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(54) **COMPOSITIONS AND METHODS RELATED TO PROTEIN A (SPA) VARIANTS**

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See application file for complete search history.

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(57) **ABSTRACT**

Disclosed are methods and compositions for treating or preventing a *Staphylococcus* bacterial infection using a non-toxigenic Protein A (SpA) variant.

14 Claims, 15 Drawing Sheets

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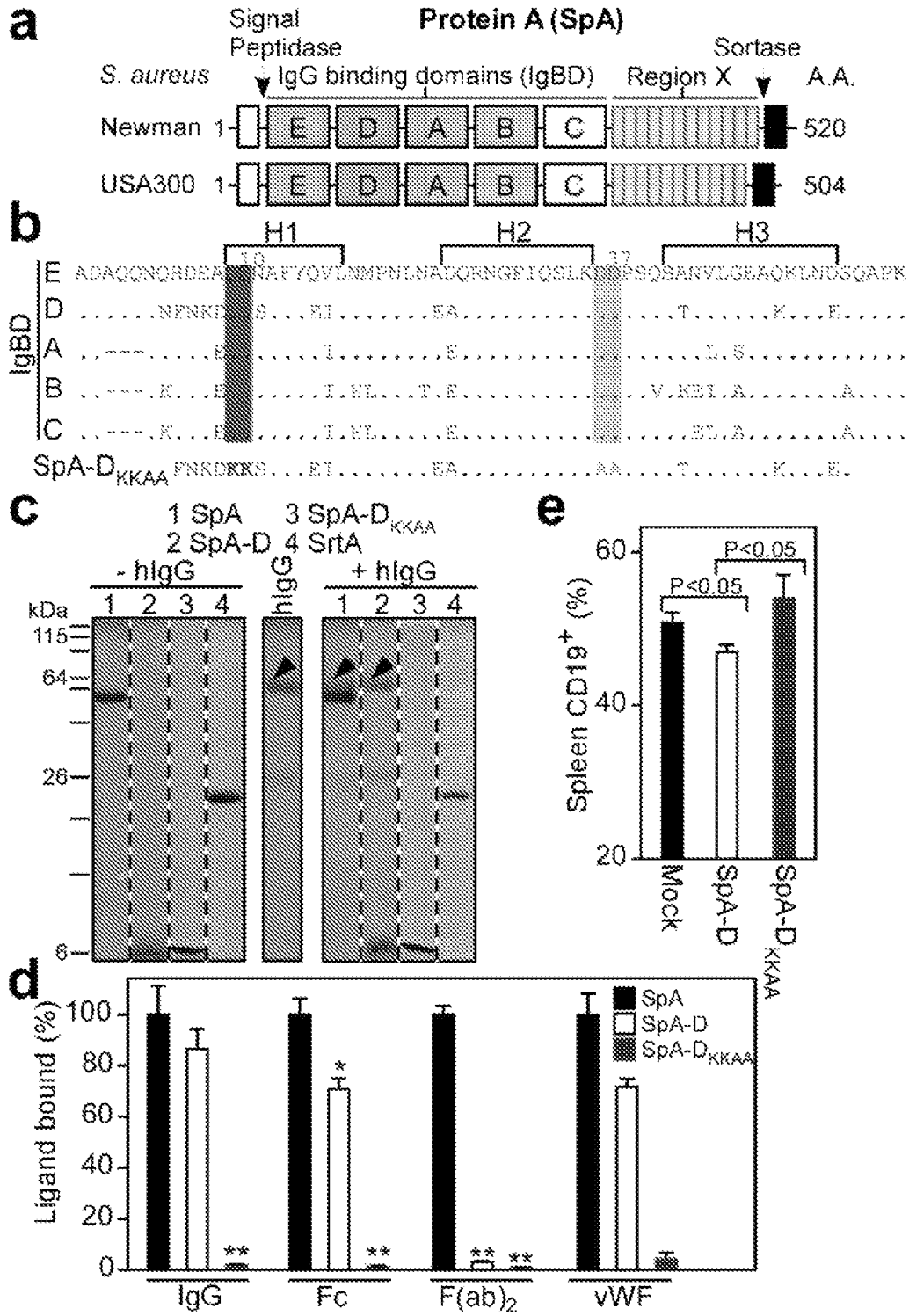


FIG. 1

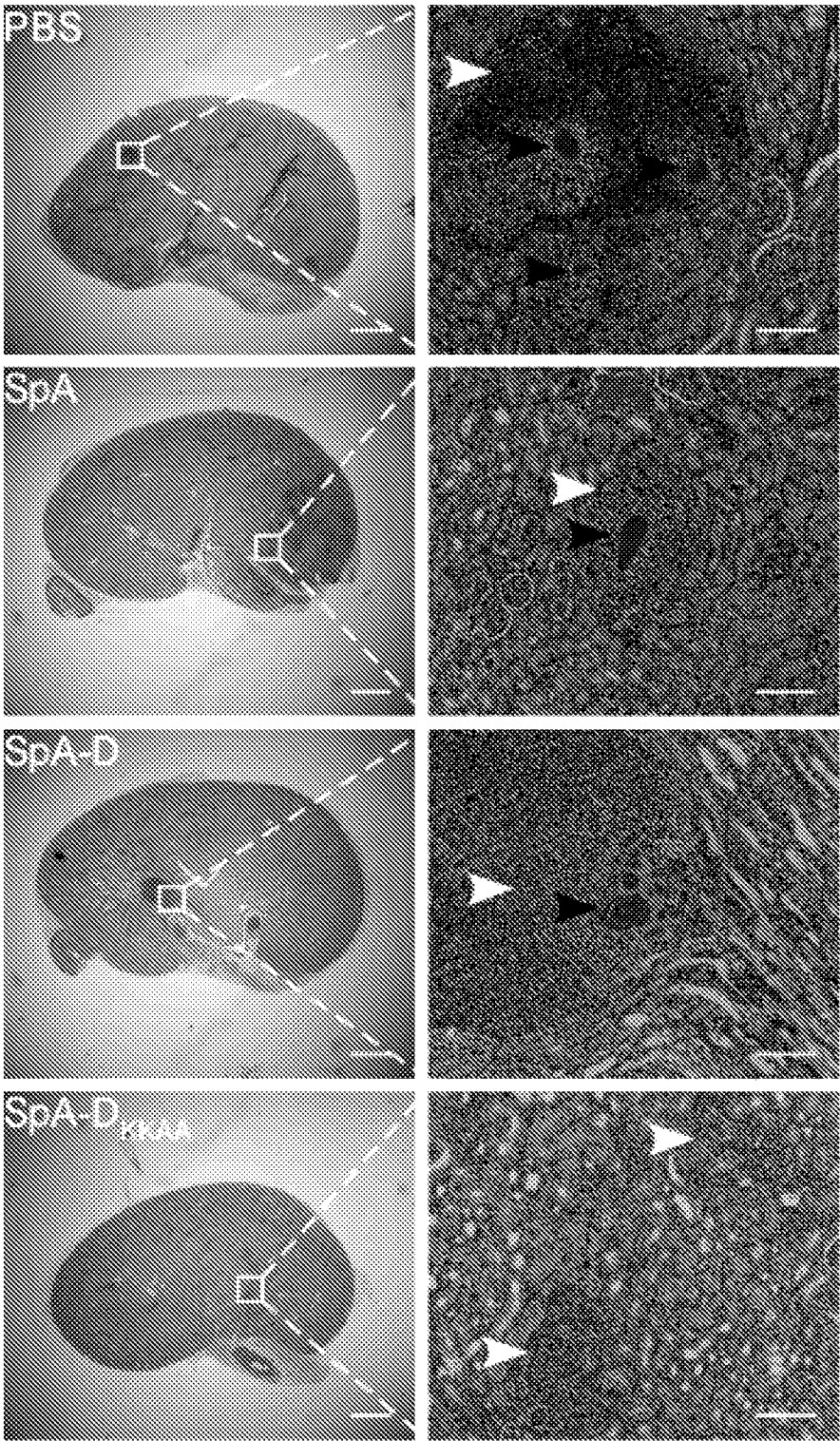


FIG. 2

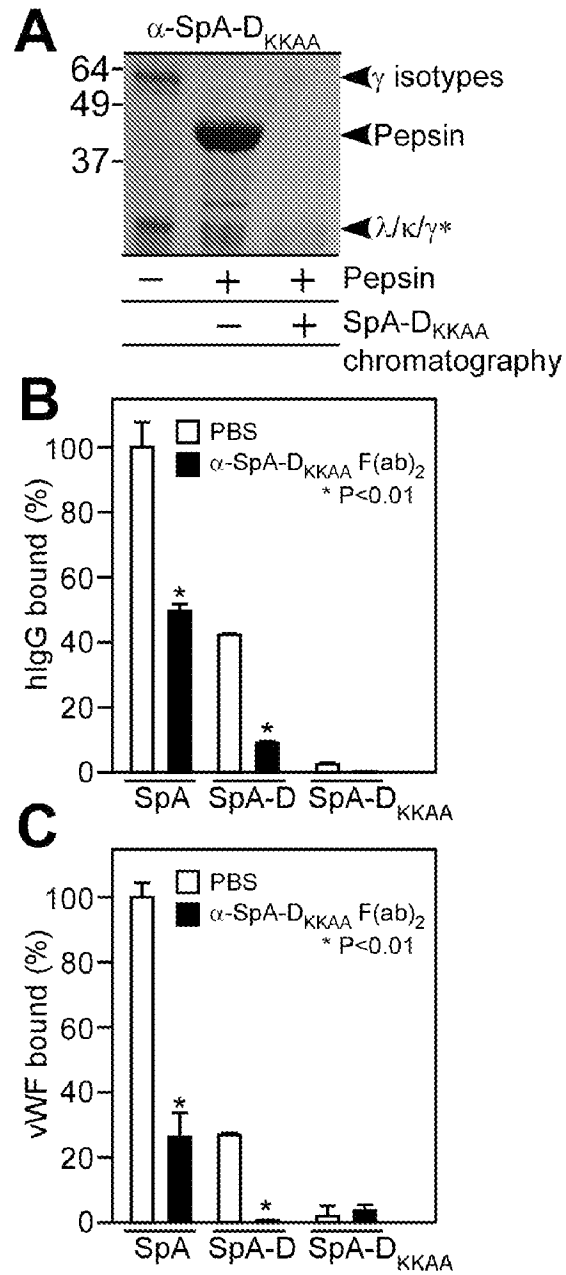


FIG. 3

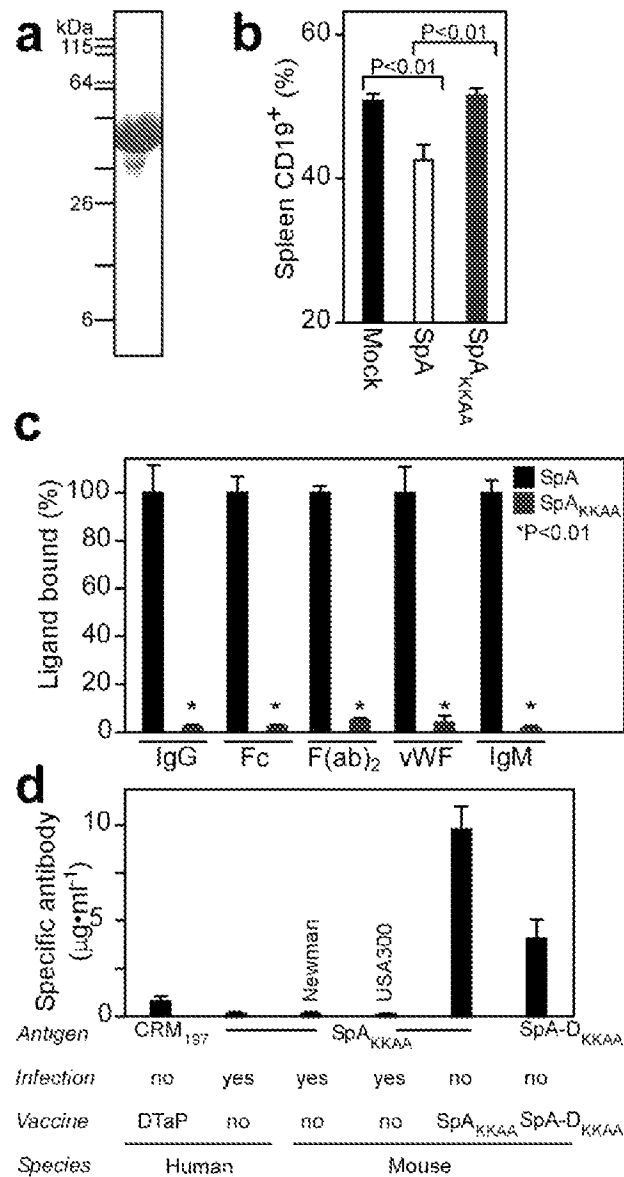


FIG. 4

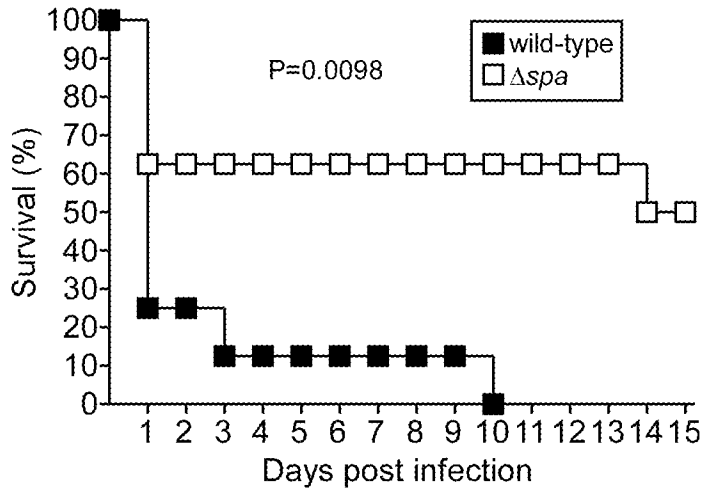


FIG. 5

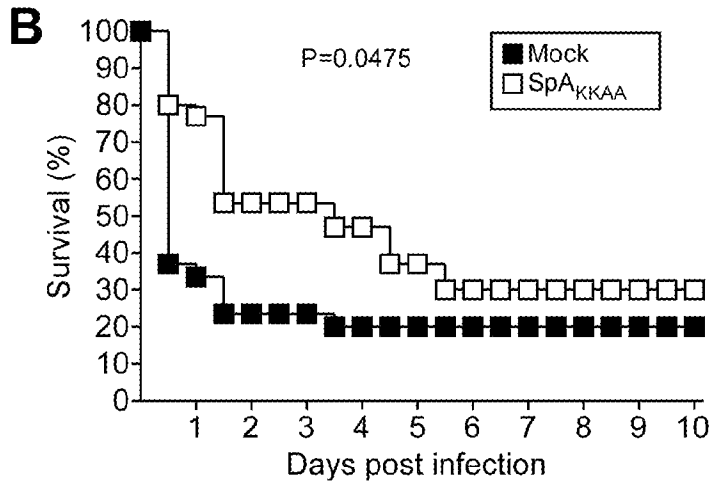
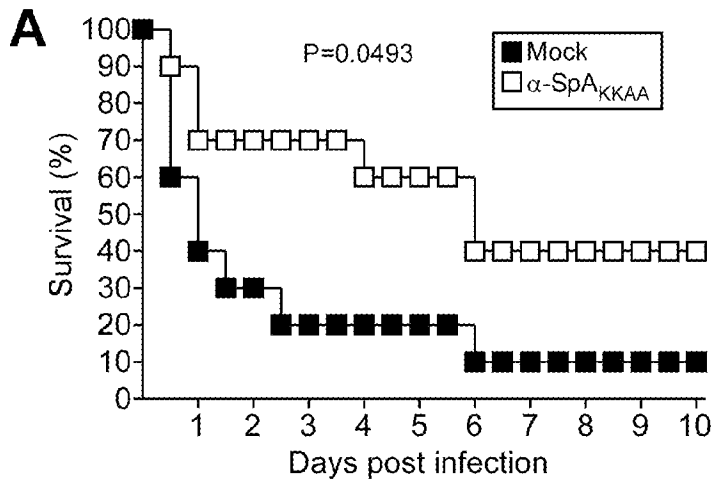


FIG. 6

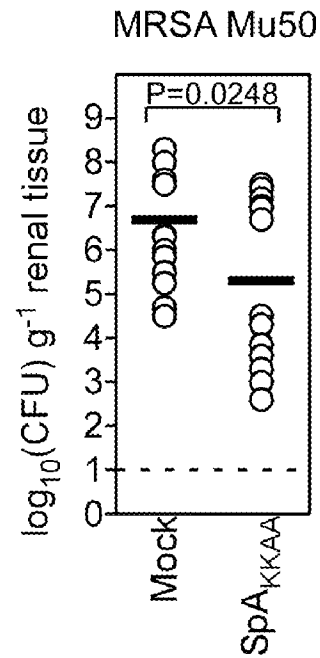


FIG. 7

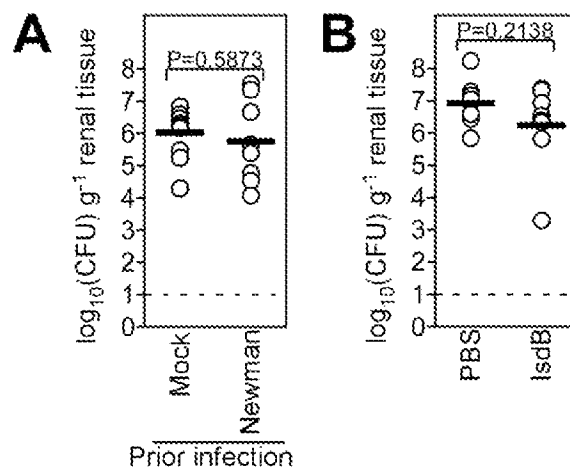


FIG. 8

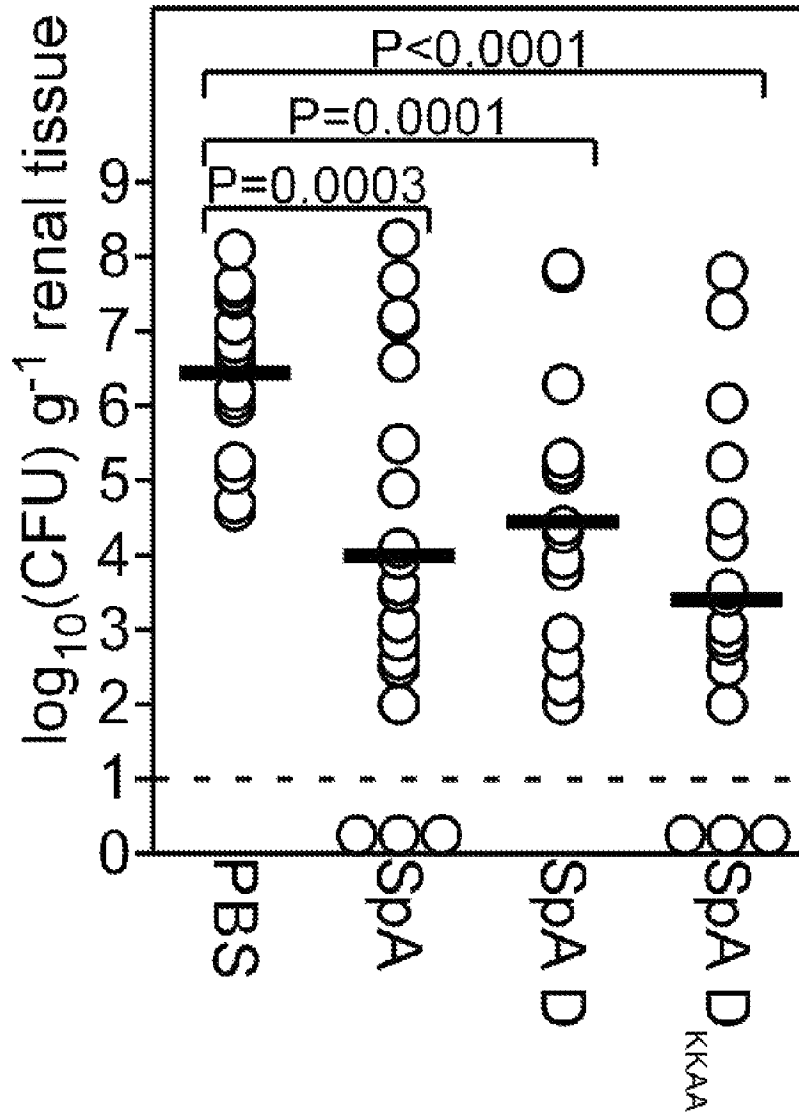


FIG. 9

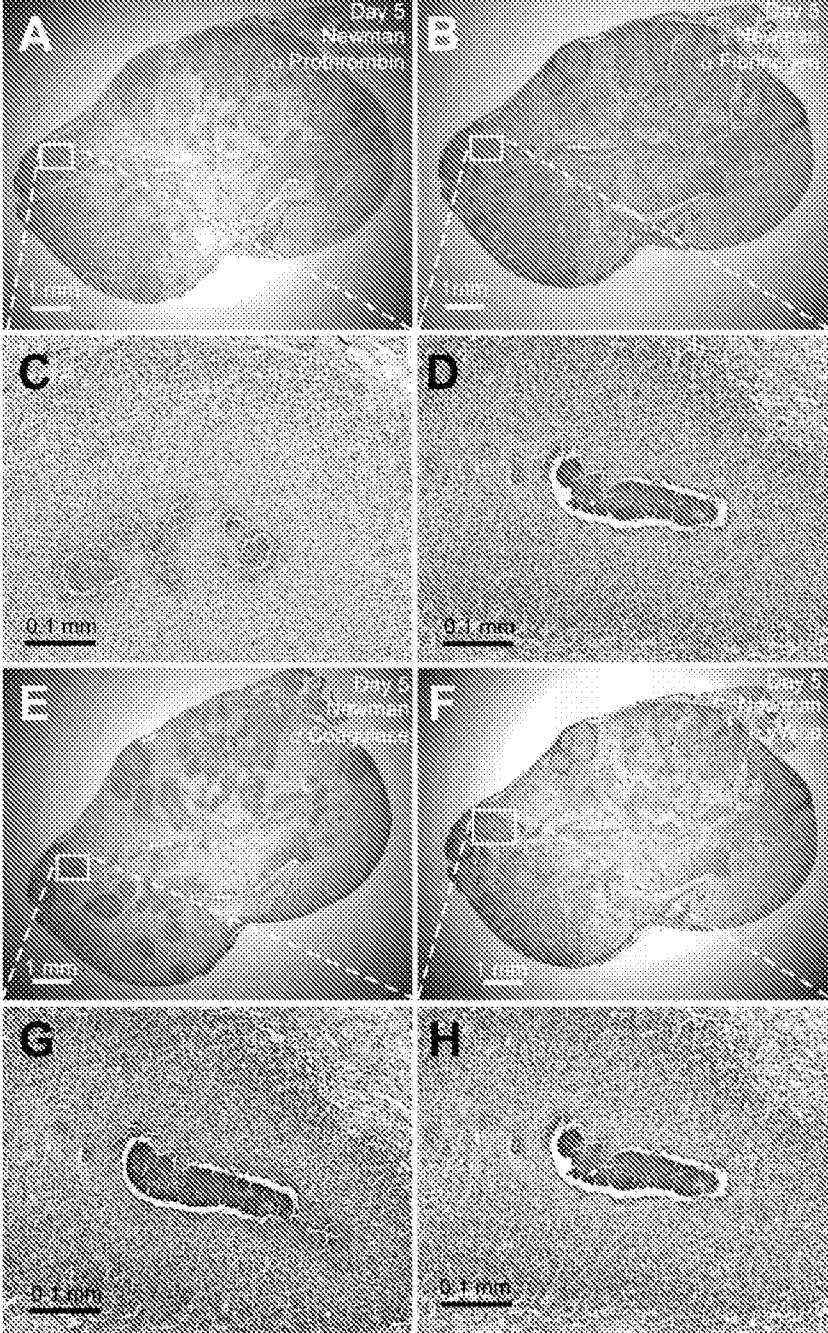


FIG. 10

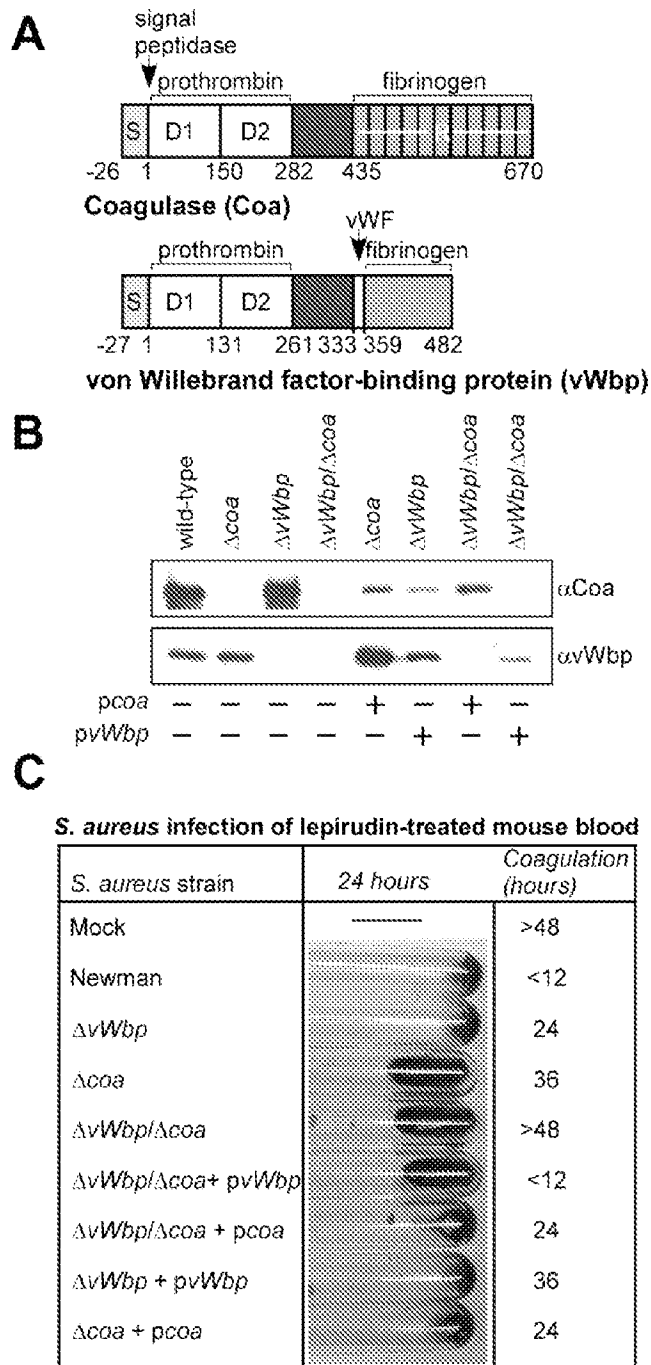


FIG. 11

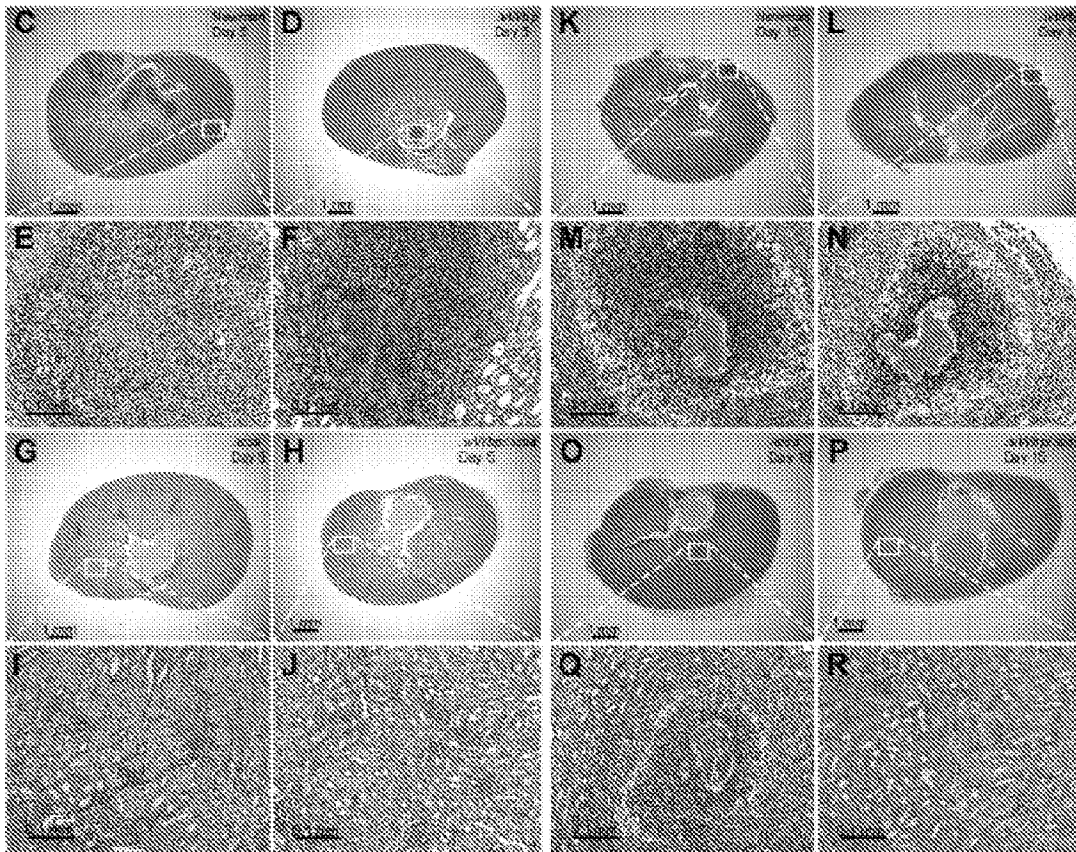
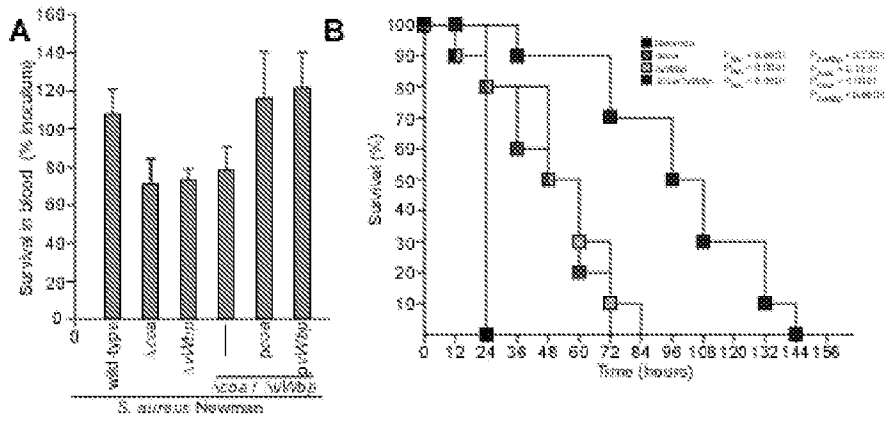


