). They are usually between 10 and 300 bp in length, and they function in cis. Enhancers function to increase transcription from nearby promoters. Enhancers also often contain response elements that mediate the regulation of transcription. Promoters can also contain response elements that mediate the regulation of transcription. Enhancers often determine the regulation of expression of a gene. While many enhancer sequences are now known from mammalian genes (globin, elastase, albumin, -fetoprotein and insulin), typically one will use an enhancer from a eukaryotic cell virus for general expression. Preferred examples are the SV40 enhancer on the late side of the replication origin (bp 100-270), the cytomegalovirus early promoter enhancer, the polyoma enhancer on the late side of the replication origin, and adenovirus enhancers.

The promoter and/or enhancer can be specifically activated either by light or specific chemical events which trigger their function. Systems can be regulated by reagents such as tetracycline and dexamethasone. There are also ways to enhance viral vector gene expression by exposure to irradiation, such as gamma irradiation, or alkylating chemotherapy drugs.

In certain embodiments the promoter and/or enhancer region can act as a constitutive promoter and/or enhancer to maximize expression of the region of the transcription unit to be transcribed. In certain constructs the promoter and/or enhancer region be active in all eukaryotic cell types, even if it is only expressed in a particular type of cell at a particular time. A preferred promoter of this type is the CMV promoter (650 bases). Other preferred promoters are SV40 promoters, cytomegalovirus (full length promoter), and retroviral vector LTF.

It has been shown that all specific regulatory elements can be cloned and used to construct expression vectors that are selectively expressed in specific cell types such as melanoma cells. The glial fibrillary acetic protein (GFAP) promoter has been used to selectively express genes in cells of glial origin.

Expression vectors used in eukaryotic host cells (yeast, fungi, insect, plant, animal, human or nucleated cells) can also contain sequences necessary for the termination of transcription which can affect mRNA expression. These regions are transcribed as polyadenylated segments in the untrans-

lated portion of the mRNA encoding tissue factor protein. The 3' untranslated regions also include transcription termination sites. It is preferred that the transcription unit also contains a polyadenylation region. One benefit of this region is that it increases the likelihood that the transcribed unit will be processed and transported like mRNA. The identification and use of polyadenylation signals in expression constructs is well established. It is preferred that homologous polyadenylation signals be used in the transgene constructs. In certain transcription units, the polyadenylation region is derived from the SV40 early polyadenylation signal and consists of about 400 bases. It is also preferred that the transcribed units contain other standard sequences alone or in combination with the above sequences improve expression from, or stability of, the construct.

b) Markers

The viral vectors can include nucleic acid sequence encoding a marker product. This marker product is used to determine if the gene has been delivered to the cell and once delivered is being expressed. Preferred marker genes are the *E. Coli* lacZ gene, which encodes β -galactosidase, and green fluorescent protein.

In some embodiments the marker can be a selectable marker. Examples of suitable selectable markers for mammalian cells are dihydrofolate reductase (DHFR), thymidine kinase, neomycin, neomycin analog G418, hygromycin, and puromycin. When such selectable markers are successfully transferred into a mammalian host cell, the transformed mammalian host cell can survive if placed under selective pressure. There are two widely used distinct categories of selective regimes. The first category is based on a cell's metabolism and the use of a mutant cell line which lacks the ability to grow independent of a supplemented media. Two examples are: CHO DHFR-cells and mouse LTK-cells. These cells lack the ability to grow without the addition of such nutrients as thymidine or hypoxanthine. Because these cells lack certain genes necessary for a complete nucleotide synthesis pathway, they cannot survive unless the missing nucleotides are provided in a supplemented media. An alternative to supplementing the media is to introduce an intact DHFR or TK gene into cells lacking the respective genes, thus altering their growth requirements. Individual cells which were not transformed with the DHFR or TK gene will not be capable of survival in non-supplemented media.

The second category is dominant selection which refers to a selection scheme used in any cell type and does not require the use of a mutant cell line. These schemes typically use a drug to arrest growth of a host cell. Those cells which have a novel gene would express a protein conveying drug resistance and would survive the selection. Examples of such dominant selection use the drugs neomycin, (Southern P. and Berg, P., *J. Molec. Appl. Genet.* 1: 327 (1982)), mycophenolic acid, (Mulligan, R. C. and Berg, P. *Science* 209: 1422 (1980)) or hygromycin, (Sugden, B. et al., *Mol. Cell. Biol.* 5: 410-413 (1985)). The three examples employ bacterial genes under eukaryotic control to convey resistance to the appropriate drug G418 or neomycin (geneticin), xgpt (mycophenolic acid) or hygromycin, respectively. Others include the neomycin analog G418 and puramycin.

5. Peptides

a) Protein Variants

As discussed herein there are numerous variants of the K9/9, DAF, CD59, CR1, MCP, Crry, K9/9-DAF, K9/9-CR1,

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K9/9-MCP, K9/9-CD59, K9/9-CD59 (human), or K9/9-Crry protein that are known and herein contemplated. In addition, to the known functional K9/9, DAF, CD59, CR1, MCP, Crry, K9/9-DAF, K9/9-CD59, K9/9-CD59 (human), K9/9-CR1, K9/9-MCP, and K9/9-Crry, strain variants, there are derivatives of the K9/9, DAF, CD59, CR1, MCP, Crry, K9/9-DAF, K9/9-CD59, K9/9-CD59 (human), K9/9-CR1, K9/9-MCP, and K9/9-Crry proteins which also function in the disclosed methods and compositions. Protein variants and derivatives are well understood to those of skill in the art and can involve amino acid sequence modifications. For example, amino acid sequence modifications typically fall into one or more of three classes: substitutional, insertional or deletional variants. Insertions include amino and/or carboxyl terminal fusions as well as intrasequence insertions of single or multiple amino acid residues. Insertions ordinarily will be smaller insertions than those of amino or carboxyl terminal fusions, for example, on the order of one to four residues. Immunogenic fusion protein derivatives, such as those described in the examples, are made by fusing a polypeptide sufficiently large to confer immunogenicity to the target sequence by cross-linking in vitro or by recombinant cell culture transformed with DNA encoding the fusion. Deletions are characterized by the removal of one or more amino acid residues from the protein sequence. Typically, no more than about from 2 to 6 residues are deleted at any one site within the protein molecule. These variants ordinarily are prepared by site specific mutagenesis of nucleotides in the DNA encoding the protein, thereby producing DNA encoding the variant, and thereafter expressing the DNA in recombinant cell culture. Techniques for making substitution mutations at predetermined sites in DNA having a known sequence are well known, for example M13 primer mutagenesis and PCR mutagenesis. Amino acid substitutions are typically of single residues, but can occur at a number of different locations at once; insertions usually will be on the order of about from 1 to 10 amino acid residues; and deletions will range about from 1 to 30 residues. Deletions or insertions preferably are made in adjacent pairs, i.e. a deletion of 2 residues or insertion of 2 residues. Substitutions, deletions, insertions or any combination thereof can be combined to arrive at a final construct. The mutations must not place the sequence out of reading frame and preferably will not create complementary regions that could produce secondary mRNA structure. Substitutional variants are those in which at least one residue has been removed and a different residue inserted in its place. Such substitutions generally are made in accordance with the following Tables 1 and 2 and are referred to as conservative substitutions.

TABLE 1

Amino Acid Abbreviations	
Amino Acid	Abbreviations
alanine	Ala A
allosoleucine	Alle
arginine	Arg R
asparagine	Asn N
aspartic acid	Asp D
cysteine	Cys C
glutamic acid	Glu E
glutamine	Gln Q
glycine	Gly G
histidine	His H
isoleucine	Ile I
leucine	Leu L
lysine	Lys K
phenylalanine	Phe F
proline	Pro P

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TABLE 1-continued

Amino Acid Abbreviations	
Amino Acid	Abbreviations
pyroglutamic acidp	pGlu
serine	Ser S
threonine	Thr T
tyrosine	Tyr Y
tryptophan	Trp W
valine	Val V

TABLE 2

Amino Acid Substitutions	
Original Residue	Exemplary Conservative Substitutions, others are known in the art.
	Ala; Ser
	Arg; Lys; Gln
	Asn; Gln; His
	Asp; Glu
	Cys; Ser
	Gln; Asn; Lys
	Glu; Asp
	Gly; Pro
	His; Asn; Gln
	Ile; Leu; Val
	Leu; Ile; Val
	Lys; Arg; Gln;
	Met; Leu; Ile
	Phe; Met; Leu; Tyr
	Ser; Thr
	Thr; Ser
	Trp; Tyr
	Tyr; Trp; Phe
	Val; Ile; Leu

Substantial changes in function or immunological identity are made by selecting substitutions that are less conservative than those in Table 2, i.e., selecting residues that differ more significantly in their effect on maintaining (a) the structure of the polypeptide backbone in the area of the substitution, for example as a sheet or helical conformation, (b) the charge or hydrophobicity of the molecule at the target site or (c) the bulk of the side chain. The substitutions which in general are expected to produce the greatest changes in the protein properties will be those in which (a) a hydrophilic residue, e.g. seryl or threonyl, is substituted for (or by) a hydrophobic residue, e.g. leucyl, isoleucyl, phenylalanyl, valyl or alanyl; (b) a cysteine or proline is substituted for (or by) any other residue; (c) a residue having an electropositive side chain, e.g., lysyl, arginyl, or histidyl, is substituted for (or by) an electronegative residue, e.g., glutamyl or aspartyl; or (d) a residue having a bulky side chain, e.g., phenylalanine, is substituted for (or by) one not having a side chain, e.g., glycine, in this case, (e) by increasing the number of sites for sulfation and/or glycosylation.

For example, the replacement of one amino acid residue with another that is biologically and/or chemically similar is known to those skilled in the art as a conservative substitution. For example, a conservative substitution would be replacing one hydrophobic residue for another, or one polar residue for another. The substitutions include combinations such as, for example, Gly, Ala; Val, Ile, Leu; Asp, Glu; Asn, Gln; Ser, Thr; Lys, Arg; and Phe, Tyr. Such conservatively substituted variations of each explicitly disclosed sequence are included within the mosaic polypeptides provided herein.

Substitutional or deletional mutagenesis can be employed to insert sites for N-glycosylation (Asn-X-Thr/Ser) or O-glycosylation (Ser or Thr). Deletions of cysteine or other labile residues also may be desirable. Deletions or substitutions of potential proteolysis sites, e.g. Arg, is accomplished for example by deleting one of the basic residues or substituting one by glutaminyl or histidyl residues.

Certain post-translational derivatizations are the result of the action of recombinant host cells on the expressed polypeptide. Glutaminyl and asparaginyl residues are frequently post-translationally deamidated to the corresponding glutamyl and asparyl residues. Alternatively, these residues are deamidated under mildly acidic conditions. Other post-translational modifications include hydroxylation of proline and lysine, phosphorylation of hydroxyl groups of seryl or threonyl residues, methylation of the o-amino groups of lysine, arginine, and histidine side chains (T. E. Creighton, *Proteins: Structure and Molecular Properties*, W. H. Freeman & Co., San Francisco pp 79-86 [1983]), acetylation of the N-terminal amine and, in some instances, amidation of the C-terminal carboxyl.

It is understood that one way to define the variants and derivatives of the disclosed proteins herein is through defining the variants and derivatives in terms of homology/identity to specific known sequences. For example, SEQ ID NO: 10 sets forth a particular sequence of K9/9 and SEQ ID NO: 2 sets forth a particular sequence of a DAF protein. Specifically disclosed are variants of these and other proteins herein disclosed which have at least, 70% or 75% or 80% or 85% or 90% or 95% homology to the stated sequence. Those of skill in the art readily understand how to determine the homology of two proteins. For example, the homology can be calculated after aligning the two sequences so that the homology is at its highest level.

Another way of calculating homology can be performed by published algorithms. Optimal alignment of sequences for comparison can be conducted by the local homology algorithm of Smith and Waterman *Adv. Appl. Math.* 2: 482 (1981), by the homology alignment algorithm of Needleman and Wunsch, *J. Mol. Biol.* 48: 443 (1970), by the search for similarity method of Pearson and Lipman, *Proc. Natl. Acad. Sci. U.S.A.* 85: 2444 (1988), by computerized implementations of these algorithms (GAP, BESTFIT, FASTA, and TFASTA in the Wisconsin Genetics Software Package, Genetics Computer Group, 575 Science Dr., Madison, Wis.), or by inspection.

The same types of homology can be obtained for nucleic acids by for example the algorithms disclosed in Zuker, *M. Science* 244:48-52, 1989, Jaeger et al. *Proc. Natl. Acad. Sci. USA* 86:7706-7710, 1989, Jaeger et al. *Methods Enzymol.* 183:281-306, 1989 which are herein incorporated by reference for at least material related to nucleic acid alignment.

It is understood that the description of conservative mutations and homology can be combined together in any combination, such as embodiments that have at least 70% homology to a particular sequence wherein the variants are conservative mutations.

As this specification discusses various proteins and protein sequences it is understood that the nucleic acids that can encode those protein sequences are also disclosed. This would include all degenerate sequences related to a specific protein sequence, i.e. all nucleic acids having a sequence that encodes one particular protein sequence as well as all nucleic acids, including degenerate nucleic acids, encoding the disclosed variants and derivatives of the protein sequences. Thus, while each particular nucleic acid sequence may not be written out herein, it is understood that each and every

sequence is in fact disclosed and described herein through the disclosed protein sequence. It is understood that for this mutation all of the nucleic acid sequences that encode this particular derivative of any of the disclosed sequences are also disclosed. It is also understood that while no amino acid sequence indicates what particular DNA sequence encodes that protein within an organism, where particular variants of a disclosed protein are disclosed herein, the known nucleic acid sequence that encodes that protein from which that protein arises is also known and herein disclosed and described.

6. Antibodies

a) Antibodies Generally

The term "antibodies" is used herein in a broad sense and includes both polyclonal and monoclonal antibodies. In addition to intact immunoglobulin molecules, also included in the term "antibodies" are fragments or polymers of those immunoglobulin molecules, and human or humanized versions of immunoglobulin molecules or fragments thereof, as described herein. The antibodies are tested for their desired activity using the in vitro assays described herein, or by analogous methods, after which their in vivo therapeutic and/or prophylactic activities are tested according to known clinical testing methods.

As used herein, the term "antibody" encompasses, but is not limited to, whole immunoglobulin (i.e., an intact antibody) of any class. Native antibodies are usually heterotetrameric glycoproteins, composed of two identical light (L) chains and two identical heavy (H) chains. Typically, each light chain is linked to a heavy chain by one covalent disulfide bond, while the number of disulfide linkages varies between the heavy chains of different immunoglobulin isotypes. Each heavy and light chain also has regularly spaced intrachain disulfide bridges. Each heavy chain has at one end a variable domain (V(H)) followed by a number of constant domains. Each light chain has a variable domain at one end (V(L)) and a constant domain at its other end; the constant domain of the light chain is aligned with the first constant domain of the heavy chain, and the light chain variable domain is aligned with the variable domain of the heavy chain. Particular amino acid residues are believed to form an interface between the light and heavy chain variable domains. The light chains of antibodies from any vertebrate species can be assigned to one of two clearly distinct types, called kappa (κ) and lambda (λ), based on the amino acid sequences of their constant domains. Depending on the amino acid sequence of the constant domain of their heavy chains, immunoglobulins can be assigned to different classes. There are five major classes of human immunoglobulins: IgA, IgD, IgE, IgG and IgM, and several of these can be further divided into subclasses (isotypes), e.g., IgG-1, IgG-2, IgG-3, and IgG-4; IgA-1 and IgA-2. One skilled in the art would recognize the comparable classes for mouse. The heavy chain constant domains that correspond to the different classes of immunoglobulins are called alpha, delta, epsilon, gamma, and mu, respectively.

The term "variable" is used herein to describe certain portions of the variable domains that differ in sequence among antibodies and are used in the binding and specificity of each particular antibody for its particular antigen. However, the variability is not usually evenly distributed through the variable domains of antibodies. It is typically concentrated in three segments called complementarity determining regions (CDRs) or hypervariable regions both in the light chain and the heavy chain variable domains. The more highly conserved portions of the variable domains are called the framework (FR). The variable domains of native heavy and light chains

each comprise four FR regions, largely adopting a b-sheet configuration, connected by three CDRs, which form loops connecting, and in some cases forming part of, the b-sheet structure. The CDRs in each chain are held together in close proximity by the FR regions and, with the CDRs from the other chain, contribute to the formation of the antigen binding site of antibodies (see Kabat E. A. et al., "Sequences of Proteins of Immunological Interest," National Institutes of Health, Bethesda, Md. (1987)). The constant domains are not involved directly in binding an antibody to an antigen, but exhibit various effector functions, such as participation of the antibody in antibody-dependent cellular toxicity.

As used herein, the term "antibody or fragments thereof" encompasses chimeric antibodies and hybrid antibodies, with dual or multiple antigen or epitope specificities, and fragments, such as scFv, sFv, F(ab')₂, Fab', Fab and the like, including hybrid fragments. Thus, fragments of the antibodies that retain the ability to bind their specific antigens are provided. For example, fragments of antibodies which maintain complement binding activity binding activity are included within the meaning of the term "antibody or fragment thereof." Such antibodies and fragments can be made by techniques known in the art and can be screened for specificity and activity according to the methods set forth in the Examples and in general methods for producing antibodies and screening antibodies for specificity and activity (See Harlow and Lane. *Antibodies, A Laboratory Manual*. Cold Spring Harbor Publications, New York, (1988)).

Also included within the meaning of "antibody or fragments thereof" are conjugates of antibody fragments and antigen binding proteins (single chain antibodies) as described, for example, in U.S. Pat. No. 4,704,692, the contents of which are hereby incorporated by reference.

As used herein, the term "antibody" or "antibodies" can also refer to a human antibody and/or a humanized antibody. Many non-human antibodies (e.g., those derived from mice, rats, or rabbits) are naturally antigenic in humans, and thus can give rise to undesirable immune responses when administered to humans. Therefore, the use of human or humanized antibodies in the methods of the invention serves to lessen the chance that an antibody administered to a human will evoke an undesirable immune response.

Optionally, the antibodies are generated in other species and "humanized" for administration in humans. Humanized forms of non-human (e.g., murine) antibodies are chimeric immunoglobulins, immunoglobulin chains or fragments thereof (such as Fc, scFv, sFv, Fv, Fab, Fab', F(ab')₂, or other antigen-binding subsequences of antibodies) which contain minimal sequence derived from non-human immunoglobulin. Humanized antibodies include human immunoglobulins (recipient antibody) in which residues from a complementary determining region (CDR) of the recipient are replaced by residues from a CDR of a non-human species (donor antibody) such as mouse, rat or rabbit having the desired specificity, affinity and capacity. In some instances, Fv framework residues of the human immunoglobulin are replaced by corresponding non-human residues. Humanized antibodies can also comprise residues that are found neither in the recipient antibody nor in the imported CDR or framework sequences. In general, the humanized antibody will comprise substantially all of at least one, and typically two, variable domains, in which all or substantially all of the CDR regions correspond to those of a non-human immunoglobulin and all or substantially all of the FR regions are those of a human immunoglobulin consensus sequence. The humanized antibody optimally also will comprise at least a portion of an immunoglobulin constant region (Fc), typically that of a

human immunoglobulin (Jones et al., *Nature*, 321:522-525 (1986); Riechmann et al., *Nature*, 332:323-327 (1988); and Presta, *Curr. Op. Struct. Biol.*, 2:593-596 (1992)).

Methods for humanizing non-human antibodies are well known in the art. Generally, a humanized antibody has one or more amino acid residues introduced into it from a source that is non-human. These non-human amino acid residues are often referred to as "import" residues, which are typically taken from an "import" variable domain. Humanization can be essentially performed following the method of Winter and co-workers (Jones et al., *Nature*, 321:522-525 (1986); Riechmann et al., *Nature*, 332:323-327 (1988); Verhoeven et al., *Science*, 239:1534-1536 (1988)), by substituting rodent CDRs or CDR sequences for the corresponding sequences of a human antibody. Accordingly, such "humanized" antibodies are chimeric antibodies (U.S. Pat. No. 4,816,567), wherein substantially less than an intact human variable domain has been substituted by the corresponding sequence from a non-human species. In practice, humanized antibodies are typically human antibodies in which some CDR residues and possibly some FR residues are substituted by residues from analogous sites in rodent antibodies.

The choice of human variable domains, both light and heavy, to be used in making the humanized antibodies is very important in order to reduce antigenicity. According to the "best-fit" method, the sequence of the variable domain of a rodent antibody is screened against the entire library of known human variable domain sequences. The human sequence which is closest to that of the rodent is then accepted as the human framework (FR) for the humanized antibody (Sims et al., *J. Immunol.*, 151:2296 (1993) and Chothia et al., *J. Mol. Biol.*, 196:901 (1987)). Another method uses a particular framework derived from the consensus sequence of all human antibodies of a particular subgroup of light or heavy chains. The same framework can be used for several different humanized antibodies (Carter et al., *Proc. Natl. Acad. Sci. USA*, 89:4285 (1992); Presta et al., *J. Immunol.*, 151:2623 (1993)).

It is further important that antibodies be humanized with retention of high affinity for the antigen and other favorable biological properties. To achieve this goal, according to a preferred method, humanized antibodies are prepared by a process of analysis of the parental sequences and various conceptual humanized products using three dimensional models of the parental and humanized sequences. Three dimensional immunoglobulin models are commonly available and are familiar to those skilled in the art. Computer programs are available which illustrate and display probable three-dimensional conformational structures of selected candidate immunoglobulin sequences. Inspection of these displays permits analysis of the likely role of the residues in the functioning of the candidate immunoglobulin sequence, i.e., the analysis of residues that influence the ability of the candidate immunoglobulin to bind its antigen. In this way, FR residues can be selected and combined from the consensus and import sequence so that the desired antibody characteristic, such as increased affinity for the target antigen(s), is achieved. In general, the CDR residues are directly and most substantially involved in influencing antigen binding (see, WO 94/04679, published 3 Mar. 1994).

The Fab fragments produced in the antibody digestion also contain the constant domains of the light chain and the first constant domain of the heavy chain. Fab' fragments differ from Fab fragments by the addition of a few residues at the carboxy terminus of the heavy chain domain including one or more cysteines from the antibody hinge region. The F(ab')₂ fragment is a bivalent fragment comprising two Fab' frag-

ments linked by a disulfide bridge at the hinge region. Fab'-SH is the designation herein for Fab' in which the cysteine residue(s) of the constant domains bear a free thiol group. Antibody fragments originally were produced as pairs of Fab' fragments which have hinge cysteines between them. Other chemical couplings of antibody fragments are also known.

An isolated immunogenically specific paratope or fragment of the antibody is also provided. A specific immunogenic epitope of the antibody can be isolated from the whole antibody by chemical or mechanical disruption of the molecule. The purified fragments thus obtained are tested to determine their immunogenicity and specificity by the methods taught herein. Immunoreactive paratopes of the antibody, optionally, are synthesized directly. An immunoreactive fragment is defined as an amino acid sequence of at least about two to five consecutive amino acids derived from the antibody amino acid sequence.

One method of producing proteins comprising the antibodies of the present invention is to link two or more peptides or polypeptides together by protein chemistry techniques. For example, peptides or polypeptides can be chemically synthesized using currently available laboratory equipment using either Fmoc (9-fluorenylmethyloxycarbonyl) or Boc (tert-butyloxycarbonyl) chemistry. (Applied Biosystems, Inc., Foster City, Calif.). One skilled in the art can readily appreciate that a peptide or polypeptide corresponding to the antibody of the present invention, for example, can be synthesized by standard chemical reactions. For example, a peptide or polypeptide can be synthesized and not cleaved from its synthesis resin whereas the other fragment of an antibody can be synthesized and subsequently cleaved from the resin, thereby exposing a terminal group which is functionally blocked on the other fragment. By peptide condensation reactions, these two fragments can be covalently joined via a peptide bond at their carboxyl and amino termini, respectively, to form an antibody, or fragment thereof. (Grant G A (1992) *Synthetic Peptides: A User Guide*. W.H. Freeman and Co., N.Y. (1992); Bodansky M and Trost B., Ed. (1993) *Principles of Peptide Synthesis*. Springer-Verlag Inc., NY. Alternatively, the peptide or polypeptide is independently synthesized *in vivo* as described above. Once isolated, these independent peptides or polypeptides can be linked to form an antibody or fragment thereof via similar peptide condensation reactions.

For example, enzymatic ligation of cloned or synthetic peptide segments allow relatively short peptide fragments to be joined to produce larger peptide fragments, polypeptides or whole protein domains (Abrahmsen L et al., *Biochemistry*, 30:4151 (1991)). Alternatively, native chemical ligation of synthetic peptides can be utilized to synthetically construct large peptides or polypeptides from shorter peptide fragments. This method consists of a two step chemical reaction (Dawson et al. *Synthesis of Proteins by Native Chemical Ligation*. *Science*, 266:776-779 (1994)). The first step is the chemoselective reaction of an unprotected synthetic peptide-alpha-thioester with another unprotected peptide segment containing an amino-terminal Cys residue to give a thioester-linked intermediate as the initial covalent product. Without a change in the reaction conditions, this intermediate undergoes spontaneous, rapid intramolecular reaction to form a native peptide bond at the ligation site. Application of this native chemical ligation method to the total synthesis of a protein molecule is illustrated by the preparation of human interleukin 8 (IL-8) (Baggiolini M et al. (1992) *FEBS Lett.* 307:97-101; Clark-Lewis I et al., *J. Biol. Chem.*, 269:16075 (1994); Clark-Lewis I et al., *Biochemistry*, 30:3128 (1991); Rajarathnam K et al., *Biochemistry* 33:6623-30 (1994)).

Alternatively, unprotected peptide segments are chemically linked where the bond formed between the peptide segments as a result of the chemical ligation is an unnatural (non-peptide) bond (Schnolzer, M et al. *Science*, 256:221 (1992)). This technique has been used to synthesize analogs of protein domains as well as large amounts of relatively pure proteins with full biological activity (deLisle Milton R C et al., *Techniques in Protein Chemistry IV*. Academic Press, New York, pp. 257-267 (1992)).

The invention also provides fragments of antibodies which have bioactivity. The polypeptide fragments of the present invention can be recombinant proteins obtained by cloning nucleic acids encoding the polypeptide in an expression system capable of producing the polypeptide fragments thereof, such as an adenovirus or baculovirus expression system. For example, one can determine the active domain of an antibody from a specific hybridoma that can cause a biological effect associated with the interaction of the antibody with an Fc receptor. For example, amino acids found to not contribute to either the activity or the binding specificity or affinity of the antibody can be deleted without a loss in the respective activity. For example, in various embodiments, amino or carboxy-terminal amino acids are sequentially removed from either the native or the modified non-immunoglobulin molecule or the immunoglobulin molecule and the respective activity assayed in one of many available assays. In another example, a fragment of an antibody comprises a modified antibody wherein at least one amino acid has been substituted for the naturally occurring amino acid at a specific position, and a portion of either amino terminal or carboxy terminal amino acids, or even an internal region of the antibody, has been replaced with a polypeptide fragment or other moiety, such as biotin, which can facilitate in the purification of the modified antibody.

The fragments, whether attached to other sequences or not, include insertions, deletions, substitutions, or other selected modifications of particular regions or specific amino acid residues, provided the activity of the fragment is not significantly altered or impaired compared to the nonmodified antibody or antibody fragment. These modifications can provide for some additional property, such as to remove or add amino acids capable of disulfide bonding, to increase its bio-longevity, to alter its secretory characteristics, etc. In any case, the fragment must possess a bioactive property, such as binding activity, regulation of binding at the binding domain, etc. Functional or active regions of the antibody can be identified by mutagenesis of a specific region of the protein, followed by expression and testing of the expressed polypeptide. Such methods are readily apparent to a skilled practitioner in the art and can include site-specific mutagenesis of the nucleic acid encoding the antigen. (Zoller M J et al. *Nucl. Acids Res.* 10:6487-500 (1982)).

A variety of immunoassay formats can be used to select antibodies that selectively bind with a particular protein, variant, or fragment. For example, solid-phase ELISA immunoassays are routinely used to select antibodies selectively immunoreactive with a protein, protein variant, or fragment thereof. See Harlow and Lane. *Antibodies, A Laboratory Manual*. Cold Spring Harbor Publications, New York, (1988), for a description of immunoassay formats and conditions that could be used to determine selective binding. The binding affinity of a monoclonal antibody can, for example, be determined by the Scatchard analysis of Munson et al., *Anal. Biochem.*, 107:220 (1980).

Also provided is an antibody reagent kit comprising containers of the monoclonal antibody or fragment thereof of the invention and one or more reagents for detecting binding of

the antibody or fragment thereof to the Fc receptor molecule. The reagents can include, for example, fluorescent tags, enzymatic tags, or other tags. The reagents can also include secondary or tertiary antibodies or reagents for enzymatic reactions, wherein the enzymatic reactions produce a product that can be visualized.

b) Human Antibodies

The human antibodies of the invention can be prepared using any technique. Examples of techniques for human monoclonal antibody production include those described by Cole et al. (*Monoclonal Antibodies and Cancer Therapy*, Alan R. Liss, p. 77, 1985) and by Boerner et al. (*J. Immunol.*, 147(1):86-95, 1991). Human antibodies of the invention (and fragments thereof) can also be produced using phage display libraries (Hoogenboom et al., *J. Mol. Biol.*, 227:381, 1991; Marks et al., *J. Mol. Biol.*, 222:581, 1991).

The human antibodies of the invention can also be obtained from transgenic animals. For example, transgenic, mutant mice that are capable of producing a full repertoire of human antibodies, in response to immunization, have been described (see, e.g., Jakobovits et al., *Proc. Natl. Acad. Sci. USA*, 90:2551-255 (1993); Jakobovits et al., *Nature*, 362:255-258 (1993); Bruggermann et al., *Year in Immunol.*, 7:33 (1993)). Specifically, the homozygous deletion of the antibody heavy chain joining region (J(H)) gene in these chimeric and germ-line mutant mice results in complete inhibition of endogenous antibody production, and the successful transfer of the human germ-line antibody gene array into such germ-line mutant mice results in the production of human antibodies upon antigen challenge. Antibodies having the desired activity are selected using Env-CD4-co-receptor complexes as described herein.

c) Administration of Antibodies

Antibodies of the invention are preferably administered to a subject in a pharmaceutically acceptable carrier. Suitable carriers and their formulations are described in *Remington: The Science and Practice of Pharmacy* (19th ed.) ed. A. R. Gennaro, Mack Publishing Company, Easton, Pa. 1995. Typically, an appropriate amount of a pharmaceutically-acceptable salt is used in the formulation to render the formulation isotonic. Examples of the pharmaceutically-acceptable carrier include, but are not limited to, saline, Ringer's solution and dextrose solution. The pH of the solution is preferably from about 5 to about 8, and more preferably from about 7 to about 7.5. Further carriers include sustained release preparations such as semipermeable matrices of solid hydrophobic polymers containing the antibody, which matrices are in the form of shaped articles, e.g., films, liposomes or microparticles. It will be apparent to those persons skilled in the art that certain carriers may be more preferable depending upon, for instance, the route of administration and concentration of antibody being administered.

The antibodies can be administered to the subject, patient, or cell by injection (e.g., intravenous, intraperitoneal, subcutaneous, intramuscular), or by other methods such as infusion that ensure its delivery to the bloodstream in an effective form. Local or intravenous injection is preferred.

Effective dosages and schedules for administering the antibodies can be determined empirically, and making such determinations is within the skill in the art. Those skilled in the art will understand that the dosage of antibodies that must be administered will vary depending on, for example, the subject that will receive the antibody, the route of administration, the

particular type of antibody used and other drugs being administered. Guidance in selecting appropriate doses for antibodies is found in the literature on therapeutic uses of antibodies, e.g., *Handbook of Monoclonal Antibodies*, Ferrone et al., eds., Noyes Publications, Park Ridge, N.J., (1985) ch. 22 and pp. 303-357; Smith et al., *Antibodies in Human Diagnosis and Therapy*, Haber et al., eds., Raven Press, New York (1977) pp. 365-389. A typical daily dosage of the antibody used alone might range from about 1 µg/kg to up to 100 mg/kg of body weight or more per day, depending on the factors mentioned above.

Following administration of an antibody for treating, inhibiting, or preventing an HIV infection, the efficacy of the therapeutic antibody can be assessed in various ways well known to the skilled practitioner. For instance, one of ordinary skill in the art will understand that an antibody of the invention is efficacious in treating or inhibiting an HIV infection in a subject by observing that the antibody reduces viral load or prevents a further increase in viral load. Viral loads can be measured by methods that are known in the art, for example, using polymerase chain reaction assays to detect the presence of HIV nucleic acid or antibody assays to detect the presence of HIV protein in a sample (e.g., but not limited to, blood) from a subject or patient, or by measuring the level of circulating anti-HIV antibody levels in the patient. Efficacy of the antibody treatment can also be determined by measuring the number of CD4⁺ T cells in the HIV-infected subject. An antibody treatment that inhibits an initial or further decrease in CD4⁺ T cells in an HIV-positive subject or patient, or that results in an increase in the number of CD4⁺ T cells in the HIV-positive subject, is an efficacious antibody treatment.

d) Nucleic Acid Approaches for Antibody Delivery

The compositions of the invention can also be administered to patients or subjects as a nucleic acid preparation (e.g., DNA or RNA) that encodes the antibody or antibody fragment, such that the patient's or subject's own cells take up the nucleic acid and produce and secrete the encoded composition (e.g., K9/9-DAF, K9/9-CD59, K9/9-CR1, K9/9-MCP, K9/9-Crry, K9/9-IgG1 Fc (human), K9/9-IgM Fc, K9/9-IgG3 Fc (murine), or K9/9-CVF).

e) Nucleic Acid Delivery

In the methods described above which include the administration and uptake of exogenous DNA into the cells of a subject (i.e., gene transduction or transfection), the nucleic acids of the present invention can be in the form of naked DNA or RNA, or the nucleic acids can be in a vector for delivering the nucleic acids to the cells, whereby the antibody-encoding DNA fragment is under the transcriptional regulation of a promoter, as would be well understood by one of ordinary skill in the art. The vector can be a commercially available preparation, such as an adenovirus vector (Quantum Biotechnologies, Inc. (Laval, Quebec, Canada). Delivery of the nucleic acid or vector to cells can be via a variety of mechanisms. As one example, delivery can be via a liposome, using commercially available liposome preparations such as LIPOFECTIN, LIPOFECTAMINE (GIBCO-BRL, Inc., Gaithersburg, Md.), SUPERFECT (Qiagen, Inc. Hilden, Germany) and TRANSFECTAM (Promega Biotec, Inc., Madison, Wis.), as well as other liposomes developed according to procedures standard in the art. In addition, the nucleic acid or vector of this invention can be delivered in vivo by electroporation, the technology for which is available from Genetron

ics, Inc. (San Diego, Calif.) as well as by means of a SONOPORATION machine (ImaRx Pharmaceutical Corp., Tucson, Ariz.).

As one example, vector delivery can be via a viral system, such as a retroviral vector system which can package a recombinant retroviral genome (see e.g., Pastan et al., *Proc. Natl. Acad. Sci. U.S.A.* 85:4486, 1988; Miller et al., *Mol. Cell. Biol.* 6:2895, 1986). The recombinant retrovirus can then be used to infect and thereby deliver to the infected cells nucleic acid encoding a broadly neutralizing antibody (or active fragment thereof) of the invention. The exact method of introducing the altered nucleic acid into mammalian cells is, of course, not limited to the use of retroviral vectors. Other techniques are widely available for this procedure including the use of adenoviral vectors (Mitani et al., *Hum. Gene Ther.* 5:941-948, 1994), adeno-associated viral (AAV) vectors (Goodman et al., *Blood* 84:1492-1500, 1994), lentiviral vectors (Naidini et al., *Science* 272:263-267, 1996), pseudotyped retroviral vectors (Agrawal et al., *Exper. Hematol.* 24:738-747, 1996). Physical transduction techniques can also be used, such as liposome delivery and receptor-mediated and other endocytosis mechanisms (see, for example, Schwartzberger et al., *Blood* 87:472-478, 1996). This invention can be used in conjunction with any of these or other commonly used gene transfer methods.