FIG. 12

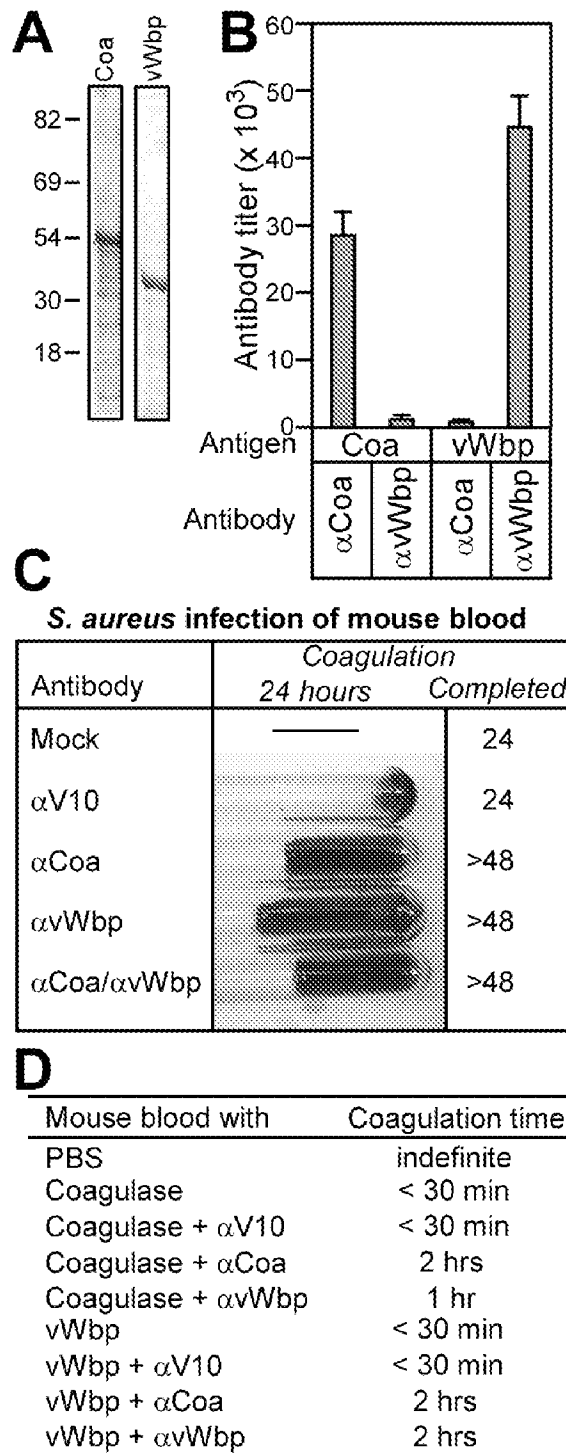


FIG. 13

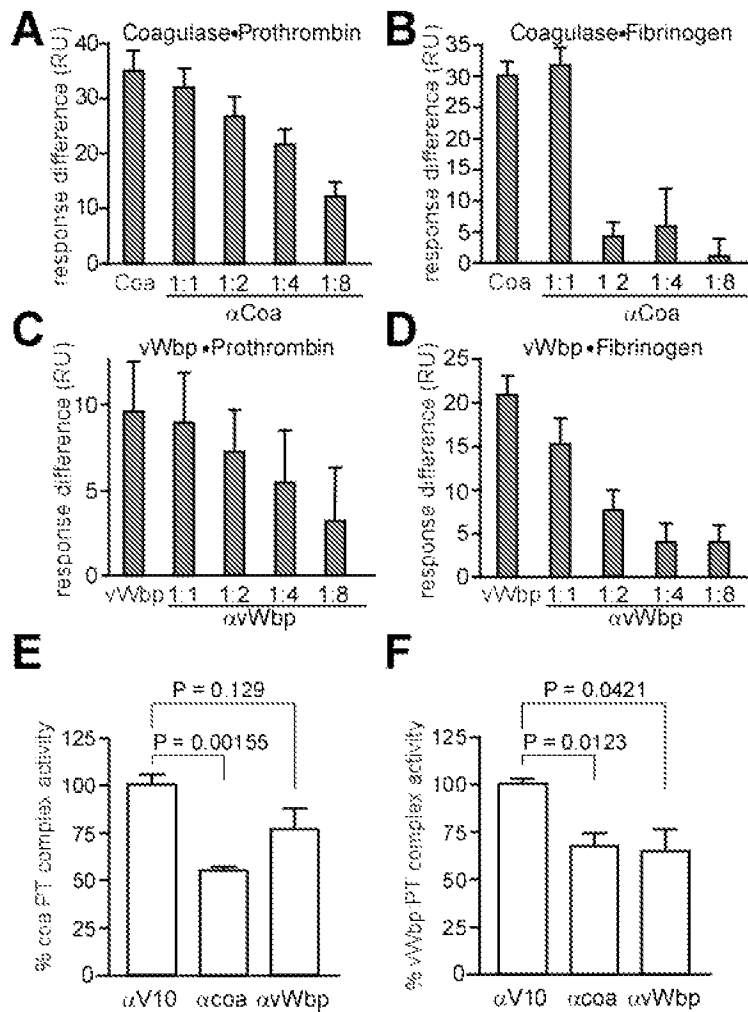


FIG. 14

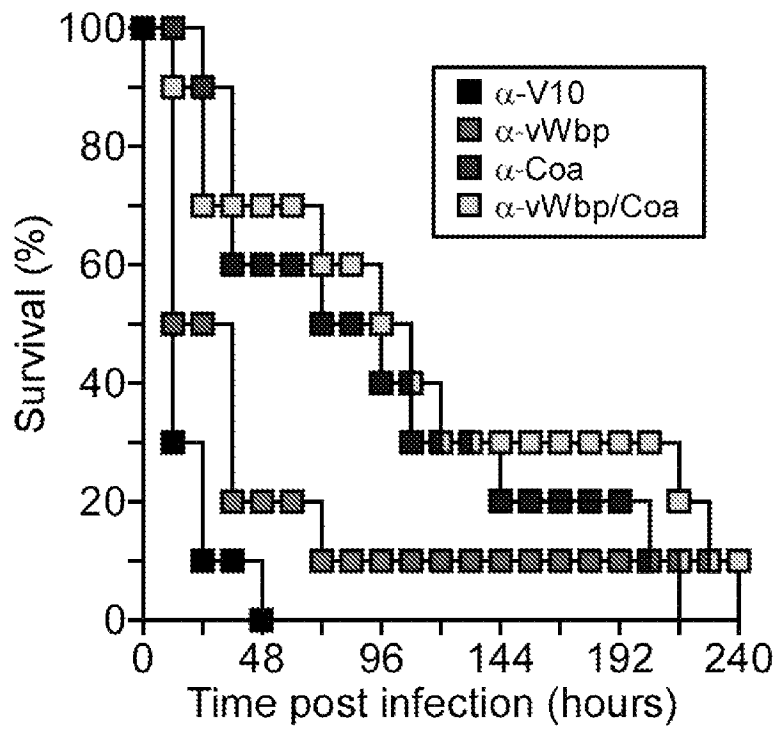


FIG. 15

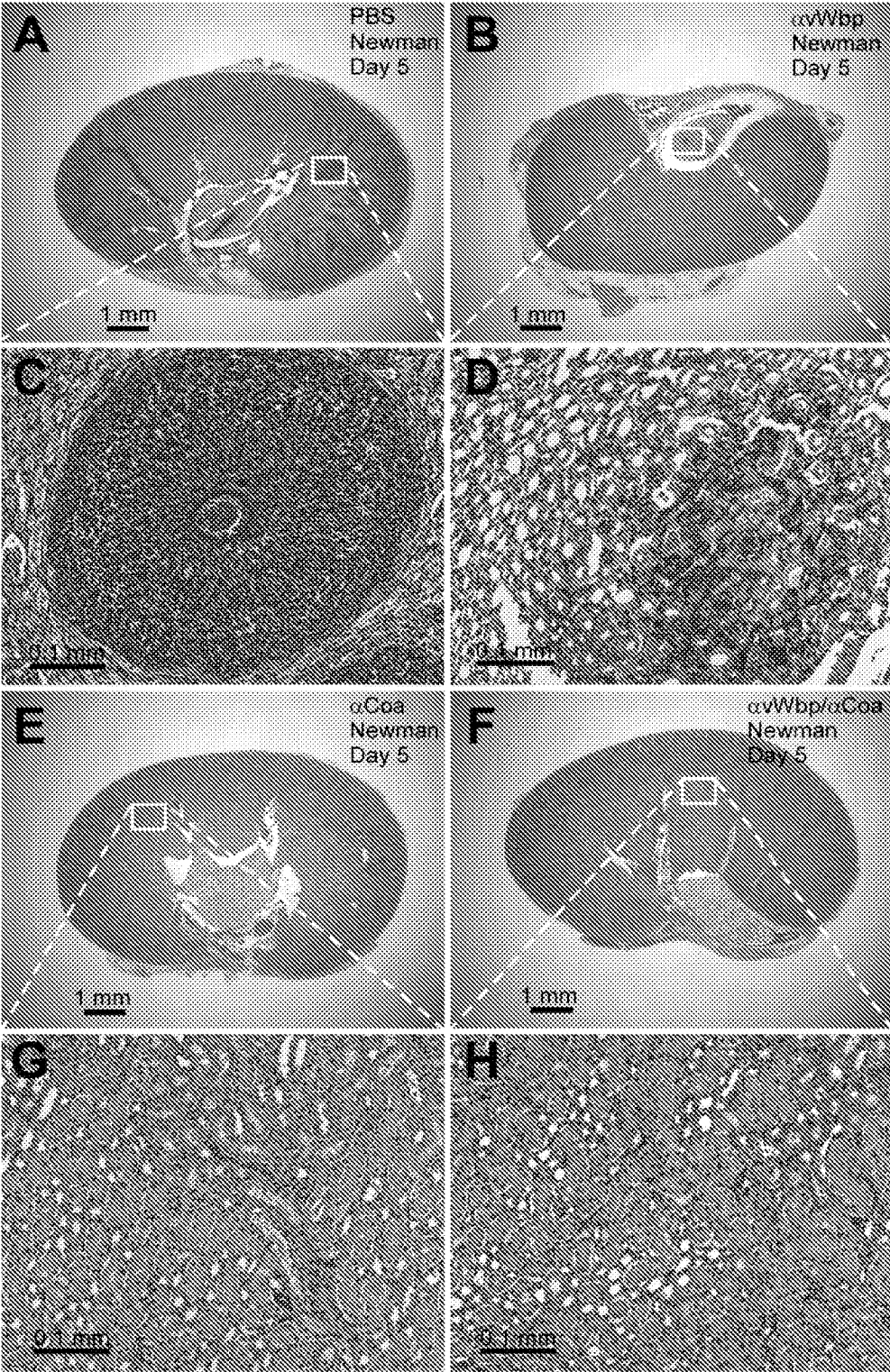


FIG. 16

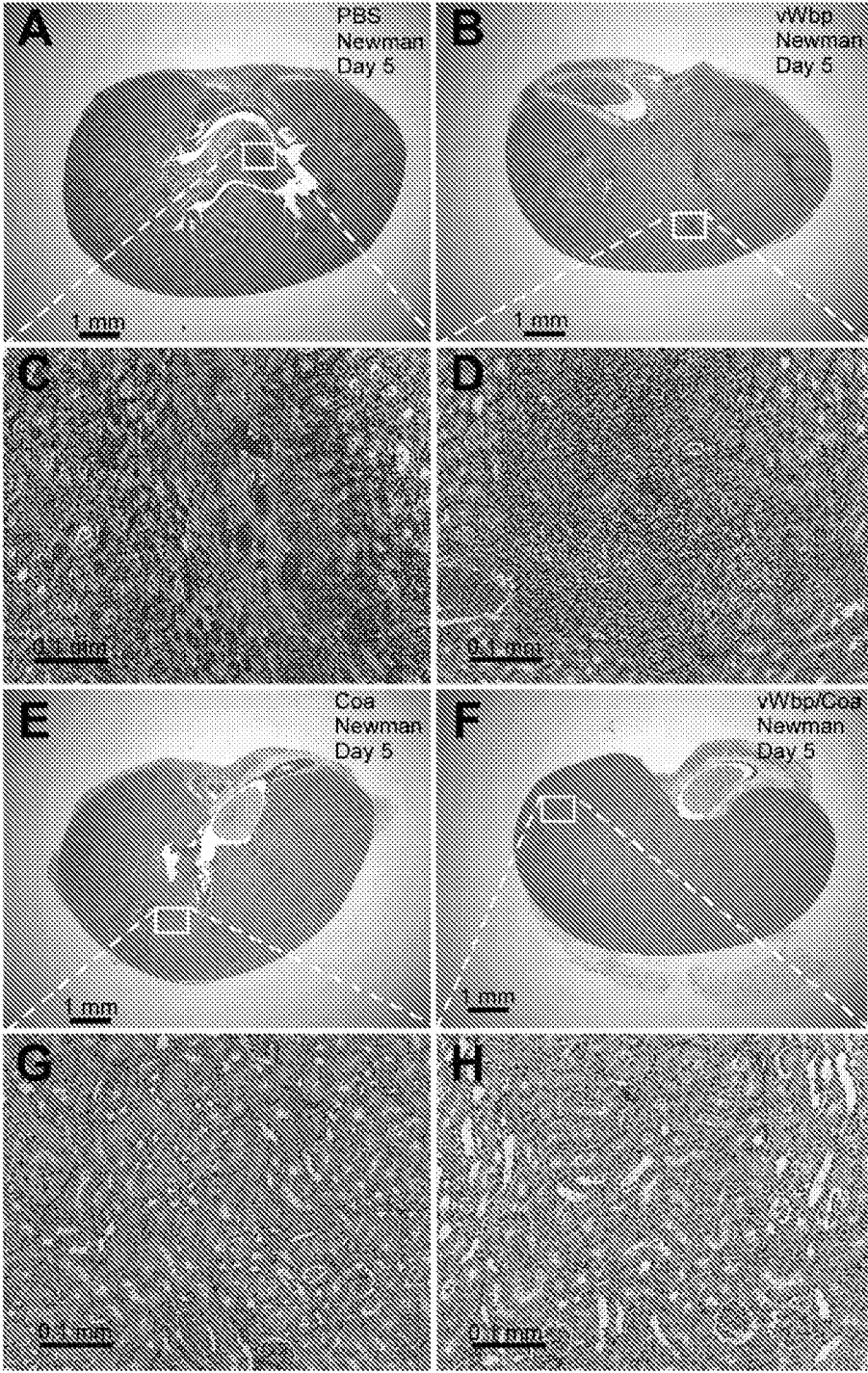


FIG. 17

COMPOSITIONS AND METHODS RELATED TO PROTEIN A (SPA) VARIANTS

The present application is a national phase application under 35 U.S.C. §371 of International Patent Application No. PCT/US2010/029959 filed Apr. 5, 2010, which claims benefit of priority to U.S. Provisional Application Ser. No. 61/166,432, filed Apr. 3, 2009, U.S. Provisional Application Ser. No. 61/237,956, filed Aug. 28, 2009, and U.S. Provisional Application Ser. No. 61/287,996, filed Dec. 18, 2009, the entire contents of each of which are hereby incorporated by reference in their entirety without disclaimer.

This invention was made with government support under AI057153, AI75258, AI052474, and GM007281 awarded by the National Institutes of Health. The government has certain rights in the invention.

BACKGROUND OF THE INVENTION

I. Field of the Invention

The present invention relates generally to the fields of immunology, microbiology, and pathology. More particularly, it concerns methods and compositions involving bacterial Protein A variants, which can be used to invoke an immune response against the bacteria.

II. Background

The number of both community acquired and hospital acquired infections have increased over recent years with the increased use of intravascular devices. Hospital acquired (nosocomial) infections are a major cause of morbidity and mortality, more particularly in the United States, where it affects more than 2 million patients annually. The most frequent infections are urinary tract infections (33% of the infections), followed by pneumonia (15.5%), surgical site infections (14.8%) and primary bloodstream infections (13%) (Emorl and Gaynes, 1993).

The major nosocomial pathogens include *Staphylococcus aureus*, coagulase-negative Staphylococci (mostly *Staphylococcus epidermidis*), *enterococcus* spp., *Escherichia coli* and *Pseudomonas aeruginosa*. Although these pathogens cause approximately the same number of infections, the severity of the disorders they can produce combined with the frequency of antibiotic resistant isolates balance this ranking towards *S. aureus* and *S. epidermidis* as being the most significant nosocomial pathogens.

Staphylococci can cause a wide variety of diseases in humans and other animals through either toxin production or invasion. Staphylococcal toxins are also a common cause of food poisoning, as the bacteria can grow in improperly-stored food.

Staphylococcus epidermidis is a normal skin commensal which is also an important opportunistic pathogen responsible for infections of impaired medical devices and infections at sites of surgery. Medical devices infected by *S. epidermidis* include cardiac pacemakers, cerebrospinal fluid shunts, continuous ambulatory peritoneal dialysis catheters, orthopedic devices and prosthetic heart valves.

Staphylococcus aureus is the most common cause of nosocomial infections with a significant morbidity and mortality. It is the cause of some cases of osteomyelitis, endocarditis, septic arthritis, pneumonia, abscesses, and toxic shock syndrome. *S. aureus* can survive on dry surfaces, increasing the chance of transmission. Any *S. aureus* infection can cause the staphylococcal scalded skin syndrome, a cutaneous reaction to exotoxin absorbed into the bloodstream. It can also cause a type of septicemia called pyaemia that can be life-threatening. Problematically, Methicillin-

resistant *Staphylococcus aureus* (MRSA) has become a major cause of hospital-acquired infections.

S. aureus and *S. epidermidis* infections are typically treated with antibiotics, with penicillin being the drug of choice, whereas vancomycin is used for methicillin resistant isolates. The percentage of staphylococcal strains exhibiting wide-spectrum resistance to antibiotics has become increasingly prevalent, posing a threat for effective antimicrobial therapy. In addition, the recent emergence of vancomycin resistant *S. aureus* strain has aroused fear that MRSA strains are emerging and spreading for which no effective therapy is available.

An alternative to antibiotic treatment for staphylococcal infections is under investigation that uses antibodies directed against staphylococcal antigens. This therapy involves administration of polyclonal antisera (WO00/15238, WO00/12132) or treatment with monoclonal antibodies against lipoteichoic acid (WO98/57994).

An alternative approach would be the use of active vaccination to generate an immune response against staphylococci. The *S. aureus* genome has been sequenced and many of the coding sequences have been identified (WO02/094868, EP0786519), which can lead to the identification of potential antigens. The same is true for *S. epidermidis* (WO01/34809). As a refinement of this approach, others have identified proteins that are recognized by hyperimmune sera from patients who have suffered staphylococcal infection (WO01/98499, WO02/059148).

S. aureus secretes a plethora of virulence factors into the extracellular milieu (Archer, 1998; Dinges et al., 2000; Foster, 2005; Shaw et al., 2004; Sibbald et al., 2006). Like most secreted proteins, these virulence factors are translocated by the Sec machinery across the plasma membrane. Proteins secreted by the Sec machinery bear an N-terminal leader peptide that is removed by leader peptidase once the pre-protein is engaged in the Sec translocon (Dalbey and Wickner, 1985; van Wely et al., 2001). Recent genome analysis suggests that Actinobacteria and members of the Firmicutes encode an additional secretion system that recognizes a subset of proteins in a Sec-independent manner (Pallen, 2002). ESAT-6 (early secreted antigen target 6 kDa) and CFP-10 (culture filtrate antigen 10 kDa) of *Mycobacterium tuberculosis* represent the first substrates of this novel secretion system termed ESX-1 or Snm in *M. tuberculosis* (Andersen et al., 1995; Hsu et al., 2003; Pym et al., 2003; Stanley et al., 2003). In *S. aureus*, two ESAT-6 like factors designated EsxA and EsxB are secreted by the Ess pathway (ESAT-6 secretion system) (Burts et al., 2005).

The first generation of vaccines targeted against *S. aureus* or against the exoproteins it produces have met with limited success (Lee, 1996). There remains a need to develop effective vaccines against staphylococcal infections. Additional compositions for treating staphylococcal infections are also needed.

SUMMARY OF THE INVENTION

Protein A (SpA)(SEQ ID NO:33), a cell wall anchored surface protein of *Staphylococcus aureus*, provides for bacterial evasion from innate and adaptive immune responses. Protein A binds immunoglobulins at their Fc portion, interacts with the VH3 domain of B cell receptors inappropriately stimulating B cell proliferation and apoptosis, binds to von Willebrand factor A1 domains to activate intracellular clotting, and also binds to the TNF Receptor-1 to contribute to the pathogenesis of staphylococcal pneumonia. Due to the fact that Protein A captures immunoglobulin and displays

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toxic attributes, the possibility that this surface molecule may function as a vaccine in humans has not been rigorously pursued. Here the inventors demonstrate that Protein A variants no longer able to bind to immunoglobulins, which are thereby removed of their toxigenic potential, i.e., are

non-toxicogenic, stimulate humoral immune responses that protect against staphylococcal disease.

In certain embodiments the SpA variant is a full length SpA variant comprising a variant A, B, C, D, and E domain. In certain aspects, the SpA variant comprises or consists of the amino acid sequence that is 80, 90, 95, 98, 99, or 100% identical to the amino acid sequence of SEQ ID NO:34. In other embodiments the SpA variant comprises a segment of SpA. The SpA segment can comprise at least or at most 1, 2, 3, 4, 5 or more IgG binding domains. The IgG domains can be at least or at most 1, 2, 3, 4, 5 or more variant A, B, C, D, or E domains. In certain aspects the SpA variant comprises at least or at most 1, 2, 3, 4, 5, or more variant A domains. In a further aspect the SpA variant comprises at least or at most 1, 2, 3, 4, 5, or more variant B domains. In still a further aspect the SpA variant comprises at least or at most 1, 2, 3, 4, 5, or more variant C domains. In yet a further aspect the SpA variant comprises at least or at most 1, 2, 3, 4, 5, or more variant D domains. In certain aspects the SpA variant comprises at least or at most 1, 2, 3, 4, 5, or more variant E domains. In a further aspect the SpA variant comprises a combination of A, B, C, D, and E domains in various combinations and permutations. The combinations can include all or part of a SpA signal peptide segment, a SpA region X segment, and/or a SpA sorting signal segment. In other aspects the SpA variant does not include a SpA signal peptide segment, a SpA region X segment, and/or a SpA sorting signal segment. In certain aspects a variant A domain comprises a substitution at position(s) 7, 8, 34, and/or 35 of SEQ ID NO:4. In another aspect a variant B domain comprises a substitution at position(s) 7, 8, 34, and/or 35 of SEQ ID NO:6. In still another aspect a variant C domain comprises a substitution at position(s) 7, 8, 34, and/or 35 of SEQ ID NO:5. In certain aspects a variant D domain comprises a substitution at position(s) 9, 10, 37, and/or 38 of SEQ ID NO:2. In a further aspect a variant E domain comprises a substitution at position(s) 6, 7, 33, and/or 34 of SEQ ID NO:3.

In certain aspects the SpA variant includes a substitution of (a) one or more amino acid substitution in an IgG Fc binding sub-domain of SpA domain A, B, C, D, and/or E that disrupts or decreases binding to IgG Fc, and (b) one or more amino acid substitution in a V_H3 binding sub-domain of SpA domain A, B, C, D, and/or E that disrupts or decreases binding to V_H3 . In still further aspects the amino acid sequence of a SpA variant comprises an amino acid sequence that is at least 50%, 60%, 70%, 80%, 90%, 95%, or 100% identical, including all values and ranges there between, to the amino acid sequence of SEQ ID NOs:2-6.

In a further aspect the SpA variant includes (a) one or more amino acid substitution in an IgG Fc binding sub-domain of SpA domain D, or at a corresponding amino acid position in other IgG domains, that disrupts or decreases binding to IgG Fc, and (b) one or more amino acid substitution in a V_H3 binding sub-domain of SpA domain D, or at a corresponding amino acid position in other IgG domains, that disrupts or decreases binding to V_H3 . In certain aspects amino acid residue F5, Q9, Q10, S11, F13, Y14, L17, N28, I31, and/or K35 (SEQ ID NO:2, QQNNFNKDDQQA-FYEILNMPNLNEAQRNGFIQSLKDDPSQSTNV-LGEAKKLNES) of the IgG Fc binding sub-domain of domain D are modified or substituted. In certain aspects

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amino acid residue Q26, G29, F30, S33, D36, D37, Q40, N43, and/or E47 (SEQ ID NO:2) of the V_H3 binding sub-domain of domain D are modified or substituted such that binding to Fc or V_H3 is attenuated. In further aspects corresponding modifications or substitutions can be engineered in corresponding positions of the domain A, B, C, and/or E. Corresponding positions are defined by alignment of the domain D amino acid sequence with one or more of the amino acid sequences from other IgG binding domains of SpA, for example see FIG. 1. In certain aspects the amino acid substitution can be any of the other 20 amino acids. In a further aspect conservative amino acid substitutions can be specifically excluded from possible amino acid substitutions. In other aspects only non-conservative substitutions are included. In any event, any substitution or combination of substitutions that reduces the binding of the domain such that SpA toxicity is significantly reduced is contemplated. The significance of the reduction in binding refers to a variant that produces minimal to no toxicity when introduced into a subject and can be assessed using in vitro methods described herein.

In certain embodiments, a variant SpA comprises at least or at most 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, or more variant SpA domain D peptides. In certain aspects 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, or 19 or more amino acid residues of the variant SpA are substituted or modified—including but not limited to amino acids F5, Q9, Q10, S11, F13, Y14, L17, N28, I31, and/or K35 (SEQ ID NO:2) of the IgG Fc binding sub-domain of domain D and amino acid residue Q26, G29, F30, S33, D36, D37, Q40, N43, and/or E47 (SEQ ID NO:2) of the V_H3 binding sub-domain of domain D. In one aspect of the invention glutamine residues at position 9 and/or 10 of SEQ ID NO:2 (or corresponding positions in other domains) are mutated. In another aspect, aspartic acid residues 36 and/or 37 of SEQ ID NO:2 (or corresponding positions in other domains) are mutated. In a further aspect, glutamine 9 and 10, and aspartic acid residues 36 and 37 are mutated. Purified non-toxicogenic SpA or SpA-D mutants/variants described herein are no longer able to significantly bind (i.e., demonstrate attenuated or disrupted binding affinity) Fcγ or F(ab)₂ V_H3 and also do not stimulate B cell apoptosis. These non-toxicogenic Protein A variants can be used as subunit vaccines and raise humoral immune responses and confer protective immunity against *S. aureus* challenge. Compared to wild-type full-length Protein A or the wild-type SpA-domain D, immunization with SpA-D variants resulted in an increase in Protein A specific antibody. Using a mouse model of staphylococcal challenge and abscess formation, it was observed that immunization with the non-toxicogenic Protein A variants generated significant protection from staphylococcal infection and abscess formation. As virtually all *S. aureus* strains express Protein A, immunization of humans with the non-toxicogenic Protein A variants can neutralize this virulence factor and thereby establish protective immunity. In certain aspects the protective immunity protects or ameliorates infection by drug resistant strains of *Staphylococcus*, such as USA300 and other MRSA strains.

Embodiments include the use of Protein A variants in methods and compositions for the treatment of bacterial and/or staphylococcal infection. This application also provides an immunogenic composition comprising a Protein A variant or immunogenic fragment thereof. In certain aspects, the immunogenic fragment is a Protein A domain D segment. Furthermore, the present invention provides methods and compositions that can be used to treat (e.g., limiting staphylococcal abscess formation and/or persistence in a

subject) or prevent bacterial infection. In some cases, methods for stimulating an immune response involve administering to the subject an effective amount of a composition including or encoding all or part of a Protein A variant polypeptide or antigen, and in certain aspects other bacterial proteins. Other bacterial proteins include, but are not limited to (i) a secreted virulence factor, and/or a cell surface protein or peptide, or (ii) a recombinant nucleic acid molecule encoding a secreted virulence factor, and/or a cell surface protein or peptide.

In other aspects, the subject can be administered all or part of a Protein A variant, such as a variant Protein A domain D segment. The polypeptide of the invention can be formulated in a pharmaceutically acceptable composition. The composition can further comprise one or more of at least or at most 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, or 19 additional staphylococcal antigen or immunogenic fragment thereof (e.g., Eap, Ebh, Emp, EsaB, EsaC, EsxA, EsxB, SdrC, SdrD, SdrE, IsdA, IsdB, ClfA, CHB, Coa, Hla (e.g., H35 mutants), IsdC, SasF, vWbp, or vWh). Additional staphylococcal antigens that can be used in combination with a Protein A variant include, but are not limited to 52 kDa vitronectin binding protein (WO 01/60852), Aaa (GenBank CAC80837), Aap (GenBank accession AJ249487), Ant (GenBank accession NP_372518), autolysin glucosaminidase, autolysin amidase, Cna, collagen binding protein (U.S. Pat. No. 6,288,214), EFB (FIB), Elastin binding protein (EbpS), EPB, FbpA, fibrinogen binding protein (U.S. Pat. No. 6,008,341), Fibronectin binding protein (U.S. Pat. No. 5,840,846), FnbA, FnbB, GehD (US 2002/0169288), HarA, HBP, Immunodominant ABC transporter, IsaA/PisA, laminin receptor, Lipase GehD, MAP, Mg²⁺ transporter, MHC II analogue (U.S. Pat. No. 5,648,240), MRPII, Npase, RNA III activating protein (RAP), SasA, SasB, SasC, SasD, SasK, SBI, SdrF (WO 00/12689), SdrG/ Fig (WO 00/12689), SdrH (WO 00/12689), SEA exotoxins (WO 00/02523), SEB exotoxins (WO 00/02523), SitC and Ni ABC transporter, SitC/MntC/saliva binding protein (U.S. Pat. No. 5,801,234), SsaA, SSP-1, SSP-2, and/or Vitronectin binding protein (see PCT publications WO2007/113222, WO2007/113223, WO2006/032472, WO2006/032475, WO2006/032500, each of which is incorporated herein by reference in their entirety). The staphylococcal antigen or immunogenic fragment can be administered concurrently with the Protein A variant. The staphylococcal antigen or immunogenic fragment and the Protein A variant can be administered in the same composition. The Protein A variant can also be a recombinant nucleic acid molecule encoding a Protein A variant. A recombinant nucleic acid molecule can encode the Protein A variant and at least one staphylococcal antigen or immunogenic fragment thereof. As used herein, the term "modulate" or "modulation" encompasses the meanings of the words "enhance," or "inhibit." "Modulation" of activity may be either an increase or a decrease in activity. As used herein, the term "modulator" refers to compounds that effect the function of a moiety, including up-regulation, induction, stimulation, potentiation, inhibition, down-regulation, or suppression of a protein, nucleic acid, gene, organism or the like.

In certain embodiments the methods and compositions use or include or encode all or part of the Protein A variant or antigen. In other aspects, the Protein A variant may be used in combination with secreted factors or surface antigens including, but not limited to one or more of an isolated Eap, Ebh, Emp, EsaB, EsaC, EsxA, EsxB, SdrC, SdrD, SdrE, IsdA, IsdB, ClfA, ClfB, Coa, Hla, IsdC, SasF, vWbp, or vWh polypeptide or immunogenic segment thereof. Addi-

tional staphylococcal antigens that can be used in combination with a Protein A variant include, but are not limited to 52 kDa vitronectin binding protein (WO 01/60852), Aaa, Aap, Ant, autolysin glucosaminidase, autolysin amidase, Cna, collagen binding protein (U.S. Pat. No. 6,288,214), EFB (FIB), Elastin binding protein (EbpS), EPB, FbpA, fibrinogen binding protein (U.S. Pat. No. 6,008,341), Fibronectin binding protein (U.S. Pat. No. 5,840,846), FnbA, FnbB, GehD (US 2002/0169288), HarA, HBP, Immunodominant ABC transporter, IsaA/PisA, laminin receptor, Lipase GehD, MAP, Mg²⁺ transporter, MEW II analogue (U.S. Pat. No. 5,648,240), MRPII, Npase, RNA III activating protein (RAP), SasA, SasB, SasC, SasD, SasK, SBI, SdrF (WO 00/12689), SdrG/ Fig (WO 00/12689), SdrH (WO 00/12689), SEA exotoxins (WO 00/02523), SEB exotoxins (WO 00/02523), SitC and Ni ABC transporter, SitC/MntC/saliva binding protein (U.S. Pat. No. 5,801,234), SsaA, SSP-1, SSP-2, and/or Vitronectin binding protein. In certain embodiments, 1, 2, 3, 4, 5, 6, 7, 8, 9, 10 or more of Eap, Ebh, Emp, EsaB, EsaC, EsxA, EsxB, SdrC, SdrD, SdrE, IsdA, IsdB, ClfA, CHB, Coa, Hla, IsdC, SasF, vWbp, vWh, 52 kDa vitronectin binding protein (WO 01/60852), Aaa, Aap, Ant, autolysin glucosaminidase, autolysin amidase, Cna, collagen binding protein (U.S. Pat. No. 6,288,214), EFB (FIB), Elastin binding protein (EbpS), EPB, FbpA, fibrinogen binding protein (U.S. Pat. No. 6,008,341), Fibronectin binding protein (U.S. Pat. No. 5,840,846), FnbA, FnbB, GehD (US 2002/0169288), HarA, HBP, Immunodominant ABC transporter, IsaA/PisA, laminin receptor, Lipase GehD, MAP, Mg²⁺ transporter, MHC II analogue (U.S. Pat. No. 5,648,240), MRPII, Npase, RNA III activating protein (RAP), SasA, SasB, SasC, SasD, SasK, SBI, SdrF (WO 00/12689), SdrG/ Fig (WO 00/12689), SdrH (WO 00/12689), SEA exotoxins (WO 00/02523), SEB exotoxins (WO 00/02523), SitC and Ni ABC transporter, SitC/MntC/saliva binding protein (U.S. Pat. No. 5,801,234), SsaA, SSP-1, SSP-2, and/or Vitronectin binding protein. can be specifically excluded from a formulation of the invention.

In still further aspects, the isolated Protein A variant is multimerized, e.g., dimerized or a linear fusion of two or more polypeptides or peptide segments. In certain aspects of the invention, a composition comprises multimers or concatamers of 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20 or more isolated cell surface proteins or segments thereof. Concatamers are linear polypeptides having one or more repeating peptide units. SpA polypeptides or fragments can be consecutive or separated by a spacer or other peptide sequences, e.g., one or more additional bacterial peptide. In a further aspect, the other polypeptides or peptides contained in the multimer or concatamer can include, but are not limited to 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19 of Eap, Ebh, Emp, EsaB, EsaC, EsxA, EsxB, SdrC, SdrD, SdrE, IsdA, IsdB, ClfA, CHB, Coa, Hla, IsdC, SasF, vWbp, vWh or immunogenic fragments thereof. Additional staphylococcal antigens that can be used in combination with a Protein A variant include, but are not limited to 52 kDa vitronectin binding protein (WO 01/60852), Aaa, Aap, Ant, autolysin glucosaminidase, autolysin amidase, Cna, collagen binding protein (U.S. Pat. No. 6,288,214), EFB (FIB), Elastin binding protein (EbpS), EPB, FbpA, fibrinogen binding protein (U.S. Pat. No. 6,008,341), Fibronectin binding protein (U.S. Pat. No. 5,840,846), FnbA, FnbB, GehD (US 2002/0169288), HarA, HBP, Immunodominant ABC transporter, IsaA/PisA, laminin receptor, Lipase GehD, MAP, Mg²⁺ transporter, MHC II analogue (U.S. Pat. No. 5,648,240), MRPII, Npase, RNA III activating protein (RAP), SasA, SasB, SasC, SasD, SasK,

SBI, SdrF (WO 00/12689), SdrG/Fig (WO 00/12689), SdrH (WO 00/12689), SEA exotoxins (WO 00/02523), SEB exotoxins (WO 00/02523), SitC and Ni ABC transporter, SitC/MntC/saliva binding protein (U.S. Pat. No. 5,801,234), SsaA, SSP-1, SSP-2, and/or Vitronectin binding protein.

The term "Protein A variant" or "SpA variant" refers to polypeptides that include a SpA IgG domain having two or more amino acid substitutions that disrupt binding to Fc and V_H3 . In certain aspect, a SpA variant includes a variant domain D peptide, as well as variants of SpA polypeptides and segments thereof that are non-toxicogenic and stimulate an immune response against *staphylococcus* bacteria Protein A and/or bacteria expressing such.

Embodiments of the present invention include methods for eliciting an immune response against a *staphylococcus* bacterium or staphylococci in a subject comprising providing to the subject an effective amount of a Protein A variant or a segment thereof. In certain aspects, the methods for eliciting an immune response against a *staphylococcus* bacterium or staphylococci in a subject comprising providing to the subject an effective amount of 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19 or more secreted proteins and/or cell surface proteins or segments/fragments thereof. A secreted protein or cell surface protein includes, but is not limited to Eap, Ebh, Emp, EsaB, EsaC, EsxA, EsxB, SdrC, SdrD, SdrE, IsdA, IsdB, ClfA, ClfB, Coa, Hla, IsdC, SasF, vWbp, and/or vWh proteins and immunogenic fragments thereof. Additional staphylococcal antigens that can be used in combination with a Protein A variant include, but are not limited to 52 kDa vitronectin binding protein (WO 01/60852), Aaa, Aap, Ant, autolysin glucosaminidase, autolysin amidase, Cna, collagen binding protein (U.S. Pat. No. 6,288,214), EFB (FIB), Elastin binding protein (EbpS), EPB, FbpA, fibrinogen binding protein (U.S. Pat. No. 6,008,341), Fibronectin binding protein (U.S. Pat. No. 5,840,846), FnbA, FnbB, GehD (US 2002/0169288), HarA, HBP, Immunodominant ABC transporter, IsaA/PisA, laminin receptor, Lipase GehD, MAP, Mg2+ transporter, MHC II analogue (U.S. Pat. No. 5,648,240), MRPII, Npase, RNA III activating protein (RAP), SasA, SasB, SasC, SasD, SasK, SBI, SdrF (WO 00/12689), SdrG/Fig (WO 00/12689), SdrH (WO 00/12689), SEA exotoxins (WO 00/02523), SEB exotoxins (WO 00/02523), SitC and Ni ABC transporter, SitC/MntC/saliva binding protein (U.S. Pat. No. 5,801,234), SsaA, SSP-1, SSP-2, and/or Vitronectin binding protein.

Embodiments of the invention include compositions that include a polypeptide, peptide, or protein that is or is at least 70%, 75%, 80%, 85%, 90%, 95%, 96%, 97%, 98%, or 99% identical or similar to Protein A, or a second protein or peptide that is a secreted bacterial protein or a bacterial cell surface protein. In a further embodiment of the invention a composition may include a polypeptide, peptide, or protein that is or is at least 70%, 75%, 80%, 85%, 90%, 95%, 96%, 97%, 98%, or 99% identical or similar to a Protein A domain D polypeptide (SEQ ID NO:2), domain E (SEQ ID NO:3), domain A (SEQ ID NO:4), domain C (SEQ ID NO:5), domain B (SEQ ID NO:6), or a nucleic acid sequence encoding a Protein A domain D, domain E, domain A, domain C, or domain B polypeptide. In certain aspects a Protein A polypeptide segment will have an amino acid sequence of SEQ ID NO:8. Similarity or identity, with identity being preferred, is known in the art and a number of different programs can be used to identify whether a protein (or nucleic acid) has sequence identity or similarity to a known sequence. Sequence identity and/or similarity is determined using standard techniques known in the art, including, but not limited to, the local sequence identity

algorithm of Smith & Waterman (1981), by the sequence identity alignment algorithm of Needleman & Wunsch (1970), by the search for similarity method of Pearson & Lipman (1988), by computerized implementations of these algorithms (GAP, BESTFIT, FASTA, and TFASTA in the Wisconsin Genetics Software Package, Genetics Computer Group, 575 Science Drive, Madison, Wis.), the Best Fit sequence program described by Devereux et al. (1984), preferably using the default settings, or by inspection. Preferably, percent identity is calculated by using alignment tools known to and readily ascertainable to those of skill in the art. Percent identity is essentially the number of identical amino acids divided by the total number of amino acids compared times one hundred.

Still further embodiments include methods for stimulating in a subject a protective or therapeutic immune response against a *staphylococcus* bacterium comprising administering to the subject an effective amount of a composition including (i) a SpA variant, e.g., a variant SpA domain D polypeptide or peptide thereof; or, (ii) a nucleic acid molecule encoding such a SpA variant polypeptide or peptide thereof, or (iii) administering a SpA variant domain D polypeptide with any combination or permutation of bacterial proteins described herein. In a preferred embodiment the composition is not a *staphylococcus* bacterium. In certain aspects the subject is a human or a cow. In a further aspect the composition is formulated in a pharmaceutically acceptable formulation. The staphylococci may be *Staphylococcus aureus*.

Yet still further embodiments include vaccines comprising a pharmaceutically acceptable composition having an isolated SpA variant polypeptide, or any other combination or permutation of protein(s) or peptide(s) described herein, wherein the composition is capable of stimulating an immune response against a *staphylococcus* bacterium. The vaccine may comprise an isolated SpA variant polypeptide, or any other combination or permutation of protein(s) or peptide(s) described. In certain aspects of the invention the isolated SpA variant polypeptide, or any other combination or permutation of protein(s) or peptide(s) described are multimerized, e.g., dimerized or concatamerized. In a further aspect, the vaccine composition is contaminated by less than about 10, 9, 8, 7, 6, 5, 4, 3, 2, 1, 0.5, 0.25, 0.05% (or any range derivable therein) of other Staphylococcal proteins. A composition may further comprise an isolated non-SpA peptide. Typically the vaccine comprises an adjuvant. In certain aspects a protein or peptide of the invention is linked (covalently or non-covalently) to the adjuvant, preferably the adjuvant is chemically conjugated to the protein.

In still yet further embodiments, a vaccine composition is a pharmaceutically acceptable composition having a recombinant nucleic acid encoding all or part of a SpA variant polypeptide, or any other combination or permutation of protein(s) or peptide(s) described herein, wherein the composition is capable of stimulating an immune response against a *staphylococcus* bacteria. The vaccine composition may comprise a recombinant nucleic acid encoding all or part of a SpA variant polypeptide, or any other combination or permutation of protein(s) or peptide(s) described herein. In certain embodiments the recombinant nucleic acid contains a heterologous promoter. Preferably the recombinant nucleic acid is a vector. More preferably the vector is a plasmid or a viral vector. In some 97597664.1 aspects the vaccine includes a recombinant, non-*staphylococcus* bacterium containing the nucleic acid. The recombinant non-staphylococci may be *Salmonella* or another gram-positive

bacteria. The vaccine may comprise a pharmaceutically acceptable excipient, more preferably an adjuvant.

Still further embodiments include methods for stimulating in a subject a protective or therapeutic immune response against a *staphylococcus* bacterium comprising administering to the subject an effective amount of a composition of a SpA variant polypeptide or segment/fragment thereof and further comprising one or more of a Eap, Ebh, Emp, EsaB, EsaC, EsxA, EsxB, SdrC, SdrD, SdrE, IsdA, IsdB, ClfA, ClfB, Coa, Hla, IsdC, SasF, vWbp, or vWh protein or peptide thereof. In a preferred embodiment the composition comprises a non-*staphylococcus* bacterium. In a further aspect the composition is formulated in a pharmaceutically acceptable formulation. The staphylococci for which a subject is being treated may be *Staphylococcus aureus*. Methods of the invention also include SpA variant compositions that contain 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19 or more secreted virulence factors and/or cell surface proteins, such as Eap, Ebh, Emp, EsaC, EsxA, EsxB, SdrC, SdrD, SdrE, IsdA, IsdB, ClfA, ClfB, Coa, Hla, IsdC, SasF, vWbp, or vWh in various combinations. In certain aspects a vaccine formulation includes Eap, Ebh, Emp, EsaC, EsxA, EsxB, SdrC, SdrD, SdrE, IsdA, IsdB, ClfA, ClfB, Coa, Hla, IsdC, SasF, vWbp, and vWh. In certain aspects an antigen combination can include (1) a SpA variant and IsdA; (2) SpA variant and ClfB; (3) SpA variant and SdrD; (4) SpA variant and Hla or Hla variant; (5) SpA variant and ClfB, SdrD, and Hla or Hla variant; (6) SpA variant, IsdA, SdrD, and Hla or Hla variant; (7) SpA variant, IsdA, ClfB, and Hla or Hla variant; (8) SpA variant, IsdA, ClfB, and SdrD; (9) SpA variant, IsdA, ClfB, SdrD and Hla or Hla variant; (10) SpA variant, IsdA, ClfB, and SdrD; (11) SpA variant, IsdA, SdrD, and Hla or Hla variant; (12) SpA variant, IsdA, and Hla or Hla variant; (13) SpA variant, IsdA, ClfB, and Hla or Hla variant; (14) SpA variant, ClfB, and SdrD; (15) SpA variant, ClfB, and Hla or Hla variant; or (16) SpA variant, SdrD, and Hla or Hla variant.

In certain aspects, a bacterium delivering a composition of the invention will be limited or attenuated with respect to prolonged or persistent growth or abscess formation. In yet a further aspect, SpA variant(s) can be overexpressed in an attenuated bacterium to further enhance or supplement an immune response or vaccine formulation.

Certain embodiments are directed to methods for eliciting an immune response against a *staphylococcus* bacterium in a subject comprising providing to the subject an effective amount of a peptide comprising a coagulase polypeptide or an immunogenic segment thereof having an amino acid sequence that is at least 80, 85, 90, 95, 98, to 100% identical to SEQ ID NO:27 or a segment thereof or at least 80, 85, 90, 95, 98, to 100% identical to amino acids 27-508 of SEQ ID NO:32 or a segment thereof.

In certain aspects, the subject is provided with an effective amount of an coagulase polypeptide by administering to the subject a composition comprising: (i) an isolated coagulase polypeptide or segment thereof having an amino acid sequence that is at least 90% identical to SEQ ID NO:27 or a segment thereof or is at least 90% identical to amino acids 27-508 of SEQ ID NO:32 or a segment thereof; or (ii) at least one isolated recombinant nucleic acid molecule encoding a coagulase polypeptide or a segment thereof having an amino acid sequence that is at least 90% identical to SEQ ID NO:27 or a segment thereof or is at least 90% identical to amino acids 27-508 of SEQ ID NO:32 or a segment thereof. In a further aspect, the composition comprises an isolated

coagulase polypeptide having the amino acid sequence of SEQ ID NO:27 or the amino acid sequence of amino acids 27-508 of SEQ ID NO:32.

Certain embodiments are directed to methods for treating a staphylococcal infection in a subject comprising providing to a subject having or suspected of having or at risk of developing a staphylococcal infection an effective amount of an isolated peptide comprising a coagulase polypeptide having an amino acid sequence that is at least 80, 85, 90, 95, 98, to 100% identical to SEQ ID NO:27 or is at least 80, 85, 90, 95, 98, to 100% identical to amino acids 27-508 of SEQ ID NO:32. In a particular aspect, the coagulase polypeptide has an amino acid sequence of SEQ ID NO:27 or has an amino acid identical to amino acids 27-508 of SEQ ID NO:32. In certain aspects, the subject is diagnosed with a persistent staphylococcal infection. In a further aspect, the coagulase polypeptide elicits production of an antibody that binds Coa or vWbpvWh in the subject.

Embodiments include methods of preventing or treating staphylococcal infection comprising the step of administering an immunogenic composition comprising a Staphylococcal coagulase or an immunogenic segment thereof.

Certain embodiments are directed to methods of preparing an immunoglobulin for use in prevention or treatment of staphylococcal infection comprising the steps of immunizing a recipient with a coagulase polypeptide and isolating immunoglobulin from the recipient.

A further embodiment is directed to an immunoglobulin prepared by the method described herein.

A further embodiment is directed to methods for treatment or prevention of staphylococcal infection comprising a step of administering to a patient an effective amount of pharmaceutical preparation of immunoglobulin that binds a coagulase.

Other embodiments are directed to a use of the pharmaceutical preparation of coagulase immunoglobulins in the manufacture of a medicament for the treatment or prevention of staphylococcal infection.

Yet still further embodiments include vaccines comprising a pharmaceutically acceptable composition having an isolated coagulase polypeptide, or any other combination or permutation of protein(s) or peptide(s) described herein, wherein the composition is capable of stimulating an immune response against a *staphylococcus* bacterium. The vaccine may comprise an isolated coagulase polypeptide, or any other combination or permutation of protein(s) or peptide(s) described. In certain aspects of the invention the isolated coagulase polypeptide, or any other combination or permutation of protein(s) or peptide(s) described are multimerized, e.g., dimerized or concatamerized. In a further aspect, the vaccine composition is contaminated by less than about 10, 9, 8, 7, 6, 5, 4, 3, 2, 1, 0.5, 0.25, 0.05% (or any range derivable therein) of other Staphylococcal proteins. A composition may further comprise an isolated non-coagulase polypeptide. Typically the vaccine comprises an adjuvant. In certain aspects a protein or peptide of the invention is linked (covalently or non-covalently) to the adjuvant, preferably the adjuvant is chemically conjugated to the protein.

In still yet further embodiments, a vaccine composition is a pharmaceutically acceptable composition having a recombinant nucleic acid encoding all or part of a coagulase polypeptide, or any other combination or permutation of protein(s) or peptide(s) described herein, wherein the composition is capable of stimulating an immune response against a *staphylococcus* bacterium. The vaccine composition may comprise a recombinant nucleic acid encoding all

or part of a coagulase polypeptide, or any other combination or permutation of protein(s) or peptide(s) described herein. In certain embodiments the recombinant nucleic acid contains a heterologous promoter. Preferably the recombinant nucleic acid is a vector. More preferably the vector is a plasmid or a viral vector. In some aspects the vaccine includes a recombinant, non-*staphylococcus* bacterium containing the nucleic acid. The recombinant non-staphylococci may be *Salmonella* or another gram-positive bacteria. The vaccine may comprise a pharmaceutically acceptable excipient, more preferably an adjuvant.

Still further embodiments include methods for stimulating in a subject a protective or therapeutic immune response against a *staphylococcus* bacterium comprising administering to the subject an effective amount of a composition of a coagulase polypeptide or segment/fragment thereof and further comprising one or more of a Eap, Ebh, Emp, EsaB, EsaC, EsxA, EsxB, SdrC, SdrD, SdrE, IsdA, IsdB, ClfA, ClfB, Coa, Hla, IsdC, SasF, vWbp, or vWh protein or peptide thereof. In a preferred embodiment the composition comprises a non-*staphylococcus* bacterium. In a further aspect the composition is formulated in a pharmaceutically acceptable formulation. The staphylococci for which a subject is being treated may be *Staphylococcus aureus*. Methods of the invention also include coagulase compositions that contain one or more of 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19 or more secreted virulence factors and/or cell surface proteins, such as Eap, Ebh, Emp, EsaC, EsxA, EsxB, SdrC, SdrD, SdrE, IsdA, IsdB, ClfA, ClfB, Coa, Hla, IsdC, SasF, SpA and variants thereof, vWbp, or vWh in various combinations. In certain aspects a vaccine formulation includes Eap, Ebh, Emp, EsaC, EsxA, EsxB, SdrC, SdrD, SdrE, IsdA, IsdB, ClfA, ClfB, Coa, Hla, IsdC, SasF, vWbp, and vWh. In certain aspects an antigen combination can include (1) a Coa and/or vWbp, and IsdA; (2) a Coa and/or vWbp, and ClfB; (3) a Coa and/or vWbp, and SdrD; (4) a Coa and/or vWbp, and Hla or Hla variant; (5) a Coa and/or vWbp, and ClfB, SdrD, and Hla or Hla variant; (6) a Coa and/or vWbp, and IsdA, SdrD, and Hla or Hla variant; (7) a Coa and/or vWbp, and IsdA, ClfB, and Hla or Hla variant; (8) a Coa and/or vWbp, and IsdA, ClfB, and SdrD; (9) a Coa and/or vWbp, and IsdA, ClfB, SdrD and Hla or Hla variant; (10) a Coa and/or vWbp, and IsdA, ClfB, and SdrD; (11) a Coa and/or vWbp, and IsdA, SdrD, and Hla or Hla variant; (12) a Coa and/or vWbp, and IsdA, and Hla or Hla variant; (13) a Coa and/or vWbp, and IsdA, ClfB, and Hla or Hla variant; (14) a Coa and/or vWbp, and ClfB, and SdrD; (15) a Coa and/or vWbp, and ClfB, and Hla or Hla variant; or (16) a Coa and/or vWbp, and SdrD, and Hla or Hla variant.

The term “EsxA protein” refers to a protein that includes isolated wild-type EsxA polypeptides from *staphylococcus* bacteria and segments thereof, as well as variants that stimulate an immune response against *staphylococcus* bacteria EsxA proteins.

The term “EsxB protein” refers to a protein that includes isolated wild-type EsxB polypeptides from *staphylococcus* bacteria and segments thereof, as well as variants that stimulate an immune response against *staphylococcus* bacteria EsxB proteins.

The term “SdrD protein” refers to a protein that includes isolated wild-type SdrD polypeptides from *staphylococcus* bacteria and segments thereof, as well as variants that stimulate an immune response against *staphylococcus* bacteria SdrD proteins.

The term “SdrE protein” refers to a protein that includes isolated wild-type SdrE polypeptides from *staphylococcus*

bacteria and segments thereof, as well as variants that stimulate an immune response against *staphylococcus* bacteria SdrE proteins.

The term “IsdA protein” refers to a protein that includes isolated wild-type IsdA polypeptides from *staphylococcus* bacteria and segments thereof, as well as variants that stimulate an immune response against *staphylococcus* bacteria IsdA proteins.

The term “IsdB protein” refers to a protein that includes isolated wild-type IsdB polypeptides from *staphylococcus* bacteria and segments thereof, as well as variants that stimulate an immune response against *staphylococcus* bacteria IsdB proteins.

The term “Eap protein” refers to a protein that includes isolated wild-type Eap polypeptides from *staphylococcus* bacteria and segments thereof, as well as variants that stimulate an immune response against *staphylococcus* bacteria Eap proteins.

The term “Ebh protein” refers to a protein that includes isolated wild-type Ebh polypeptides from *staphylococcus* bacteria and segments thereof, as well as variants that stimulate an immune response against *staphylococcus* bacteria Ebh proteins.

The term “Emp protein” refers to a protein that includes isolated wild-type Emp polypeptides from *staphylococcus* bacteria and segments thereof, as well as variants that stimulate an immune response against *staphylococcus* bacteria Emp proteins.

The term “EsaB protein” refers to a protein that includes isolated wild-type EsaB polypeptides from *staphylococcus* bacteria and segments thereof, as well as variants that stimulate an immune response against *staphylococcus* bacteria EsaB proteins.

The term “EsaC protein” refers to a protein that includes isolated wild-type EsaC polypeptides from *staphylococcus* bacteria and segments thereof, as well as variants that stimulate an immune response against *staphylococcus* bacteria EsaC proteins.

The term “SdrC protein” refers to a protein that includes isolated wild-type SdrC polypeptides from *staphylococcus* bacteria and segments thereof, as well as variants that stimulate an immune response against *staphylococcus* bacteria SdrC proteins.

The term “ClfA protein” refers to a protein that includes isolated wild-type ClfA polypeptides from *staphylococcus* bacteria and segments thereof, as well as variants that stimulate an immune response against *staphylococcus* bacteria ClfA proteins.

The term “ClfB protein” refers to a protein that includes isolated wild-type ClfB polypeptides from *staphylococcus* bacteria and segments thereof, as well as variants that stimulate an immune response against *staphylococcus* bacteria ClfB proteins.

The term “Coa protein” refers to a protein that includes isolated wild-type Coa polypeptides from *staphylococcus* bacteria and segments thereof, as well as variants that stimulate an immune response against *staphylococcus* bacteria Coa proteins.

The term “Hla protein” refers to a protein that includes isolated wild-type Hla polypeptides from *staphylococcus* bacteria and segments thereof, as well as variants that stimulate an immune response against *staphylococcus* bacteria Hla proteins.

The term “IsdC protein” refers to a protein that includes isolated wild-type IsdC polypeptides from *staphylococcus*

bacteria and segments thereof, as well as variants that stimulate an immune response against *staphylococcus* bacteria IsdC proteins.

The term "SasF protein" refers to a protein that includes isolated wild-type SasF polypeptides from *staphylococcus* bacteria and segments thereof, as well as variants that stimulate an immune response against *staphylococcus* bacteria SasF proteins.

The term "vWbp protein" refers to a protein that includes isolated wild-type vWbp (von Willebrand factor binding protein) polypeptides from *staphylococcus* bacteria and segments thereof, as well as variants that stimulate an immune response against *staphylococcus* bacteria vWbp proteins.

The term "vWh protein" refers to a protein that includes isolated wild-type vWh (von Willebrand factor binding protein homolog) polypeptides from *staphylococcus* bacteria and segments thereof, as well as variants that stimulate an immune response against *staphylococcus* bacteria vWh proteins.

An immune response refers to a humoral response, a cellular response, or both a humoral and cellular response in an organism. An immune response can be measured by assays that include, but are not limited to, assays measuring the presence or amount of antibodies that specifically recognize a protein or cell surface protein, assays measuring T-cell activation or proliferation, and/or assays that measure modulation in terms of activity or expression of one or more cytokines.

In still further embodiments of the invention a composition may include a polypeptide, peptide, or protein that is or is at least 70%, 75%, 80%, 85%, 90%, 95%, 96%, 97%, 98%, or 99% identical or similar to an EsxA protein. In certain aspects the EsxA protein will have all or part of the amino acid sequence of SEQ ID NO:11.

In still further embodiments of the invention a composition may include a polypeptide, peptide, or protein that is or is at least 70%, 75%, 80%, 85%, 90%, 95%, 96%, 97%, 98%, or 99% identical or similar to an EsxB protein. In certain aspects the EsxB protein will have all or part of the amino acid sequence of SEQ ID NO:12.

In yet still further embodiments of the invention a composition may include a polypeptide, peptide, or protein that is or is at least 70%, 75%, 80%, 85%, 90%, 95%, 96%, 97%, 98%, or 99% identical or similar to an SdrD protein. In certain aspects the SdrD protein will have all or part of the amino acid sequence of SEQ ID NO:13.

In further embodiments of the invention a composition may include a polypeptide, peptide, or protein that is or is at least 70%, 75%, 80%, 85%, 90%, 95%, 96%, 97%, 98%, or 99% identical or similar to an SdrE protein. In certain aspects the SdrE protein will have all or part of the amino acid sequence of SEQ ID NO:14.