As one example, if the complement modulating construct-encoding nucleic acid of the invention is delivered to the cells of a subject in an adenovirus vector, the dosage for administration of adenovirus to humans can range from about 10^7 to 10^9 plaque forming units (pfu) per injection but can be as high as 10^{12} pfu per injection (Crystal, *Hum. Gene Ther.* 8:985-1001, 1997; Alvarez and Curiel, *Hum. Gene Ther.* 8:597-613, 1997). A subject can receive a single injection, or, if additional injections are necessary, they can be repeated at six month intervals (or other appropriate time intervals, as determined by the skilled practitioner) for an indefinite period and/or until the efficacy of the treatment has been established.

Parenteral administration of the nucleic acid or vector of the present invention, if used, is generally characterized by injection. Injectables can be prepared in conventional forms, either as liquid solutions or suspensions, solid forms suitable for solution of suspension in liquid prior to injection, or as emulsions. A more recently revised approach for parenteral administration involves use of a slow release or sustained release system such that a constant dosage is maintained. See, e.g., U.S. Pat. No. 3,610,795, which is incorporated by reference herein. For additional discussion of suitable formulations and various routes of administration of therapeutic compounds, see, e.g., *Remington: The Science and Practice of Pharmacy* (19th ed.) ed. A. R. Gennaro, Mack Publishing Company, Easton, Pa. 1995.

7. Pharmaceutical Carriers/Delivery of Pharmaceutical Products

As described above, the compositions can also be administered in vivo in a pharmaceutically acceptable carrier. By "pharmaceutically acceptable" is meant a material that is not biologically or otherwise undesirable, i.e., the material can be administered to a subject, along with the nucleic acid or vector, without causing any undesirable biological effects or interacting in a deleterious manner with any of the other components of the pharmaceutical composition in which it is contained. The carrier would naturally be selected to minimize any degradation of the active ingredient and to minimize any adverse side effects in the subject, as would be well known to one of skill in the art.

The compositions can be administered orally, parenterally (e.g., intravenously), by intramuscular injection, by intraperi-

toneal injection, transdermally, extracorporeally, topically or the like, although topical intranasal administration or administration by inhalant is typically preferred. As used herein, "topical intranasal administration" means delivery of the compositions into the nose and nasal passages through one or both of the nares and can comprise delivery by a spraying mechanism or droplet mechanism, or through aerosolization of the nucleic acid or vector. The latter is effective when a large number of animals is to be treated simultaneously. Administration of the compositions by inhalant can be through the nose or mouth via delivery by a spraying or droplet mechanism. Delivery can also be directly to any area of the respiratory system (e.g., lungs) via intubation. The exact amount of the compositions required will vary from subject to subject, depending on the species, age, weight and general condition of the subject, the severity of the allergic disorder being treated, the particular nucleic acid or vector used, its mode of administration and the like. Thus, it is not possible to specify an exact amount for every composition. However, an appropriate amount can be determined by one of ordinary skill in the art using only routine experimentation given the teachings herein.

Parenteral administration of the composition, if used, is generally characterized by injection. Injectables can be prepared in conventional forms, either as liquid solutions or suspensions, solid forms suitable for solution of suspension in liquid prior to injection, or as emulsions. A more recently revised approach for parenteral administration involves use of a slow release or sustained release system such that a constant dosage is maintained. See, e.g., U.S. Pat. No. 3,610,795, which is incorporated by reference herein.

The materials can be in solution, suspension (for example, incorporated into microparticles, liposomes, or cells). These can be targeted to a particular cell type via antibodies, receptors, or receptor ligands. The following references are examples of the use of this technology to target specific proteins to tumor tissue (Senter, et al., *Bioconjugate Chem.*, 2:447-451, (1991); Bagshawe, K. D., *Br. J. Cancer*, 60:275-281, (1989); Bagshawe, et al., *Br. J. Cancer*, 58:700-703, (1988); Senter, et al., *Bioconjugate Chem.*, 4:3-9, (1993); Battelli, et al., *Cancer Immunol. Immunother.*, 35:421-425, (1992); Pietersz and McKenzie, *Immunolog. Reviews*, 129: 57-80, (1992); and Roffler, et al., *Biochem. Pharmacol.*, 42:2062-2065, (1991)). Vehicles such as "stealth" and other antibody conjugated liposomes (including lipid mediated drug targeting to colonic carcinoma), receptor mediated targeting of DNA through cell specific ligands, lymphocyte directed tumor targeting, and highly specific therapeutic retroviral targeting of murine glioma cells in vivo. The following references are examples of the use of this technology to target specific proteins to tumor tissue (Hughes et al., *Cancer Research*, 49:6214-6220, (1989); and Litzinger and Huang, *Biochimica et Biophysica Acta*, 1104:179-187, (1992)). In general, receptors are involved in pathways of endocytosis, either constitutive or ligand induced. These receptors cluster in clathrin-coated pits, enter the cell via clathrin-coated vesicles, pass through an acidified endosome in which the receptors are sorted, and then either recycle to the cell surface, become stored intracellularly, or are degraded in lysosomes. The internalization pathways serve a variety of functions, such as nutrient uptake, removal of activated proteins, clearance of macromolecules, opportunistic entry of viruses and toxins, dissociation and degradation of ligand, and receptor-level regulation. Many receptors follow more than one intracellular pathway, depending on the cell type, receptor concentration, type of ligand, ligand valency, and ligand concentration. Molecular and cellular mechanisms of recep-

tor-mediated endocytosis has been reviewed (Brown and Greene, *DNA and Cell Biology* 10:6, 399-409 (1991)).

a) Pharmaceutically Acceptable Carriers

The compositions, including antibodies, can be used therapeutically in combination with a pharmaceutically acceptable carrier.

Pharmaceutical carriers are known to those skilled in the art. These most typically would be standard carriers for administration of drugs to humans, including solutions such as sterile water, saline, and buffered solutions at physiological pH. The compositions can be administered intramuscularly or subcutaneously. Other compounds will be administered according to standard procedures used by those skilled in the art.

Pharmaceutical compositions can include carriers, thickeners, diluents, buffers, preservatives, surface active agents and the like in addition to the molecule of choice. Pharmaceutical compositions can also include one or more active ingredients such as antimicrobial agents, antiinflammatory agents, anesthetics, and the like.

The pharmaceutical composition can be administered in a number of ways depending on whether local or systemic treatment is desired, and on the area to be treated. Administration can be topically (including ophthalmically, vaginally, rectally, intranasally), orally, by inhalation, or parenterally, for example by intravenous drip, subcutaneous, intraperitoneal or intramuscular injection. The disclosed antibodies can be administered intravenously, intraperitoneally, intramuscularly, subcutaneously, intracavity, or transdermally.

Preparations for parenteral administration include sterile aqueous or non-aqueous solutions, suspensions, and emulsions. Examples of non-aqueous solvents are propylene glycol, polyethylene glycol, vegetable oils such as olive oil, and injectable organic esters such as ethyl oleate. Aqueous carriers include water, alcoholic/aqueous solutions, emulsions or suspensions, including saline and buffered media. Parenteral vehicles include sodium chloride solution, Ringer's dextrose, dextrose and sodium chloride, lactated Ringer's, or fixed oils. Intravenous vehicles include fluid and nutrient replenishers, electrolyte replenishers (such as those based on Ringer's dextrose), and the like. Preservatives and other additives can also be present such as, for example, antimicrobials, antioxidants, chelating agents, and inert gases and the like.

Formulations for topical administration can include ointments, lotions, creams, gels, drops, suppositories, sprays, liquids and powders. Conventional pharmaceutical carriers, aqueous, powder or oily bases, thickeners and the like may be necessary or desirable.

Compositions for oral administration include powders or granules, suspensions or solutions in water or non-aqueous media, capsules, sachets, or tablets. Thickeners, flavorings, diluents, emulsifiers, dispersing aids or binders may be desirable.

Some of the compositions can be administered as a pharmaceutically acceptable acid- or base-addition salt, formed by reaction with inorganic acids such as hydrochloric acid, hydrobromic acid, perchloric acid, nitric acid, thiocyanic acid, sulfuric acid, and phosphoric acid, and organic acids such as formic acid, acetic acid, propionic acid, glycolic acid, lactic acid, pyruvic acid, oxalic acid, malonic acid, succinic acid, maleic acid, and fumaric acid, or by reaction with an inorganic base such as sodium hydroxide, ammonium

hydroxide, potassium hydroxide, and organic bases such as mono-, di-, trialkyl and aryl amines and substituted ethanamines.

b) Therapeutic Uses

The dosage ranges for the administration of the compositions are those large enough to produce the desired effect in which the symptoms of the disorder are effected. The dosage should not be so large as to cause adverse side effects, such as unwanted cross-reactions, anaphylactic reactions, and the like. Generally, the dosage will vary with the age, condition, sex and extent of the disease in the patient and can be determined by one of skill in the art. The dosage can be adjusted by the individual physician in the event of any counterindications. Dosage can vary, and can be administered in one or more dose administrations daily, for one or several days.

8. Kits

Disclosed herein are kits that are drawn to reagents that can be used in practicing the methods disclosed herein. The kits can include any reagent or combination of reagent discussed herein or that would be understood to be required or beneficial in the practice of the disclosed methods. For example, the kits could include primers to perform the amplification reactions discussed in certain embodiments of the methods, as well as the buffers and enzymes required to use the primers as intended. For example, disclosed is a kit for assessing a subject's risk for cancer, asthma, systemic lupus erythematosus, rheumatoid arthritis, reactive arthritis, spondylarthritis, systemic vasculitis, insulin dependent diabetes mellitus, multiple sclerosis, experimental allergic encephalomyelitis, Sjögren's syndrome, graft versus host disease, inflammatory bowel disease including Crohn's disease, ulcerative colitis, Ischemia reperfusion injury, myocardial infarction, Alzheimer's disease, transplant rejection (allogeneic and xenogeneic), thermal trauma, any immune complex-induced inflammation, glomerulonephritis, myasthenia gravis, multiple sclerosis, cerebral lupus, Guillain-Barre syndrome, vasculitis, systemic sclerosis, anaphylaxis, catheter reactions, atheroma, infertility, thyroiditis, ARDS, post-bypass syndrome, hemodialysis, juvenile rheumatoid, Behcets syndrome, hemolytic anemia, pemphigus, bullous pemphigoid, stroke, atherosclerosis, and scleroderma.

9. Compositions with Similar Functions

It is understood that the compositions disclosed herein have certain functions, such as modulating complement activity or binding glycoproteins expressed on rat glomerular epithelial cells and PTEC. Disclosed herein are certain structural requirements for performing the disclosed functions, and it is understood that there are a variety of structures which can perform the same function which are related to the disclosed structures, and that these structures will ultimately achieve the same result, for example stimulation or inhibition complement activity.

E. EXAMPLES

The following examples are put forth so as to provide those of ordinary skill in the art with a complete disclosure and description of how the compounds, compositions, articles, devices and/or methods claimed herein are made and evaluated, and are intended to be purely exemplary of the invention and are not intended to limit the scope of what the inventors regard as their invention. Efforts have been made to ensure accuracy with respect to numbers (e.g., amounts, temperature, etc.), but some errors and deviations should be accounted for. Unless indicated otherwise, parts are parts by

weight, temperature is in ° C. or is at ambient temperature, and pressure is at or near atmospheric.

1. Example 1

Complement Inhibitors Targeted to the Proximal Tubule

a) Results

In glomerular diseases of diverse etiologies, dysfunction of the glomerular barrier to protein passage results in proteinuria. There is a link between proteinuria and progressive renal damage, and proteinuria is considered an independent risk factor that plays a direct role in interstitial fibrosis and inflammation. The mechanism by which proteinuria leads to nephrotoxic injury is unclear, but an important role for complement in mediating interstitial damage appears likely (1-5). During glomerular proteinuria, complement proteins are present in the glomerular filtrate, and proximal tubular epithelial cells (PTEC) from both rat and human kidneys activate complement (6, 7). There is also evidence that tubular cells are an important local source of complement, and proteinuric filtrate may contain nephrotic factors that activate tubular cells and upregulate expression of complement proteins (4). Complement activation results in the formation of the C3 convertase, a central enzymatic complex which cleaves C3 leading to the generation of C3 opsonins and C3a. C3 convertase is also involved in the formation of the C5 convertase, an enzymatic complex that cleaves C5 to generate C5a and ultimately C5b-9 or the membrane attack complex (MAC). C3 opsonins, the proinflammatory C3a and C5a peptides and the cytolytic and proinflammatory MAC are variously implicated in pathogenic mechanisms when complement is inappropriately activated.

Under normal circumstances, host cells are effectively protected from complement attack by membrane complement regulators that function by inhibiting either C3 convertase or MAC formation. Human C3 inhibitors are complement receptor 1 (CR1), decay accelerating factor (DAF) and membrane cofactor protein (MCP). Rodents possess an additional C3 inhibitor termed Crry, a structural and functional analogue of the human C3 inhibitors (8). MAC formation is controlled by CD59 in both humans and rodents. Since complement inhibitors display species selective activity, the use of Crry and rodent CD59 in rodent models is necessary.

There is a link between proteinuria and progressive renal damage and there is data to support the hypothesis that proteinuria itself results in interstitial fibrosis and inflammation. The mechanism by which proteinuria leads to nephrotoxic injury is not known, but there is evidence that complement plays a key role and that the MAC is the principal mediator of tubulointerstitial injury (TI) due to proteinuria. The availability of an inhibitor that can specifically block MAC formation would allow an assessment of the role of MAC in tubulointerstitial injury under clinically relevant conditions.

A panel of well characterized mouse anti-rat kidney monoclonal antibodies was used (14, 32, incorporated herein by reference for their teaching regarding these antibodies and their sequences). The variable region DNA from a total of 5 antibodies was isolated by standard PCR techniques (31, incorporated herein by reference for its teachings regarding PCR). All were successfully cloned and some were expressed as single chain antibodies. All single chain antibodies recognized either a rat kidney epithelial or endothelial cell line in vitro. One of the mAbs, K9/9, binds in vivo to an unidentified glycoprotein expressed on rat glomerular epithelial cells and

PTEC (14). This antibody was chosen as a targeting vehicle for investigation of targeted Crry- and CD59-mediated complement inhibition in a rat model of acute tubulointerstitial injury. Although the K9/9 mAb was previously shown to induce glomerular damage, the antibody was only pathogenic when administered together with Freund's adjuvant. In fact the pathogenic nature of K9/9 mAb (with adjuvant) was not reproduced.

To generate recombinant targeted complement inhibitors, the extracellular regions of the rat membrane complement inhibitors Crry and CD59 were linked at their amino-termini to a single chain antibody (scFv) targeting moiety derived from K9/9 mAb (FIG. 5). In addition to targeted Crry and CD59 (tCrry, tCD59), soluble untargeted forms of Crry and CD59 (sCrry, sCD59) were also prepared, along with a scFv targeting moiety only (t-vehicle). All recombinant proteins were expressed by *Pichia* fermentation and all purified proteins migrated as a single band of expected molecular weight following SDS-PAGE.

The targeting and complement inhibitory function of recombinant proteins were analyzed in vitro using rat kidney PTEC as target cells. To assess the cell surface binding of the recombinant proteins, the binding of radiolabeled proteins to PTEC was measured. Iodinated tCrry, tCD59 and targeting vehicle all bound PTEC, and scFv specificity was demonstrated by the ability of unlabeled scFv to inhibit binding of labeled proteins (FIG. 1a). To determine functional activity, PTEC were sensitized to complement with a polyclonal antibody and the effect of the recombinant proteins on complement-mediated cell lysis (FIGS. 1b and c) and on C3 deposition (FIG. 1d) was measured. tCD59, tCrry and sCrry were equally effective at inhibiting rat complement-mediated lysis of PTEC. Untargeted sCD59 was not effective at inhibiting complement, and this result is consistent with previous data on sCD59 activity (15, 16). If complement inhibitors were removed following incubation with PTEC, but before the addition of rat serum, only tCrry and tCD59 were protective demonstrating that the complement inhibitors were functional when membrane bound. tCrry also inhibited complement activation and C3 deposition on PTEC, and as expected tCD59 had no effect on C3 deposition (FIG. 1d). The scFv targeting vehicle alone had no complement inhibitory activity. Since sCD59 did not effectively inhibit complement in vitro, further in vivo characterization of sCD59 was not pursued.

To determine whether tCrry and tCD59 target to the rat kidney, a biodistribution study was performed using iodinated proteins. Forty eight hours after tail vein injection of radiolabeled tCrry, tCD59 or t-vehicle, a significantly higher proportion of radioactivity (between 45 and 60% infected dose) was localized to the kidney compared to the other organs that were examined (FIGS. 2a and b). The radiolabeled proteins targeted the kidney in both healthy and PAN-treated proteinuric rats (described below) with somewhat higher levels of binding in proteinuric rats, perhaps reflecting increased access to targeting antigen on tubular cells (K9/9 mAb was shown previously to bind glomerular and tubular antigens in vivo (14)). Binding of sCrry in the kidney was not detected. Although a high concentration of targeted complement inhibitors remained in the kidneys at 48 hr relative to other organs, the circulatory half life ($t_{1/2}$) of the proteins was short; the $t_{1/2}$ in healthy rats for tCrry, tCD59, sCrry and t-vehicle was 38, 33, 34 and 18 minutes, respectively (FIG. 2c). The $t_{1/2}$ in proteinuric rats was not significantly different. Both tCrry and sCrry blocked complement activity in sera following injection, but in correlation with their short $t_{1/2}$, systemic complement inhibitory activity rapidly declined;

serum complement activity was at 85% of normal by 3-4 hr ($7\times t_{1/2}$). Negligible systemic activity of sCD59 was also confirmed (FIG. 2d). The short $t_{1/2}$ of targeted and untargeted inhibitors, together with biodistribution data and the fact that sCrry is not protective, indicate that the kidney-bound complement inhibitors would be effective at inhibiting complement locally and for a prolonged period.

The recombinant complement inhibitors were investigated therapeutically in a well characterized rat model of non-immunologic proteinuria-associated tubulointerstitial injury. Groups of 4 rats were injected with puromycin (PAN) to induce proteinuria on day 0, followed by intra-peritoneal injection of either PBS, complement inhibitor or targeting vehicle on days 4, 6, 8 and 10 (refer to table 3).

TABLE 3

Treatment groups and resulting proteinuria.			
Group	PAN	Therapy	Urinary protein mg/24 h
I	150 mg/kg	PBS	134 ± 16.4
II	150 mg/kg	t-Vehicle	150 ± 15.8
III	150 mg/kg	sCrry	131 ± 7.5
IV	150 mg/kg	tCrry	142 ± 18.0
V	150 mg/kg	tCD59	148 ± 24.0
VI	saline	PBS	0.56 ± 0.02

Urinary protein was determined during 24 h period following therapy and just prior to sacrifice.