In still further embodiments of the invention a composition may include a polypeptide, peptide, or protein that is or is at least 70%, 75%, 80%, 85%, 90%, 95%, 96%, 97%, 98%, or 99% identical or similar to an IsdA protein. In certain aspects the IsdA protein will have all or part of the amino acid sequence of SEQ ID NO:15.

In yet still further embodiments of the invention a composition may include a polypeptide, peptide, or protein that is or is at least 70%, 75%, 80%, 85%, 90%, 95%, 96%, 97%, 98%, or 99% identical or similar to an IsdB protein. In certain aspects the IsdB protein will have all or part of the amino acid sequence of SEQ ID NO:16.

Embodiments of the invention include compositions that include a polypeptide, peptide, or protein that is or is at least 70%, 75%, 80%, 85%, 90%, 95%, 96%, 97%, 98%, or 99%

identical or similar to a EsaB protein. In certain aspects the EsaB protein will have all or part of the amino acid sequence of SEQ ID NO:17.

In a further embodiments of the invention a composition may include a polypeptide, peptide, or protein that is or is at least 70%, 75%, 80%, 85%, 90%, 95%, 96%, 97%, 98%, or 99% identical or similar to a ClfB protein. In certain aspects the ClfB protein will have all or part of the amino acid sequence of SEQ ID NO:18.

In still further embodiments of the invention a composition may include a polypeptide, peptide, or protein that is or is at least 70%, 75%, 80%, 85%, 90%, 95%, 96%, 97%, 98%, or 99% identical or similar to an IsdC protein. In certain aspects the IsdC protein will have all or part of the amino acid sequence of SEQ ID NO:19.

In yet further embodiments of the invention a composition may include a polypeptide, peptide, or protein that is or is at least 70%, 75%, 80%, 85%, 90%, 95%, 96%, 97%, 98%, or 99% identical or similar to a SasF protein. In certain aspects the SasF protein will have all or part of the amino acid sequence of SEQ ID NO:20.

In yet still further embodiments of the invention a composition may include a polypeptide, peptide, or protein that is or is at least 70%, 75%, 80%, 85%, 90%, 95%, 96%, 97%, 98%, or 99% identical or similar to a SdrC protein. In certain aspects the SdrC protein will have all or part of the amino acid sequence of SEQ ID NO:21.

In yet still further embodiments of the invention a composition may include a polypeptide, peptide, or protein that is or is at least 70%, 75%, 80%, 85%, 90%, 95%, 96%, 97%, 98%, or 99% identical or similar to a ClfA protein. In certain aspects the ClfA protein will have all or part of the amino acid sequence of SEQ ID NO:22.

In yet still further embodiments of the invention a composition may include a polypeptide, peptide, or protein that is or is at least 70%, 75%, 80%, 85%, 90%, 95%, 96%, 97%, 98%, or 99% identical or similar to an Eap protein. In certain aspects the Eap protein will have all or part of the amino acid sequence of SEQ ID NO:23.

In yet still further embodiments of the invention a composition may include a polypeptide, peptide, or protein that is or is at least 70%, 75%, 80%, 85%, 90%, 95%, 96%, 97%, 98%, or 99% identical or similar to an Ebh protein. In certain aspects the Ebh protein will have all or part of the amino acid sequence of SEQ ID NO:24.

In yet still further embodiments of the invention a composition may include a polypeptide, peptide, or protein that is or is at least 70%, 75%, 80%, 85%, 90%, 95%, 96%, 97%, 98%, or 99% identical or similar to an Emp protein. In certain aspects the Emp protein will have all or part of the amino acid sequence of SEQ ID NO:25.

In yet still further embodiments of the invention a composition may include a polypeptide, peptide, or protein that is or is at least 70%, 75%, 80%, 85%, 90%, 95%, 96%, 97%, 98%, or 99% identical or similar to an EsaC protein. In certain aspects the EsaC protein will have all or part of the amino acid sequence of SEQ ID NO:26. Sequence of EsaC polypeptides can be found in the protein databases and include, but are not limited to accession numbers ZP_02760162 (GI:168727885), NP_645081.1 (GI:21281993), and NP_370813.1 (GI:15923279), each of which is incorporated herein by reference as of the priority date of this application.

In yet still further embodiments of the invention a composition may include a polypeptide, peptide, or protein that is or is at least 70%, 75%, 80%, 85%, 90%, 95%, 96%, 97%,

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98%, or 99% identical or similar to a Coa protein. In certain aspects the Coa protein will have all or part of the amino acid sequence of SEQ ID NO:27.

In yet still further embodiments of the invention a composition may include a polypeptide, peptide, or protein that is or is at least 70%, 75%, 80%, 85%, 90%, 95%, 96%, 97%, 98%, or 99% identical or similar to a Hla protein. In certain aspects the Hla protein will have all or part of the amino acid sequence of SEQ ID NO:28.

In yet still further embodiments of the invention a composition may include a polypeptide, peptide, or protein that is or is at least 70%, 75%, 80%, 85%, 90%, 95%, 96%, 97%, 98%, or 99% identical or similar to a vWa protein. In certain aspects the vWa protein will have all or part of the amino acid sequence of SEQ ID NO:29.

In yet still further embodiments of the invention a composition may include a polypeptide, peptide, or protein that is or is at least 70%, 75%, 80%, 85%, 90%, 95%, 96%, 97%, 98%, or 99% identical or similar to a vWbp protein. In certain aspects the vWbp protein will have all or part of the amino acid sequence of SEQ ID NO:32.

In certain aspects, a polypeptide or segment/fragment can have a sequence that is at least 85%, at least 90%, at least 95%, at least 98%, or at least 99% or more identical to the amino acid sequence of the reference polypeptide. The term "similarity" refers to a polypeptide that has a sequence that has a certain percentage of amino acids that are either identical with the reference polypeptide or constitute conservative substitutions with the reference polypeptides.

The polypeptides described herein may include 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, or more variant amino acids within at least, or at most 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 21, 22, 23, 24, 25, 26, 27, 28, 29, 30, 31, 32, 33, 34, 35, 36, 37, 38, 39, 40, 41, 42, 43, 44, 45, 46, 47, 48, 49, 50, 51, 52, 53, 54, 55, 56, 57, 58, 59, 60, 61, 62, 63, 64, 65, 66, 67, 68, 69, 70, 71, 72, 73, 74, 75, 76, 77, 78, 79, 80, 81, 82, 83, 84, 85, 86, 87, 88, 89, 90, 91, 92, 93, 94, 95, 96, 97, 98, 99, 100, 101, 102, 103, 104, 105, 106, 107, 108, 109, 110, 111, 112, 113, 114, 115, 116, 117, 118, 119, 120, 121, 122, 123, 124, 125, 126, 127, 128, 129, 130, 131, 132, 133, 134, 135, 136, 137, 138, 139, 140, 141, 142, 143, 144, 145, 146, 147, 148, 149, 150, 151, 152, 153, 154, 155, 156, 157, 158, 159, 160, 161, 162, 163, 164, 165, 166, 167, 168, 169, 170, 171, 172, 173, 174, 175, 176, 177, 178, 179, 180, 181, 182, 183, 184, 185, 186, 187, 188, 189, 190, 191, 192, 193, 194, 195, 196, 197, 198, 199, 200, 201, 202, 203, 204, 205, 206, 207, 208, 209, 210, 211, 212, 213, 214, 215, 216, 217, 218, 219, 220, 221, 222, 223, 224, 225, 226, 227, 228, 229, 230, 231, 232, 233, 234, 235, 236, 237, 238, 239, 240, 241, 242, 243, 244, 245, 246, 247, 248, 249, 250, 300, 400, 500, 550, 1000 or more contiguous amino acids, or any range derivable therein, of SEQ ID NO:2-30, or SEQ ID NO:32-34.

A polypeptide segment as described herein may include 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 21, 22, 23, 24, 25, 26, 27, 28, 29, 30, 31, 32, 33, 34, 35, 36, 37, 38, 39, 40, 41, 42, 43, 44, 45, 46, 47, 48, 49, 50, 51, 52, 53, 54, 55, 56, 57, 58, 59, 60, 61, 62, 63, 64, 65, 66, 67, 68, 69, 70, 71, 72, 73, 74, 75, 76, 77, 78, 79, 80, 81, 82, 83, 84, 85, 86, 87, 88, 89, 90, 91, 92, 93, 94, 95, 96, 97, 98, 99, 100, 101, 102, 103, 104, 105, 106, 107, 108, 109, 110, 111, 112, 113, 114, 115, 116, 117, 118, 119, 120, 121, 122, 123, 124, 125, 126, 127, 128, 129, 130, 131, 132, 133, 134, 135, 136, 137, 138, 139, 140, 141, 142, 143, 144, 145, 146, 147, 148, 149, 150, 151, 152, 153, 154, 155, 156, 157, 158, 159, 160, 161, 162, 163, 164, 165, 166, 167, 168, 169, 170, 171, 172, 173, 174, 175, 176, 177, 178, 179, 180, 181, 182, 183, 184, 185,

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The compositions may be formulated in a pharmaceutically acceptable composition. In certain aspects of the invention the *staphylococcus* bacterium is an *S. aureus* bacterium.

In further aspects, a composition may be administered more than one time to the subject, and may be administered 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 15, 20 or more times. The administration of the compositions include, but is not limited to oral, parenteral, subcutaneous, intramuscular, intravenous, or various combinations thereof, including inhalation or aspiration.

In still further embodiments, a composition comprises a recombinant nucleic acid molecule encoding a polypeptide described herein or segments/fragments thereof. Typically a recombinant nucleic acid molecule encoding a polypeptide described herein contains a heterologous promoter. In certain aspects, a recombinant nucleic acid molecule of the invention is a vector, in still other aspects the vector is a plasmid. In certain embodiments the vector is a viral vector. In certain aspects a composition includes a recombinant, non-*staphylococcus* bacterium containing or expressing a polypeptide described herein. In particular aspects the recombinant non-*staphylococcus* bacteria is *Salmonella* or another gram-positive bacteria. A composition is typically administered to mammals, such as human subjects, but administration to other animals that are capable of eliciting an immune response is contemplated. In further aspects the *staphylococcus* bacterium containing or expressing the polypeptide is *Staphylococcus aureus*. In further embodiments the immune response is a protective immune response.

In further embodiments a composition comprises a recombinant nucleic acid molecule encoding all or part of one or more of a Eap, Ebh, Emp, EsaB, EsaC, EsxA, EsxB, SdrC, SdrD, SdrE, IsdA, IsdB, ClfA, ClfB, Coa, Hla, IsdC, SasF, SpA, vWbp, or vWh protein or peptide or variant thereof. Additional staphylococcal antigens that can be used in combination with the polypeptides described herein include, but are not limited to 52 kDa vitronectin binding protein (WO 01/60852), Aaa, Aap, Ant, autolysin glucosaminidase, autolysin amidase, Cna, collagen binding protein (U.S. Pat. No. 6,288,214), EFB (FIB), Elastin binding protein (EbpS), EPB, FbpA, fibrinogen binding protein (U.S. Pat. No. 6,008,341), Fibronectin binding protein (U.S. Pat. No. 5,840,846), FnbA, FnbB, GehD (US 2002/0169288), HarA, HBP, Immunodominant ABC transporter, IsaA/PisA, laminin receptor, Lipase GehD, MAP, Mg2+ transporter, MHC II analogue (U.S. Pat. No. 5,648,240), MRPII, Npase, RNA III activating protein (RAP), SasA, SasB, SasC, SasD, SasK, SBI, SdrF (WO 00/12689), SdrG/Fig (WO 00/12689), SdrH (WO 00/12689), SEA exotoxins (WO 00/02523), SEB exotoxins (WO 00/02523), SitC and Ni ABC transporter, SitC/MntC/saliva binding protein (U.S. Pat. No. 5,801,234), SsaA, SSP-1, SSP-2, and/or Vitronectin binding protein. In particular aspects, a bacteria is a recombinant non-*staphylococcus* bacteria, such as a *Salmonella* or other gram-positive bacteria.

Compositions of the invention are typically administered to human subjects, but administration to other animals that are capable of eliciting an immune response to a *staphylo-*

coccus bacterium is contemplated, particularly cattle, horses, goats, sheep and other domestic animals, i.e., mammals.

In certain aspects the *staphylococcus* bacterium is a *Staphylococcus aureus*. In further embodiments the immune response is a protective immune response. In still further aspects, the methods and compositions of the invention can be used to prevent, ameliorate, reduce, or treat infection of tissues or glands, e.g., mammary glands, particularly mastitis and other infections. Other methods include, but are not limited to prophylactically reducing bacterial burden in a subject not exhibiting signs of infection, particularly those subjects suspected of or at risk of being colonized by a target bacteria, e.g., patients that are or will be at risk or susceptible to infection during a hospital stay, treatment, and/or recovery.

Any embodiment discussed with respect to one aspect of the invention applies to other aspects of the invention as well. In particular, any embodiment discussed in the context of a SpA variant polypeptide or peptide or nucleic acid may be implemented with respect to other antigens, such as Eap, Ebh, Emp, EsaC, EsxA, EsxB, SdrC, SdrD, SdrE, IsdA, IsdB, ClfA, ClfB, Coa, Hla, IsdC, SasF, vWbp, vWh, 52 kDa vitronectin binding protein (WO 01/60852), Aaa, Aap, Ant, autolysin glucosaminidase, autolysin amidase, Cna, collagen binding protein (U.S. Pat. No. 6,288,214), EFB (FIB), Elastin binding protein (EbpS), EPB, FbpA, fibrinogen binding protein (U.S. Pat. No. 6,008,341), Fibronectin binding protein (U.S. Pat. No. 5,840,846), FnbA, FnbB, GehD (US 2002/0169288), HarA, HBP, Immunodominant ABC transporter, IsaA/PisA, laminin receptor, Lipase GehD, MAP, Mg2+ transporter, MEW II analogue (U.S. Pat. No. 5,648,240), MRPII, Npase, RNA III activating protein (RAP), SasA, SasB, SasC, SasD, SasK, SBI, SdrF (WO 00/12689), SdrG/Fig (WO 00/12689), SdrH (WO 00/12689), SEA exotoxins (WO 00/02523), SEB exotoxins (WO 00/02523), SitC and Ni ABC transporter, SitC/MntC/saliva binding protein (U.S. Pat. No. 5,801,234), SsaA, SSP-1, SSP-2, and/or Vitronectin binding protein (or nucleic acids), and vice versa. It is also understood that any one or more of Eap, Ebh, Emp, EsaC, EsxA, EsxB, SdrC, SdrD, SdrE, IsdA, IsdB, ClfA, CHB, Coa, Hla, IsdC, SasF, vWbp, vWh, 52 kDa vitronectin binding protein (WO 01/60852), Aaa, Aap, Ant, autolysin glucosaminidase, autolysin amidase, Cna, collagen binding protein (U.S. Pat. No. 6,288,214), EFB (FIB), Elastin binding protein (EbpS), EPB, FbpA, fibrinogen binding protein (U.S. Pat. No. 6,008,341), Fibronectin binding protein (U.S. Pat. No. 5,840,846), FnbA, FnbB, GehD (US 2002/0169288), HarA, HBP, Immunodominant ABC transporter, IsaA/PisA, laminin receptor, Lipase GehD, MAP, Mg2+ transporter, MEW II analogue (U.S. Pat. No. 5,648,240), MRPII, Npase, RNA III activating protein (RAP), SasA, SasB, SasC, SasD, SasK, SBI, SdrF (WO 00/12689), SdrG/Fig (WO 00/12689), SdrH (WO 00/12689), SEA exotoxins (WO 00/02523), SEB exotoxins (WO 00/02523), SitC and Ni ABC transporter, SitC/MntC/saliva binding protein (U.S. Pat. No. 5,801,234), SsaA, SSP-1, SSP-2, and/or Vitronectin binding protein can be specifically excluded from a claimed composition.

Embodiments of the invention include compositions that contain or do not contain a bacterium. A composition may or may not include an attenuated or viable or intact staphylococcal bacterium. In certain aspects, the composition comprises a bacterium that is not a staphylococcal bacterium or does not contain staphylococcal bacteria. In certain embodiments a bacterial composition comprises an isolated or recombinantly expressed staphylococcal Protein A variant or

a nucleotide encoding the same. The composition may be or include a recombinantly engineered *staphylococcus* bacterium that has been altered in a way that comprises specifically altering the bacterium with respect to a secreted virulence factor or cell surface protein. For example, the bacteria may be recombinantly modified to express more of the virulence factor or cell surface protein than it would express if unmodified.

The term “isolated” can refer to a nucleic acid or polypeptide that is substantially free of cellular material, bacterial material, viral material, or culture medium (when produced by recombinant DNA techniques) of their source of origin, or chemical precursors or other chemicals (when chemically synthesized). Moreover, an isolated compound refers to one that can be administered to a subject as an isolated compound; in other words, the compound may not simply be considered “isolated” if it is adhered to a column or embedded in an agarose gel. Moreover, an “isolated nucleic acid fragment” or “isolated peptide” is a nucleic acid or protein fragment that is not naturally occurring as a fragment and/or is not typically in the functional state.

Moieties of the invention, such as polypeptides, peptides, antigens, or immunogens, may be conjugated or linked covalently or noncovalently to other moieties such as adjuvants, proteins, peptides, supports, fluorescence moieties, or labels. The term “conjugate” or “immunoconjugate” is broadly used to define the operative association of one moiety with another agent and is not intended to refer solely to any type of operative association, and is particularly not limited to chemical “conjugation.” Recombinant fusion proteins are particularly contemplated. Compositions of the invention may further comprise an adjuvant or a pharmaceutically acceptable excipient. An adjuvant may be covalently or non-covalently coupled to a polypeptide or peptide of the invention. In certain aspects, the adjuvant is chemically conjugated to a protein, polypeptide, or peptide.

The term “providing” is used according to its ordinary meaning to indicate “to supply or furnish for use.” In some embodiments, the protein is provided directly by administering the protein, while in other embodiments, the protein is effectively provided by administering a nucleic acid that encodes the protein. In certain aspects the invention contemplates compositions comprising various combinations of nucleic acid, antigens, peptides, and/or epitopes.

The subject will have (e.g., are diagnosed with a staphylococcal infection), will be suspected of having, or will be at risk of developing a staphylococcal infection. Compositions of the present invention include immunogenic compositions wherein the antigen(s) or epitope(s) are contained in an amount effective to achieve the intended purpose. More specifically, an effective amount means an amount of active ingredients necessary to stimulate or elicit an immune response, or provide resistance to, amelioration of, or mitigation of infection. In more specific aspects, an effective amount prevents, alleviates or ameliorates symptoms of disease or infection, or prolongs the survival of the subject being treated. Determination of the effective amount is well within the capability of those skilled in the art, especially in light of the detailed disclosure provided herein. For any preparation used in the methods of the invention, an effective amount or dose can be estimated initially from in vitro studies, cell culture, and/or animal model assays. For example, a dose can be formulated in animal models to achieve a desired immune response or circulating antibody concentration or titer. Such information can be used to more accurately determine useful doses in humans.

The embodiments in the Example section are understood to be embodiments of the invention that are applicable to all aspects of the invention.

The use of the term “or” in the claims is used to mean “and/or” unless explicitly indicated to refer to alternatives only or the alternatives are mutually exclusive, although the disclosure supports a definition that refers to only alternatives and “and/or.” It is also contemplated that anything listed using the term “or” may also be specifically excluded.

Throughout this application, the term “about” is used to indicate that a value includes the standard deviation of error for the device or method being employed to determine the value.

Following long-standing patent law, the words “a” and “an,” when used in conjunction with the word “comprising” in the claims or specification, denotes one or more, unless specifically noted.

Other objects, features and advantages of the present invention will become apparent from the following detailed description. It should be understood, however, that the detailed description and the specific examples, while indicating specific embodiments of the invention, are given by way of illustration only, since various changes and modifications within the spirit and scope of the invention will become apparent to those skilled in the art from this detailed description.

DESCRIPTION OF THE DRAWINGS

So that the matter in which the above-recited features, advantages and objects of the invention as well as others which will become clear are attained and can be understood in detail, more particular descriptions and certain embodiments of the invention briefly summarized above are illustrated in the appended drawings. These drawings form a part of the specification. It is to be noted, however, that the appended drawings illustrate certain embodiments of the invention and therefore are not to be considered limiting in their scope.

FIGS. 1A-1E Generation of a non-toxicogenic protein A vaccine. FIG. 1A Translational protein A (SpA) product of *S. aureus* Newman and USA300 LAC with an N-terminal signal peptide (white box), five immunoglobulin binding domains (IgBDs designated E, D, A, B, and C), variable region X and C-terminal sorting signal (black box). FIG. 1B (SEQ ID NOs. 64-69), Amino acid sequence of the five IgBDs as well as nontoxicogenic SpA-D_{KKAA} (SEQ ID NO. 69), with the positions of triple α -helical bundles (H1, H2 and H3) as well as glutamine (Q) 9, 10 and aspartate (D) 36, 37 indicated. FIG. 1C, COOMASSIE-BLUE-stained SDS-PAGE of SpA, SpA-D, SpA-D_{KKAA} (SEQ ID NO. 69) or SrtA purified on Ni-NTA sepharose in the presence or absence of human immunoglobulin (hIgG). FIG. 1D, ELISA examining the association of immobilized SpA, SpA-D or SpA-D_{KKAA} (SEQ ID NO. 69) with human IgG as well as its Fc or F(ab)₂ fragments and von Willebrand factor (vWF). FIG. 1E, CD19+ B lymphocytes in splenic tissue of BALB/c mice that had been mock immunized or treated with SpA-D or SpA-D_{KKAA} (SEQ ID NO. 69) were quantified by FACS.

FIG. 2 Non-toxicogenic protein A vaccine prevents abscess formation. Histopathology of renal tissue isolated during necropsy of BALB/c mice that had been mock immunized (PBS) or vaccinated with SpA, SpA-D as well as SpA-D_{KKAA} (SEQ ID NO. 69) and challenged with *S. aureus* Newman. Thin sectioned tissues were stained with hema-

toxylin-eosin. White arrows identify polymorphonuclear leukocyte (PMN) infiltrates. Dark arrows identify staphylococcal abscess communities.

FIGS. 3A-C Antibodies raised by the non-toxicogenic protein A vaccine block the B cell superantigen function of SpA. FIG. 3A, Rabbit antibodies raised against SpA-D_{KKAA} (SEQ ID NO. 69) were purified on a matrix with immobilized antigen and analyzed by COOMASSIE-BLUE-stained SDS-PAGE. Antibodies were cleaved with pepsin and F(ab)₂ fragments were purified by a second round of affinity chromatography on SpA-D_{KKAA} (SEQ ID NO. 69) matrix. FIG. 3B, SpA-D_{KKAA} (SEQ ID NO. 69) specific F(ab)₂ interfere with the binding of SpA or SpA-D to human immunoglobulin (hIgG) or, FIG. 3C, to von Willebrand Factor (vWF).

FIGS. 4A-D Full-length non-toxicogenic protein A generates improved immune responses. FIG. 4A, Full-length SpA_{KKAA} (SEQ ID NO. 34) was purified on Ni-NTA sepharose and analyzed by COOMASSIE-BLUE stained SDS-PAGE. FIG. 4B, CD19+B lymphocytes in splenic tissue of BALB/c mice that had been mock immunized or treated with SpA or SpA_{KKAA} (SEQ ID NO. 34) were quantified by FACS. FIG. 4C, ELISA examining the association of immobilized SpA or SpA_{KKAA} (SEQ ID NO. 34) with human IgG as well as its Fc or F(ab)₂ fragments or von Willebrand factor (vWF). FIG. 4D, Human or mouse serum antibody titers to diphtheria toxoid (CRM197) and non-toxicogenic SpA_{KKAA} (SEQ ID NO. 34) or SpA-D_{KKAA} (SEQ ID NO. 69). Human volunteers with a history of DTaP immunization and staphylococcal infection (n=16) as well as mice (n=20) that had been infected with *S. aureus* Newman or USA 300 LAC or immunized with SpA_{KKAA} (SEQ ID NO. 34) or SpA-D_{KKAA} (SEQ ID NO. 69) were examined by quantitative dot blot.

FIG. 5 Protein A is required for the pathogenesis of lethal *S. aureus* infections in mice. Cohorts of BALB/c mice (n=8) were injected with suspensions of 2×10^8 CFU *S. aureus* Newman or its isogenic protein A deletion variant (Δ spa) in PBS. Infected animals were monitored for survival over a period of 15 days.

FIGS. 6A-B Antibodies against protein A protect mice against lethal *S. aureus* infections. FIG. 6A Cohorts of BALB/c mice (n=10) were injected with 5 mg kg⁻¹ affinity purified rabbit IgG specific for SpA_{KKAA} (α -Sp_{AKKAA}) (SEQ ID NO. 34) or the plague vaccine antigen rV10 (DeBord et al., 2006) (mock). Four hours later, each animal was infected by intraperitoneal injection with a suspension of 3×10^8 CFU *S. aureus* Newman and monitored for survival over a period of 10 days. Data are representative of three independent experiments FIG. 6B Cohorts of BALB/c mice (n=10) were prime-booster immunized with SpA_{KKAA} (SEQ ID NO. 34) or PBS/adjuvant control (mock). Each animal was subsequently infected by intraperitoneal injection with a suspension of 6×10^8 CFU *S. aureus* Newman and monitored for survival over a period of 10 days. Statistical significance (P) was analyzed with the unpaired two-tailed log-rank test. Data are representative of all three independent experiments.

FIG. 7 SpA_{KKAA} (SEQ ID NO. 34) immunization protects mice against challenge with the vancomycin-resistant MRSA isolated Mu50. Cohorts of BALB/c mice (n=15) were prime-booster immunized with SpA_{KKAA} (SEQ ID NO. 34) or PBS/adjuvant control (mock). Each animal was subsequently infected by intravenous injection with a suspension of 3×10^7 CFU *S. aureus* Mu50. Staphylococcal load, calculated as log₁₀ CFU g⁻¹, was determined in homogenized renal tissues 4 days following infection. Sta-

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tistical significance was calculated with the unpaired two-tailed Students t-test and P-value recorded.

FIGS. 8A-B Lack of protective immune responses to staphylococcal infections. FIG. 8A Staphylococcal infection does not generate protective immunity. BALB/c mice (n=10) were infected with *S. aureus* Newman or mock challenged (PBS) for thirty days and infection cleared with chloramphenicol treatment. Both cohorts of animals were then challenged with *S. aureus* Newman and bacterial load (CFU) in kidney tissue homogenate analyzed following necropsy on day 4. Data are representative of three independent analyses. FIG. 8B IsdB immunization does not protect mice against *S. aureus* USA300 (LAC) challenge. BALB/c mice (n=10) were immunized with IsdB (100 µg IsdB emulsified in CFA followed by IFA/IsdB booster on day 11) and challenged by retro-orbital injection with 5×10^6 CFU *S. aureus* USA300 (LAC) on day 21. Four days following challenge, kidneys were removed during necropsy and staphylococcal load per gram of homogenized tissue enumerated by colony formation on agar plates. Compared to mock immunized (PBS/adjuvant) animals with $6.93 (\pm 0.24) \log_{10}$ CFU g^{-1} , IsdB vaccination was associated with $6.25 (\pm 0.46) \log_{10}$ CFU g^{-1} and did not generate statistically significant protection ($P=0.2138$, two-tailed Student's t-test) from USA300 (LAC) challenge. Data are representative of three independent analyses.

FIG. 9 Comparison of abscess formation in mice treated with PBS, SpA, SpA-D and SpA-D_{KKAA} (SEQ ID NO. 69).

FIGS. 10A-10H Localization of prothrombin, fibrinogen, coagulase (Coa), and von Willebrand factor binding protein (vWbp) in staphylococcal abscesses. BALB/c mice infected by intravenous inoculation with 1×10^7 CFU *S. aureus* Newman were killed 5 days post infection. Kidneys were removed, embedded in paraffin, thin-sectioned and stained by immunochemistry using rabbit antibodies (a) specific for mouse prothrombin (FIG. 10A, 10C), mouse fibrinogen/fibrin (FIG. 10B, 10D), *S. aureus* Coa (FIG. 10E, 10G) or *S. aureus* vWbp (FIG. 10F, 10H). Displayed images are representative of three sampled kidneys. Panels FIGS. 10C, 10D, 10G, and 10H illustrate antibody staining within a single abscess analyzed as four sequential sections, enlarged from an area in panels FIGS. 10A, 10B, 10E, and 10F that is defined by box with white margins.

FIGS. 11A-11C *Staphylococcus aureus* coa and vWbp mutants display defects in blood clotting. (FIG. 11A) Diagram illustrating the primary translational product of coa and vWbp including signal peptide (S), the D1 and D2 domain from prothrombin binding, a domain of unknown function, von Willebrand factor (vWF) binding site on vWbp, and the fibrinogen binding repeats (R) of Coa. Numbers indicate amino acid residues. (FIG. 11B) Culture supernatants from *S. aureus* Newman (wild-type) or isogenic variants lacking coa (Δ coa), vWbp (Δ vWbp) or both genes (Δ coa, Δ vWbp) were examined by immunoblotting with antibodies specific for Coa (α Coa) or vWbp (α vWbp). For complementation studies, plasmids expressing the wild-type alleles of coa (pcoa) or vWbp (pvWbp) were electroporated into staphylococcal strains and subsequently analyzed by immunoblotting. (FIG. 11C) Lepirudin-treated mouse blood was mock treated or infected with *S. aureus* Newman or its isogenic coagulase variants and incubated for up to 48 hours at 25° C. Tubes were tilted to assess for coagulation. Data are representative of four independent determinations.

FIGS. 12A-12R Contributions of coa and vWbp to bacterial survival in blood and *S. aureus* induced lethal bacteremia of mice. (FIG. 12A) Staphylococcal strains Newman, Δ coa, Δ vWbp or Δ coa, Δ vWbp and the complemented

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variants were incubated with lepirudin anticoagulated mouse blood for 30 minutes and bacterial survival assessed by colony formation on agar plates. Data were generated from three separate trials. (FIG. 12B) Cohorts of 10 mice were injected into the retro-orbital plexus with 1×10^8 CFU of *S. aureus* Newman (wild-type) as well as Δ coa, Δ vWbp or Δ coa, Δ vWbp. Animal survival over time was recorded over 10 days. Similar to B, mice were given 1×10^7 CFU of staphylococcal strains Newman (FIGS. 12C, E and K, M), Δ vWbp (FIGS. 14D, F and M, L), Δ coa (FIGS. 14G, I and O, Q) or Δ coa, Δ vWbp (FIGS. 12H, J and P, R), harvested on days 5 (FIG. 12C-J) or 15 (FIG. 12K-R) and assessed for bacterial load in the renal tissue (Table 7) and histopathological abscess formation. All animal data are representative of two independent experiments.

FIGS. 13A-13D Antibodies against Coa and vWbp block the clotting of blood by staphylococcal coagulases. (FIG. 13A) His₆-Coa and His₆-vWbp were purified by affinity chromatography from *E. coli* and analyzed on COOMASSIE-stained SDS-PAGE. (FIG. 13B) Rabbit antibodies raised against His₆-Coa or His₆-vWbp were affinity purified and analyzed by ELISA for immune reactivity with purified coagulases. Data are averaged from three independent experimental determinations. (FIG. 13C) Lepirudin-treated mouse blood was treated with PBS (mock), irrelevant antibodies (α V10) or antibodies directed against Coa (α Coa), vWbp (α vWbp) or both coagulases (α Coa/ α vWbp) prior to infection with *S. aureus* Newman and incubation for 48 hours at 25° C. (FIG. 13D) Lepirudin-treated mouse blood was treated with antibodies as above. Blood samples were then incubated with functionally active Coa or vWbp and coagulation time recorded.

FIGS. 14A-14F Biological effects of antibodies directed against staphylococcal coagulases. Surface plasmon resonance measurement of antibody perturbing the association between Coa or vWbp and prothrombin or fibrinogen. Response differences upon addition of coagulase (Coa) to either prothrombin (FIG. 14A) or fibrinogen (FIG. 14B) were compared to response differences in the presence of increasing amounts of antibodies (α Coa—1:1, 1:2, 1:4, 1:8). Response differences upon addition of vWbp to either prothrombin (FIG. 16A) or fibrinogen (FIG. 14B) were compared to response differences in the presence of increasing amounts of antibodies (α vWbp—1:1, 1:2, 1:4, 1:8). (FIG. 14E, F) Purified active Coa or vWbp was incubated in a 1:1 molar ratio with human prothrombin. The enzymatic ability of the complex was assessed by monitoring the rate of S-2238 cleavage (fibrinogen substitute chromogenic substrate, given in excess). The assay was repeated in presence of specific or cross antibodies added in 3M excess and the data was normalized to the % average activity without inhibition. Data are an average of three independent trials.

FIG. 15 Contribution of coagulase specific antibodies to the survival of mice with staphylococcal bacteremia. Twenty-four hours prior to infection, BALB/c mice (n=15) were injected into the peritoneum with purified rabbit antibodies (5 mg antibody/kg body weight). Animals were then challenged with 1×10^8 CFU *S. aureus* Newman injected into the retro-orbital plexus and monitored for survival. Data are representative of two independent experiments.

FIGS. 16A-16H Passive transfer of coagulase antibodies confers protection against *S. aureus* abscess formation. An experimental mock (PBS, FIGS. 18A and 18C) or purified rabbit antibodies directed against vWbp (α vWbp, FIGS. 18B and 18D), Coa (α Coa, FIGS. 18E and 18G) or both coagulases (α Coa/ α vWbp, FIGS. 18F and 18H) were injected into the peritoneal cavity of BALB/c mice (n=10)

and antibody titers analyzed by ELISA (Table 8). Passively immunized animals were infected by injecting 1×10^7 CFU *S. aureus* Newman into the retro-orbital plexus. Bacterial load and abscess formation were determined following necropsy in the kidneys of animals that had been killed five days following infection. Renal tissues were fixed with paraformaldehyde, embedded in paraffin, thin sectioned, stained with hematoxylin-eosin and histopathology images acquired by light microscopy. Data are representative of two separate experiments.

FIG. 17s A-H Immunization with coagulases protects mice against *S. aureus* abscess formation. BALB/c mice (n=15) were immunized with 50 μ g His₆-Coa, His₆-vWbp, His₆-Coa and His₆-vWbp or mock (PBS) emulsified with adjuvant on day 0 and 11 and antibody titers analyzed by ELISA on day 21 (Table 8). On day 21, animals were challenged by injecting 1×10^7 CFU *S. aureus* Newman into the retro-orbital plexus. Bacterial load and abscess formation were determined following necropsy in the kidneys of animals that had been killed five days following infection. Renal tissues were fixed with paraformaldehyde, embedded in paraffin, thin sectioned, stained with hematoxylin-eosin and histopathology images acquired by light microscopy. Data are representative of two separate experiments.

DETAILED DESCRIPTION

Staphylococcus aureus is a commensal of the human skin and nares, and the leading cause of bloodstream, skin and soft tissue infections (Klebens et al., 2007). Recent dramatic increases in the mortality of staphylococcal diseases are attributed to the spread of methicillin-resistant *S. aureus* (MRSA) strains often not susceptible to antibiotics (Kennedy et al., 2008). In a large retrospective study, the incidence of MRSA infections was 4.6% of all hospital admissions in the United States (Klebens et al., 2007). The annual health care costs for 94,300 MRSA infected individuals in the United States exceed \$2.4 billion (Klebens et al., 2007). The current MRSA epidemic has precipitated a public health crisis that needs to be addressed by development of a preventive vaccine (Boucher and Corey, 2008). To date, an FDA licensed vaccine that prevents *S. aureus* diseases is not available.

The inventors describe here the use of Protein A, a cell wall anchored surface protein of staphylococci, for the generation of variants that can serve as subunit vaccines. The pathogenesis of staphylococcal infections is initiated as bacteria invade the skin or blood stream via trauma, surgical wounds, or medical devices (Lowy, 1998). Although the invading pathogen may be phagocytosed and killed, staphylococci can also escape innate immune defenses and seed infections in organ tissues, inducing inflammatory responses that attract macrophages, neutrophils, and other phagocytes (Lowy, 1998). The responsive invasion of immune cells to the site of infection is accompanied by liquefaction necrosis as the host seeks to prevent staphylococcal spread and allow for removal of necrotic tissue debris (Lam et al., 1963). Such lesions can be observed by microscopy as hypercellular areas containing necrotic tissue, leukocytes, and a central nidus of bacteria (Lam et al., 1963). Unless staphylococcal abscesses are surgically drained and treated with antibiotics, disseminated infection and septicemia produce a lethal outcome (Sheagren, 1984).

III. STAPHYLOCOCCAL ANTIGENS

A. Staphylococcal Protein A (SpA)

All *Staphylococcus aureus* strains express the structural gene for Protein A (spa) (Jensen, 1958; Said-Salim et al., 2003), a well characterized virulence factor whose cell wall anchored surface protein product (SpA) encompasses five highly homologous immunoglobulin binding domains designated E, D, A, B, and C (Sjodahl, 1977). These domains display ~80% identity at the amino acid level, are 56 to 61 residues in length, and are organized as tandem repeats (Uhlen et al., 1984). SpA is synthesized as a precursor protein with an N-terminal YSIRK/GS (SEQ ID NO.: 70) signal peptide and a C-terminal LPXTG (SEQ ID NO.: 71) motif sorting signal (DeDent et al., 2008; Schneewind et al., 1992). Cell wall anchored Protein A is displayed in great abundance on the staphylococcal surface (DeDent et al., 2007; Sjoquist et al., 1972). Each of its immunoglobulin binding domains is composed of anti-parallel α -helices that assemble into a three helix bundle and bind the Fc domain of immunoglobulin G (IgG) (Deisenhofer, 1981; Deisenhofer et al., 1978), the VH3 heavy chain (Fab) of IgM (i.e., the B cell receptor) (Graille et al., 2000), the von Willebrand factor at its A1 domain [vWF A1 is a ligand for platelets] (O'Seaghda et al., 2006) and the tumor necrosis factor α (TNF- α) receptor I (TNFRI) (Gomez et al., 2006), which is displayed on surfaces of airway epithelia (Gomez et al., 2004; Gomez et al., 2007).

SpA impedes neutrophil phagocytosis of staphylococci through its attribute of binding the Fc component of IgG (Jensen, 1958; Uhlen et al., 1984). Moreover, SpA is able to activate intravascular clotting via its binding to von Willebrand factor A1 domains (Hartleib et al., 2000). Plasma proteins such as fibrinogen and fibronectin act as bridges between staphylococci (ClfA and ClfB) and the platelet integrin GPIIb/IIIa (O'Brien et al., 2002), an activity that is supplemented through Protein A association with vWF A1, which allows staphylococci to capture platelets via the GPIIb- α platelet receptor (Foster, 2005; O'Seaghda et al., 2006). SpA also binds TNFRI and this interaction contributes to the pathogenesis of staphylococcal pneumonia (Gomez et al., 2004). SpA activates proinflammatory signaling through TNFR1 mediated activation of TRAF2, the p38/c-Jun kinase, mitogen activate protein kinase (MAPK) and the Rel-transcription factor NF-KB. SpA binding further induces TNFR1 shedding, an activity that appears to require the TNF-converting enzyme (TACE) (Gomez et al., 2007). All of the aforementioned SpA activities are mediated through its five IgG binding domains and can be perturbed by the same amino acid substitutions, initially defined by their requirement for the interaction between Protein A and human IgG1 (Cedergren et al., 1993).

SpA also functions as a B cell superantigen by capturing the Fab region of VH3 bearing IgM, the B cell receptor (Gomez et al., 2007; Goodyear et al., 2003; Goodyear and Silverman, 2004; Roben et al., 1995). Following intravenous challenge, staphylococcal Protein A (SpA) mutations show a reduction in staphylococcal load in organ tissues and dramatically diminished ability to form abscesses (described herein). During infection with wildtype *S. aureus*, abscesses are formed within forty-eight hours and are detectable by light microscopy of hematoxylin-eosin stained, thin-sectioned kidney tissue, initially marked by an influx of polymorphonuclear leukocytes (PMNs). On day 5 of infection, abscesses increase in size and enclosed a central population of staphylococci, surrounded by a layer of eosinophilic, amorphous material and a large cuff of PMNs. Histopathology revealed massive necrosis of PMNs in proximity to the staphylococcal nidus at the center of abscess lesions as well as a mantle of healthy phagocytes. The inventors also

observed a rim of necrotic PMNs at the periphery of abscess lesions, bordering the eosinophilic pseudocapsule that separated healthy renal tissue from the infectious lesion. Staphylococcal variants lacking Protein A are unable to establish the histopathology features of abscesses and are cleared during infection.

In previous studies, Cedergren et al. (1993) engineered five individual substitutions in the Fc fragment binding sub-domain of the B domain of SpA, L17D, N28A, I31A and K35A. These authors created these proteins to test data gathered from a three dimensional structure of a complex between one domain of SpA and Fc₁. Cedergren et al. determined the effects of these mutations on stability and binding, but did not contemplate use of such substitutions for the production of a vaccine antigen.

Brown et al. (1998) describe studies designed to engineer new proteins based on SpA that allow the use of more favorable elution conditions when used as affinity ligands. The mutations studied included single mutations of Q13A, Q14H, N15A, N15H, F17H, Y18F, L21H, N32H, or K39H. Brown et al. report that Q13A, N15A, N15H, and N32H substitutions made little difference to the dissociation constant values and that the Y18F substitution resulted in a 2 fold decrease in binding affinity as compared to wild type SpA. Brown et al. also report that L21H and F17H substitutions decrease the binding affinity by five-fold and a hundred-fold respectively. The authors also studied analogous substitutions in two tandem domains. Thus, the Brown et al. studies were directed to generating a SpA with a more favorable elution profile, hence the use of His substitutions to provide a pH sensitive alteration in the binding affinity. Brown et al. is silent on the use of SpA as a vaccine antigen.

Graille et al. (2000) describe a crystal structure of domain D of SpA and the Fab fragment of a human IgM antibody. Graille et al. define by analysis of a crystal structure the D domain amino acid residues that interact with the Fab fragment as residues Q26, G29, F30, Q32, S33, D36, D37, Q40, N43, E47, or L51, as well as the amino acid residues that form the interface between the domain D sub-domains. Graille et al. define the molecular interactions of these two proteins, but is silent in regard to any use of substitutions in the interacting residues in producing a vaccine antigen.

O'Seaghda et al. (2006) describe studies directed at elucidating which sub-domain of domain D binds vWF. The authors generated single mutations in either the Fc or VH3 binding sub-domains, i.e., amino acid residues F5A, Q9A, Q10A, F13A, Y14A, L17A, N28A, I31A, K35A, G29A, F30A, S33A, D36A, D37A, Q40A, E47A, or Q32A. The authors discovered that vWF binds the same sub-domain that binds Fc. O'Seaghda et al. define the sub-domain of domain D responsible for binding vWF, but is silent in regard to any use of substitutions in the interacting residues in producing a vaccine antigen.

Gomez et al. (2006) describe the identification of residues responsible for activation of the TNFR1 by using single mutations of F5A, F13A, Y14A, L17A, N21A, I31A, Q32A, and K35A. Gomez et al. is silent in regard to any use of substitutions in the interacting residues in producing a vaccine antigen.

Recombinant affinity tagged Protein A, a polypeptide encompassing the five IgG domains (EDCAB) (Sjodahl, 1977) but lacking the C-terminal Region X (Guss et al., 1984), was purified from recombinant *E. coli* and used as a vaccine antigen (Stranger-Jones et al., 2006). Because of the attributes of SpA in binding the Fc portion of IgG, a specific humoral immune response to Protein A could not be measured (Stranger-Jones et al., 2006). The inventors have

overcome this obstacle through the generation of SpA-DQ9, 10K;D36,37A. BALB/c mice immunized with recombinant Protein A (SpA) displayed significant protection against intravenous challenge with *S. aureus* strains: a 2.951 log reduction in staphylococcal load as compared to the wild-type (P>0.005; Student's t-test) (Stranger-Jones et al., 2006). SpA specific antibodies may cause phagocytic clearance prior to abscess formation and/or impact the formation of the aforementioned eosinophilic barrier in abscesses that separate staphylococcal communities from immune cells since these do not form during infection with Protein A mutant strains. Each of the five SpA domains (i.e., domains formed from three helix bundles designated E, D, A, B, and C) exerts similar binding properties (Jansson et al., 1998). The solution and crystal structure of the domain D has been solved both with and without the Fc and VH3 (Fab) ligands, which bind Protein A in a non-competitive manner at distinct sites (Graille et al., 2000). Mutations in residues known to be involved in IgG binding (F5, Q9, Q10, S11, F13, Y14, L17, N28, I31 and K35) are also required for vWF AI and TNFR1 binding (Cedergren et al., 1993; Gomez et al., 2006; O'Seaghda et al., 2006), whereas residues important for the VH3 interaction (Q26, G29, F30, S33, D36, D37, Q40, N43, E47) appear to have no impact on the other binding activities (Graille et al., 2000; Jansson et al., 1998). SpA specifically targets a subset of B cells that express VH3 family related IgM on their surface, i.e., VH3 type B cell receptors (Roben et al., 1995). Upon interaction with SpA, these B cells proliferate and commit to apoptosis, leading to preferential and prolonged deletion of innate-like B lymphocytes (i.e., marginal zone B cells and follicular B2 cells)(Goodyear et al., 2003; Goodyear et al., 2004).

Molecular Basis of Protein A Surface Display and Function.

Protein A is synthesized as a precursor in the bacterial cytoplasm and secreted via its YSIRK signal peptide at the cross wall, i.e. the cell division septum of staphylococci (FIG. 1) (DeDent et al., 2007; DeDent et al., 2008). Following cleavage of the C-terminal LPXTG sorting signal, Protein A is anchored to bacterial peptidoglycan cross-bridges by sortase A (Mazmanian et al., 1999; Schneewind et al., 1995; Mazmanian et al., 2000). Protein A is the most abundant surface protein of staphylococci; the molecule is expressed by virtually all *S. aureus* strains (Cespedes et al., 2005; Kennedy et al., 2008; Said-Salim et al., 2003). Staphylococci turn over 15-20% of their cell wall per division cycle (Navarre and Schneewind, 1999). Murine hydrolases cleave the glycan strands and wall peptides of peptidoglycan, thereby releasing Protein A with its attached C-terminal cell wall disaccharide tetrapeptide into the extracellular medium (Ton-That et al., 1999). Thus, by physiological design, Protein A is both anchored to the cell wall and displayed on the bacterial surface but also released into surrounding tissues during host infection (Marraffini et al., 2006).

Protein A captures immunoglobulins on the bacterial surface and this biochemical activity enables staphylococcal escape from host innate and acquired immune responses (Jensen, 1958; Goodyear et al., 2004). Interestingly, region X of Protein A (Guss et al., 1984), a repeat domain that tethers the IgG binding domains to the LPXTG sorting signal/cell wall anchor, is perhaps the most variable portion of the staphylococcal genome (Said-Salim, 2003; Schneewind et al., 1992). Each of the five immunoglobulin binding domains of Protein A (SpA), formed from three helix bundles and designated E, D, A, B, and C, exerts similar structural and functional properties (Sjodahl, 1977; Jansson et al., 1998). The solution and crystal structure of the domain

D has been solved both with and without the Fc and V_H3 (Fab) ligands, which bind Protein A in a non-competitive manner at distinct sites (Graille 2000).

In the crystal structure complex, the Fab interacts with helix II and helix III of domain D via a surface composed of four VH region β -strands (Graille 2000). The major axis of helix II of domain D is approximately 50° to the orientation of the strands, and the interhelical portion of domain D is most proximal to the CO strand. The site of interaction on Fab is remote from the Ig light chain and the heavy chain constant region. The interaction involves the following domain D residues: Asp-36 of helix II, Asp-37 and Gln-40 in the loop between helix II and helix III and several other residues (Graille 2000). Both interacting surfaces are composed predominantly of polar side chains, with three negatively charged residues on domain D and two positively charged residues on the 2A2 Fab buried by the interaction, providing an overall electrostatic attraction between the two molecules. Of the five polar interactions identified between Fab and domain D, three are between side chains. A salt bridge is formed between Arg-H19 and Asp-36 and two hydrogen bonds are made between Tyr-H59 and Asp-37 and between Asn-H82a and Ser-33. Because of the conservation of Asp-36 and Asp-37 in all five IgG binding domains of Protein A, the inventors mutated these residues.

The SpA-D sites responsible for Fab binding are structurally separate from the domain surface that mediates Fc γ binding. The interaction of Fc γ with domain D primarily involves residues in helix I with lesser involvement of helix II (Gouda et al., 1992; Deisenhofer, 1981). With the exception of the Gln-32, a minor contact in both complexes, none of the residues that mediate the Fc γ interaction are involved in Fab binding. To examine the spatial relationship between these different Ig-binding sites, the SpA domains in these complexes have been superimposed to construct a model of a complex between Fab, the SpA-domain D, and the Fc γ molecule. In this ternary model, Fab and Fc γ form a sandwich about opposite faces of the helix II without evidence of steric hindrance of either interaction. These findings illustrate how, despite its small size (i.e., 56-61 aa), an SpA domain can simultaneously display both activities, explaining experimental evidence that the interactions of Fab with an individual domain are noncompetitive. Residues for the interaction between SpA-D and Fc γ are Gln-9 and Gln-10.

In contrast, occupancy of the Fc portion of IgG on the domain D blocks its interaction with vWF A1 and probably also TNFR1 (O'Seaghda et al., 2006). Mutations in residues essential for IgG Fc binding (F5, Q9, Q10, S11, F13, Y14, L17, N28, I31 and K35) are also required for vWF A1 and TNFR1 binding (O'Seaghda et al., 2006; Cedergren et al., 1993; Gomez et al., 2006), whereas residues critical for the VH3 interaction (Q26, G29, F30, S33, D36, D37, Q40, N43, E47) have no impact on the binding activities of IgG Fc, vWF A1 or TNFR1 (Jansson et al., 1998; Graille et al., 2000). The Protein A immunoglobulin Fab binding activity targets a subset of B cells that express V_H3 family related IgM on their surface, i.e., these molecules function as VH3type B cell receptors (Roben et al., 1995). Upon interaction with SpA, these B cells rapidly proliferate and then commit to apoptosis, leading to preferential and prolonged deletion of innate-like B lymphocytes (i.e., marginal zone B cells and follicular B2 cells) (Goodyear and Silverman, 2004; Goodyear and Silverman, 2003). More than 40% of circulating B cells are targeted by the Protein A interaction and the V_H3 family represents the largest family of human B cell receptors to impart protective humoral responses against pathogens (Goodyear and Silverman, 2004; Good-

year and Silverman, 2003). Thus, Protein A functions analogously to staphylococcal superantigens (Roben et al., 1995), albeit that the latter class of molecules, for example SEB, TSST-1, TSST-2, form complexes with the T cell receptor to inappropriately stimulate host immune responses and thereby precipitating characteristic disease features of staphylococcal infections (Roben et al., 1995; Tiedemann et al., 1995). Together these findings document the contributions of Protein A in establishing staphylococcal infections and in modulating host immune responses.

In sum, Protein A domains can viewed as displaying two different interfaces for binding with host molecules and any development of Protein A based vaccines must consider the generation of variants that do not perturb host cell signaling, platelet aggregation, sequestration of immunoglobulins or the induction of B cell proliferation and apoptosis. Such Protein A variants should also be useful in analyzing vaccines for the ability of raising antibodies that block the aforementioned SpA activities and occupy the five repeat domains at their dual binding interfaces. This goal is articulated and pursued here for the first time and methods are described in detail for the generation of Protein A variants that can be used as a safe vaccine for humans. To perturb IgG Fc γ , vWF A1 and TNFR1 binding, glutamine (Q) 9 and 10 [numbering derived from the SpA domain D as described in Uhlen et al., 1984] were mutated, and generated lysine substitutions for both glutamines with the expectation that these abolish the ligand attributes at the first binding interface. To perturb IgM Fab VH3 binding, aspartate (D) 36 and 37 were mutated, each of which is required for the association with the B cell receptor. D36 and D37 were both substituted with alanine. Q9,10K and D36,37A mutations are here combined in the recombinant molecule SpA-DQ9,10K;D36,37A and tested for the binding attributes of Protein A. Further, SpA-D and SpA-DQ9,10K;D36,37A are subjected to immunization studies in mice and rabbits and analyzed for [1] the production of specific antibodies (SpA-D Ab); [2] the ability of SpA-D Ab to block the association between Protein A and its four different ligands; and, [3] the attributes of SpA-D Ab to generate protective immunity against staphylococcal infections. (See Examples section below).

B. Staphylococcal Coagulases

Coagulases are enzymes produced by *Staphylococcus* bacteria that convert fibrinogen to fibrin. Coa and vW_h activate prothrombin without proteolysis (Friedrich et al., 2003). The coagulase•prothrombin complex recognizes fibrinogen as a specific substrate, converting it directly into fibrin. The crystal structure of the active complex revealed binding of the D1 and D2 domains to prothrombin and insertion of its Ile1-Val² N-terminus into the Ile¹⁶ pocket, inducing a functional active site in the zymogen through conformational change (Friedrich et al., 2003). Exosite I of α -thrombin, the fibrinogen recognition site, and proexosite I on prothrombin are blocked by the D2 of Coa (Friedrich et al., 2003). Nevertheless, association of the tetrameric (Coa•prothrombin)₂ complex binds fibrinogen at a new site with high affinity (Panizzi et al., 2006). This model explains the coagulant properties and efficient fibrinogen conversion by coagulase (Panizzi et al., 2006).

Fibrinogen is a large glycoprotein (Mr~340,000), formed by three pairs of α -, β -, and γ -chains covalently linked to form a "dimer of trimers," where A and B designate the fibrinopeptides released by thrombin cleavage (Panizzi et al., 2006). The elongated molecule folds into three separate domains, a central fragment E that contains the N-termini of all six chains and two flanking fragments D formed mainly

by the C-termini of the β - and γ -chains. These globular domains are connected by long triple-helical structures. Coagulase-prothrombin complexes, which convert human fibrinogen to the self-polymerizing fibrin, are not targeted by circulating thrombin inhibitors (Panizzi et al., 2006). Thus, staphylococcal coagulases bypass the physiological blood coagulation pathway.

All *S. aureus* strains secrete coagulase and vWbp (Bjerketorp et al., 2004; Field and Smith, 1945). Although early work reported important contributions of coagulase to the pathogenesis of staphylococcal infections (Ekstedt and Yotis, 1960; Smith et al., 1947), more recent investigations with molecular genetics tools challenged this view by observing no virulence phenotypes with endocarditis, skin abscess and mastitis models in mice (Moreillon et al., 1995; Phonimdaeng et al., 1990). Generating isogenic variants of *S. aureus* Newman, a fully virulent clinical isolate (Duthie et al., 1952), it is described herein that coa mutants indeed display virulence defects in a lethal bacteremia and renal abscess model in mice. In the inventors experience, *S. aureus* 8325-4 is not fully virulent and it is presumed that mutational lesions in this strain may not be able to reveal virulence defects in vivo. Moreover, antibodies raised against Coa or vWbp perturb the pathogenesis of *S. aureus* Newman infections to a degree mirroring the impact of gene deletions. Coa and vWbp contribute to staphylococcal abscess formation and lethal bacteremia and may also function as protective antigens in subunit vaccines.

Biochemical studies document the biological value of antibodies against Coa and vWbp. By binding to antigen and blocking its association with clotting factors, the antibodies prevent the formation of Coa prothrombin and vWbp-prothrombin complexes. Passive transfer studies revealed protection of experimental animals against staphylococcal abscess formation and lethal challenge by Coa and vWbp antibodies. Thus, Coa and vWbp neutralizing antibodies generate immune protection against staphylococcal disease.

Earlier studies revealed a requirement of coagulase for resisting phagocytosis in blood (Smith et al., 1947) and the inventors observed a similar phenotype for Δ coa mutants in lepirudin-treated mouse blood (see Example 3 below). As vWbp displays higher affinity for human prothrombin than the mouse counterpart, it is suspected the same may be true for Δ vWbp variants in human blood. Further, expression of Coa and vWbp in abscess lesions as well as their striking distribution in the eosinophilic pseudocapsule surrounding (staphylococcal abscess communities (SACs) or the peripheral fibrin wall, suggest that secreted coagulases contribute to the establishment of these lesions. This hypothesis was tested and, indeed, Δ coa mutants were defective in the establishment of abscesses. A corresponding test, blocking Coa function with specific antibodies, produced the same effect. Consequently, it is proposed that the clotting of fibrin is a critical event in the establishment of staphylococcal abscesses that can be targeted for the development of protective vaccines. Due to their overlapping function on human prothrombin, both Coa and vWbp are considered excellent candidates for vaccine development.

C. Other Staphylococcal Antigens

Research over the past several decades identified *S. aureus* exotoxins, surface proteins and regulatory molecules as important virulence factors (Foster, 2005; Mazmanian et al., 2001; Novick, 2003). Much progress has been achieved regarding the regulation of these genes. For example, staphylococci perform a bacterial census via the secretion of auto-inducing peptides that bind to a cognate receptor at

threshold concentration, thereby activating phospho-relay reactions and transcriptional activation of many of the exotoxin genes (Novick, 2003). The pathogenesis of staphylococcal infections relies on these virulence factors (secreted exotoxins, exopolysaccharides, and surface adhesins). The development of staphylococcal vaccines is hindered by the multifaceted nature of staphylococcal invasion mechanisms. It is well established that live attenuated microorganisms are highly effective vaccines; immune responses elicited by such vaccines are often of greater magnitude and of longer duration than those produced by non-replicating immunogens. One explanation for this may be that live attenuated strains establish limited infections in the host and mimic the early stages of natural infection. Embodiments of the invention are directed to compositions and methods including variant SpA polypeptides and peptides, as well as other immunogenic extracellular proteins, polypeptides, and peptides (including both secreted and cell surface proteins or peptides) of gram positive bacteria for the use in mitigating or immunizing against infection. In particular embodiments the bacteria is a *staphylococcus* bacteria. Extracellular proteins, polypeptides, or peptides include, but are not limited to secreted and cell surface proteins of the targeted bacteria.

The human pathogen *S. aureus* secretes EsxA and EsxB, two ESAT-6 like proteins, across the bacterial envelope (Burts et al., 2005, which is incorporated herein by reference). Staphylococcal esxA and esxB are clustered with six other genes in the order of transcription: esxA esaA essA esaB essB essC esaC esxB. The acronyms esa, ess, and esx stand for ESAT-6 secretion accessory, system, and extracellular, respectively, depending whether the encoded proteins play an accessory (esa) or direct (ess) role for secretion, or are secreted (esx) in the extracellular milieu. The entire cluster of eight genes is herein referred to as the Ess cluster. EsxA, esxB, essA, essB, and essC are all required for synthesis or secretion of EsxA and EsxB. Mutants that fail to produce EsxA, EsxB, and EssC display defects in the pathogenesis of *S. aureus* murine abscesses, suggesting that this specialized secretion system may be a general strategy of human bacterial pathogenesis. Secretion of non-WXG100 substrates by the ESX-1 pathway has been reported for several antigens including EspA, EspB, Rv3483c, and Rv3615c (Fortune et al., 2005; MacGurn et al., 2005; McLaughlin et al., 2007; Xu et al., 2007). The alternate ESX-5 pathway has also been shown to secrete both WXG100 and non-WXG100 proteins in pathogenic mycobacteria (Abdallah et al., 2007; Abdallah et al., 2006).