Proteinuria appeared prior to the first dose of complement inhibitor (day 4) and the model is thus clinically relevant. Urine was collected over a 24 hr period following the final injection, and on day 11 blood was collected, the rats sacrificed, and the kidneys removed for analysis. Using this protocol, a dose-response study was performed using tCrry and sCrry (FIG. 6). tCrry, but not sCrry, was therapeutically effective at the higher doses tested, and both creatinine clearance and tissue injury score were statistically different ($p < 0.05$ comparing treated and control groups).

Based on this data, a larger therapeutic study ($n=4$) was undertaken using all of the recombinant proteins at a dose of 40 mg/kg (Table 3). The results showed that PAN induced heavy proteinuria, and proteinuria appeared before the first dose of complement inhibitor (day 4), verifying that the model is clinically relevant. Also, to address the most clinically relevant situation, individual inhibitors were administered rather than the two combined at a time when proteinuria was evident. There was no significant difference in urinary protein levels between the treatment groups after the final therapeutic injection (Table 3), and PAN treatment significantly impaired renal function as measured by creatinine clearance. In proteinuric rats, relative creatinine clearance was 40% of normal, and clearance was not significantly improved with sCrry treatment (FIG. 3C). In contrast, creatinine clearance in proteinuric rats receiving either tCrry or tCD59 was significantly improved and was 89 and 84% of normal, respectively ($p < 0.01$). Both tCrry and tCD59 individually provided a level of protection that was not statistically different from injury seen in the control group (rats not receiving puromycin). Targeting vehicle alone had no effect on creatinine clearance. Histological examination of kidney sections prepared from rats treated with PAN and receiving no therapy revealed dilation of tubular lumina and tubular and epithelial cell degeneration as assessed by loss of brush border (FIGS. 3A and B). Minimal improvement was seen with sCrry therapy. However, tubular dilation and degeneration was significantly and similarly suppressed in proteinuric rats

receiving either tCrry or tCD59 (FIGS. 3A and B). Creatinine clearances were not measured nor survival renal biopsies performed to assess tubular injury scores before administration of targeted inhibitors, but it is understood that these measurements are expected to be relatively normal at this time even though proteinuria was present. The data showed that the observed pathophysiological effects seen in control treated proteinuric animals were prevented.

To investigate the functional consequence of the complement inhibitors in vivo, C3 and MAC deposition were examined on proximal tubules from rats in the different treatment groups. Proximal tubules of proteinuric rats that received no therapy stained strongly positive for both C3 and MAC (C9) (FIG. 4, A and B). C3 and C9 staining was apparent within the tubular cells. Tubular cells are metabolically active, and the apparent cytosolic location of C3 and C9 is likely a result of endocytosis of membrane fixed complement. In this regard, analysis of C3 and C9 deposition was performed on tissue sections prepared from kidneys removed 11 days after PAN treatment, or ~7 days after onset of proteinuria and initial complement deposition at the tubular surface. In contrast to untreated proteinuric rats, rats receiving tCrry therapy had markedly reduced levels of both C3 and MAC deposition. tCD59 therapy, in contrast, reduced MAC but not C3 deposition. Untargeted sCrry had a minimal effect on C3 and MAC deposition. Thus, the measured parameters of tubulointerstitial injury after therapy correlated with MAC deposition, but not C3 deposition, on proximal tubules. Together, the data demonstrate that the MAC is the primary mediator of tissue injury in this model and that neither C3 opsonins, nor C3a, nor C5a make significant contributions to pathogenesis. Tubular binding of the targeted complement inhibitors was confirmed by immunofluorescence (FIG. 4C). Targeting vehicle alone gave a binding pattern similar to those of the targeted complement inhibitors. Neutrophil infiltration was also examined, but there was no difference between the different treatment groups (as determined on day 11 by immunohistochemistry and myeloperoxidase determinations).

Both tCrry and sCrry blocked complement activity in sera after i.p. injection at 40 mg/kg (route and dose used in therapeutic studies). However, in correlation with their short $t_{1/2}$ after tail vein injection (see above), systemic complement-inhibitory activity rapidly declined; serum complement activity was at 85% of normal by 3-4 h ($7\times t_{1/2}$). Negligible systemic activity of sCD59 was also confirmed (FIG. 2D). The short $t_{1/2}$ of targeted and untargeted inhibitors, together with biodistribution data and the fact that sCrry is not protective, indicate that the kidney-bound complement inhibitors are effective at inhibiting complement locally and for a prolonged period. Furthermore, 24 h after the final injection of inhibitor or PBS, there was no difference in serum complement activity between healthy control mice and proteinuric mice treated with either PBS or the different inhibitors. Thus, neither ongoing disease nor inhibitor treatment affected serum complement levels (determined by hemolytic assay).

Together the data demonstrate that the MAC is the primary mediator of tissue injury and that neither C3 opsonins, C3a nor C5a make significant contributions to pathogenesis. Soluble untargeted CD59 did not effectively inhibit complement, and to function effectively CD59 must be bound at the site of MAC formation. This functional constraint on CD59 has been shown in vitro (15, 16) and more recently in vivo by the demonstration that a membrane inserting form of CD59 when injected intra-articularly ameliorated disease in a model of rheumatoid arthritis (18). However, this study represents the first demonstration of effective and specific MAC inhibition in vivo following the systemic administration of inhibi-

tor, and has important implications for proteinuric disorders and other disease conditions in which the MAC has a major pathogenic role; a CD59-based inhibitor will not interfere with the generation of the complement activation products that are important in normal physiological processes (9, 12, 13).

Specific targeting of systemically administered CD59 provides the means to investigate the pathologic role of the MAC in various complement-dependent diseases under clinically relevant conditions. In this context, complement deficient animals have provided for a much better understanding of complement-associated disease mechanisms, but the data can be at odds with results obtained from studies in which complement has been temporarily inhibited. Complement deficiency may influence development of other immune functions, with the possible modulation of compensatory pathways. Also, there is likely to be some remaining functionality following the administration of a complement inhibitor and this may have a bearing on therapeutic outcome, since complement can have both protective and injurious effects during disease. Since most patients will have normal or only partially depleted complement levels, the availability of complement inhibitors that can be targeted to a site of disease and that can inhibit complement at different points in the pathway, provides a clinically relevant means to study the role of complement in experimental disease.

For complement-dependent disease processes involving early complement activation fragments, the targeting of C3 (and potentially C5) inhibitors offers advantages of improved efficacy and of safety due to low levels of systemic inhibition. A short circulatory half life of a targeted complement inhibitor (as was the case with the constructs used here) may be advantageous, and effective therapy was achieved in this model with very low levels of systemic inhibition. The C3 inhibitor sCR1 has shown therapeutic benefit in a variety of animal models of disease, and therapeutically effective doses maintain systemic inhibition for >24 hr (19). A sialyl Lewis x (sLex) glycosylated form of sCR1 has been shown to bind selectins in vitro, albeit with low affinity, and to localize to the vasculature in some models of inflammatory disease (20-22). However sLex sCR1 and sCR1 have similar pharmacokinetics and serum inhibitory activity (23) and therapeutically effective doses maintain systemic inhibition. Recently, in vivo targeting of human DAF to sites of C3 deposition and to erythrocytes have been demonstrated in mice using a soluble complement receptor 2 (CR2) (16) and an antibody fragment (24) respectively, but without functional characterization of activity in vivo.

With regard to the current study, the glomerulus is an effective barrier preventing the passage of protein into the nearly 170 liters of ultrafiltrate formed daily in humans. This restriction occurs in a size-selective manner. Since almost all proteins of the complement activation cascade are large (MW ~200 kDa), few of these proteins will appear in this ultrafiltrate and reach tubular lumina under normal circumstances. As such, the apical aspects of tubules are not endowed with significant quantities of proteins that regulate the cleavage of C3/C5 or the formation of C5b-9 (6, 7, 25). Consistent with this, complement can be spontaneously activated on tubular cells in vitro (6, 7). In any disease in which the glomerular barrier to protein passage is disrupted, complement proteins can appear in the tubular fluid and lead to complement activation. Such complement activation can occur in any condition accompanied by proteinuria, including immunological diseases as well as those not considered to have an immunological basis, such as the common disease diabetic nephropathy.

thy. The current data establish the benefits of targeted complement inhibition with regard to both efficacy and safety.

b) Materials and Methods

(1) Animals, Cells, and Reagents

Normal rat serum was obtained from Cocalico Biologicals, and C6-deficient rat serum was a gift from Dr. W. M. Baldwin (Johns Hopkins University School of Medicine, Baltimore, Md.). Monoclonal mouse anti-rat Crry, 5I2 (36), and monoclonal mouse anti-rat CD59, 6D1 (37), are described. Antiserum against rat C9 that recognizes MAC in rat tissue was a gift from Dr. Paul Morgan (University of Wales, Cardiff, U.K.). Anti-rat C3 serum was from MP Biomedicals. Rabbit antiserum to rat Crry, human CD59 (cross-reactive with rat CD59), and K9/9 single-chain Fv (scFv) were prepared by Cocalico Biologicals. All secondary Abs were purchased from Sigma-Aldrich. Female Sprague-Dawley rats weighing 45-50 g were purchased from Harlan and were housed with free access to food and water. Experimental protocols were conducted according to current guidelines presented in the National Institute of Health Guide for the care and use of laboratory animals. The rat proximal tubule epithelial cell (PTEC) line was obtained from American Type Culture Collection and cultured in DMEM with 10% FCS.

(2) Recombinant DNA Techniques

For the cloning of Ab variable region cDNA, total RNA was extracted from the hybridoma cell line expressing K9/9 mAb (14) using an RNA Extract Kit (Qiagen). Variable region cDNAs (VL and VH chains) were amplified by RT-PCR. The 5'- and 3'-primers used for VL chain were ATGAAGTTGCTGTTAGGCTGTTGGTGCTG (SEQ ID NO: 16) and ACTGGATGGTGGGAAGATGG (SEQ ID NO: 17). The 5'- and 3'-primers used for VH chain were ATGAAATGCAGCTGGGGCATGTTCTTC (SEQ ID NO: 18) and CAGTGGATAGACCGATGGGCC (SEQ ID NO: 19). PCR fragments were subcloned into TA cloning vector (Invitrogen) and VL and VH chains joined by a (G4S)₃ linker. For construction of targeted complement inhibitors, the K9/9 targeting scFv was linked to cDNA encoding an extracellular domain of rat Crry (five N-terminal short consensus repeats, residues 1-981) (36) or mature rat CD59 (residues 1-77) (38). scFv and complement inhibitors were joined by a G4S linker in the sequence VL-VH inhibitor by standard PCR techniques. cDNA was cloned into pPICZα *Pichia pastoris* expression plasmid (Invitrogen). All sequences were verified.

(3) Expression and Purification of Recombinant proteins

P. pastoris strain X-33 was used for expression of recombinant proteins. After transformation, positive clones were selected by PCR, and protein expression was verified by Western blot. Yeast was grown in a 7-liter fermentor (New Brunswick Scientific), and recombinant proteins were purified from culture supernatants by standard bulk anion exchange separation followed by Q Sepharose and Mono Q FPLC (Amersham Biosciences). Protein concentration was determined by bicinchoninic acid assay (Pierce Chemical), and purified proteins were analyzed by SDS-PAGE and Western blotting.

(4) In Vitro Analysis of Binding

Recombinant proteins were iodinated using the iodogen method according to manufacturer's instructions (Pierce

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Chemical) to approximate specific activities of 1×10^7 cpm/ μ g and incubated with PTEC in the absence or presence of 100-fold excess of unlabeled proteins. The recombinant proteins (0.1 mg/ml) were added to a monolayer of PTEC (80% confluent in 24-well plates) for 30 min at 4° C. Cells were then washed with 1% BSA in culture medium and treated with 0.5 M NaOH to remove bound protein; then radioactivity in the supernatant was determined.

(5) In Vitro Complement Lysis and C3 Deposition Assays

Rat PTEC at 60-80% confluency were detached with 0.05 MEDTA in PBS, washed twice, and resuspended to 1×10^6 /ml in DMEM. Cells were then sensitized to complement by incubation (4° C. for 30 min) in 10% rabbit antiserum raised against rat 13762 mammary adenocarcinoma cells (39). After incubation, an equal volume of rat serum diluted in DMEM was added, and cell viability was determined after 45 min at 37° C. using the CytoTox 96 Non-Radioactive Cytotoxicity Assay Kit (Promega). Cell lysis assays were typically performed in 96-well ELISA plates in a final volume of 100 μ l. To determine the effect of complement inhibitors on complement-mediated lysis, the recombinant proteins in PBS were added after Ab sensitization, and incubations were continued for a further 30 min at 4° C. before addition of rat serum. In some experiments, cells were washed twice with DMEM before addition of rat serum. Percentage of inhibition of lysis was calculated as $(A-B)/A$, where A=percentage of cell lysis in 10% normal rat serum and B=percentage of cell lysis in 10% normal rat serum with recombinant protein. The effect of the complement-inhibitory proteins on C3 deposition was determined by flow cytometry. For flow cytometry, the procedure described above for cell lysis determinations was followed except that 10% C6-deficient rat serum was used in place of normal rat serum. After incubation in serum, the cells were washed, and C3 deposition was detected by means of anti-rat C3 Ab-FITC by standard procedures (40).

(6) In Vivo Blood Clearance and Biodistribution of Radiolabeled Recombinant Proteins

For determination of rate of clearance from the circulation, iodinated recombinant proteins (1×10^8 cpm) were injected into the tail vein; blood was collected at 30 min, 1 h, 2 h, 4 h, and 8 h after injection; and radioactivity in blood was counted. Circulatory half-life ($t_{1/2}$) was calculated by standard methods (41). Biodistribution studies were performed using standard procedures for determining tissue distribution of injected radiolabeled proteins (41, 33). Briefly, 1×10^8 cpm of 125 I-labeled soluble Crry (sCrry; 1.2×10^7 cpm/ μ g), targeted Crry (tCrry) (1.3×10^7 cpm/ μ g), targeted CD59 (tCD59; 1.2×10^7 cpm/ μ g), or targeting vehicle only (t-vehicle) (1.0×10^7 cpm/ μ g) were injected into the tail vein of 40- to 55-g female rats. After 48 h, a blood sample was taken, and major organs were removed, shredded, washed in PBS containing 10 mM EDTA, weighed, and counted. Targeting specificity was determined as percentage of cpm per gram of tissue divided by total injected cpm per gram of body weight. Percentage = $A/B \times 100$, where A=tissue cpm per gram of tissue weight and B=total cpm per gram of body weight. All animal studies were approved by the Institutional Animal Care and Use Committee institutional review board at Medical University of South Carolina.

(7) Serum Complement—Inhibitory Activity

Ab-sensitized sheep erythrocytes (1×10^7 ; Advanced Research Technologies) were incubated in serum dilutions in

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a final volume of 300 μ l at 37° C. for 1 h. Gelatin-veronal buffer-EDTA (Sigma-Aldrich) was then added, and hemolysis was determined by measuring OD414 of supernatants (42). Total complement hemolytic activity (CH50) was defined as the reciprocal of the dilution of serum that lysed 50% of the erythrocytes; this measurement was used to compare serum complement activity. After i.p. injection of recombinant complement inhibitors (40 mg/kg), serum was collected at different times and percentage of complement inhibitory activity in sera was defined as the difference between CH50 of normal serum and CH50 of sample divided by CH50 of normal serum.

(8) Rat Model of Puromycin Aminonucleoside-Induced Nephrosis and Experimental Protocol

Proteinuria-induced tubulointerstitial injury was induced in female Sprague-Dawley rats (Harlan) weighing 45-50 g by single tail vein injection of 150 mg/kg puromycin aminonucleoside (PAN). Proteinuria develops by day 4. In a dose-response pilot study, 20 rats were divided into 10 groups of 2 animals. Nine groups received PAN, and a control group received saline. On days 4, 6, 8, and 10, control rats received an i.p. injection of PBS, and each group of PAN-treated rats received i.p. injections of varying doses (5-40 mg/kg) of sCrry or tCrry. After the final injection, rats were placed in metabolic cages, and urine was collected for 24 h. On day 11, blood was collected, rats were sacrificed, and kidneys were removed. In a larger therapeutic study, 24 rats were divided into 6 groups of 4 animals. Five groups received PAN, and a control group received saline. Control rats received i.p. injections of PBS, and PAN-treated rats received i.p. injections of PBS, sCrry, tCrry, tCD59, or targeting vehicle only (Table 3) at 40 mg/kg using the same protocol described above for pilot study.

(9) Renal Function and Histology

Urinary protein was measured using a bicinchoninic acid protein assay kit (Pierce). Creatinine clearance, a measure of renal function, was calculated after the measurement of rat serum and urine creatinine with a creatinine reagent kit (Roche Pharmaceuticals) according to manufacturer's instructions. For histological assessment of tubulointerstitial injury, paraffin-embedded kidney sections were stained with periodic acid-Schiff reagent, and tubulointerstitial injury was assessed in a blinded manner. Tubulointerstitial injury is defined as tubular dilation, tubular atrophy, tubular cast formation, sloughing of tubular epithelial cells, or thickening of the tubular basement membrane and was scored on a scale of 0-4, as follows: 0, no injury; 1, <25% of the tubulointerstitium injured; 2, 25-50% of the tubulointerstitium injured; 3, 51-75% of the tubulointerstitium injured; 4, >75% of the tubulointerstitium injured.

(10) Immunofluorescence Microscopy

To investigate C3 and MAC deposition, frozen kidney sections were stained with anti-rat C3 or anti-rat C9 antiserum together with appropriate FITC-labeled secondary Abs as described (16). The binding of targeted complement inhibitors in the kidney was similarly analyzed using rabbit antiserum raised against K9/9 scFv. Digital images were acquired and optimized with Adobe Photoshop using identical settings. Neutrophil infiltration was also assessed by immunofluorescence microscopy using polymorphonuclear leuko-

cyte-specific mouse anti-rat granulocyte mAb (BD Biosciences) and by measuring myeloperoxidase activity in kidney tissue samples as described (43).

(11) Statistics

Data are presented as mean \pm SD. Data were analyzed by global ANOVA followed by the post hoc test (Scheffé's procedure). For dose-response experiments, group comparisons were done with the Kruskal-Wallis test.

2. Example 2

Model Complement Activation in Proteinuric Disease

The following events of complement activation occurring from the fluid phase are relevant to what occurs in vivo on the PTEC. The initial alternative pathway C3 convertase requires factor D to cleave factor B present in a trimolecular complex containing hydrolyzed C3 (C3(H₂O)), factor B, and Mg⁺⁺. The resulting C3(H₂O)Bb can recruit and cleave native C3 to generate alternative pathway C3 and C5 convertases, C3bBb and C3bBbC3b, respectively. This whole process can occur spontaneously because factor D is an active serine esterase (unlike most others that exist as zymogens) while the formation of C3(H₂O) is continuous, and can be accelerated by conditions present in tubular fluid, such as exposure to ammonia and acidic pH (28, 51, 57). The generation of C5b-9 can proceed simply when its composite proteins are present in the vicinity. Besides the constraints of component availability, and time and space (that is, the proteins must find each other and a receptive surface prior to spontaneous inactivation, which varies depending on the intermediate), there are competing influences of complement regulatory proteins, including DAF, MCP, CR1, and clusterin, protein S and CD59 for C5b-9. The balance between pro- and anti-activating influences in normal tubular fluid is unfavorable for alternative pathway activation. However, this shifts towards activation in proteinuric conditions, leading to C5b-9 generation, which over time results in tubulointerstitial injury pathology. The explanation for this is that C3 and C5, which are key for this activation to occur are large (185-190 kDa) which leads to their being restricted from normal but not abnormal glomerular ultrafiltrate; based upon previously discussed models, $1_{C3/C5}$ can be estimated as 0.0005 in controls and 0.0125 in nephrotics (45), reflecting in a 50-fold increase in their appearance into tubular fluid (which would be 20 and 1 mg/L for C3 and C5, respectively). Other aspects include local PTEC production of complement components and CRPs under disease conditions (49, 8-60).

These studies can be performed with cPTEC, in which PTEC is exposed to various concentrations of normal human serum (a stock of pooled serum from several normal laboratory donors). To confirm that the observed effects are due to alternative pathway-mediated C5b-9 formation, controls can include heat-inactivated serum, serum in Mg⁺⁺-EGTA to allow only alternative pathway activation, and serum lacking specific complement components. Complement activation is reflected by cell injury as indicated by release from intracellular pools of previously cell-loaded with 2',7'-bis-(carboxyethyl)-5(6)-carboxyfluorescein (BCECF) and the intrinsic cellular enzyme, LDH. Release of the former but not the latter is typical of so-called sublytic cell damage and seems most relevant to complement activation in disease states (54, 61, 62). In addition, the quantities of C3b and C5b-9 on cell

membranes can be evaluated by western blotting using mAbs specific for the iC3b chain of cleaved C3 and for neoantigens on C5b-9 (50, 63).

Also of interest is the phenotypic response of the PTEC model system to sublytic complement activation. mRNA for inflammatory mediators known to be produced by PTEC can be examined, including upon complement activation and/or exposure to excess concentrations of proteins (such as albumin), which are likely to be relevant to human proteinuric diseases (48, 64, 65). These can include CRPs (to evaluate whether PTECs respond "defensively" to complement activation as other cells do (66) and as appears to occur in vivo (49)), anaphylatoxin receptors (55, 56), complement proteins (C3, factor B), cytokines (IL-1 and IL-6) and chemokines (CCL2, CCL5 and CX₃CL1). In addition, potential activation of phospholipases, protein kinase C, and NF- κ B can be assessed as relevant to complement activation and/or PTEC stress (48, 67).

3. Example 3

mAbs to PTEC Antigens

It is disclosed herein that PTEC antigens facilitate targeted delivery of CD59 providing protection against the spontaneous formation of C5b-9. In studies, CD59 has been functionally active to prevent C5b-9 generation on cells when targeted to dansyl (15), C3 (16) and DPPIV (44) on the surface of cells. The latter is an intrinsic membrane protein of the PTEC, and it was fortuitous the mAb clone selected (K9/9) was effective. DPPIV and iC3b/C3d can be used as targets for mAbs and CR2, respectively. Other potential target antigens include at least 18 members of the solute carrier superfamily present in the PTEC brush border (from Slc1a1 to Slc34a1; see list above), other transporters such as water and chloride channels, and members of the ATP binding cassette family (Abcb1b, Abcc2 and 4). The previously discussed proteins megalin, NEP, and KIM-1, as well as Mucin 20 (MUC20) (68, 69) are also potential target antigens. As noted above, the advantages of these molecules are that they have high expression in the apical membrane of the PTEC, their protein structure is known, and in the case of the latter two they are highly expressed in states in which the PTEC is injured (52, 53, 68). In addition, mAbs are available for KIM-1 which can be exploited.

Megalyn and MUC20 each are type I transmembrane proteins, with relatively extended extracellular domains (68-70). mAbs can be raised against recombinant proteins containing defined extracellular regions of megalin and MUC20 fused to C terminal human 1 Fc domains and produced in CHO cells. A MUC20 recombinant is created containing 4-6 mucin repeats using a PCR strategy with the 5' primer upstream from the repeating region and the 3' primer ending just outside the transmembrane domain at L202 (68). For megalin, a similar approach is taken to produce a recombinant containing amino acids 4330-4420, which encompasses EGF-like domains 16 and 17 of this protein, 7 predicted disulfide bonds within this fragment (and none extending outside of it), excludes a potential glycosylation site at N4327, and terminates just before the transmembrane domain.

DPPIV and NEP are type II transmembrane proteins also with large extracellular domains but more complicated tertiary structures than megalin, MUC20 or KIM-1. Although soluble recombinants containing the entire extracellular domains of each that retained activity and structure have been produced using standard methodology (71, 72), α -helical regions modeled to be close to the membrane are of interest.

In the case of DPPIV, these are all contained in the α/β hydrolase domain (extending from Q508 to the C-terminal P766) in the final five β propellers (71, 73), while in NEP, they are contained in the larger catalytic domain (72). The peptides are listed below, and are utilized to raise mAbs using standard multiple antigenic peptide (MAP) technology (74).

NEP			DPPIV		
AAs	Peptide		AAs	Peptide	
59-71	SDCIKSAARLIQN	(SEQ ID NO: 20)	601-614	FEVEDQIEAARQFS	(SEQ ID NO: 21)
82-93	FFKYACGGWLKR	(SEQ ID NO: 22)	631-640	YGGYVTSML	(SEQ ID NO: 23)
435-456	SKHVVEDLIAQIREVFIQTLDD	(SEQ ID NO: 24)	713-726	FQQAQISKALVDV	(SEQ ID NO: 25)
462-474	AETKKRAEEKALA	(SEQ ID NO: 26)	745-763	STAHQHIYTHMSHFQKQCF	(SEQ ID NO: 27)

mAbs are raised in mice using standard techniques. Four mice each for megalin, MUC20, DPPIV and NEP are immunized and those making high titer antibodies boosted and used for hybridoma production. Screening for mAbs at each step is done by ELISA using the immunogens. In addition, the ability to bind native protein on PTECs in culture (using a high throughput in situ fluorescence assay) and in normal kidney tissue (by standard indirect immunofluorescence microscopy) can be assessed. These latter abilities can be used as guides to selection of hybridoma clones.

4. Example 4

mAb/CR2-CD59 Chimeras to Target Human PTECs

Herein disclosed are several scFv-CD59 proteins each for megalin, MUC20, DPPIV, NEP, and KIM-1. These will bind to their respective proteins on the PTEC surface. In addition, CR2-CD59 can be constructed to bind C3 fragments on PTEC. By virtue of their localization to epitopes close to the plasma membrane, the CD59 in scFv-CD59 (or CR2-CD59) limit C5b-9 generation occurring on PTECs from fluid phase complement activation.

cDNA encoding the variable light and heavy chain domains from hybridomas producing mAbs against the selected PTEC antigens (DPPIV, NEP, Megalin, KIM-1 and MUC20) is isolated and the sequences used to construct plasmids encoding scFvs by standard techniques (75, 76). The scFvs are prepared as His-tagged constructs for ease of purification and in vitro characterization of scFv binding to cultured PTECs. scFv binding to cultured PTECs is quantified using FITC-conjugated anti-His-tag antibodies. Standard controls are used to confirm the specificity of binding in these in situ assays. As with scFv's alone, the scFv-CD59 proteins are tested for their ability to bind their respective PTEC antigens in culture and in kidney tissue. Western blotting is performed to evaluate specificity of protein binding.

5. Example 5

Disclosed herein are examples of constructs of the present invention made in accordance with the teaching herein. The terminology used has the following meaning: SCR=short consensus repeats; LP=Leader Peptide. The constructs all have the basic formula of K9/9-linker-complement modulator. Notations in parenthesis indicate details within a particular section of the composition. It is understood that a linker

can be a chemical linker, a natural linker peptide, or amino acid linking sequences (e.g., (Gly₄Ser)₂). It is understood that this list is not limiting and only provides examples of some of the constructs disclosed in the present application.

K9/9-(Gly₄Ser)₂--DAF
K9/9-(Gly₄Ser)₂-human CD59

20 K9/9-(Gly₄Ser)₂--MCP
K9/9-(Gly₄Ser)₂--CR1
K9/9-(Gly₄Ser)₂--Crry
K9/9-(Gly₄Ser)₂--mouse CD59
25 K9/9-(Gly₄Ser)₂-human IgG1 Fc
K9/9-(Gly₄Ser)₂-human IgM Fc
K9/9-(Gly₄Ser)₂-murine IgG3 Fc
K9/9-(Gly₄Ser)₂-murine IgM Fc
K9/9-(Gly₄Ser)₂--CVF
30 K9/9-(Gly₄Ser)₃--DAF
K9/9-(Gly₄Ser)₃-human CD59
K9/9-(Gly₄Ser)₃--MCP
K9/9-(Gly₄Ser)₃--CR1
K9/9-(Gly₄Ser)₃--Crry
35 K9/9-(Gly₄Ser)₃-mouse CD59
K9/9-(Gly₄Ser)₃-human IgG1 Fc
K9/9-(Gly₄Ser)₃-human IgM Fc
K9/9-(Gly₄Ser)₃-murine IgG3 Fc
K9/9-(Gly₄Ser)₃-murine IgM Fc
40 K9/9-(Gly₄Ser)₃--CVF
K9/9-(Gly₃Ser)₄--DAF
K9/9-(Gly₃Ser)₄-human CD59
K9/9-(Gly₃Ser)₄--MCP
K9/9-(Gly₃Ser)₄--CR1
K9/9-(Gly₃Ser)₄--Crry
45 K9/9-(Gly₃Ser)₄-mouse CD59
K9/9-(Gly₃Ser)₄-human IgG1 Fc
K9/9-(Gly₃Ser)₄-human IgM Fc
K9/9-(Gly₃Ser)₄-murine IgG3 Fc
K9/9-(Gly₃Ser)₄-murine IgM Fc
50 K9/9-(Gly₃Ser)₄--CVF
K9/9-(Gly₄Ser)₃--DAF (SCRs 2-4)
K9/9-(Gly₃Ser)₄--DAF (SCRs 2-4)
K9/9-(Gly₄Ser)₃--CR1 (LP--SCR1-4-SCR8-11-SCR15-
55 18)
K9/9-(Gly₄Ser)₃--Crry (5 N-terminal SCRS)
DPPIV-(Gly₄Ser)₃--DAF
DPPIV-(Gly₄Ser)₃-human CD59
DPPIV-(Gly₄Ser)₃--MCP
60 DPPIV-(Gly₄Ser)₃--CR1
DPPIV-(Gly₄Ser)₃--Crry
DPPIV-(Gly₄Ser)₃-mouse CD59
DPPIV-(Gly₄Ser)₃-human IgG1 Fc
DPPIV-(Gly₄Ser)₃-human IgM Fc
DPPIV-(Gly₄Ser)₃-murine IgG3 Fc
65 DPPIV-(Gly₄Ser)₃-murine IgM Fc
DPPIV-(Gly₄Ser)₃--CVF

Kim-1-(Gly₄Ser)₃--DAF
 Kim-1-(Gly₄Ser)₃-human CD59
 Kim-1-(Gly₄Ser)₃--MCP
 Kim-1-(Gly₄Ser)₃--CR1
 Kim-1-(Gly₄Ser)₃--Crry
 Kim-1-(Gly₄Ser)₃-mouse CD59
 Kim-1-(Gly₄Ser)₃-human IgG1 Fc
 Kim-1-(Gly₄Ser)₃-human IgM Fc
 Kim-1-(Gly₄Ser)₃-murine IgG3 Fc
 Kim-1-(Gly₄Ser)₃-murine IgM Fc
 Kim-1-(Gly₄Ser)₃--CVF
 NED-(Gly₄Ser)₃--DAF
 NED-(Gly₄Ser)₃-human CD59
 NED-(Gly₄Ser)₃--MCP
 NED-(Gly₄Ser)₃--CR1
 NED-(Gly₄Ser)₃--Crry
 NED-(Gly₄Ser)₃-mouse CD59
 NED-(Gly₄Ser)₃-human IgG1 Fc
 NED-(Gly₄Ser)₃-human IgM Fc
 NED-(Gly₄Ser)₃-murine IgG3 Fc
 NED-(Gly₄Ser)₃-murine IgM Fc
 NED-(Gly₄Ser)₃--CVF
 K9/9-VSVFPLE--DAF
 K9/9-VSVFPLE-human CD59
 K9/9-VSVFPLE--MCP
 K9/9-VSVFPLE--CR1
 K9/9-VSVFPLE--Crry
 K9/9-VSVFPLE-mouse CD59
 K9/9-VSVFPLE-human IgG1 Fc
 K9/9-VSVFPLE-human IgM Fc
 K9/9-VSVFPLE-murine IgG3 Fc
 K9/9-VSVFPLE-murine IgM Fc
 K9/9-VSVFPLE--CVF
 DPPIV-VSVFPLE--DAF
 DPPIV-VSVFPLE-human CD59
 DPPIV-VSVFPLE--MCP
 DPPIV-VSVFPLE--CR1
 DPPIV-VSVFPLE--Crry
 DPPIV-VSVFPLE-mouse CD59
 DPPIV-VSVFPLE-human IgG1 Fc
 DPPIV-VSVFPLE-human IgM Fc
 DPPIV-VSVFPLE-murine IgG3 Fc
 DPPIV-VSVFPLE-murine IgM Fc
 DPPIV-VSVFPLE--CVF
 K9/9---m-Maleimidobenzoyl-N-hydroxysuccinimide ester--DAF
 K9/9---m-Maleimidobenzoyl-N-hydroxysuccinimide ester-human CD59
 K9/9-m-Maleimidobenzoyl-N-hydroxysuccinimide ester--MCP
 K9/9-m-Maleimidobenzoyl-N-hydroxysuccinimide ester--CR1
 K9/9-m-Maleimidobenzoyl-N-hydroxysuccinimide ester--Crry
 K9/9-m-Maleimidobenzoyl-N-hydroxysuccinimide ester--mouse CD59
 K9/9-m-Maleimidobenzoyl-N-hydroxysuccinimide ester--human IgG1 Fc
 K9/9-m-Maleimidobenzoyl-N-hydroxysuccinimide ester--human IgM Fc
 K9/9-m-Maleimidobenzoyl-N-hydroxysuccinimide ester--murine IgG3 Fc
 K9/9-m-Maleimidobenzoyl-N-hydroxysuccinimide ester--murine IgM Fc
 K9/9-m-Maleimidobenzoyl-N-hydroxysuccinimide ester--CVF

DPPIV---m-Maleimidobenzoyl-N-hydroxysuccinimide ester--DAF
 DPPIV---m-Maleimidobenzoyl-N-hydroxysuccinimide ester-human CD59
 5 DPPIV---m-Maleimidobenzoyl-N-hydroxysuccinimide ester--MCP
 DPPIV-m-Maleimidobenzoyl-N-hydroxysuccinimide ester--CR1
 10 DPPIV-m-Maleimidobenzoyl-N-hydroxysuccinimide ester--Crry
 DPPIV-m-Maleimidobenzoyl-N-hydroxysuccinimide ester--mouse CD59
 DPPIV-m-Maleimidobenzoyl-N-hydroxysuccinimide ester--human IgG1 Fc
 15 DPPIV-m-Maleimidobenzoyl-N-hydroxysuccinimide ester--human IgM Fc
 DPPIV-m-Maleimidobenzoyl-N-hydroxysuccinimide ester--murine IgG3 Fc
 20 DPPIV-m-Maleimidobenzoyl-N-hydroxysuccinimide ester--murine IgM Fc
 DPPIV-m-Maleimidobenzoyl-N-hydroxysuccinimide ester--CVF
 K9/9-bismaleimidohexane-DAF
 25 K9/9-bismaleimidohexane-human CD59
 K9/9-bismaleimidohexane--MCP
 K9/9-bismaleimidohexane--CR1
 K9/9-bismaleimidohexane--Crry
 K9/9-bismaleimidohexane-mouse CD59
 30 K9/9-bismaleimidohexane-human IgG1 Fc
 K9/9-bismaleimidohexane-human IgM Fc
 K9/9-bismaleimidohexane-murine IgG3 Fc
 K9/9-bismaleimidohexane-murine IgM Fc
 K9/9-bismaleimidohexane--CVF
 35 DPPIV-bismaleimidohexane--DAF
 DPPIV-bismaleimidohexane-human CD59
 DPPIV-bismaleimidohexane--MCP
 DPPIV-bismaleimidohexane--CR1
 DPPIV-bismaleimidohexane--Crry
 40 DPPIV-bismaleimidohexane-mouse CD59
 DPPIV-bismaleimidohexane-human IgG1 Fc
 DPPIV-bismaleimidohexane-human IgM Fc
 DPPIV-bismaleimidohexane-murine IgG3 Fc
 DPPIV-bismaleimidohexane-murine IgM Fc
 45 DPPIV-bismaleimidohexane--CVF
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- G. Sequences
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6. K9/9
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8. K9/9-CD59 (tCD59)
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- Amino acid Sequence corresponds to SEQ ID NO: 13
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<213> ORGANISM: Artificial Sequence

<220> FEATURE:

<223> OTHER INFORMATION: Description of Artificial Sequence;