The *Staphylococcus aureus* Ess pathway can be viewed as a secretion module equipped with specialized transport components (Ess), accessory factors (Esa) and cognate secretion substrates (Esx). EssA, EssB and EssC are required for EsxA and EsxB secretion. Because EssA, EssB and EssC are predicted to be transmembrane proteins, it is contemplated that these proteins form a secretion apparatus. Some of the proteins in the ess gene cluster may actively transport secreted substrates (acting as motor) while others may regulate transport (regulator). Regulation may be achieved, but need not be limited to, transcriptional or post-translational mechanisms for secreted polypeptides, sorting of specific substrates to defined locations (e.g., extracellular medium or host cells), or timing of secretion events during infection. At this point, it is unclear whether all secreted Esx proteins function as toxins or contribute indirectly to pathogenesis.

Staphylococci rely on surface protein mediated-adhesion to host cells or invasion of tissues as a strategy for escape from immune defenses. Furthermore, *S. aureus* utilize sur-

face proteins to sequester iron from the host during infection. The majority of surface proteins involved in staphylococcal pathogenesis carry C-terminal sorting signals, i.e., they are covalently linked to the cell wall envelope by sortase. Further, staphylococcal strains lacking the genes required for surface protein anchoring, i.e., sortase A and B, display a dramatic defect in the virulence in several different mouse models of disease. Thus, surface protein antigens represent a validated vaccine target as the corresponding genes are essential for the development of staphylococcal disease and can be exploited in various embodiments of the invention. The sortase enzyme superfamily are Gram-positive transpeptidases responsible for anchoring surface protein virulence factors to the peptidoglycan cell wall layer. Two sortase isoforms have been identified in *Staphylococcus aureus*, SrtA and SrtB. These enzymes have been shown to recognize a LPXTG motif in substrate proteins. The SrtB isoform appears to be important in heme iron acquisition and iron homeostasis, whereas the SrtA isoform plays a critical role in the pathogenesis of Gram-positive bacteria by modulating the ability of the bacterium to adhere to host tissue via the covalent anchoring of adhesins and other proteins to the cell wall peptidoglycan. In certain embodiments the SpA variants described herein can be used in combination with other staphylococcal proteins such as Coa, Eap, Ebh, Emp, EsaC, EsaB, EsxA, EsxB, Hla, SdrC, SdrD, SdrE, IsdA, IsdB, ClfA, ClfB, IsdC, SasF, vWbp, and/or vWh proteins.

Certain aspects of the invention include methods and compositions concerning proteinaceous compositions including polypeptides, peptides, or nucleic acid encoding SpA variant(s) and other staphylococcal antigens such as other proteins transported by the Ess pathway, or sortase substrates. These proteins may be modified by deletion, insertion, and/or substitution.

The Esx polypeptides include the amino acid sequence of Esx proteins from bacteria in the *Staphylococcus* genus. The Esx sequence may be from a particular *staphylococcus* species, such as *Staphylococcus aureus*, and may be from a particular strain, such as Newman. In certain embodiments, the EsxA sequence is SAV0282 from strain Mu50 (which is the same amino acid sequence for Newman) and can be accessed using Genbank Accession Number Q99WU4 (gi|68565539), which is hereby incorporated by reference. In other embodiments, the EsxB sequence is SAV0290 from strain Mu50 (which is the same amino acid sequence for Newman) and can be accessed using Genbank Accession Number Q99WT7 (gi|68565532), which is hereby incorporated by reference. In further embodiments, other polypeptides transported by the Ess pathway may be used, the sequences of which may be identified by one of skill in the art using databases and internet accessible resources.

The sortase substrate polypeptides include, but are not limited to the amino acid sequence of SdrC, SdrD, SdrE, IsdA, IsdB, ClfA, ClfB, IsdC or SasF proteins from bacteria in the *Staphylococcus* genus. The sortase substrate polypeptide sequence may be from a particular *staphylococcus* species, such as *Staphylococcus aureus*, and may be from a particular strain, such as Newman. In certain embodiments, the SdrD sequence is from strain N315 and can be accessed using Genbank Accession Number NP_373773.1 (gi|15926240), which is incorporated by reference. In other embodiments, the SdrE sequence is from strain N315 and can be accessed using Genbank Accession Number NP_373774.1 (gi|15926241), which is incorporated by reference. In other embodiments, the IsdA sequence is SAV1130 from strain Mu50 (which is the same amino acid sequence for Newman) and can be accessed using Genbank

Accession Number NP_371654.1 (gi|15924120), which is incorporated by reference. In other embodiments, the IsdB sequence is SAV1129 from strain Mu50 (which is the same amino acid sequence for Newman) and can be accessed using Genbank Accession Number NP_371653.1 (gi|15924119), which is incorporated by reference. In further embodiments, other polypeptides transported by the Ess pathway or processed by sortase may be used, the sequences of which may be identified by one of skill in the art using databases and internet accessible resources.

Examples of various proteins that can be used in the context of the present invention can be identified by analysis of database submissions of bacterial genomes, including but not limited to accession numbers NC_002951 (GI:57650036 and GenBank CP000046), NC_002758 (GI:57634611 and GenBank BA000017), NC_002745 (GI:29165615 and GenBank BA000018), NC_003923 (GI:21281729 and GenBank BA000033), NC_002952 (GI:49482253 and GenBank BX571856), NC_002953 (GI:49484912 and GenBank BX571857), NC_007793 (GI:87125858 and GenBank CP000255), NC_007795 (GI:87201381 and GenBank CP000253) each of which are incorporated by reference.

As used herein, a "protein" or "polypeptide" refers to a molecule comprising at least ten amino acid residues. In some embodiments, a wild-type version of a protein or polypeptide are employed, however, in many embodiments of the invention, a modified protein or polypeptide is employed to generate an immune response. The terms described above may be used interchangeably. A "modified protein" or "modified polypeptide" or a "variant" refers to a protein or polypeptide whose chemical structure, particularly its amino acid sequence, is altered with respect to the wild-type protein or polypeptide. In some embodiments, a modified/variant protein or polypeptide has at least one modified activity or function (recognizing that proteins or polypeptides may have multiple activities or functions). It is specifically contemplated that a modified/variant protein or polypeptide may be altered with respect to one activity or function yet retain a wild-type activity or function in other respects, such as immunogenicity.

In certain embodiments the size of a protein or polypeptide (wild-type or modified) may comprise, but is not limited to, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 21, 22, 23, 24, 25, 26, 27, 28, 29, 30, 31, 32, 33, 34, 35, 36, 37, 38, 39, 40, 41, 42, 43, 44, 45, 46, 47, 48, 49, 50, 51, 52, 53, 54, 55, 56, 57, 58, 59, 60, 61, 62, 63, 64, 65, 66, 67, 68, 69, 70, 71, 72, 73, 74, 75, 76, 77, 78, 79, 80, 81, 82, 83, 84, 85, 86, 87, 88, 89, 90, 91, 92, 93, 94, 95, 96, 97, 98, 99, 100, 110, 120, 130, 140, 150, 160, 170, 180, 190, 200, 210, 220, 230, 240, 250, 275, 300, 325, 350, 375, 400, 425, 450, 475, 500, 525, 550, 575, 600, 625, 650, 675, 700, 725, 750, 775, 800, 825, 850, 875, 900, 925, 950, 975, 1000, 1100, 1200, 1300, 1400, 1500, 1750, 2000, 2250, 2500 amino molecules or greater, and any range derivable therein, or derivative of a corresponding amino sequence described or referenced herein. It is contemplated that polypeptides may be mutated by truncation, rendering them shorter than their corresponding wild-type form, but also they might be altered by fusing or conjugating a heterologous protein sequence with a particular function (e.g., for targeting or localization, for enhanced immunogenicity, for purification purposes, etc.).

As used herein, an "amino molecule" refers to any amino acid, amino acid derivative, or amino acid mimic known in the art. In certain embodiments, the residues of the proteinaceous molecule are sequential, without any non-amino molecule interrupting the sequence of amino molecule residues. In other embodiments, the sequence may comprise one or

more non-amino molecule moieties. In particular embodiments, the sequence of residues of the proteinaceous molecule may be interrupted by one or more non-amino molecule moieties.

Accordingly, the term "proteinaceous composition" encompasses amino molecule sequences comprising at least one of the 20 common amino acids in naturally synthesized proteins, or at least one modified or unusual amino acid.

Proteinaceous compositions may be made by any technique known to those of skill in the art, including (i) the expression of proteins, polypeptides, or peptides through standard molecular biological techniques, (ii) the isolation of proteinaceous compounds from natural sources, or (iii) the chemical synthesis of proteinaceous materials. The nucleotide as well as the protein, polypeptide, and peptide sequences for various genes have been previously disclosed, and may be found in the recognized computerized databases. One such database is the National Center for Biotechnology Information's Genbank and GenPept databases (on the World Wide Web at ncbi.nlm.nih.gov/). The coding regions for these genes may be amplified and/or expressed using the techniques disclosed herein or as would be known to those of ordinary skill in the art.

Amino acid sequence variants of SpA, coagulases and other polypeptides of the invention can be substitutional, insertional, or deletion variants. A variation in a polypeptide of the invention may affect 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 21, 22, 23, 24, 25, 26, 27, 28, 29, 30, 31, 32, 33, 34, 35, 36, 37, 38, 39, 40, 41, 42, 43, 44, 45, 46, 47, 48, 49, 50, or more non-contiguous or contiguous amino acids of the polypeptide, as compared to wild-type. A variant can comprise an amino acid sequence that is at least 50%, 60%, 70%, 80%, or 90%, including all values and ranges there between, identical to any sequence provided or referenced herein, e.g., SEQ ID NO:2-8 or SEQ ID NO:11-30, A variant can include 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13,

from any *staphylococcus* species and strain are contemplated for use in compositions and methods described herein.

Deletion variants typically lack one or more residues of the native or wild-type protein. Individual residues can be deleted or a number of contiguous amino acids can be deleted. A stop codon may be introduced (by substitution or insertion) into an encoding nucleic acid sequence to generate a truncated protein. Insertional mutants typically involve the addition of material at a non-terminal point in the polypeptide. This may include the insertion of one or more residues. Terminal additions, called fusion proteins, may also be generated. These fusion proteins include multimers or concatamers of one or more peptide or polypeptide described or referenced herein.

Substitutional variants typically contain the exchange of one amino acid for another at one or more sites within the protein, and may be designed to modulate one or more properties of the polypeptide, with or without the loss of other functions or properties. Substitutions may be conservative, that is, one amino acid is replaced with one of similar shape and charge. Conservative substitutions are well known in the art and include, for example, the changes of: alanine to serine; arginine to lysine; asparagine to glutamine or histidine; aspartate to glutamate; cysteine to serine; glutamine to asparagine; glutamate to aspartate; glycine to proline; histidine to asparagine or glutamine; isoleucine to leucine or valine; leucine to valine or isoleucine; lysine to arginine; methionine to leucine or isoleucine; phenylalanine to tyrosine, leucine or methionine; serine to threonine; threonine to serine; tryptophan to tyrosine; tyrosine to tryptophan or phenylalanine; and valine to isoleucine or leucine. Alternatively, substitutions may be non-conservative such that a function or activity of the polypeptide is affected. Non-conservative changes typically involve substituting a residue with one that is chemically dissimilar, such as a polar or charged amino acid for a nonpolar or uncharged amino acid, and vice versa.

TABLE 1

Exemplary surface proteins of <i>S. aureus</i> strains.								
SAV #	SA#	Surface	MW2	Mu50	N315	Newman	MRSA252*	MSSA476*
SAV0111	SA0107	Spa	492	450	450	520	516	492
SAV2503	SA2291	FnBPA	1015	1038	1038	741	—	1015
SAV2502	SA2290	FnBPB	943	961	961	677	965	957
SAV0811	SA0742	ClfA	946	935	989	933	1029	928
SAV2630	SA2423	ClfB	907	877	877	913	873	905
Np	Np	Cna	1183	—	—	—	1183	1183
SAV0561	SA0519	SdrC	955	953	953	947	906	957
SAV0562	SA0520	SdrD	1347	1385	1385	1315	—	1365
SAV0563	SA0521	SdrE	1141	1141	1141	1166	1137	1141
Np	Np	Pls	—	—	—	—	—	—
SAV2654	SA2447	SasA	2275	2271	2271	2271	1351	2275
SAV2160	SA1964	SasB	686	2481	2481	2481	2222	685
	SA1577	SasC	2186	213	2186	2186	2189	2186
SAV0134	SA0129	SasD	241	241	241	241	221	241
SAV1130	SA0977	SasE/IsdA	350	350	350	350	354	350
SAV2646	SA2439	SasF	635	635	635	635	627	635
SAV2496		SasG	1371	525	927	—	—	1371
SAV0023	SA0022	SasH	772	—	772	772	786	786
SAV1731	SA1552	SasI	895	891	891	891	534	895
SAV1129	SA0976	SasJ/IsdB	645	645	645	645	652	645
	SA2381	SasK	198	211	211	—	—	197
	Np	SasL	—	232	—	—	—	—
SAV1131	SA0978	IsdC	227	227	227	227	227	227

14, 15, 16, 17, 18, 19, 20, or more substitute amino acids. A polypeptide processed or secreted by the Ess pathway or other surface proteins (see Table 1) or sortase substrates

Proteins of the invention may be recombinant, or synthesized in vitro. Alternatively, a non-recombinant or recombinant protein may be isolated from bacteria. It is also

contemplated that a bacteria containing such a variant may be implemented in compositions and methods of the invention. Consequently, a protein need not be isolated.

The term "functionally equivalent codon" is used herein to refer to codons that encode the same amino acid, such as the six codons for arginine or serine, and also refers to codons that encode biologically equivalent amino acids (see Table 2, below).

TABLE 2

Codon Table			
Amino Acids		Codons	
Alanine	Ala	A	GCA GCC GCG GCU
Cysteine	Cys	C	UGC UGU
Aspartic acid	Asp	D	GAC GAU
Glutamic acid	Glu	E	GAA GAG
Phenylalanine	Phe	F	UUC UUU
Glycine	Gly	G	GGA GGC GGG GGU
Histidine	His	H	CAC CAU
Isoleucine	Ile	I	AUA AUC AUU
Lysine	Lys	K	AAA AAG
Leucine	Leu	L	UUA UUG CUA CUC CUG CUU
Methionine	Met	M	AUG
Asparagine	Asn	N	AAC AAU
Proline	Pro	P	CCA CCC CCG CCU
Glutamine	Gln	Q	CAA CAG
Arginine	Arg	R	AGA AGG CGA CGC CGG CGU
Serine	Ser	S	AGC AGU UCA UCC UCG UCU
Threonine	Thr	T	ACA ACC ACG ACU
Valine	Val	V	GUA GUC GUG GUU
Tryptophan	Trp	W	UGG
Tyrosine	Tyr	Y	UAC UAU

It also will be understood that amino acid and nucleic acid sequences may include additional residues, such as additional N- or C-terminal amino acids, or 5' or 3' sequences, respectively, and yet still be essentially as set forth in one of the sequences disclosed herein, so long as the sequence meets the criteria set forth above, including the maintenance of biological protein activity (e.g., immunogenicity) where protein expression is concerned. The addition of terminal sequences particularly applies to nucleic acid sequences that may, for example, include various non-coding sequences flanking either of the 5' or 3' portions of the coding region.

The following is a discussion based upon changing of the amino acids of a protein to create a variant polypeptide or peptide. For example, certain amino acids may be substituted for other amino acids in a protein structure with or without appreciable loss of interactive binding capacity with structures such as, for example, antigen-binding regions of antibodies or binding sites on substrate molecules. Since it is the interactive capacity and nature of a protein that defines that protein's functional activity, certain amino acid substi-

tutions can be made in a protein sequence, and in its underlying DNA coding sequence, and nevertheless produce a protein with a desirable property. It is thus contemplated by the inventors that various changes may be made in the DNA sequences of genes.

It is contemplated that in compositions of the invention, there is between about 0.001 mg and about 10 mg of total polypeptide, peptide, and/or protein per ml. The concentration of protein in a composition can be about, at least about or at most about 0.001, 0.010, 0.050, 0.1, 0.2, 0.3, 0.4, 0.5, 0.6, 0.7, 0.8, 0.9, 1.0, 1.5, 2.0, 2.5, 3.0, 3.5, 4.0, 4.5, 5.0, 5.5, 6.0, 6.5, 7.0, 7.5, 8.0, 8.5, 9.0, 9.5, 10.0 mg/ml or more (or any range derivable therein). Of this, about, at least about, or at most about 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 21, 22, 23, 24, 25, 26, 27, 28, 29, 30, 31, 32, 33, 34, 35, 36, 37, 38, 39, 40, 41, 42, 43, 44, 45, 46, 47, 48, 49, 50, 51, 52, 53, 54, 55, 56, 57, 58, 59, 60, 61, 62, 63, 64, 65, 66, 67, 68, 69, 70, 71, 72, 73, 74, 75, 76, 77, 78, 79, 80, 81, 82, 83, 84, 85, 86, 87, 88, 89, 90, 91, 92, 93, 94, 95, 96, 97, 98, 99, 100% may be an SpA variant or a coagulase, and may be used in combination with other peptides or polypeptides, such as other bacterial peptides and/or antigens.

The present invention contemplates the administration of variant SpA polypeptides or peptides to effect a preventative therapy or therapeutic effect against the development of a disease or condition associated with infection by a *staphylococcus* pathogen.

In certain aspects, combinations of staphylococcal antigens are used in the production of an immunogenic composition that is effective at treating or preventing staphylococcal infection. Staphylococcal infections progress through several different stages. For example, the staphylococcal life cycle involves commensal colonization, initiation of infection by accessing adjoining tissues or the bloodstream, and/or anaerobic multiplication in the blood. The interplay between *S. aureus* virulence determinants and the host defense mechanisms can induce complications such as endocarditis, metastatic abscess formation, and sepsis syndrome. Different molecules on the surface of the bacterium are involved in different steps of the infection cycle. Combinations of certain antigens can elicit an immune response which protects against multiple stages of staphylococcal infection. The effectiveness of the immune response can be measured either in animal model assays and/or using an opsonophagocytic assay.

D. Polypeptides and Polypeptide Production

The present invention describes polypeptides, peptides, and proteins and immunogenic fragments thereof for use in various embodiments of the present invention. For example, specific polypeptides are assayed for or used to elicit an immune response. In specific embodiments, all or part of the proteins of the invention can also be synthesized in solution or on a solid support in accordance with conventional techniques. Various automatic synthesizers are commercially available and can be used in accordance with known protocols. See, for example, Stewart and Young, (1984); Tam et al., (1983); Merrifield, (1986); and Barany and Merrifield (1979), each incorporated herein by reference.

Alternatively, recombinant DNA technology may be employed wherein a nucleotide sequence which encodes a peptide of the invention is inserted into an expression vector, transformed or transfected into an appropriate host cell and cultivated under conditions suitable for expression.

One embodiment of the invention includes the use of gene transfer to cells, including microorganisms, for the production and/or presentation of polypeptides or peptides. The

gene for the polypeptide or peptide of interest may be transferred into appropriate host cells followed by culture of cells under the appropriate conditions. The generation of recombinant expression vectors, and the elements included therein, are well known in the art and briefly discussed herein. Alternatively, the protein to be produced may be an endogenous protein normally synthesized by the cell that is isolated and purified.

Another embodiment of the present invention uses autologous B lymphocyte cell lines, which are transfected with a viral vector that expresses an immunogen product, and more specifically, a protein having immunogenic activity. Other examples of mammalian host cell lines include, but are not limited to Vero and HeLa cells, other B- and T-cell lines, such as CEM, 721.221, H9, Jurkat, Raji, as well as cell lines of Chinese hamster ovary, W138, BHK, COS-7, 293, HepG2, 3T3, RIN and MDCK cells. In addition, a host cell strain may be chosen that modulates the expression of the inserted sequences, or that modifies and processes the gene product in the manner desired. Such modifications (e.g., glycosylation) and processing (e.g., cleavage) of protein products may be important for the function of the protein. Different host cells have characteristic and specific mechanisms for the post-translational processing and modification of proteins. Appropriate cell lines or host systems can be chosen to ensure the correct modification and processing of the foreign protein expressed.

A number of selection systems may be used including, but not limited to HSV thymidine kinase, hypoxanthine-guanine phosphoribosyltransferase, and adenine phosphoribosyltransferase genes, in tk-, hgp^rt- or ap^rt-cells, respectively. Also, anti-metabolite resistance can be used as the basis of selection: for dhfr, which confers resistance to trimethoprim and methotrexate; gpt, which confers resistance to mycophenolic acid; neo, which confers resistance to the aminoglycoside G418; and hyg^r, which confers resistance to hygromycin.

Animal cells can be propagated in vitro in two modes: as non-anchorage-dependent cells growing in suspension throughout the bulk of the culture or as anchorage-dependent cells requiring attachment to a solid substrate for their propagation (i.e., a monolayer type of cell growth).

Non-anchorage dependent or suspension cultures from continuous established cell lines are the most widely used means of large scale production of cells and cell products. However, suspension cultured cells have limitations, such as tumorigenic potential and lower protein production than adherent cells.

Where a protein is specifically mentioned herein, it is preferably a reference to a native or recombinant protein or optionally a protein in which any signal sequence has been removed. The protein may be isolated directly from the staphylococcal strain or produced by recombinant DNA techniques. Immunogenic fragments of the protein may be incorporated into the immunogenic composition of the invention. These are fragments comprising at least 10 amino acids, 20 amino acids, 30 amino acids, 40 amino acids, 50 amino acids, or 100 amino acids, including all values and ranges there between, taken contiguously from the amino acid sequence of the protein. In addition, such immunogenic fragments are immunologically reactive with antibodies generated against the Staphylococcal proteins or with antibodies generated by infection of a mammalian host with Staphylococci. Immunogenic fragments also include fragments that when administered at an effective dose, (either alone or as a hapten bound to a carrier), elicit a protective or therapeutic immune response against Staphylococcal infec-

tion, in certain aspects it is protective against *S. aureus* and/or *S. epidermidis* infection. Such an immunogenic fragment may include, for example, the protein lacking an N-terminal leader sequence, and/or a transmembrane domain and/or a C-terminal anchor domain. In a preferred aspect the immunogenic fragment according to the invention comprises substantially all of the extracellular domain of a protein which has at least 80% identity, at least 85% identity, at least 90% identity, at least 95% identity, or at least 97-99% identity, including all values and ranges there between, to a sequence selected segment of a polypeptide described or referenced herein.

Also included in immunogenic compositions of the invention are fusion proteins composed of one or more Staphylococcal proteins, or immunogenic fragments of staphylococcal proteins. Such fusion proteins may be made recombinantly and may comprise one portion of at least 1, 2, 3, 4, 5, or 6 staphylococcal proteins or segments. Alternatively, a fusion protein may comprise multiple portions of at least 1, 2, 3, 4 or 5 staphylococcal proteins. These may combine different Staphylococcal proteins and/or multiples of the same protein or protein fragment, or immunogenic fragments in the same protein (forming a multimer or a concatamer). Alternatively, the invention also includes individual fusion proteins of Staphylococcal proteins or immunogenic fragments thereof, as a fusion protein with heterologous sequences such as a provider of T-cell epitopes or purification tags, for example: β -galactosidase, glutathione-S-transferase, green fluorescent proteins (GFP), epitope tags such as FLAG, myc tag, poly histidine, or viral surface proteins such as influenza virus haemagglutinin, or bacterial proteins such as tetanus toxoid, diphtheria toxoid, or CRM197.

IV. NUCLEIC ACIDS

In certain embodiments, the present invention concerns recombinant polynucleotides encoding the proteins, polypeptides, peptides of the invention. The nucleic acid sequences for SpA, coagulases and other bacterial proteins are included, all of which are incorporated by reference, and can be used to prepare peptides or polypeptides.

As used in this application, the term "polynucleotide" refers to a nucleic acid molecule that either is recombinant or has been isolated free of total genomic nucleic acid. Included within the term "polynucleotide" are oligonucleotides (nucleic acids of 100 residues or less in length), recombinant vectors, including, for example, plasmids, cosmids, phage, viruses, and the like. Polynucleotides include, in certain aspects, regulatory sequences, isolated substantially away from their naturally occurring genes or protein encoding sequences. Polynucleotides may be single-stranded (coding or antisense) or double-stranded, and may be RNA, DNA (genomic, cDNA or synthetic), analogs thereof, or a combination thereof. Additional coding or non-coding sequences may, but need not, be present within a polynucleotide.

In this respect, the term "gene," "polynucleotide," or "nucleic acid" is used to refer to a nucleic acid that encodes a protein, polypeptide, or peptide (including any sequences required for proper transcription, post-translational modification, or localization). As will be understood by those in the art, this term encompasses genomic sequences, expression cassettes, cDNA sequences, and smaller engineered nucleic acid segments that express, or may be adapted to express, proteins, polypeptides, domains, peptides, fusion proteins, and mutants. A nucleic acid encoding all or part of a

polypeptide may contain a contiguous nucleic acid sequence of: 10, 20, 30, 40, 50, 60, 70, 80, 90, 100, 110, 120, 130, 140, 150, 160, 170, 180, 190, 200, 210, 220, 230, 240, 250, 260, 270, 280, 290, 300, 310, 320, 330, 340, 350, 360, 370, 380, 390, 400, 410, 420, 430, 440, 441, 450, 460, 470, 480, 490, 500, 510, 520, 530, 540, 550, 560, 570, 580, 590, 600, 610, 620, 630, 640, 650, 660, 670, 680, 690, 700, 710, 720, 730, 740, 750, 760, 770, 780, 790, 800, 810, 820, 830, 840, 850, 860, 870, 880, 890, 900, 910, 920, 930, 940, 950, 960, 970, 980, 990, 1000, 1010, 1020, 1030, 1040, 1050, 1060, 1070, 1080, 1090, 1095, 1100, 1500, 2000, 2500, 3000, 3500, 4000, 4500, 5000, 5500, 6000, 6500, 7000, 7500, 8000, 9000, 10000, or more nucleotides, nucleosides, or base pairs, including all values and ranges therebetween, of a polynucleotide encoding one or more amino acid sequence described or referenced herein. It also is contemplated that a particular polypeptide may be encoded by nucleic acids containing variations having slightly different nucleic acid sequences but, nonetheless, encode the same or substantially similar protein (see Table 2 above).

In particular embodiments, the invention concerns isolated nucleic acid segments and recombinant vectors incorporating nucleic acid sequences that encode a variant SpA or coagulase. The term “recombinant” may be used in conjunction with a polynucleotide or polypeptide and generally refers to a polypeptide or polynucleotide produced and/or manipulated *in vitro* or that is a replication product of such a molecule.

In other embodiments, the invention concerns isolated nucleic acid segments and recombinant vectors incorporating nucleic acid sequences that encode a variant SpA or coagulase polypeptide or peptide to generate an immune response in a subject. In various embodiments the nucleic acids of the invention may be used in genetic vaccines.

The nucleic acid segments used in the present invention can be combined with other nucleic acid sequences, such as promoters, polyadenylation signals, additional restriction enzyme sites, multiple cloning sites, other coding segments, and the like, such that their overall length may vary considerably. It is therefore contemplated that a nucleic acid fragment of almost any length may be employed, with the total length preferably being limited by the ease of preparation and use in the intended recombinant nucleic acid protocol. In some cases, a nucleic acid sequence may encode a polypeptide sequence with additional heterologous coding sequences, for example to allow for purification of the polypeptide, transport, secretion, post-translational modification, or for therapeutic benefits such as targeting or efficacy. As discussed above, a tag or other heterologous polypeptide may be added to the modified polypeptide-encoding sequence, wherein “heterologous” refers to a polypeptide that is not the same as the modified polypeptide.

In certain other embodiments, the invention concerns isolated nucleic acid segments and recombinant vectors that include within their sequence a contiguous nucleic acid sequence from SEQ ID NO:1 (SpA domain D) or SEQ ID NO:3 (SpA) or any other nucleic acid sequences encoding coagulases or other secreted virulence factors and/or surface proteins including proteins transported by the Ess pathway, processed by sortase, or proteins incorporated herein by reference.

In certain embodiments, the present invention provides polynucleotide variants having substantial identity to the sequences disclosed herein; those comprising at least 70%, 75%, 80%, 85%, 90%, 95%, 96%, 97%, 98%, or 99% or higher sequence identity, including all values and ranges there between, compared to a polynucleotide sequence of

this invention using the methods described herein (e.g., BLAST analysis using standard parameters).

The invention also contemplates the use of polynucleotides which are complementary to all the above described polynucleotides.

E. Vectors

Polypeptides of the invention may be encoded by a nucleic acid molecule comprised in a vector. The term “vector” is used to refer to a carrier nucleic acid molecule into which a heterologous nucleic acid sequence can be inserted for introduction into a cell where it can be replicated and expressed. A nucleic acid sequence can be “heterologous,” which means that it is in a context foreign to the cell in which the vector is being introduced or to the nucleic acid in which is incorporated, which includes a sequence homologous to a sequence in the cell or nucleic acid but in a position within the host cell or nucleic acid where it is ordinarily not found. Vectors include DNAs, RNAs, plasmids, cosmids, viruses (bacteriophage, animal viruses, and plant viruses), and artificial chromosomes (e.g., YACs). One of skill in the art would be well equipped to construct a vector through standard recombinant techniques (for example Sambrook et al., 2001; Ausubel et al., 1996, both incorporated herein by reference). In addition to encoding a variant SpA polypeptide the vector can encode other polypeptide sequences such as a one or more other bacterial peptide, a tag, or an immunogenicity enhancing peptide. Useful vectors encoding such fusion proteins include pIN vectors (Inouye et al., 1985), vectors encoding a stretch of histidines, and pGEX vectors, for use in generating glutathione S-transferase (GST) soluble fusion proteins for later purification and separation or cleavage.

The term “expression vector” refers to a vector containing a nucleic acid sequence coding for at least part of a gene product capable of being transcribed. In some cases, RNA molecules are then translated into a protein, polypeptide, or peptide. Expression vectors can contain a variety of “control sequences,” which refer to nucleic acid sequences necessary for the transcription and possibly translation of an operably linked coding sequence in a particular host organism. In addition to control sequences that govern transcription and translation, vectors and expression vectors may contain nucleic acid sequences that serve other functions as well and are described herein.