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note=synthetic construct

<400> SEQUENCE: 6

Met Cys Leu Gly Arg Met Gly Ala Ser Ser Pro Arg Ser Pro Glu Pro
 1 5 10 15
 Val Gly Pro Pro Ala Pro Gly Leu Pro Phe Cys Cys Gly Gly Ser Leu
 20 25 30
 Leu Ala Val Val Val Leu Leu Ala Leu Pro Val Ala Trp Gly Gln Cys
 35 40 45
 Asn Ala Gln Cys Asn Ala Pro Glu Trp Leu Pro Phe Ala Arg Pro Thr
 50 55 60
 Asn Leu Thr Asp Glu Phe Glu Phe Pro Ile Gly Thr Tyr Leu Asn Tyr
 65 70 75 80
 Glu Cys Arg Pro Gly Tyr Ser Gly Arg Pro Phe Ser Ile Ile Cys Leu
 85 90 95
 Lys Asn Ser Val Trp Thr Gly Ala Lys Asp Arg Cys Arg Arg Lys Ser
 100 105 110
 Cys Arg Asn Pro Pro Asp Pro Val Asn Gly Met Val His Val Ile Lys
 115 120 125
 Gly Ile Gln Phe Gly Ser Gln Ile Lys Tyr Ser Cys Thr Lys Gly Tyr
 130 135 140
 Arg Leu Ile Gly Ser Ser Ser Ala Thr Cys Ile Ile Ser Gly Asp Thr
 145 150 155 160
 Val Ile Trp Asp Asn Glu Thr Pro Ile Cys Asp Arg Ile Pro Cys Gly
 165 170 175
 Leu Pro Pro Thr Ile Thr Asn Gly Asp Phe Ile Ser Thr Asn Arg Glu
 180 185 190
 Asn Phe His Tyr Gly Ser Val Val Thr Tyr Arg Cys Asn Pro Gly Ser
 195 200 205
 Gly Gly Arg Lys Val Phe Glu Leu Val Gly Glu Pro Ser Ile Tyr Cys
 210 215 220
 Thr Ser Asn Asp Asp Gln Val Gly Ile Trp Ser Gly Pro Ala Pro Gln
 225 230 235 240
 Cys Ile Ile Pro Asn Lys Cys Thr Pro Pro Asn Val Glu Asn Gly Ile
 245 250 255
 Leu Val Ser Asp Asn Arg Ser Leu Phe Ser Leu Asn Glu Val Val Glu
 260 265 270
 Phe Arg Cys Gln Pro Gly Phe Val Met Lys Gly Pro Arg Arg Val Lys
 275 280 285
 Cys Gln Ala Leu Asn Lys Trp Glu Pro Glu Leu Pro Ser Cys Ser Arg
 290 295 300
 Val Cys Gln Pro Pro Pro Asp Val Leu His Ala Glu Arg Thr Gln Arg
 305 310 315 320
 Asp Lys Asp Asn Phe Ser Pro Gly Gln Glu Val Phe Tyr Ser Cys Glu
 325 330 335
 Pro Gly Tyr Asp Leu Arg Gly Ala Ala Ser Met Arg Cys Thr Pro Gln
 340 345 350
 Gly Asp Trp Ser Pro Ala Ala Pro Thr Cys Glu Val Lys Ser Cys Asp
 355 360 365
 Asp Phe Met Gly Gln Leu Leu Asn Gly Arg Val Leu Phe Pro Val Asn
 370 375 380
 Leu Gln Leu Gly Ala Lys Val Asp Phe Val Cys Asp Glu Gly Phe Gln
 385 390 395 400
 Leu Lys Gly Ser Ser Ala Ser Tyr Cys Val Leu Ala Gly Met Glu Ser

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405					410					415					
Leu	Trp	Asn	Ser	Ser	Val	Pro	Val	Cys	Glu	Gln	Ile	Phe	Cys	Pro	Ser
			420					425					430		
Pro	Pro	Val	Ile	Pro	Asn	Gly	Arg	His	Thr	Gly	Lys	Pro	Leu	Glu	Val
		435					440					445			
Phe	Pro	Phe	Gly	Lys	Ala	Val	Asn	Tyr	Thr	Cys	Asp	Pro	His	Pro	Asp
	450					455					460				
Arg	Gly	Thr	Ser	Phe	Asp	Leu	Ile	Gly	Glu	Ser	Thr	Ile	Arg	Cys	Thr
465					470						475				480
Ser	Asp	Pro	Gln	Gly	Asn	Gly	Val	Trp	Ser	Ser	Pro	Ala	Pro	Arg	Cys
			485						490					495	
Gly	Ile	Leu	Gly	His	Cys	Gln	Ala	Pro	Asp	His	Phe	Leu	Phe	Ala	Lys
			500					505					510		
Leu	Lys	Thr	Gln	Thr	Asn	Ala	Ser	Asp	Phe	Pro	Ile	Gly	Thr	Ser	Leu
		515					520					525			
Lys	Tyr	Glu	Cys	Arg	Pro	Glu	Tyr	Tyr	Gly	Arg	Pro	Phe	Ser	Ile	Thr
	530					535					540				
Cys	Leu	Asp	Asn	Leu	Val	Trp	Ser	Ser	Pro	Lys	Asp	Val	Cys	Lys	Arg
545					550					555					560
Lys	Ser	Cys	Lys	Thr	Pro	Pro	Asp	Pro	Val	Asn	Gly	Met	Val	His	Val
			565						570					575	
Ile	Thr	Asp	Ile	Gln	Val	Gly	Ser	Arg	Ile	Asn	Tyr	Ser	Cys	Thr	Thr
		580						585					590		
Gly	His	Arg	Leu	Ile	Gly	His	Ser	Ser	Ala	Glu	Cys	Ile	Leu	Ser	Gly
		595					600					605			
Asn	Ala	Ala	His	Trp	Ser	Thr	Lys	Pro	Pro	Ile	Cys	Gln	Arg	Ile	Pro
	610						615					620			
Cys	Gly	Leu	Pro	Pro	Thr	Ile	Ala	Asn	Gly	Asp	Phe	Ile	Ser	Thr	Asn
625					630					635					640
Arg	Glu	Asn	Phe	His	Tyr	Gly	Ser	Val	Val	Thr	Tyr	Arg	Cys	Asn	Pro
				645					650					655	
Gly	Ser	Gly	Gly	Arg	Lys	Val	Phe	Glu	Leu	Val	Gly	Glu	Pro	Ser	Ile
			660					665					670		
Tyr	Cys	Thr	Ser	Asn	Asp	Asp	Gln	Val	Gly	Ile	Trp	Ser	Gly	Pro	Ala
		675					680						685		
Pro	Gln	Cys	Ile	Ile	Pro	Asn	Lys	Cys	Thr	Pro	Pro	Asn	Val	Glu	Asn
	690					695					700				
Gly	Ile	Leu	Val	Ser	Asp	Asn	Arg	Ser	Leu	Phe	Ser	Leu	Asn	Glu	Val
705					710					715					720
Val	Glu	Phe	Arg	Cys	Gln	Pro	Gly	Phe	Val	Met	Lys	Gly	Pro	Arg	Arg
				725					730					735	
Val	Lys	Cys	Gln	Ala	Leu	Asn	Lys	Trp	Glu	Pro	Glu	Leu	Pro	Ser	Cys
			740					745					750		
Ser	Arg	Val	Cys	Gln	Pro	Pro	Pro	Asp	Val	Leu	His	Ala	Glu	Arg	Thr
		755					760					765			
Gln	Arg	Asp	Lys	Asp	Asn	Phe	Ser	Pro	Gly	Gln	Glu	Val	Phe	Tyr	Ser
	770					775					780				
Cys	Glu	Pro	Gly	Tyr	Asp	Leu	Arg	Gly	Ala	Ala	Ser	Met	Arg	Cys	Thr
785					790					795					800
Pro	Gln	Gly	Asp	Trp	Ser	Pro	Ala	Ala	Pro	Thr	Cys	Glu	Val	Lys	Ser
				805					810					815	
Cys	Asp	Asp	Phe	Met	Gly	Gln	Leu	Leu	Asn	Gly	Arg	Val	Leu	Phe	Pro
			820					825						830	

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Lys Ser Cys Asp Asp Phe Leu Gly Gln Leu Pro His Gly Arg Val Leu
 1265 1270 1275 1280
 Phe Pro Leu Asn Leu Gln Leu Gly Ala Lys Val Ser Phe Val Cys Asp
 1285 1290 1295
 Glu Gly Phe Arg Leu Lys Gly Ser Ser Val Ser His Cys Val Leu Val
 1300 1305 1310
 Gly Met Arg Ser Leu Trp Asn Asn Ser Val Pro Val Cys Glu His Ile
 1315 1320 1325
 Phe Cys Pro Asn Pro Pro Ala Ile Leu Asn Gly Arg His Thr Gly Thr
 1330 1335 1340
 Pro Ser Gly Asp Ile Pro Tyr Gly Lys Glu Ile Ser Tyr Thr Cys Asp
 1345 1350 1355 1360
 Pro His Pro Asp Arg Gly Met Thr Phe Asn Leu Ile Gly Glu Ser Thr
 1365 1370 1375
 Ile Arg Cys Thr Ser Asp Pro His Gly Asn Gly Val Trp Ser Ser Pro
 1380 1385 1390
 Ala Pro Arg Cys Glu Leu Ser Val Arg Ala Gly His Cys Lys Thr Pro
 1395 1400 1405
 Glu Gln Phe Pro Phe Ala Ser Pro Thr Ile Pro Ile Asn Asp Phe Glu
 1410 1415 1420
 Phe Pro Val Gly Thr Ser Leu Asn Tyr Glu Cys Arg Pro Gly Tyr Phe
 1425 1430 1435 1440
 Gly Lys Met Phe Ser Ile Ser Cys Leu Glu Asn Leu Val Trp Ser Ser
 1445 1450 1455
 Val Glu Asp Asn Cys Arg Arg Lys Ser Cys Gly Pro Pro Pro Glu Pro
 1460 1465 1470
 Phe Asn Gly Met Val His Ile Asn Thr Asp Thr Gln Phe Gly Ser Thr
 1475 1480 1485
 Val Asn Tyr Ser Cys Asn Glu Gly Phe Arg Leu Ile Gly Ser Pro Ser
 1490 1495 1500
 Thr Thr Cys Leu Val Ser Gly Asn Asn Val Thr Trp Asp Lys Lys Ala
 1505 1510 1515 1520
 Pro Ile Cys Glu Ile Ile Ser Cys Glu Pro Pro Pro Thr Ile Ser Asn
 1525 1530 1535
 Gly Asp Phe Tyr Ser Asn Asn Arg Thr Ser Phe His Asn Gly Thr Val
 1540 1545 1550
 Val Thr Tyr Gln Cys His Thr Gly Pro Asp Gly Glu Gln Leu Phe Glu
 1555 1560 1565
 Leu Val Gly Glu Arg Ser Ile Tyr Cys Thr Ser Lys Asp Asp Gln Val
 1570 1575 1580
 Gly Val Trp Ser Ser Pro Pro Pro Arg Cys Ile Ser Thr Asn Lys Cys
 1585 1590 1595 1600
 Thr Ala Pro Glu Val Glu Asn Ala Ile Arg Val Pro Gly Asn Arg Ser
 1605 1610 1615
 Phe Phe Ser Leu Thr Glu Ile Ile Arg Phe Arg Cys Gln Pro Gly Phe
 1620 1625 1630
 Val Met Val Gly Ser His Thr Val Gln Cys Gln Thr Asn Gly Arg Trp
 1635 1640 1645
 Gly Pro Lys Leu Pro His Cys Ser Arg Val Cys Gln Pro Pro Pro Glu
 1650 1655 1660
 Ile Leu His Gly Glu His Thr Leu Ser His Gln Asp Asn Phe Ser Pro
 1665 1670 1675 1680
 Gly Gln Glu Val Phe Tyr Ser Cys Glu Pro Ser Tyr Asp Leu Arg Gly

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1685							1690							1695								
Ala	Ala	Ser	Leu	His	Cys	Thr	Pro	Gln	Gly	Asp	Trp	Ser	Pro	Glu	Ala							
			1700						1705					1710								
Pro	Arg	Cys	Thr	Val	Lys	Ser	Cys	Asp	Asp	Phe	Leu	Gly	Gln	Leu	Pro							
		1715						1720				1725										
His	Gly	Arg	Val	Leu	Leu	Pro	Leu	Asn	Leu	Gln	Leu	Gly	Ala	Lys	Val							
		1730						1735				1740										
Ser	Phe	Val	Cys	Asp	Glu	Gly	Phe	Arg	Leu	Lys	Gly	Arg	Ser	Ala	Ser							
				1750						1755					1760							
His	Cys	Val	Leu	Ala	Gly	Met	Lys	Ala	Leu	Trp	Asn	Ser	Ser	Val	Pro							
			1765						1770					1775								
Val	Cys	Glu	Gln	Ile	Phe	Cys	Pro	Asn	Pro	Pro	Ala	Ile	Leu	Asn	Gly							
			1780						1785					1790								
Arg	His	Thr	Gly	Thr	Pro	Phe	Gly	Asp	Ile	Pro	Tyr	Gly	Lys	Glu	Ile							
			1795					1800					1805									
Ser	Tyr	Ala	Cys	Asp	Thr	His	Pro	Asp	Arg	Gly	Met	Thr	Phe	Asn	Leu							
		1810						1815				1820										
Ile	Gly	Glu	Ser	Ser	Ile	Arg	Cys	Thr	Ser	Asp	Pro	Gln	Gly	Asn	Gly							
		1825			1830					1835				1840								
Val	Trp	Ser	Ser	Pro	Ala	Pro	Arg	Cys	Glu	Leu	Ser	Val	Pro	Ala	Ala							
				1845					1850					1855								
Cys	Pro	His	Pro	Pro	Lys	Ile	Gln	Asn	Gly	His	Tyr	Ile	Gly	Gly	His							
			1860						1865					1870								
Val	Ser	Leu	Tyr	Leu	Pro	Gly	Met	Thr	Ile	Ser	Tyr	Thr	Cys	Asp	Pro							
		1875						1880					1885									
Gly	Tyr	Leu	Leu	Val	Gly	Lys	Gly	Phe	Ile	Phe	Cys	Thr	Asp	Gln	Gly							
		1890				1895						1900										
Ile	Trp	Ser	Gln	Leu	Asp	His	Tyr	Cys	Lys	Glu	Val	Asn	Cys	Ser	Phe							
		1905			1910					1915				1920								
Pro	Leu	Phe	Met	Asn	Gly	Ile	Ser	Lys	Glu	Leu	Glu	Met	Lys	Lys	Val							
				1925					1930					1935								
Tyr	His	Tyr	Gly	Asp	Tyr	Val	Thr	Leu	Lys	Cys	Glu	Asp	Gly	Tyr	Thr							
			1940					1945					1950									
Leu	Glu	Gly	Ser	Pro	Trp	Ser	Gln	Cys	Gln	Ala	Asp	Asp	Arg	Trp	Asp							
		1955						1960					1965									
Pro	Pro	Leu	Ala	Lys	Cys	Thr	Ser	Arg	Ala	His	Asp	Ala	Leu	Ile	Val							
		1970						1975				1980										
Gly	Thr	Leu	Ser	Gly	Thr	Ile	Phe	Phe	Ile	Leu	Leu	Ile	Ile	Phe	Leu							
		1985			1990					1995				2000								
Ser	Trp	Ile	Ile	Leu	Lys	His	Arg	Lys	Gly	Asn	Asn	Ala	His	Glu	Asn							
				2005					2010					2015								
Pro	Lys	Glu	Val	Ala	Ile	His	Leu	His	Ser	Gln	Gly	Gly	Ser	Ser	Val							
			2020						2025				2030									
His	Pro	Arg	Thr	Leu	Gln	Thr	Asn	Glu	Glu	Asn	Ser	Arg	Val	Leu	Pro							
			2035					2040					2045									

<210> SEQ ID NO 7
 <211> LENGTH: 1029
 <212> TYPE: DNA
 <213> ORGANISM: Artificial Sequence
 <220> FEATURE:
 <223> OTHER INFORMATION: Description of Artificial Sequence;
 note=synthetic construct

<400> SEQUENCE: 7

tgtaggagc caccaacatt tgaagctatg gagctcattg gtaaaccaaa accctactat 60

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gagattggtg aacgagtaga ttataagtgt aaaaaggat acttctatat acctcctctt 120
gccaccata ctatttgtga tcggaatcat acatggctac ctgtctcaga tgacgcctgt 180
tatagagaaa catgtccata tatacgggat cctttaaag gccaagcagt cctgcaaat 240
gggacttacg agtttggtta tcagatgcac tttatttga atgagggtta ttacttaatt 300
ggtgaagaaa ttctatattg tgaacttaaa ggatcagtag caatttgag cggtaaagccc 360
ccaatatgtg aaaaggtttt gtgtacacca cctccaaaaa taaaaaatgg aaaacacacc 420
tttagtgaag tagaagtatt tgagtatctt gatgcagtaa cttatagttg tgactctgca 480
cctggaccag atccatttcc acttattgga gagagcacga tttattgtgg tgacaattca 540
gtgtggagtc gtgctgctcc agagtgtaaa gtggtcaaat gtcgatttcc agtagtcgaa 600
aatgaaaaac agataticagg atttgaaaaa aaattttact acaagcaac agttatgttt 660
gaatgcgata agggttttta cctcgatggc agcgacacaa ttgtctgtga cagtaacagt 720
acttgggagc ccccgattcc aaagtgtctt aaagtgtcga cttcttccac tacaaaatct 780
ccagcgtcca gtgcctcagg tcctaggcct acttacaagc ctccagtcctc aaattatcca 840
ggatataccta aacctgagga aggaatactt gacagtttgg atgtttgggt cattgctgtg 900
attgttattg ccatagttgt tggagttgca gtaatttggg ttgtcccgtc cagatatctt 960
caaaggagga agaagaaagg cacataccta actgatgaga cccacagaga agtaaaattt 1020
acttctctc 1029

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<210> SEQ ID NO 8
<211> LENGTH: 376
<212> TYPE: PRT
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Description of Artificial Sequence;
note=synthetic construct