1. Promoters and Enhancers

A “promoter” is a control sequence. The promoter is typically a region of a nucleic acid sequence at which initiation and rate of transcription are controlled. It may contain genetic elements at which regulatory proteins and molecules may bind such as RNA polymerase and other transcription factors. The phrases “operatively positioned,” “operatively linked,” “under control,” and “under transcriptional control” mean that a promoter is in a correct functional location and/or orientation in relation to a nucleic acid sequence to control transcriptional initiation and expression of that sequence. A promoter may or may not be used in conjunction with an “enhancer,” which refers to a cis-acting regulatory sequence involved in the transcriptional activation of a nucleic acid sequence.

Naturally, it may be important to employ a promoter and/or enhancer that effectively directs the expression of the DNA segment in the cell type or organism chosen for expression. Those of skill in the art of molecular biology generally know the use of promoters, enhancers, and cell type combinations for protein expression (see Sambrook et al., 2001, incorporated herein by reference). The promoters employed may be constitutive, tissue-specific, or inducible

and in certain embodiments may direct high level expression of the introduced DNA segment under specified conditions, such as large-scale production of recombinant proteins or peptides.

Various elements/promoters may be employed in the context of the present invention to regulate the expression of a gene. Examples of such inducible elements, which are regions of a nucleic acid sequence that can be activated in response to a specific stimulus, include but are not limited to Immunoglobulin Heavy Chain (Banerji et al., 1983; Gilles et al., 1983; Grosschedl et al., 1985; Atchinson et al., 1986, 1987; Imler et al., 1987; Weinberger et al., 1984; Kiledjian et al., 1988; Porton et al., 1990), Immunoglobulin Light Chain (Queen et al., 1983; Picard et al., 1984), T Cell Receptor (Luria et al., 1987; Winoto et al., 1989; Redondo et al., 1990), HLA DQ α and/or DQ β (Sullivan et al., 1987), β Interferon (Goodbourn et al., 1986; Fujita et al., 1987; Goodbourn et al., 1988), Interleukin-2 (Greene et al., 1989), Interleukin-2 Receptor (Greene et al., 1989; Lin et al., 1990), MHC Class II 5 (Koch et al., 1989), MEW Class II HLA-DR α (Sherman et al., 1989), β -Actin (Kawamoto et al., 1988; Ng et al., 1989), Muscle Creatine Kinase (MCK) (Jaynes et al., 1988; Horlick et al., 1989; Johnson et al., 1989), Prealbumin (Transferrin) (Costa et al., 1988), Elastase I (Ornitz et al., 1987), Metallothionein (MTII) (Karin et al., 1987; Culotta et al., 1989), Collagenase (Pinkert et al., 1987; Angel et al., 1987), Albumin (Pinkert et al., 1987; Tronche et al., 1989, 1990), α -Fetoprotein (Godbout et al., 1988; Campere et al., 1989), γ -Globin (Bodine et al., 1987; Perez-Stable et al., 1990), β -Globin (Trudel et al., 1987), c-fos (Cohen et al., 1987), c-Ha-Ras (Triesman, 1986; Deschamps et al., 1985), Insulin (Edlund et al., 1985), Neural Cell Adhesion Molecule (NCAM) (Hirsh et al., 1990), α 1-Antitrypsin (Latimer et al., 1990), H2B (TH2B) Histone (Hwang et al., 1990), Mouse and/or Type I Collagen (Ripe et al., 1989), Glucose-Regulated Proteins (GRP94 and GRP78) (Chang et al., 1989), Rat Growth Hormone (Larsen et al., 1986), Human Serum Amyloid A (SAA) (Edbrooke et al., 1989), Troponin I (TN I) (Yutzey et al., 1989), Platelet-Derived Growth Factor (PDGF) (Pech et al., 1989), Duchenne Muscular Dystrophy (Klamut et al., 1990), SV40 (Banerji et al., 1981; Moreau et al., 1981; Sleight et al., 1985; Firak et al., 1986; Herr et al., 1986; Imbra et al., 1986; Kadesch et al., 1986; Wang et al., 1986; Ondek et al., 1987; Kuhl et al., 1987; Schaffner et al., 1988), Polyoma (Swartzendruber et al., 1975; Vasseur et al., 1980; Katinka et al., 1980, 1981; Tyndell et al., 1981; Dandolo et al., 1983; de Villiers et al., 1984; Hen et al., 1986; Satake et al., 1988; Campbell et al., 1988), Retroviruses (Kriegler et al., 1982, 1983; Levinson et al., 1982; Kriegler et al., 1983, 1984a, b, 1988; Bosze et al., 1986; Miksicek et al., 1986; Celandier et al., 1987; Thiesen et al., 1988; Celandier et al., 1988; Choi et al., 1988; Reisman et al., 1989), Papilloma Virus (Campo et al., 1983; Lusky et al., 1983; Spandidos and Wilkie, 1983; Spalholz et al., 1985; Lusky et al., 1986; Cripe et al., 1987; Gloss et al., 1987; Hirochika et al., 1987; Stephens et al., 1987), Hepatitis B Virus (Bulla et al., 1986; Jameel et al., 1986; Shaul et al., 1987; Spandau et al., 1988; Vannice et al., 1988), Human Immunodeficiency Virus (Muesing et al., 1987; Hauber et al., 1988; Jakobovits et al., 1988; Feng et al., 1988; Takebe et al., 1988; Rosen et al., 1988; Berkhout et al., 1989; Laspia et al., 1989; Sharp et al., 1989; Braddock et al., 1989), Cytomegalovirus (CMV) IE (Weber et al., 1984; Boshart et al., 1985; Foecking et al., 1986), Gibbon Ape Leukemia Virus (Holbrook et al., 1987; Quinn et al., 1989).

Inducible elements include, but are not limited to MT II—Phorbol Ester (TFA)/Heavy metals (Palmiter et al., 1982; Haslinger et al., 1985; Searle et al., 1985; Stuart et al., 1985; Imagawa et al., 1987; Karin et al., 1987; Angel et al., 1987b; McNeall et al., 1989); MMTV (mouse mammary tumor virus)—Glucocorticoids (Huang et al., 1981; Lee et al., 1981; Majors et al., 1983; Chandler et al., 1983; Lee et al., 1984; Ponta et al., 1985; Sakai et al., 1988); β -Interferon—poly(rI)/poly(rc) (Tavernier et al., 1983); Adenovirus 5 E2—E1A (Imperiale et al., 1984); Collagenase—Phorbol Ester (TPA) (Angel et al., 1987a); Stromelysin—Phorbol Ester (TPA) (Angel et al., 1987b); SV40—Phorbol Ester (TPA) (Angel et al., 1987b); Murine MX Gene—Interferon, Newcastle Disease Virus (Hug et al., 1988); GRP78 Gene—A23187 (Resendez et al., 1988); α -2-Macroglobulin—IL-6 (Kunz et al., 1989); Vimentin—Serum (Rittling et al., 1989); MHC Class I Gene H-2 κ b—Interferon (Blonar et al., 1989); HSP70—E1A/SV40 Large T Antigen (Taylor et al., 1989, 1990a, 1990b); Proliferin—Phorbol Ester/TPA (Mordacq et al., 1989); Tumor Necrosis Factor—PMA (Hensel et al., 1989); and Thyroid Stimulating Hormone α Gene—Thyroid Hormone (Chatterjee et al., 1989).

The particular promoter that is employed to control the expression of peptide or protein encoding polynucleotide of the invention is not believed to be critical, so long as it is capable of expressing the polynucleotide in a targeted cell, preferably a bacterial cell. Where a human cell is targeted, it is preferable to position the polynucleotide coding region adjacent to and under the control of a promoter that is capable of being expressed in a human cell. Generally speaking, such a promoter might include either a bacterial, human or viral promoter.

In embodiments in which a vector is administered to a subject for expression of the protein, it is contemplated that a desirable promoter for use with the vector is one that is not down-regulated by cytokines or one that is strong enough that even if down-regulated, it produces an effective amount of a variant SpA for eliciting an immune response. Non-limiting examples of these are CMV IE and RSV LTR. Tissue specific promoters can be used, particularly if expression is in cells in which expression of an antigen is desirable, such as dendritic cells or macrophages. The mammalian MHC I and MHC II promoters are examples of such tissue-specific promoters.

2. Initiation Signals and Internal Ribosome Binding Sites (IRES)

A specific initiation signal also may be required for efficient translation of coding sequences. These signals include the ATG initiation codon or adjacent sequences. Exogenous translational control signals, including the ATG initiation codon, may need to be provided. One of ordinary skill in the art would readily be capable of determining this and providing the necessary signals.

In certain embodiments of the invention, the use of internal ribosome entry sites (IRES) elements are used to create multigene, or polycistronic, messages. IRES elements are able to bypass the ribosome scanning model of 5' methylated Cap dependent translation and begin translation at internal sites (Pelletier and Sonenberg, 1988; Macejak and Sarnow, 1991). IRES elements can be linked to heterologous open reading frames. Multiple open reading frames can be transcribed together, each separated by an IRES, creating polycistronic messages. Multiple genes can be efficiently expressed using a single promoter/enhancer to transcribe a single message (see U.S. Pat. Nos. 5,925,565 and 5,935,819, herein incorporated by reference).

3. Selectable and Screenable Markers

In certain embodiments of the invention, cells containing a nucleic acid construct of the present invention may be identified *in vitro* or *in vivo* by encoding a screenable or selectable marker in the expression vector. When transcribed and translated, a marker confers an identifiable change to the cell permitting easy identification of cells containing the expression vector. Generally, a selectable marker is one that confers a property that allows for selection. A positive selectable marker is one in which the presence of the marker allows for its selection, while a negative selectable marker is one in which its presence prevents its selection. An example of a positive selectable marker is a drug resistance marker.

F. Host Cells

As used herein, the terms “cell,” “cell line,” and “cell culture” may be used interchangeably. All of these terms also include their progeny, which is any and all subsequent generations. It is understood that all progeny may not be identical due to deliberate or inadvertent mutations. In the context of expressing a heterologous nucleic acid sequence, “host cell” refers to a prokaryotic or eukaryotic cell, and it includes any transformable organism that is capable of replicating a vector or expressing a heterologous gene encoded by a vector. A host cell can, and has been, used as a recipient for vectors or viruses. A host cell may be “transfected” or “transformed,” which refers to a process by which exogenous nucleic acid, such as a recombinant protein-encoding sequence, is transferred or introduced into the host cell. A transformed cell includes the primary subject cell and its progeny.

Host cells may be derived from prokaryotes or eukaryotes, including bacteria, yeast cells, insect cells, and mammalian cells for replication of the vector or expression of part or all of the nucleic acid sequence(s). Numerous cell lines and cultures are available for use as a host cell, and they can be obtained through the American Type Culture Collection (ATCC), which is an organization that serves as an archive for living cultures and genetic materials (www.atcc.org).

G. Expression Systems

Numerous expression systems exist that comprise at least a part or all of the compositions discussed above. Prokaryote- and/or eukaryote-based systems can be employed for use with the present invention to produce nucleic acid sequences, or their cognate polypeptides, proteins and peptides. Many such systems are commercially and widely available.

The insect cell/baculovirus system can produce a high level of protein expression of a heterologous nucleic acid segment, such as described in U.S. Pat. Nos. 5,871,986, 4,879,236, both herein incorporated by reference, and which can be bought, for example, under the name MAXBAC® 2.0 from INVITROGEN® and BACPACK™ BACULOVIRUS EXPRESSION SYSTEM FROM CLONTECH®.

In addition to the disclosed expression systems of the invention, other examples of expression systems include STRATAGENE®'s COMPLETE CONTROL™ Inducible Mammalian Expression System, which involves a synthetic ecdysone-inducible receptor, or its pET Expression System, an *E. coli* expression system. Another example of an inducible expression system is available from INVITROGEN®, which carries the T-REX™ (tetracycline-regulated expression) System, an inducible mammalian expression system that uses the full-length CMV promoter. INVITROGEN® also provides a yeast expression system called the *Pichia methanolica* Expression System, which is designed for

high-level production of recombinant proteins in the methylophilic yeast *Pichia methanolica*. One of skill in the art would know how to express a vector, such as an expression construct, to produce a nucleic acid sequence or its cognate polypeptide, protein, or peptide.

V. POLYSACCHARIDES

The immunogenic compositions of the invention may further comprise capsular polysaccharides including one or more of PIA (also known as PNAG) and/or *S. aureus* Type V and/or type VIII capsular polysaccharide and/or *S. epidermidis* Type I, and/or Type II and/or Type III capsular polysaccharide.

15 H. PIA (PNAG)

It is now clear that the various forms of staphylococcal surface polysaccharides identified as PS/A, PIA and SAA are the same chemical entity—PNAG (Maira-Litran et al., 2004). Therefore the term PIA or PNAG encompasses all these polysaccharides or oligosaccharides derived from them.

PIA is a polysaccharide intercellular adhesin and is composed of a polymer of β -(1→6)-linked glucosamine substituted with N-acetyl and O-succinyl constituents. This polysaccharide is present in both *S. aureus* and *S. epidermidis* and can be isolated from either source (Joyce et al., 2003; Maira-Litran et al., 2002). For example, PNAG may be isolated from *S. aureus* strain MN8m (WO04/43407). PIA isolated from *S. epidermidis* is an integral constituent of biofilm. It is responsible for mediating cell-cell adhesion and probably also functions to shield the growing colony from the host's immune response. The polysaccharide previously known as poly-N-succinyl- β -(1→6)-glucosamine (PNSG) was recently shown not to have the expected structure since the identification of N-succinylation was incorrect (Maira-Litran et al., 2002). Therefore the polysaccharide formally known as PNSG and now found to be PNAG is also encompassed by the term PIA.

PIA (or PNAG) may be of different sizes varying from over 400 kDa to between 75 and 400 kDa to between 10 and 75 kDa to oligosaccharides composed of up to 30 repeat units (of β -(1→6)-linked glucosamine substituted with N-acetyl and O-succinyl constituents). Any size of PIA polysaccharide or oligosaccharide may be used in an immunogenic composition of the invention, in one aspect the polysaccharide is over 40 kDa. Sizing may be achieved by any method known in the art, for instance by microfluidization, ultrasonic irradiation or by chemical cleavage (WO 03/53462, EP497524, EP497525). In certain aspects PIA (PNAG) is at least or at most 40-400 kDa, 40-300 kDa, 50-350 kDa, 60-300 kDa, 50-250 kDa and 60-200 kDa.

PIA (PNAG) can have different degree of acetylation due to substitution on the amino groups by acetate. PIA produced *in vitro* is almost fully substituted on amino groups (95-100%). Alternatively, a deacetylated PIA (PNAG) can be used having less than 60%, 50%, 40%, 30%, 20%, 10% acetylation. Use of a deacetylated PIA (PNAG) is preferred since non-acetylated epitopes of PNAG are efficient at mediating opsonic killing of Gram positive bacteria, preferably *S. aureus* and/or *S. epidermidis*. In certain aspects, the PIA (PNAG) has a size between 40 kDa and 300 kDa and is deacetylated so that less than 60%, 50%, 40%, 30% or 20% of amino groups are acetylated.

The term deacetylated PNAG (dPNAG) refers to a PNAG polysaccharide or oligosaccharide in which less than 60%, 50%, 40%, 30%, 20% or 10% of the amino groups are acetylated. In certain aspects, PNAG is deacetylated to form

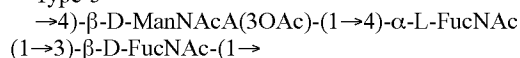
dPNAG by chemically treating the native polysaccharide. For example, the native PNAG is treated with a basic solution such that the pH rises to above 10. For instance the PNAG is treated with 0.1-5 M, 0.2-4 M, 0.3-3 M, 0.5-2 M, 0.75-1.5 M or 1 M NaOH, KOH or NH₄OH. Treatment is for at least 10 to 30 minutes, or 1, 2, 3, 4, 5, 10, 15 or 20 hours at a temperature of 20-100, 25-80, 30-60 or 30-50 or 35-45° C. dPNAG may be prepared as described in WO 04/43405.

The polysaccharide(s) can be conjugated or unconjugated to a carrier protein.

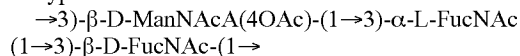
I. Type 5 and Type 8 Polysaccharides from *S. aureus*

Most strains of *S. aureus* that cause infection in man contain either Type 5 or Type 8 polysaccharides. Approximately 60% of human strains are Type 8 and approximately 30% are Type 5. The structures of Type 5 and Type 8 capsular polysaccharide antigens are described in Moreau et al., (1990) and Fournier et al., (1984). Both have FucNAcP in their repeat unit as well as ManNAcA which can be used to introduce a sulfhydryl group. The structures are:

Type 5

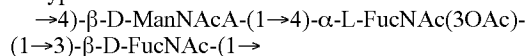


Type 8

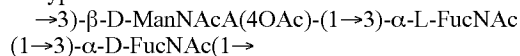


Recently (Jones, 2005) NMR spectroscopy revised the structures to:

Type 5



Type 8



Polysaccharides may be extracted from the appropriate strain of *S. aureus* using method well known to of skill in the art, See U.S. Pat. No. 6,294,177. For example, ATCC 12902 is a Type 5 *S. aureus* strain and ATCC 12605 is a Type 8 *S. aureus* strain.

Polysaccharides are of native size or alternatively may be sized, for instance by microfluidisation, ultrasonic irradiation, or by chemical treatment. The invention also covers oligosaccharides derived from the type 5 and 8 polysaccharides from *S. aureus*. The type 5 and 8 polysaccharides included in the immunogenic composition of the invention are preferably conjugated to a carrier protein as described below or are alternatively unconjugated. The immunogenic compositions of the invention alternatively contains either type 5 or type 8 polysaccharide.

J. *S. aureus* 336 Antigen

In an embodiment, the immunogenic composition of the invention comprises the *S. aureus* 336 antigen described in U.S. Pat. No. 6,294,177. The 336 antigen comprises β -linked hexosamine, contains no O-acetyl groups, and specifically binds to antibodies to *S. aureus* Type 336 deposited under ATCC 55804. In an embodiment, the 336 antigen is a polysaccharide which is of native size or alternatively may be sized, for instance by microfluidisation, ultrasonic irradiation, or by chemical treatment. The invention also covers oligosaccharides derived from the 336 antigen. The 336 antigen can be unconjugated or conjugated to a carrier protein.

K. Type I, II and III Polysaccharides from *S. epidermidis*

Amongst the problems associated with the use of polysaccharides in vaccination, is the fact that polysaccharides per se are poor immunogens. It is preferred that the polysaccharides utilized in the invention are linked to a protein carrier which provide bystander T-cell help to improve

immunogenicity. Examples of such carriers which may be conjugated to polysaccharide immunogens include the Diphtheria and Tetanus toxoids (DT, DT CRM197 and TT respectively), Keyhole Limpet Haemocyanin (KLH), and the purified protein derivative of Tuberculin (PPD), *Pseudomonas aeruginosa* exoprotein A (rEPA), protein D from *Haemophilus influenzae*, pneumolysin or fragments of any of the above. Fragments suitable for use include fragments encompassing T-helper epitopes. In particular the protein D fragment from *H. influenzae* will preferably contain the N-terminal 1/3 of the protein. Protein D is an IgD-binding protein from *Haemophilus influenzae* (EP 0 594 610 B1) and is a potential immunogen. In addition, staphylococcal proteins may be used as a carrier protein in the polysaccharide conjugates of the invention.

A carrier protein that would be particularly advantageous to use in the context of a staphylococcal vaccine is staphylococcal alpha toxoid. The native form may be conjugated to a polysaccharide since the process of conjugation reduces toxicity. Preferably genetically detoxified alpha toxins such as the His35Leu or His35Arg variants are used as carriers since residual toxicity is lower. Alternatively the alpha toxin is chemically detoxified by treatment with a cross-linking reagent, formaldehyde or glutaraldehyde. A genetically detoxified alpha toxin is optionally chemically detoxified, preferably by treatment with a cross-linking reagent, formaldehyde or glutaraldehyde to further reduce toxicity.

The polysaccharides may be linked to the carrier protein(s) by any known method (for example those methods described in U.S. Pat. Nos. 4,372,945, 4,474,757, and 4,356,170). Preferably, CDAP conjugation chemistry is carried out (see WO95/08348). In CDAP, the cyanating reagent 1-cyano-dimethylaminopyridinium tetrafluoroborate (CDAP) is preferably used for the synthesis of polysaccharide-protein conjugates. The cyanation reaction can be performed under relatively mild conditions, which avoids hydrolysis of the alkaline sensitive polysaccharides. This synthesis allows direct coupling to a carrier protein.

Conjugation preferably involves producing a direct linkage between the carrier protein and polysaccharide. Optionally a spacer (such as adipic dihydride (ADH)) may be introduced between the carrier protein and the polysaccharide.

IV. IMMUNE RESPONSE AND ASSAYS

As discussed above, the invention concerns evoking or inducing an immune response in a subject against a variant SpA or coagulase peptide. In one embodiment, the immune response can protect against or treat a subject having, suspected of having, or at risk of developing an infection or related disease, particularly those related to staphylococci. One use of the immunogenic compositions of the invention is to prevent nosocomial infections by inoculating a subject prior to undergoing procedures in a hospital or other environment having an increased risk of infection.

A. Immunoassays

The present invention includes the implementation of serological assays to evaluate whether and to what extent an immune response is induced or evoked by compositions of the invention. There are many types of immunoassays that can be implemented. Immunoassays encompassed by the present invention include, but are not limited to, those described in U.S. Pat. No. 4,367,110 (double monoclonal antibody sandwich assay) and U.S. Pat. No. 4,452,901

(western blot). Other assays include immunoprecipitation of labeled ligands and immunocytochemistry, both *in vitro* and *in vivo*.

Immunoassays generally are binding assays. Certain preferred immunoassays are the various types of enzyme linked immunosorbent assays (ELISAs) and radioimmunoassays (RIA) known in the art. Immunohistochemical detection using tissue sections is also particularly useful. In one example, antibodies or antigens are immobilized on a selected surface, such as a well in a polystyrene microtiter plate, dipstick, or column support. Then, a test composition suspected of containing the desired antigen or antibody, such as a clinical sample, is added to the wells. After binding and washing to remove non specifically bound immune complexes, the bound antigen or antibody may be detected. Detection is generally achieved by the addition of another antibody, specific for the desired antigen or antibody, that is linked to a detectable label. This type of ELISA is known as a "sandwich ELISA." Detection also may be achieved by the addition of a second antibody specific for the desired antigen, followed by the addition of a third antibody that has binding affinity for the second antibody, with the third antibody being linked to a detectable label.

Competition ELISAs are also possible implementations in which test samples compete for binding with known amounts of labeled antigens or antibodies. The amount of reactive species in the unknown sample is determined by mixing the sample with the known labeled species before or during incubation with coated wells. The presence of reactive species in the sample acts to reduce the amount of labeled species available for binding to the well and thus reduces the ultimate signal. Irrespective of the format employed, ELISAs have certain features in common, such as coating, incubating or binding, washing to remove non specifically bound species, and detecting the bound immune complexes.

Antigen or antibodies may also be linked to a solid support, such as in the form of plate, beads, dipstick, membrane, or column matrix, and the sample to be analyzed is applied to the immobilized antigen or antibody. In coating a plate with either antigen or antibody, one will generally incubate the wells of the plate with a solution of the antigen or antibody, either overnight or for a specified period. The wells of the plate will then be washed to remove incompletely-adsorbed material. Any remaining available surfaces of the wells are then "coated" with a nonspecific protein that is antigenically neutral with regard to the test antisera. These include bovine serum albumin (BSA), casein, and solutions of milk powder. The coating allows for blocking of nonspecific adsorption sites on the immobilizing surface and thus reduces the background caused by nonspecific binding of antisera onto the surface.

B. Diagnosis of Bacterial Infection

In addition to the use of proteins, polypeptides, and/or peptides, as well as antibodies binding these polypeptides, proteins, and/or peptides, to treat or prevent infection as described above, the present invention contemplates the use of these polypeptides, proteins, peptides, and/or antibodies in a variety of ways, including the detection of the presence of Staphylococci to diagnose an infection, whether in a patient or on medical equipment which may also become infected. In accordance with the invention, a preferred method of detecting the presence of infections involves the steps of obtaining a sample suspected of being infected by one or more staphylococcal bacteria species or strains, such as a sample taken from an individual, for example, from one's blood, saliva, tissues, bone, muscle, cartilage, or skin.

Following isolation of the sample, diagnostic assays utilizing the polypeptides, proteins, peptides, and/or antibodies of the present invention may be carried out to detect the presence of staphylococci, and such assay techniques for determining such presence in a sample are well known to those skilled in the art and include methods such as radioimmunoassay, western blot analysis and ELISA assays. In general, in accordance with the invention, a method of diagnosing an infection is contemplated wherein a sample suspected of being infected with staphylococci has added to it the polypeptide, protein, peptide, antibody, or monoclonal antibody in accordance with the present invention, and staphylococci are indicated by antibody binding to the polypeptides, proteins, and/or peptides, or polypeptides, proteins, and/or peptides binding to the antibodies in the sample.

Accordingly, antibodies in accordance with the invention may be used for the prevention of infection from staphylococcal bacteria (i.e., passive immunization), for the treatment of an ongoing infection, or for use as research tools. The term "antibodies" as used herein includes monoclonal, polyclonal, chimeric, single chain, bispecific, simianized, and humanized or primatized antibodies as well as Fab fragments, such as those fragments which maintain the binding specificity of the antibodies, including the products of an Fab immunoglobulin expression library. Accordingly, the invention contemplates the use of single chains such as the variable heavy and light chains of the antibodies. Generation of any of these types of antibodies or antibody fragments is well known to those skilled in the art. Specific examples of the generation of an antibody to a bacterial protein can be found in U.S. Patent Application Pub. No. 20030153022, which is incorporated herein by reference in its entirety.

Any of the above described polypeptides, proteins, peptides, and/or antibodies may be labeled directly with a detectable label for identification and quantification of staphylococcal bacteria. Labels for use in immunoassays are generally known to those skilled in the art and include enzymes, radioisotopes, and fluorescent, luminescent and chromogenic substances, including colored particles such as colloidal gold or latex beads. Suitable immunoassays include enzyme-linked immunosorbent assays (ELISA).

C. Protective Immunity

In some embodiments of the invention, proteinaceous compositions confer protective immunity to a subject. Protective immunity refers to a body's ability to mount a specific immune response that protects the subject from developing a particular disease or condition that involves the agent against which there is an immune response. An immunogenically effective amount is capable of conferring protective immunity to the subject.

As used herein in the specification and in the claims section that follows, the term polypeptide or peptide refer to a stretch of amino acids covalently linked there amongst via peptide bonds. Different polypeptides have different functionalities according to the present invention. While according to one aspect, a polypeptide is derived from an immunogen designed to induce an active immune response in a recipient, according to another aspect of the invention, a polypeptide is derived from an antibody which results following the elicitation of an active immune response in, for example, an animal, and which can serve to induce a passive immune response in the recipient. In both cases, however, the polypeptide is encoded by a polynucleotide according to any possible codon usage.

As used herein the phrase “immune response” or its equivalent “immunological response” refers to the development of a humoral (antibody mediated), cellular (mediated by antigen-specific T cells or their secretion products) or both humoral and cellular response directed against a protein, peptide, carbohydrate, or polypeptide of the invention in a recipient patient. Such a response can be an active response induced by administration of immunogen or a passive response induced by administration of antibody, antibody containing material, or primed T-cells. A cellular immune response is elicited by the presentation of polypeptide epitopes in association with Class I or Class II MHC molecules, to activate antigen-specific CD4 (+) T helper cells and/or CD8 (+) cytotoxic T cells. The response may also involve activation of monocytes, macrophages, NK cells, basophils, dendritic cells, astrocytes, microglia cells, eosinophils or other components of innate immunity. As used herein “active immunity” refers to any immunity conferred upon a subject by administration of an antigen.

As used herein “passive immunity” refers to any immunity conferred upon a subject without administration of an antigen to the subject. “Passive immunity” therefore includes, but is not limited to, administration of activated immune effectors including cellular mediators or protein mediators (e.g., monoclonal and/or polyclonal antibodies) of an immune response. A monoclonal or polyclonal antibody composition may be used in passive immunization for the prevention or treatment of infection by organisms that carry the antigen recognized by the antibody. An antibody composition may include antibodies that bind to a variety of antigens that may in turn be associated with various organisms. The antibody component can be a polyclonal antiserum. In certain aspects the antibody or antibodies are affinity purified from an animal or second subject that has been challenged with an antigen(s). Alternatively, an antibody mixture may be used, which is a mixture of monoclonal and/or polyclonal antibodies to antigens present in the same, related, or different microbes or organisms, such as gram-positive bacteria, gram-negative bacteria, including but not limited to *staphylococcus* bacteria.

Passive immunity may be imparted to a patient or subject by administering to the patient immunoglobulins (Ig) and/or other immune factors obtained from a donor or other non-patient source having a known immunoreactivity. In other aspects, an antigenic composition of the present invention can be administered to a subject who then acts as a source or donor for globulin, produced in response to challenge with the antigenic composition (“hyperimmune globulin”), that contains antibodies directed against *Staphylococcus* or other organism. A subject thus treated would donate plasma from which hyperimmune globulin would then be obtained, via conventional plasma-fractionation methodology, and administered to another subject in order to impart resistance against or to treat *staphylococcus* infection. Hyperimmune globulins according to the invention are particularly useful for immune-compromised individuals, for individuals undergoing invasive procedures or where time does not permit the individual to produce their own antibodies in response to vaccination. See U.S. Pat. Nos. 6,936,258, 6,770,278, 6,756,361, 5,548,066, 5,512,282, 4,338,298, and 4,748,018, each of which is incorporated herein by reference in its entirety, for exemplary methods and compositions related to passive immunity.