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<400> SEQUENCE: 8

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Met Glu Pro Pro Gly Arg Arg Glu Cys Pro Phe Pro Ser Trp Arg Phe
 1          5          10          15
Phe Pro Gly Leu Leu Leu Ala Ala Met Val Leu Leu Lys Phe Ser Asp
 20          25          30
Ala Cys Glu Glu Pro Pro Thr Phe Glu Ala Met Glu Leu Ile Gly Lys
 35          40          45
Pro Lys Pro Tyr Tyr Glu Ile Gly Glu Arg Val Asp Tyr Lys Cys Lys
 50          55          60
Lys Gly Tyr Phe Tyr Ile Pro Pro Leu Ala Thr His Thr Ile Cys Asp
 65          70          75          80
Arg Asn His Thr Trp Leu Pro Val Ser Asp Asp Ala Cys Tyr Arg Glu
 85          90          95
Thr Cys Pro Tyr Ile Arg Asp Pro Leu Asn Gly Gln Ala Val Pro Ala
100          105          110
Asn Gly Thr Tyr Glu Phe Gly Tyr Gln Met His Phe Ile Cys Asn Glu
115          120          125
Gly Tyr Tyr Leu Ile Gly Glu Glu Ile Leu Tyr Cys Glu Leu Lys Gly
130          135          140
Ser Val Ala Ile Trp Ser Gly Lys Pro Pro Ile Cys Glu Lys Val Leu
145          150          155          160
Cys Thr Pro Pro Pro Lys Ile Lys Asn Gly Lys His Thr Phe Ser Glu
165          170          175
Val Glu Val Phe Glu Tyr Leu Asp Ala Val Thr Tyr Ser Cys Asp Pro

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Ala	Pro	Gly	Pro	Asp	Pro	Phe	Ser	Leu	Ile	Gly	Glu	Ser	Thr	Ile	Tyr
	195						200					205			
Cys	Gly	Asp	Asn	Ser	Val	Trp	Ser	Arg	Ala	Ala	Pro	Glu	Cys	Lys	Val
	210					215					220				
Val	Lys	Cys	Arg	Phe	Pro	Val	Val	Glu	Asn	Gly	Lys	Gln	Ile	Ser	Gly
	225				230					235					240
Phe	Gly	Lys	Lys	Phe	Tyr	Tyr	Lys	Ala	Thr	Val	Met	Phe	Glu	Cys	Asp
				245					250					255	
Lys	Gly	Phe	Tyr	Leu	Asp	Gly	Ser	Asp	Thr	Ile	Val	Cys	Asp	Ser	Asn
			260					265					270		
Ser	Thr	Trp	Asp	Pro	Pro	Val	Pro	Lys	Cys	Leu	Lys	Val	Ser	Thr	Ser
		275					280					285			
Ser	Thr	Thr	Lys	Ser	Pro	Ala	Ser	Ser	Ala	Ser	Gly	Pro	Arg	Pro	Thr
	290					295					300				
Tyr	Lys	Pro	Pro	Val	Ser	Asn	Tyr	Pro	Gly	Tyr	Pro	Lys	Pro	Glu	Glu
	305					310				315					320
Gly	Ile	Leu	Asp	Ser	Leu	Asp	Val	Trp	Val	Ile	Ala	Val	Ile	Val	Ile
				325					330					335	
Ala	Ile	Val	Val	Gly	Val	Ala	Val	Ile	Cys	Val	Val	Pro	Tyr	Arg	Tyr
		340						345						350	
Leu	Gln	Arg	Arg	Lys	Lys	Lys	Gly	Thr	Tyr	Leu	Thr	Asp	Glu	Thr	His
		355					360					365			
Arg	Glu	Val	Lys	Phe	Thr	Ser	Leu								
	370						375								

<210> SEQ ID NO 9

<211> LENGTH: 436

<212> TYPE: PRT

<213> ORGANISM: Artificial Sequence

<220> FEATURE:

<223> OTHER INFORMATION: Description of Artificial Sequence;
note=synthetic construct

<400> SEQUENCE: 9

Met	Glu	Val	Ser	Ser	Arg	Ser	Ser	Glu	Pro	Leu	Asp	Pro	Val	Trp	Leu
	1			5					10					15	
Leu	Val	Ala	Phe	Gly	Arg	Gly	Gly	Val	Lys	Leu	Glu	Val	Leu	Leu	Leu
		20					25						30		
Phe	Leu	Leu	Pro	Phe	Thr	Leu	Gly	His	Cys	Pro	Ala	Pro	Ser	Gln	Leu
		35					40					45			
Pro	Ser	Ala	Lys	Pro	Ile	Asn	Leu	Thr	Asp	Glu	Ser	Met	Phe	Pro	Ile
		50				55					60				
Gly	Thr	Tyr	Leu	Leu	Tyr	Glu	Cys	Leu	Pro	Gly	Tyr	Ile	Lys	Arg	Gln
	65				70					75					80
Phe	Ser	Ile	Thr	Cys	Lys	Gln	Asp	Ser	Thr	Trp	Thr	Ser	Ala	Glu	Asp
				85					90					95	
Lys	Cys	Ile	Arg	Lys	Gln	Cys	Lys	Thr	Pro	Ser	Asp	Pro	Glu	Asn	Gly
			100						105					110	
Leu	Val	His	Val	His	Thr	Gly	Ile	Gln	Phe	Gly	Ser	Arg	Ile	Asn	Tyr
		115					120						125		
Thr	Cys	Asn	Gln	Gly	Tyr	Arg	Leu	Ile	Gly	Ser	Ser	Ser	Ala	Val	Cys
		130				135						140			
Val	Ile	Thr	Asp	Gln	Ser	Val	Asp	Trp	Asp	Thr	Glu	Ala	Pro	Ile	Cys
	145				150					155					160
Glu	Trp	Ile	Pro	Cys	Glu	Ile	Pro	Pro	Gly	Ile	Pro	Asn	Gly	Asp	Phe

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145	150	155	160
Val Ala Thr	Ala Thr Gly Val His Ser Gln Val Gln Leu Glu Gln Ser		
	165	170	175
Gly Pro Glu	Leu Val Arg Pro Gly Val Ser Val Lys Ile Ser Cys Lys		
	180	185	190
Gly Ser Gly	Tyr Thr Phe Thr Asp Tyr Ala Ile His Trp Val Lys Gln		
	195	200	205
Ser His Ala	Lys Ser Leu Glu Trp Ile Gly Val Ile Ser Thr Tyr Tyr		
	210	215	220
Gly His Thr	His Tyr Asn Gln Lys Phe Lys Gly Lys Ala Thr Leu Thr		
	225	230	235
Val Asp Lys	Ser Ser Asn Thr Ala Tyr Met Glu Leu Ala Arg Leu Thr		
	245	250	255
Ser Glu Asp	Ser Ala Ile Tyr Tyr Cys Ala Arg Pro Asn Asn Tyr Gly		
	260	265	270
Ser Ser Pro	Pro Tyr Tyr Ala Met Asp Cys Trp Gly Gln Gly Thr Ser		
	275	280	285
Val Thr Val	Ser Ser Gly Gly Gly Ser Gly Gly Gly Ser Gly		
	290	295	300
Gly Gly Gly	Ser Cys Pro Ala Pro Pro Leu Phe Pro Tyr Ala Lys Pro		
	305	310	315
Ile Asn Pro	Thr Asp Glu Ser Thr Phe Pro Val Gly Thr Ser Leu Lys		
	325	330	335
Tyr Glu Cys	Arg Pro Gly Tyr Ile Lys Arg Gln Phe Ser Ile Thr Cys		
	340	345	350
Glu Val Asn	Ser Val Trp Thr Ser Pro Gln Asp Val Cys Ile Arg Lys		
	355	360	365
Gln Cys Glu	Thr Pro Leu Asp Pro Gln Asn Gly Ile Val His Val Asn		
	370	375	380
Thr Asp Ile	Arg Phe Gly Ser Ser Ile Thr Tyr Thr Cys Asn Glu Gly		
	385	390	395
Tyr Arg Leu	Ile Gly Ser Ser Ser Ala Met Cys Ile Ile Ser Asp Gln		
	405	410	415
Ser Val Ala	Trp Asp Ala Glu Ala Pro Ile Cys Glu Ser Ile Pro Cys		
	420	425	430
Glu Ile Pro	Pro Ser Ile Pro Asn Gly Asp Phe Phe Ser Pro Asn Arg		
	435	440	445
Glu Asp Phe	His Tyr Gly Met Val Val Thr Tyr Gln Cys Asn Thr Asp		
	450	455	460
Arg Lys Lys	Leu Phe Asn Leu Val Gly Glu Pro Ser Ile His Cys Thr		
	465	470	475
Ser Ile Asp	Gly Gln Val Gly Val Trp Ser Gly Pro Pro Pro Gln Cys		
	485	490	495
Ile Glu Leu	Asn Lys Cys Thr Pro Pro His Val Glu Asn Ala Val Ile		
	500	505	510
Val Ser Lys	Asn Lys Ser Leu Phe Ser Leu Arg Asp Met Val Glu Phe		
	515	520	525
Arg Cys Gln	Asp Gly Phe Met Met Lys Gly Asp Ser Ser Val Tyr Cys		
	530	535	540
Arg Ser Leu	Asn Arg Trp Glu Pro Gln Leu Pro Ser Cys Phe Lys Val		
	545	550	555
Lys Ser Cys	Gly Ala Phe Leu Gly Glu Leu Pro Asn Gly His Val Phe		
	565	570	575

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Val Pro Gln Asn Leu Gln Leu Gly Ala Lys Val Thr Phe Val Cys Asn
 580 585 590
 Thr Gly Tyr Gln Leu Lys Gly Asn Ser Ser Ser His Cys Val Leu Asp
 595 600 605
 Gly Val Glu Ser Ile Trp Asn Ser Ser Val Pro Val Cys Glu Gln Val
 610 615 620
 Ile Cys Lys Leu Pro Gln Asp Met Ser Gly Phe Gln Lys Gly Leu Gln
 625 630 635 640
 Met Lys Lys Asp Tyr Tyr Tyr Gly Asp Asn Val Glu Cys Glu Asp Gly
 645 650 655
 Tyr Thr Leu Glu Gly Ser Ser Gln Ser Gln Cys Gln Ser Asp Ala Ser
 660 665 670
 Trp Asp Pro Pro Leu Pro Lys Cys Val Ser Gln Val Ile Cys Lys Leu
 675 680 685
 Pro Gln Asp Met Ser Gly Phe Gln Lys Gly Leu Gln Met Lys Lys Asp
 690 695 700
 Tyr Tyr Tyr Gly Asp Asn Val Glu Cys Glu Asp Gly Tyr Thr Leu Glu
 705 710 715 720
 Gly Ser Ser Gln Ser Gln Cys Gln Ser Asp Ala Ser Trp Asp Pro Pro
 725 730 735
 Leu Pro Lys Cys Val Ser Arg Ser Asn Ser Gly Leu Ile Ala Gly Ile
 740 745 750
 Phe Ile Gly Ile Ile Val Leu Ile Leu Phe Ile Ile Phe Ser Tyr Trp
 755 760 765
 Met Ile Met Lys Phe Lys Lys Arg Asn Ser Thr Asn Glu Lys Cys Lys
 770 775 780
 Glu Val Gly Ile Tyr Leu Asn Ser Lys Glu Asp Ser Cys Val Gln Pro
 785 790 795 800
 Gln Ser Leu Leu Thr Ser Gln Glu Asn Asn Ser Thr Ser Ser Pro Ala
 805 810 815
 Arg Asn Ser Leu Thr Gln Glu Val
 820

<210> SEQ ID NO 12
 <211> LENGTH: 412
 <212> TYPE: PRT
 <213> ORGANISM: Artificial Sequence
 <220> FEATURE:
 <223> OTHER INFORMATION: Description of Artificial Sequence;
 note=synthetic construct

<400> SEQUENCE: 12

Met Gly Phe Lys Met Glu Ser His Ser Gln Val Leu Met Leu Leu Leu
 1 5 10 15
 Leu Trp Val Ser Gly Thr Cys Gly Asp Ile Val Met Ser Gln Ser Pro
 20 25 30
 Ser Ser Leu Ala Val Ser Val Gly Glu Lys Val Thr Met Asn Cys Lys
 35 40 45
 Ser Ser Gln Ser Leu Lys Thr Asn Gln Lys Asn Tyr Leu Ala Trp Tyr
 50 55 60
 Gln Gln Lys Pro Gly Gln Ser Pro Lys Leu Leu Ile Tyr Trp Ala Ser
 65 70 75 80
 Ser Arg Glu Ser Gly Val Pro Asp Arg Phe Thr Gly Ser Gly Ser Gly
 85 90 95
 Thr Asp Phe Thr Leu Thr Ile Ser Ser Val Lys Ala Glu Asp Leu Ala
 100 105 110

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Val Tyr Tyr Cys Gln Gln Tyr Tyr Ser Tyr Pro Leu Thr Phe Gly Ala
 115 120 125
 Gly Thr Lys Leu Glu Leu Lys Gly Gly Gly Gly Ser Gly Gly Gly Gly
 130 135 140
 Ser Gly Gly Gly Gly Ser Met Gly Trp Ser Cys Val Met Leu Phe Leu
 145 150 155 160
 Val Ala Thr Ala Thr Gly Val His Ser Gln Val Gln Leu Glu Gln Ser
 165 170 175
 Gly Pro Glu Leu Val Arg Pro Gly Val Ser Val Lys Ile Ser Cys Lys
 180 185 190
 Gly Ser Gly Tyr Thr Phe Thr Asp Tyr Ala Ile His Trp Val Lys Gln
 195 200 205
 Ser His Ala Lys Ser Leu Glu Trp Ile Gly Val Ile Ser Thr Tyr Tyr
 210 215 220
 Gly His Thr His Tyr Asn Gln Lys Phe Lys Gly Lys Ala Thr Leu Thr
 225 230 235 240
 Val Asp Lys Ser Ser Asn Thr Ala Tyr Met Glu Leu Ala Arg Leu Thr
 245 250 255
 Ser Glu Asp Ser Ala Ile Tyr Tyr Cys Ala Arg Pro Asn Asn Tyr Gly
 260 265 270
 Ser Ser Pro Pro Tyr Tyr Ala Met Asp Cys Trp Gly Gln Gly Thr Ser
 275 280 285
 Val Thr Val Ser Ser Gly Gly Gly Gly Ser Gly Gly Gly Gly Ser Gly
 290 295 300
 Gly Gly Gly Ser Leu Arg Cys Tyr Asn Cys Leu Asp Pro Val Ser Ser
 305 310 315 320
 Cys Lys Thr Asn Ser Thr Cys Ser Pro Asn Leu Asp Ala Cys Leu Val
 325 330 335
 Ala Val Ser Gly Lys Gln Val Tyr Gln Gln Cys Trp Arg Phe Ser Asp
 340 345 350
 Cys Asn Ala Lys Phe Ile Leu Ser Arg Leu Glu Ile Ala Asn Val Gln
 355 360 365
 Tyr Arg Cys Cys Gln Ala Asp Leu Cys Asn Lys Ser Phe Glu Asp Lys
 370 375 380
 Pro Asn Asn Gly Ala Ile Ser Leu Leu Gly Lys Thr Ala Leu Leu Val
 385 390 395 400
 Thr Ser Val Leu Ala Ala Ile Leu Lys Pro Cys Phe
 405 410

<210> SEQ ID NO 13
 <211> LENGTH: 762
 <212> TYPE: PRT
 <213> ORGANISM: Artificial Sequence
 <220> FEATURE:
 <223> OTHER INFORMATION: Description of Artificial Sequence;
 note=synthetic construct

<400> SEQUENCE: 13

Met Lys Thr Pro Trp Lys Val Leu Leu Gly Leu Leu Gly Ala Ala Ala
 1 5 10 15
 Leu Val Thr Ile Ile Thr Val Pro Val Val Leu Leu Asn Lys Gly Thr
 20 25 30
 Asp Asp Ala Thr Ala Asp Ser Arg Lys Thr Tyr Thr Leu Thr Asp Tyr
 35 40 45
 Leu Lys Asn Tyr Leu Lys Lys Leu Arg Trp Ile Ser Asp His Glu Tyr
 50 55 60

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Leu	Tyr	Lys	Gln	Glu	Asn	Asn	Ile	Leu	Val	Phe	Asn	Ala	Glu	Tyr	Gly	65	70	75	80
Asn	Ser	Ser	Val	Phe	Leu	Glu	Asn	Ser	Thr	Phe	Asp	Glu	Phe	Gly	His	85	90	95	
Ser	Ile	Asn	Asp	Tyr	Ser	Ile	Ser	Pro	Asp	Gly	Gln	Phe	Ile	Leu	Leu	100	105	110	
Glu	Tyr	Asn	Tyr	Val	Lys	Gln	Trp	Arg	His	Ser	Tyr	Thr	Ala	Ser	Tyr	115	120	125	
Asp	Ile	Tyr	Asp	Leu	Asn	Lys	Arg	Gln	Leu	Ile	Thr	Glu	Glu	Arg	Ile	130	135	140	
Pro	Asn	Asn	Thr	Gln	Trp	Val	Thr	Trp	Ser	Pro	Val	Gly	His	Lys	Leu	145	150	155	160
Ala	Tyr	Val	Trp	Asn	Asn	Asp	Ile	Tyr	Val	Lys	Ile	Glu	Pro	Asn	Leu	165	170	175	
Pro	Ser	Tyr	Arg	Ile	Thr	Trp	Thr	Gly	Lys	Glu	Asp	Ile	Ile	Tyr	Asn	180	185	190	
Gly	Ile	Thr	Asp	Trp	Val	Tyr	Glu	Glu	Glu	Val	Phe	Ser	Ala	Tyr	Ser	195	200	205	
Ala	Leu	Trp	Trp	Ser	Pro	Asn	Gly	Thr	Phe	Leu	Ala	Tyr	Ala	Gln	Phe	210	215	220	
Asn	Asp	Thr	Glu	Val	Pro	Leu	Ile	Glu	Tyr	Ser	Phe	Tyr	Ser	Asp	Glu	225	230	235	240
Ser	Leu	Gln	Tyr	Pro	Lys	Thr	Val	Arg	Val	Pro	Tyr	Pro	Lys	Ala	Gly	245	250	255	
Ala	Val	Asn	Pro	Thr	Val	Lys	Phe	Phe	Val	Val	Asn	Thr	Asp	Ser	Leu	260	265	270	
Ser	Ser	Val	Thr	Asn	Ala	Thr	Ser	Ile	Gln	Ile	Thr	Ala	Pro	Ala	Ser	275	280	285	
Met	Leu	Ile	Gly	Asp	His	Tyr	Leu	Cys	Asp	Val	Thr	Trp	Ala	Thr	Gln	290	295	300	
Glu	Arg	Ile	Ser	Leu	Gln	Trp	Leu	Arg	Arg	Ile	Gln	Asn	Tyr	Ser	Val	305	310	315	320
Met	Asp	Ile	Cys	Asp	Tyr	Asp	Glu	Ser	Ser	Gly	Arg	Trp	Asn	Cys	Leu	325	330	335	
Val	Ala	Arg	Gln	His	Ile	Glu	Met	Ser	Thr	Thr	Gly	Trp	Val	Gly	Arg	340	345	350	
Phe	Arg	Pro	Ser	Glu	Pro	His	Phe	Thr	Leu	Asp	Gly	Asn	Ser	Phe	Tyr	355	360	365	
Lys	Ile	Ile	Ser	Asn	Glu	Glu	Gly	Tyr	Arg	His	Ile	Cys	Tyr	Phe	Gln	370	375	380	
Ile	Asp	Lys	Lys	Asp	Cys	Thr	Phe	Ile	Thr	Lys	Gly	Thr	Trp	Glu	Val	385	390	395	400
Ile	Gly	Ile	Glu	Ala	Leu	Thr	Ser	Asp	Tyr	Leu	Tyr	Tyr	Ile	Ser	Asn	405	410	415	
Glu	Tyr	Lys	Gly	Met	Pro	Gly	Gly	Arg	Asn	Leu	Tyr	Lys	Ile	Gln	Leu	420	425	430	
Ser	Asp	Tyr	Thr	Lys	Val	Thr	Cys	Leu	Ser	Cys	Glu	Leu	Asn	Pro	Glu	435	440	445	
Arg	Cys	Gln	Tyr	Tyr	Ser	Val	Ser	Phe	Ser	Lys	Glu	Ala	Lys	Tyr	Tyr	450	455	460	
Gln	Leu	Arg	Cys	Ser	Gly	Pro	Gly	Leu	Pro	Leu	Tyr	Thr	Leu	His	Ser	465	470	475	480
Ser	Val	Asn	Asp	Lys	Gly	Leu	Arg	Val	Leu	Glu	Asp	Asn	Ser	Ala	Leu	485	490	495	

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Asp Lys Met Leu Gln Asn Val Gln Met Pro Ser Lys Lys Leu Asp Phe
 500 505 510
 Ile Ile Leu Asn Glu Thr Lys Phe Trp Tyr Gln Met Ile Leu Pro Pro
 515 520 525
 His Phe Asp Lys Ser Lys Lys Tyr Pro Leu Leu Leu Asp Val Tyr Ala
 530 535 540
 Gly Pro Cys Ser Gln Lys Ala Asp Thr Val Phe Arg Leu Asn Trp Ala
 545 550 555 560
 Thr Tyr Leu Ala Ser Thr Glu Asn Ile Ile Val Ala Ser Phe Asp Gly
 565 570 575
 Arg Gly Ser Gly Tyr Gln Gly Asp Lys Ile Met His Ala Ile Asn Arg
 580 585 590
 Arg Leu Gly Thr Phe Glu Val Glu Asp Gln Ile Glu Ala Ala Arg Gln
 595 600 605
 Phe Ser Lys Met Gly Phe Val Asp Asn Lys Arg Ile Ala Ile Trp Gly
 610 615 620
 Trp Ser Tyr Gly Gly Tyr Val Thr Ser Met Val Leu Gly Ser Gly Ser
 625 630 635 640
 Gly Val Phe Lys Cys Gly Ile Ala Val Ala Pro Val Ser Arg Trp Glu
 645 650
 Tyr Tyr Asp Ser Val Tyr Thr Glu Arg Tyr Met Gly Leu Pro Thr Pro
 660 665 670
 Glu Asp Asn Leu Asp His Tyr Arg Asn Ser Thr Val Met Ser Arg Ala
 675 680 685
 Glu Asn Phe Lys Gln Val Glu Tyr Leu Leu Ile His Gly Thr Ala Asp
 690 695 700
 Asp Asn Val His Phe Gln Gln Ser Ala Gln Ile Ser Lys Ala Leu Val
 705 710 715 720
 Asp Val Gly Val Asp Phe Gln Ala Met Trp Tyr Thr Asp Glu Asp His
 725 730 735
 Gly Ile Ala Ser Ser Thr Ala His Gln His Ile Tyr Thr His Met Ser
 740 745 750
 His Phe Ile Lys Gln Cys Phe Ser Leu Pro
 755 760

<210> SEQ ID NO 14

<211> LENGTH: 481

<212> TYPE: PRT

<213> ORGANISM: Artificial Sequence

<220> FEATURE:

<223> OTHER INFORMATION: Description of Artificial Sequence;
 note=synthetic construct

<400> SEQUENCE: 14

Met Gly Cys Leu Trp Gly Leu Ala Leu Pro Leu Phe Phe Phe Cys Trp
 1 5 10 15
 Glu Val Gly Val Ser Gly Ser Ser Ala Gly Pro Ser Thr Arg Arg Ala
 20 25 30
 Asp Thr Ala Met Thr Thr Asp Asp Thr Glu Val Pro Ala Met Thr Leu
 35 40 45
 Ala Pro Gly His Ala Ala Leu Glu Thr Gln Thr Leu Ser Ala Glu Thr
 50 55 60
 Ser Ser Arg Ala Ser Thr Pro Ala Gly Pro Ile Pro Glu Ala Glu Thr
 65 70 75 80
 Arg Gly Ala Lys Arg Ile Ser Pro Ala Arg Glu Thr Arg Ser Phe Thr
 85 90 95

-continued

Lys Thr Ser Pro Asn Phe Met Val Leu Ile Ala Thr Ser Val Glu Thr
 100 105 110
 Ser Ala Ala Ser Gly Ser Pro Glu Gly Ala Gly Met Thr Thr Val Gln
 115 120 125
 Thr Ile Thr Gly Ser Asp Pro Glu Glu Ala Ile Phe Asp Thr Leu Cys
 130 135 140
 Thr Asp Asp Ser Ser Glu Glu Ala Lys Thr Leu Thr Met Asp Ile Leu
 145 150 155 160
 Thr Leu Ala His Thr Ser Thr Glu Ala Lys Gly Leu Ser Ser Glu Ser
 165 170 175
 Ser Ala Ser Ser Asp Gly Pro His Pro Val Ile Thr Pro Ser Arg Ala
 180 185 190
 Ser Glu Ser Ser Ala Ser Ser Asp Gly Pro His Pro Val Ile Thr Pro
 195 200 205
 Ser Arg Ala Ser Glu Ser Ser Ala Ser Ser Asp Gly Pro His Pro Val
 210 215 220
 Ile Thr Pro Ser Trp Ser Pro Gly Ser Asp Val Leu Ala Glu Ala Leu
 225 230 235 240
 Val Thr Val Thr Asn Ile Glu Val Ile Asn Cys Ser Ile Thr Glu Ile
 245 250 255
 Glu Thr Thr Thr Ser Ser Ile Pro Gly Ala Ser Asp Ile Asp Leu Ile
 260 265 270
 Pro Thr Glu Gly Val Lys Ala Ser Ser Thr Ser Asp Pro Pro Ala Leu
 275 280 285
 Pro Asp Ser Thr Glu Ala Lys Pro His Ile Thr Glu Val Thr Ala Ser
 290 295 300
 Ala Glu Thr Leu Ser Ser Ala Leu Ser Val Glu Thr Pro Ser Tyr Val
 305 310 315 320
 Lys Val Ser Gly Ala Ala Pro Val Ser Ile Glu Ala Gly Ser Ala Val
 325 330 335
 Gly Lys Thr Thr Ser Phe Ala Gly Ser Ser Ala Ser Ser Tyr Ser Pro
 340 345 350
 Ser Glu Ala Ala Leu Lys Asn Phe Thr Pro Ser Glu Thr Pro Thr Met
 355 360 365
 Asp Ile Ala Thr Lys Gly Pro Phe Pro Thr Ser Arg Asp Pro Leu Pro
 370 375 380
 Ser Val Pro Pro Thr Thr Thr Asn Ser Ser Arg Gly Thr Asn Ser Thr
 385 390 395 400
 Leu Ala Lys Ile Thr Thr Ser Ala Lys Thr Thr Met Lys Pro Pro Thr
 405 410 415
 Ala Thr Pro Thr Thr Ala Arg Trp Thr Thr Asp Val Ser Ala Gly Glu
 420 425 430
 Asn Gly Gly Phe Leu Leu Leu Arg Leu Ser Val Asp Glu Asp Leu Thr
 435 440 445
 Asp Pro Arg Val Ala Glu Arg Leu Met Gln Gln Leu His Arg Glu Leu
 450 455 460
 His Ala His Ala Pro His Phe Gln Val Ser Leu Leu Arg Val Arg Arg
 465 470 475 480
 Gly

<210> SEQ ID NO 15

<211> LENGTH: 359

<212> TYPE: PRT

<213> ORGANISM: Artificial Sequence

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<220> FEATURE:

<223> OTHER INFORMATION: Description of Artificial Sequence;
note=synthetic construct

<400> SEQUENCE: 15

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Met His Pro Gln Val Val Ile Leu Ser Leu Ile Leu His Leu Ala Asp
 1          5          10          15
Ser Val Ala Gly Ser Val Lys Val Gly Gly Glu Ala Gly Pro Ser Val
 20          25          30
Thr Leu Pro Cys His Tyr Ser Gly Ala Val Thr Ser Met Cys Trp Asn
 35          40          45
Arg Gly Ser Cys Ser Leu Phe Thr Cys Gln Asn Gly Ile Val Trp Thr
 50          55          60
Asn Gly Thr His Val Thr Tyr Arg Lys Asp Thr Arg Tyr Lys Leu Leu
 65          70          75          80
Gly Asp Leu Ser Arg Arg Asp Val Ser Leu Thr Ile Glu Asn Thr Ala
 85          90          95
Val Ser Asp Ser Gly Val Tyr Cys Cys Arg Val Glu His Arg Gly Trp
 100         105         110
Phe Asn Asp Met Lys Ile Thr Val Ser Leu Glu Ile Val Pro Pro Lys
 115         120         125
Val Thr Thr Thr Pro Ile Val Thr Thr Val Pro Thr Val Thr Thr Val
 130         135         140
Arg Thr Ser Thr Thr Val Pro Thr Thr Thr Thr Val Pro Thr Thr Thr
 145         150         155         160
Val Pro Thr Thr Met Ser Ile Pro Thr Thr Thr Thr Val Pro Thr Thr
 165         170         175
Met Thr Val Ser Thr Thr Thr Ser Val Pro Thr Thr Thr Ser Ile Pro
 180         185         190
Thr Thr Thr Ser Val Pro Val Thr Thr Thr Val Ser Thr Phe Val Pro
 195         200         205
Pro Met Pro Leu Pro Arg Gln Asn His Glu Pro Val Ala Thr Ser Pro
 210         215         220
Ser Ser Pro Gln Pro Ala Glu Thr His Pro Thr Thr Leu Gln Gly Ala
 225         230         235         240
Ile Arg Arg Glu Pro Thr Ser Ser Pro Leu Tyr Ser Tyr Thr Thr Asp
 245         250         255
Gly Asn Asp Thr Val Thr Glu Ser Ser Asp Gly Leu Trp Asn Asn Asn
 260         265         270
Gln Thr Gln Leu Phe Leu Glu His Ser Leu Leu Thr Ala Asn Thr Thr
 275         280         285
Lys Gly Ile Tyr Ala Gly Val Cys Ile Ser Val Leu Val Leu Leu Ala
 290         295         300
Leu Leu Gly Val Ile Ile Ala Lys Lys Tyr Phe Phe Lys Lys Glu Val
 305         310         315         320
Gln Gln Leu Ser Val Ser Phe Ser Ser Leu Gln Ile Lys Ala Leu Gln
 325         330         335
Asn Ala Val Glu Lys Glu Val Gln Ala Glu Asp Asn Ile Tyr Ile Glu
 340         345         350
Asn Ser Leu Tyr Ala Thr Asp
 355

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<210> SEQ ID NO 16

<211> LENGTH: 30

<212> TYPE: DNA

<213> ORGANISM: Artificial Sequence

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<220> FEATURE:
 <223> OTHER INFORMATION: Description of Artificial Sequence;
 note=synthetic construct

<400> SEQUENCE: 16

atgaagttgc ctgtaggct gttggtgctg 30

<210> SEQ ID NO 17
 <211> LENGTH: 20
 <212> TYPE: DNA
 <213> ORGANISM: Artificial Sequence
 <220> FEATURE:
 <223> OTHER INFORMATION: Description of Artificial Sequence;
 note=synthetic construct

<400> SEQUENCE: 17

actggatggt ggaagatgg 20

<210> SEQ ID NO 18
 <211> LENGTH: 27
 <212> TYPE: DNA
 <213> ORGANISM: Artificial Sequence
 <220> FEATURE:
 <223> OTHER INFORMATION: Description of Artificial Sequence;
 note=synthetic construct

<400> SEQUENCE: 18

atgaaatgca gctggggcat gttcttc 27

<210> SEQ ID NO 19
 <211> LENGTH: 21
 <212> TYPE: DNA
 <213> ORGANISM: Artificial Sequence
 <220> FEATURE:
 <223> OTHER INFORMATION: Description of Artificial Sequence;
 note=synthetic construct

<400> SEQUENCE: 19

cagtggatag accgatgggc c 21

<210> SEQ ID NO 20
 <211> LENGTH: 13
 <212> TYPE: PRT
 <213> ORGANISM: Artificial Sequence
 <220> FEATURE:
 <223> OTHER INFORMATION: Description of Artificial Sequence;
 note=synthetic construct

<400> SEQUENCE: 20

Ser Asp Cys Ile Lys Ser Ala Ala Arg Leu Ile Gln Asn
 1 5 10

<210> SEQ ID NO 21
 <211> LENGTH: 14
 <212> TYPE: PRT
 <213> ORGANISM: Artificial Sequence
 <220> FEATURE:
 <223> OTHER INFORMATION: Description of Artificial Sequence;
 note=synthetic construct

<400> SEQUENCE: 21

Phe Glu Val Glu Asp Gln Ile Glu Ala Ala Arg Gln Phe Ser
 1 5 10

<210> SEQ ID NO 22
 <211> LENGTH: 12

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<212> TYPE: PRT
 <213> ORGANISM: Artificial Sequence
 <220> FEATURE:
 <223> OTHER INFORMATION: Description of Artificial Sequence;
 note=synthetic construct

<400> SEQUENCE: 22

Phe Phe Lys Tyr Ala Cys Gly Gly Trp Leu Lys Arg
 1 5 10

<210> SEQ ID NO 23
 <211> LENGTH: 10
 <212> TYPE: PRT
 <213> ORGANISM: Artificial Sequence
 <220> FEATURE:
 <223> OTHER INFORMATION: Description of Artificial Sequence;
 note=synthetic construct

<400> SEQUENCE: 23

Tyr Gly Gly Tyr Val Thr Ser Met Val Leu
 1 5 10

<210> SEQ ID NO 24
 <211> LENGTH: 22
 <212> TYPE: PRT
 <213> ORGANISM: Artificial Sequence
 <220> FEATURE:
 <223> OTHER INFORMATION: Description of Artificial Sequence;
 note=synthetic construct

<400> SEQUENCE: 24

Ser Lys His Val Val Glu Asp Leu Ile Ala Gln Ile Arg Glu Val Phe
 1 5 10 15

Ile Gln Thr Leu Asp Asp
 20

<210> SEQ ID NO 25
 <211> LENGTH: 14
 <212> TYPE: PRT
 <213> ORGANISM: Artificial Sequence
 <220> FEATURE:
 <223> OTHER INFORMATION: Description of Artificial Sequence;
 note=synthetic construct

<400> SEQUENCE: 25

Phe Gln Gln Ser Ala Gln Ile Ser Lys Ala Leu Val Asp Val
 1 5 10

<210> SEQ ID NO 26
 <211> LENGTH: 11
 <212> TYPE: PRT
 <213> ORGANISM: Artificial Sequence
 <220> FEATURE:
 <223> OTHER INFORMATION: Description of Artificial Sequence;
 note=synthetic construct

<400> SEQUENCE: 26

Ala Glu Thr Lys Lys Arg Ala Glu Glu Lys Ala
 1 5 10

<210> SEQ ID NO 27
 <211> LENGTH: 19
 <212> TYPE: PRT
 <213> ORGANISM: Artificial Sequence
 <220> FEATURE:
 <223> OTHER INFORMATION: Description of Artificial Sequence;
 note=synthetic construct

-continued

<400> SEQUENCE: 27

Ser	Thr	Ala	His	Gln	His	Ile	Tyr	Thr	His	Met	Ser	His	Phe	Ile	Lys
1				5					10					15	

Gln	Cys	Phe
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What is claimed:

1. A composition, comprising a fusion protein or immunocojugate, wherein the fusion protein or immunocojugate comprises a proximal tubule targeting moiety and a modulator of complement activity, wherein the modulator of complement activity comprises a complement inhibitor, wherein the complement inhibitor comprises decay accelerating factor (DAF) as set forth in SEQ ID NO. 2.

2. A composition, comprising a fusion protein or immunocojugate, wherein the fusion protein or immunocojugate comprises a proximal tubule targeting moiety and a modulator of complement activity, wherein the modulator of complement activity comprises a complement inhibitor, wherein the complement inhibitor comprises human CD59 as set forth in SEQ ID NO. 4, and wherein the targeting moiety is not Complement Receptor 2 (CR2).

3. A composition, comprising a fusion protein or immunocojugate, wherein the fusion protein or immunocojugate comprises a proximal tubule targeting moiety and a modula-

tor of complement activity, wherein the modulator of complement activity comprises a complement inhibitor, wherein the complement inhibitor comprises complement receptor 1 (CR1) as set forth in SEQ ID NO. 6.

4. A composition, comprising a fusion protein or immunocojugate, wherein the fusion protein or immunocojugate comprises a proximal tubule targeting moiety and a modulator of complement activity, wherein the modulator of complement activity comprises a complement inhibitor, wherein the complement inhibitor comprises membrane cofactor protein (MCP) as set forth in SEQ ID NO. 8.

5. A composition, comprising a fusion protein or immunocojugate, wherein the fusion protein or immunocojugate comprises a proximal tubule targeting moiety and a modulator of complement activity, wherein the modulator of complement activity comprises a complement inhibitor, wherein the complement inhibitor comprises complement receptor-related protein γ (Crry) as set forth in SEQ ID NO. 9.

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