For purposes of this specification and the accompanying claims the terms “epitope” and “antigenic determinant” are used interchangeably to refer to a site on an antigen to which B and/or T cells respond or recognize. B-cell epitopes can be

formed both from contiguous amino acids or noncontiguous amino acids juxtaposed by tertiary folding of a protein. Epitopes formed from contiguous amino acids are typically retained on exposure to denaturing solvents whereas epitopes formed by tertiary folding are typically lost on treatment with denaturing solvents. An epitope typically includes at least 3, and more usually, at least 5 or 8-10 amino acids in a unique spatial conformation. Methods of determining spatial conformation of epitopes include, for example, x-ray crystallography and 2-dimensional nuclear magnetic resonance. See, e.g., Epitope Mapping Protocols (1996). Antibodies that recognize the same epitope can be identified in a simple immunoassay showing the ability of one antibody to block the binding of another antibody to a target antigen. T-cells recognize continuous epitopes of about nine amino acids for CD8 cells or about 13-15 amino acids for CD4 cells. T cells that recognize the epitope can be identified by in vitro assays that measure antigen-dependent proliferation, as determined by ³H-thymidine incorporation by primed T cells in response to an epitope (Burke et al., 1994), by antigen-dependent killing (cytotoxic T lymphocyte assay, Tigges et al., 1996) or by cytokine secretion.

The presence of a cell-mediated immunological response can be determined by proliferation assays (CD4 (+) T cells) or CTL (cytotoxic T lymphocyte) assays. The relative contributions of humoral and cellular responses to the protective or therapeutic effect of an immunogen can be distinguished by separately isolating IgG and T-cells from an immunized syngeneic animal and measuring protective or therapeutic effect in a second subject.

As used herein and in the claims, the terms “antibody” or “immunoglobulin” are used interchangeably and refer to any of several classes of structurally related proteins that function as part of the immune response of an animal or recipient, which proteins include IgG, IgD, IgE, IgA, IgM and related proteins.

Under normal physiological conditions antibodies are found in plasma and other body fluids and in the membrane of certain cells and are produced by lymphocytes of the type denoted B cells or their functional equivalent. Antibodies of the IgG class are made up of four polypeptide chains linked together by disulfide bonds. The four chains of intact IgG molecules are two identical heavy chains referred to as H-chains and two identical light chains referred to as L-chains.

In order to produce polyclonal antibodies, a host, such as a rabbit or goat, is immunized with the antigen or antigen fragment, generally with an adjuvant and, if necessary, coupled to a carrier. Antibodies to the antigen are subsequently collected from the sera of the host. The polyclonal antibody can be affinity purified against the antigen rendering it monospecific.

Monoclonal antibodies can be produced by hyperimmunization of an appropriate donor with the antigen or ex-vivo by use of primary cultures of splenic cells or cell lines derived from spleen (Anavi, 1998; Huston et al., 1991; Johnson et al., 1991; Mernaugh et al., 1995).

As used herein and in the claims the phrase “an immunological portion of an antibody” includes a Fab fragment of an antibody, a Fv fragment of an antibody, a heavy chain of an antibody, a light chain of an antibody, a heterodimer consisting of a heavy chain and a light chain of an antibody, a variable fragment of a light chain of an antibody, a variable fragment of a heavy chain of an antibody, and a single chain variant of an antibody, which is also known as scFv. In addition, the term includes chimeric immunoglobulins which are the expression products of fused genes derived

from different species, one of the species can be a human, in which case a chimeric immunoglobulin is said to be humanized. Typically, an immunological portion of an antibody competes with the intact antibody from which it was derived for specific binding to an antigen.

Optionally, an antibody or preferably an immunological portion of an antibody, can be chemically conjugated to, or expressed as, a fusion protein with other proteins. For purposes of this specification and the accompanying claims, all such fused proteins are included in the definition of antibodies or an immunological portion of an antibody.

As used herein the terms "immunogenic agent" or "immunogen" or "antigen" are used interchangeably to describe a molecule capable of inducing an immunological response against itself on administration to a recipient, either alone, in conjunction with an adjuvant, or presented on a display vehicle.

D. Treatment Methods

A method of the present invention includes treatment for a disease or condition caused by a *staphylococcus* pathogen. An immunogenic polypeptide of the invention can be given to induce an immune response in a person infected with *staphylococcus* or suspected of having been exposed to *staphylococcus*. Methods may be employed with respect to individuals who have tested positive for exposure to *staphylococcus* or who are deemed to be at risk for infection based on possible exposure.

In particular, the invention encompasses a method of treatment for staphylococcal infection, particularly hospital acquired nosocomial infections. The immunogenic compositions and vaccines of the invention are particularly advantageous to use in cases of elective surgery. Such patients will know the date of surgery in advance and could be inoculated in advance. The immunogenic compositions and vaccines of the invention are also advantageous to use to inoculate health care workers.

In some embodiments, the treatment is administered in the presence of adjuvants or carriers or other staphylococcal antigens. Furthermore, in some examples, treatment comprises administration of other agents commonly used against bacterial infection, such as one or more antibiotics.

The use of peptides for vaccination can require, but not necessarily, conjugation of the peptide to an immunogenic carrier protein, such as hepatitis B surface antigen, keyhole limpet hemocyanin, or bovine serum albumin. Methods for performing this conjugation are well known in the art.

VI. VACCINE AND OTHER PHARMACEUTICAL COMPOSITIONS AND ADMINISTRATION

E. Vaccines

The present invention includes methods for preventing or ameliorating staphylococcal infections, particularly hospital acquired nosocomial infections. As such, the invention contemplates vaccines for use in both active and passive immunization embodiments. Immunogenic compositions, proposed to be suitable for use as a vaccine, may be prepared from immunogenic SpA polypeptide(s), such as a SpA domain D variant, or immunogenic coagulases. In other embodiments SpA or coagulases can be used in combination with other secreted virulence proteins, surface proteins or immunogenic fragments thereof. In certain aspects, antigenic material is extensively dialyzed to remove undesired small molecular weight molecules and/or lyophilized for more ready formulation into a desired vehicle.

Other options for a protein/peptide-based vaccine involve introducing nucleic acids encoding the antigen(s) as DNA vaccines. In this regard, recent reports described construction of recombinant vaccinia viruses expressing either 10 contiguous minimal CTL epitopes (Thomson, 1996) or a combination of B cell, cytotoxic T-lymphocyte (CTL), and T-helper (Th) epitopes from several microbes (An, 1997), and successful use of such constructs to immunize mice for priming protective immune responses. Thus, there is ample evidence in the literature for successful utilization of peptides, peptide-pulsed antigen presenting cells (APCs), and peptide-encoding constructs for efficient in vivo priming of protective immune responses. The use of nucleic acid sequences as vaccines is exemplified in U.S. Pat. Nos. 5,958,895 and 5,620,896.

The preparation of vaccines that contain polypeptide or peptide sequence(s) as active ingredients is generally well understood in the art, as exemplified by U.S. Pat. Nos. 4,608,251; 4,601,903; 4,599,231; 4,599,230; 4,596,792; and 4,578,770, all of which are incorporated herein by reference. Typically, such vaccines are prepared as injectables either as liquid solutions or suspensions: solid forms suitable for solution in or suspension in liquid prior to injection may also be prepared. The preparation may also be emulsified. The active immunogenic ingredient is often mixed with excipients that are pharmaceutically acceptable and compatible with the active ingredient. Suitable excipients are, for example, water, saline, dextrose, glycerol, ethanol, or the like and combinations thereof. In addition, if desired, the vaccine may contain amounts of auxiliary substances such as wetting or emulsifying agents, pH buffering agents, or adjuvants that enhance the effectiveness of the vaccines. In specific embodiments, vaccines are formulated with a combination of substances, as described in U.S. Pat. Nos. 6,793,923 and 6,733,754, which are incorporated herein by reference.

Vaccines may be conventionally administered parenterally, by injection, for example, either subcutaneously or intramuscularly. Additional formulations which are suitable for other modes of administration include suppositories and, in some cases, oral formulations. For suppositories, traditional binders and carriers may include, for example, polyalkalene glycols or triglycerides: such suppositories may be formed from mixtures containing the active ingredient in the range of about 0.5% to about 10%, preferably about 1% to about 2%. Oral formulations include such normally employed excipients as, for example, pharmaceutical grades of mannitol, lactose, starch, magnesium stearate, sodium saccharine, cellulose, magnesium carbonate and the like. These compositions take the form of solutions, suspensions, tablets, pills, capsules, sustained release formulations or powders and contain about 10% to about 95% of active ingredient, preferably about 25% to about 70%.

The polypeptides and polypeptide-encoding DNA constructs may be formulated into a vaccine as neutral or salt forms. Pharmaceutically-acceptable salts include the acid addition salts (formed with the free amino groups of the peptide) and those that are formed with inorganic acids such as, for example, hydrochloric or phosphoric acids, or such organic acids as acetic, oxalic, tartaric, mandelic, and the like.

Typically, vaccines are administered in a manner compatible with the dosage formulation, and in such amount as will be therapeutically effective and immunogenic. The quantity to be administered depends on the subject to be treated, including the capacity of the individual's immune system to synthesize antibodies and the degree of protection desired.

Precise amounts of active ingredient required to be administered depend on the judgment of the practitioner. However, suitable dosage ranges are of the order of several hundred micrograms of active ingredient per vaccination. Suitable regimes for initial administration and booster shots are also

variable, but are typified by an initial administration followed by subsequent inoculations or other administrations. The manner of application may be varied widely. Any of the conventional methods for administration of a vaccine are applicable. These are believed to include oral application within a solid physiologically acceptable base or in a physiologically acceptable dispersion, parenterally, by injection and the like. The dosage of the vaccine will depend on the route of administration and will vary according to the size and health of the subject.

In certain instances, it will be desirable to have multiple administrations of the vaccine, e.g., 2, 3, 4, 5, 6 or more administrations. The vaccinations can be at 1, 2, 3, 4, 5, 6, 7, 8, to 5, 6, 7, 8, 9, 10, 11, 12 twelve week intervals, including all ranges there between. Periodic boosters at intervals of 1-5 years will be desirable to maintain protective levels of the antibodies. The course of the immunization may be followed by assays for antibodies against the antigens, as described in U.S. Pat. Nos. 3,791,932; 4,174,384 and 3,949,064.

1. Carriers

A given composition may vary in its immunogenicity. It is often necessary therefore to boost the host immune system, as may be achieved by coupling a peptide or polypeptide to a carrier. Exemplary and preferred carriers are keyhole limpet hemocyanin (KLH) and bovine serum albumin (BSA). Other albumins such as ovalbumin, mouse serum albumin, or rabbit serum albumin can also be used as carriers. Means for conjugating a polypeptide to a carrier protein are well known in the art and include glutaraldehyde, m-maleimidobencoyl-N-hydroxysuccinimide ester, carbo-diimide, and bis-biazotized benzidine.

2. Adjuvants

The immunogenicity of polypeptide or peptide compositions can be enhanced by the use of non-specific stimulators of the immune response, known as adjuvants. Suitable adjuvants include all acceptable immunostimulatory compounds, such as cytokines, toxins, or synthetic compositions. A number of adjuvants can be used to enhance an antibody response against a variant SpA polypeptide or coagulase, or any other bacterial protein or combination contemplated herein. Adjuvants can (1) trap the antigen in the body to cause a slow release; (2) attract cells involved in the immune response to the site of administration; (3) induce proliferation or activation of immune system cells; or (4) improve the spread of the antigen throughout the subject's body.

Adjuvants include, but are not limited to, oil-in-water emulsions, water-in oil emulsions, mineral salts, polynucleotides, and natural substances. Specific adjuvants that may be used include IL-1, IL-2, IL-4, IL-7, IL-12, γ -interferon, GMCSF, BCG, aluminum salts, such as aluminum hydroxide or other aluminum compound, MDP compounds, such as thur-MDP and nor-MDP, CGP (MTP-PE), lipid A, and monophosphoryl lipid A (MPL). RIBI, which contains three components extracted from bacteria, MPL, trehalose dimycolate (TDM), and cell wall skeleton (CWS) in a 2% squalene/TWEEN 80 emulsion. MEW antigens may even be used. Others adjuvants or methods are exemplified in U.S. Pat. Nos. 6,814,971, 5,084,269, 6,656,462, each of which is incorporated herein by reference).

Various methods of achieving adjuvant affect for the vaccine includes use of agents such as aluminum hydroxide or phosphate (alum), commonly used as about 0.05 to about 0.1% solution in phosphate buffered saline, admixture with synthetic polymers of sugars (Carbopol®) used as an about 0.25% solution, aggregation of the protein in the vaccine by heat treatment with temperatures ranging between about 70° to about 101° C. for a 30-second to 2-minute period, respectively. Aggregation by reactivating with pepsin-treated (Fab) antibodies to albumin; mixture with bacterial cells (e.g., *C. parvum*), endotoxins or lipopolysaccharide components of Gram-negative bacteria; emulsion in physiologically acceptable oil vehicles (e.g., mannide monooleate (Aracel A)); or emulsion with a 20% solution of a perfluorocarbon (Fluosol-DA®) used as a block substitute may also be employed to produce an adjuvant effect.

Examples of and often preferred adjuvants include complete Freund's adjuvant (a non-specific stimulator of the immune response containing killed *Mycobacterium tuberculosis*), incomplete Freund's adjuvants, and aluminum hydroxide.

In some aspects, it is preferred that the adjuvant be selected to be a preferential inducer of either a Th1 or a Th2 type of response. High levels of Th1-type cytokines tend to favor the induction of cell mediated immune responses to a given antigen, while high levels of Th2-type cytokines tend to favor the induction of humoral immune responses to the antigen.

The distinction of Th1 and Th2-type immune response is not absolute. In reality an individual will support an immune response which is described as being predominantly Th1 or predominantly Th2. However, it is often convenient to consider the families of cytokines in terms of that described in murine CD4+ T cell clones by Mosmann and Coffman (Mosmann, and Coffman, 1989). Traditionally, Th1-type responses are associated with the production of the INF- γ and IL-2 cytokines by T-lymphocytes. Other cytokines often directly associated with the induction of Th1-type immune responses are not produced by T-cells, such as IL-12. In contrast, Th2-type responses are associated with the secretion of IL-4, IL-5, IL-6, IL-10.

In addition to adjuvants, it may be desirable to co-administer biologic response modifiers (BRM) to enhance immune responses. BRMs have been shown to upregulate T cell immunity or downregulate suppresser cell activity. Such BRMs include, but are not limited to, Cimetidine (CIM; 1200 mg/d) (Smith/Kline, PA); or low-dose Cyclophosphamide (CYP; 300 mg/m²) (Johnson/Mead, NJ) and cytokines such as γ -interferon, IL-2, or IL-12 or genes encoding proteins involved in immune helper functions, such as B-7.

F. Lipid Components and Moieties

In certain embodiments, the present invention concerns compositions comprising one or more lipids associated with a nucleic acid or a polypeptide/peptide. A lipid is a substance that is insoluble in water and extractable with an organic solvent. Compounds other than those specifically described herein are understood by one of skill in the art as lipids, and are encompassed by the compositions and methods of the present invention. A lipid component and a non-lipid may be attached to one another, either covalently or non-covalently.

A lipid may be a naturally occurring lipid or a synthetic lipid. However, a lipid is usually a biological substance. Biological lipids are well known in the art, and include for example, neutral fats, phospholipids, phosphoglycerides, steroids, terpenes, lysolipids, glycosphingolipids, glucolipids, sulphatides, lipids with ether and ester-linked fatty acids and polymerizable lipids, and combinations thereof.

A nucleic acid molecule or a polypeptide/peptide, associated with a lipid may be dispersed in a solution containing a lipid, dissolved with a lipid, emulsified with a lipid, mixed with a lipid, combined with a lipid, covalently bonded to a lipid, contained as a suspension in a lipid or otherwise associated with a lipid. A lipid or lipid-poxvirus-associated composition of the present invention is not limited to any particular structure. For example, they may also simply be interspersed in a solution, possibly forming aggregates which are not uniform in either size or shape. In another example, they may be present in a bilayer structure, as micelles, or with a "collapsed" structure. In another non-limiting example, a lipofectamine (Gibco BRL)-poxvirus or Superfect (Qiagen)-poxvirus complex is also contemplated.

In certain embodiments, a composition may comprise about 1%, about 2%, about 3%, about 4% about 5%, about 6%, about 7%, about 8%, about 9%, about 10%, about 11%, about 12%, about 13%, about 14%, about 15%, about 16%, about 17%, about 18%, about 19%, about 20%, about 21%, about 22%, about 23%, about 24%, about 25%, about 26%, about 27%, about 28%, about 29%, about 30%, about 31%, about 32%, about 33%, about 34%, about 35%, about 36%, about 37%, about 38%, about 39%, about 40%, about 41%, about 42%, about 43%, about 44%, about 45%, about 46%, about 47%, about 48%, about 49%, about 50%, about 51%, about 52%, about 53%, about 54%, about 55%, about 56%, about 57%, about 58%, about 59%, about 60%, about 61%, about 62%, about 63%, about 64%, about 65%, about 66%, about 67%, about 68%, about 69%, about 70%, about 71%, about 72%, about 73%, about 74%, about 75%, about 76%, about 77%, about 78%, about 79%, about 80%, about 81%, about 82%, about 83%, about 84%, about 85%, about 86%, about 87%, about 88%, about 89%, about 90%, about 91%, about 92%, about 93%, about 94%, about 95%, about 96%, about 97%, about 98%, about 99%, or any range therebetween, of a particular lipid, lipid type, or non-lipid component such as an adjuvant, antigen, peptide, polypeptide, sugar, nucleic acid or other material disclosed herein or as would be known to one of skill in the art. In a non-limiting example, a composition may comprise about 10% to about 20% neutral lipids, and about 33% to about 34% of a cerebroside, and about 1% cholesterol. In another non-limiting example, a liposome may comprise about 4% to about 12% terpenes, wherein about 1% of the micelle is specifically lycopene, leaving about 3% to about 11% of the liposome as comprising other terpenes; and about 10% to about 35% phosphatidyl choline, and about 1% of a non-lipid component. Thus, it is contemplated that compositions of the present invention may comprise any of the lipids, lipid types or other components in any combination or percentage range.

G. Combination Therapy

The compositions and related methods of the present invention, particularly administration of a secreted virulence factor or surface protein, including a variant SpA polypeptide or peptide, and/or other bacterial peptides or proteins to a patient/subject, may also be used in combination with the administration of traditional therapies. These include, but are not limited to, the administration of antibiotics such as streptomycin, ciprofloxacin, doxycycline, gentamycin, chloramphenicol, trimethoprim, sulfamethoxazole, ampicillin, tetracycline or various combinations of antibiotics.

In one aspect, it is contemplated that a polypeptide vaccine and/or therapy is used in conjunction with antibacterial treatment. Alternatively, the therapy may precede or follow the other agent treatment by intervals ranging from minutes to weeks. In embodiments where the other agents

and/or a proteins or polynucleotides are administered separately, one would generally ensure that a significant period of time did not expire between the time of each delivery, such that the agent and antigenic composition would still be able to exert an advantageously combined effect on the subject. In such instances, it is contemplated that one may administer both modalities within about 12-24 h of each other or within about 6-12 h of each other. In some situations, it may be desirable to extend the time period for administration significantly, where several days (2, 3, 4, 5, 6 or 7) to several weeks (1, 2, 3, 4, 5, 6, 7 or 8) lapse between the respective administrations.

Various combinations may be employed, for example antibiotic therapy is "A" and the immunogenic molecule given as part of an immune therapy regime, such as an antigen, is "B":

A/B/A B/A/B B/B/A A/A/B A/B/B B/A/A A/B/B/B B/A/B/B

BBB/A BB/AB A/A/B/B AB/AB A/B/B/A B/B/A/A

B/A/B/A B/A/A/B A/A/A/B B/A/A/A A/B/A/A A/A/B/A

Administration of the immunogenic compositions of the present invention to a patient/subject will follow general protocols for the administration of such compounds, taking into account the toxicity, if any, of the SpA composition, or other compositions described herein. It is expected that the treatment cycles would be repeated as necessary. It also is contemplated that various standard therapies, such as hydration, may be applied in combination with the described therapy.

H. General Pharmaceutical Compositions

In some embodiments, pharmaceutical compositions are administered to a subject. Different aspects of the present invention involve administering an effective amount of a composition to a subject. In some embodiments of the present invention, staphylococcal antigens, members of the Ess pathway, including polypeptides or peptides of the Esa or Esx class, and/or members of sortase substrates may be administered to the patient to protect against infection by one or more *staphylococcus* pathogens. Alternatively, an expression vector encoding one or more such polypeptides or peptides may be given to a patient as a preventative treatment. Additionally, such compounds can be administered in combination with an antibiotic or an antibacterial. Such compositions will generally be dissolved or dispersed in a pharmaceutically acceptable carrier or aqueous medium.

In addition to the compounds formulated for parenteral administration, such as those for intravenous or intramuscular injection, other pharmaceutically acceptable forms include, e.g., tablets or other solids for oral administration; time release capsules; and any other form currently used, including creams, lotions, mouthwashes, inhalants and the like.

The active compounds of the present invention can be formulated for parenteral administration, e.g., formulated for injection via the intravenous, intramuscular, subcutaneous, or even intraperitoneal routes. The preparation of an aqueous composition that contains a compound or compounds that increase the expression of an MHC class I molecule will be known to those of skill in the art in light of the present disclosure. Typically, such compositions can be prepared as injectables, either as liquid solutions or suspensions; solid forms suitable for use to prepare solutions or suspensions upon the addition of a liquid prior to injection can also be prepared; and, the preparations can also be emulsified.

Solutions of the active compounds as free base or pharmacologically acceptable salts can be prepared in water

suitably mixed with a surfactant, such as hydroxypropylcellulose. Dispersions can also be prepared in glycerol, liquid polyethylene glycols, and mixtures thereof and in oils. Under ordinary conditions of storage and use, these preparations contain a preservative to prevent the growth of microorganisms.

The pharmaceutical forms suitable for injectable use include sterile aqueous solutions or dispersions; formulations including sesame oil, peanut oil, or aqueous propylene glycol; and sterile powders for the extemporaneous preparation of sterile injectable solutions or dispersions. In all cases the form must be sterile and must be fluid to the extent that it may be easily injected. It also should be stable under the conditions of manufacture and storage and must be preserved against the contaminating action of microorganisms, such as bacteria and fungi.

The proteinaceous compositions may be formulated into a neutral or salt form. Pharmaceutically acceptable salts, include the acid addition salts (formed with the free amino groups of the protein) and which are formed with inorganic acids such as, for example, hydrochloric or phosphoric acids, or such organic acids as acetic, oxalic, tartaric, mandelic, and the like. Salts formed with the free carboxyl groups can also be derived from inorganic bases such as, for example, sodium, potassium, ammonium, calcium, or ferric hydroxides, and such organic bases as isopropylamine, trimethylamine, histidine, procaine and the like.

The carrier also can be a solvent or dispersion medium containing, for example, water, ethanol, polyol (for example, glycerol, propylene glycol, and liquid polyethylene glycol, and the like), suitable mixtures thereof, and vegetable oils. The proper fluidity can be maintained, for example, by the use of a coating, such as lecithin, by the maintenance of the required particle size in the case of dispersion, and by the use of surfactants. The prevention of the action of microorganisms can be brought about by various antibacterial and antifungal agents, for example, parabens, chlorobutanol, phenol, sorbic acid, thimerosal, and the like. In many cases, it will be preferable to include isotonic agents, for example, sugars or sodium chloride. Prolonged absorption of the injectable compositions can be brought about by the use in the compositions of agents delaying absorption, for example, aluminum monostearate and gelatin.

Sterile injectable solutions are prepared by incorporating the active compounds in the required amount in the appropriate solvent with various of the other ingredients enumerated above, as required, followed by filtered sterilization. Generally, dispersions are prepared by incorporating the various sterilized active ingredients into a sterile vehicle which contains the basic dispersion medium and the required other ingredients from those enumerated above. In the case of sterile powders for the preparation of sterile injectable solutions, the preferred methods of preparation are vacuum-drying and freeze-drying techniques, which yield a powder of the active ingredient, plus any additional desired ingredient from a previously sterile-filtered solution thereof.

Administration of the compositions according to the present invention will typically be via any common route. This includes, but is not limited to oral, nasal, or buccal administration. Alternatively, administration may be by orthotopic, intradermal, subcutaneous, intramuscular, intraperitoneal, intranasal, or intravenous injection. In certain embodiments, a vaccine composition may be inhaled (e.g., U.S. Pat. No. 6,651,655, which is specifically incorporated by reference). Such compositions would normally be administered as pharmaceutically acceptable compositions that include physi-

ologically acceptable carriers, buffers or other excipients. As used herein, the term "pharmaceutically acceptable" refers to those compounds, materials, compositions, and/or dosage forms which are, within the scope of sound medical judgment, suitable for contact with the tissues of human beings and animals without excessive toxicity, irritation, allergic response, or other problem complications commensurate with a reasonable benefit/risk ratio. The term "pharmaceutically acceptable carrier," means a pharmaceutically acceptable material, composition or vehicle, such as a liquid or solid filler, diluent, excipient, solvent or encapsulating material, involved in carrying or transporting a chemical agent.

For parenteral administration in an aqueous solution, for example, the solution should be suitably buffered, if necessary, and the liquid diluent first rendered isotonic with sufficient saline or glucose. These particular aqueous solutions are especially suitable for intravenous, intramuscular, subcutaneous, and intraperitoneal administration. In this connection, sterile aqueous media which can be employed will be known to those of skill in the art in light of the present disclosure. For example, one dosage could be dissolved in isotonic NaCl solution and either added to hypodermoclysis fluid or injected at the proposed site of infusion, (see for example, Remington's Pharmaceutical Sciences, 1990). Some variation in dosage will necessarily occur depending on the condition of the subject. The person responsible for administration will, in any event, determine the appropriate dose for the individual subject.

An effective amount of therapeutic or prophylactic composition is determined based on the intended goal. The term "unit dose" or "dosage" refers to physically discrete units suitable for use in a subject, each unit containing a predetermined quantity of the composition calculated to produce the desired responses discussed above in association with its administration, i.e., the appropriate route and regimen. The quantity to be administered, both according to number of treatments and unit dose, depends on the protection desired.

Precise amounts of the composition also depend on the judgment of the practitioner and are peculiar to each individual. Factors affecting dose include physical and clinical state of the subject, route of administration, intended goal of treatment (alleviation of symptoms versus cure), and potency, stability, and toxicity of the particular composition.

Upon formulation, solutions will be administered in a manner compatible with the dosage formulation and in such amount as is therapeutically or prophylactically effective. The formulations are easily administered in a variety of dosage forms, such as the type of injectable solutions described above.

In Vitro, Ex Vivo, or In Vivo Administration

As used herein, the term *in vitro* administration refers to manipulations performed on cells removed from or outside of a subject, including, but not limited to cells in culture. The term *ex vivo* administration refers to cells which have been manipulated *in vitro*, and are subsequently administered to a subject. The term *in vivo* administration includes all manipulations performed within a subject.

In certain aspects of the present invention, the compositions may be administered either *in vitro*, *ex vivo*, or *in vivo*. In certain *in vitro* embodiments, autologous B-lymphocyte cell lines are incubated with a virus vector of the instant invention for 24 to 48 hours or with a variant SpA and/or coagulase and/or any other composition described herein for two hours. The transduced cells can then be used for *in vitro* analysis, or alternatively for *ex vivo* administration. U.S. Pat. Nos. 4,690,915 and 5,199,942, both incorporated herein

by reference, disclose methods for ex vivo manipulation of blood mononuclear cells and bone marrow cells for use in therapeutic applications.

J. Antibodies and Passive Immunization

Another aspect of the invention is a method of preparing an immunoglobulin for use in prevention or treatment of staphylococcal infection comprising the steps of immunizing a recipient or donor with the vaccine of the invention and isolating immunoglobulin from the recipient or donor. An immunoglobulin prepared by this method is a further aspect of the invention. A pharmaceutical composition comprising the immunoglobulin of the invention and a pharmaceutically acceptable carrier is a further aspect of the invention which could be used in the manufacture of a medicament for the treatment or prevention of staphylococcal disease. A method for treatment or prevention of staphylococcal infection comprising a step of administering to a patient an effective amount of the pharmaceutical preparation of the invention is a further aspect of the invention.

Inocula for polyclonal antibody production are typically prepared by dispersing the antigenic composition in a physiologically tolerable diluent such as saline or other adjuvants suitable for human use to form an aqueous composition. An immunostimulatory amount of inoculum is administered to a mammal and the inoculated mammal is then maintained for a time sufficient for the antigenic composition to induce protective antibodies.

The antibodies can be isolated to the extent desired by well known techniques such as affinity chromatography (Harlow and Lane, 1988). Antibodies can include antiserum preparations from a variety of commonly used animals, e.g. goats, primates, donkeys, swine, horses, guinea pigs, rats or man.

An immunoglobulin produced in accordance with the present invention can include whole antibodies, antibody fragments or subfragments. Antibodies can be whole immunoglobulins of any class (e.g., IgG, IgM, IgA, IgD or IgE), chimeric antibodies or hybrid antibodies with dual specificity to two or more antigens of the invention. They may also be fragments (e.g., F(ab')₂, Fab', Fab, Fv and the like) including hybrid fragments. An immunoglobulin also includes natural, synthetic, or genetically engineered proteins that act like an antibody by binding to specific antigens to form a complex.

A vaccine of the present invention can be administered to a recipient who then acts as a source of immunoglobulin, produced in response to challenge from the specific vaccine. A subject thus treated would donate plasma from which hyperimmune globulin would be obtained via conventional plasma fractionation methodology. The hyperimmune globulin would be administered to another subject in order to impart resistance against or treat staphylococcal infection. Hyperimmune globulins of the invention are particularly useful for treatment or prevention of staphylococcal disease in infants, immune compromised individuals, or where treatment is required and there is no time for the individual to produce antibodies in response to vaccination.

An additional aspect of the invention is a pharmaceutical composition comprising two or more monoclonal antibodies (or fragments thereof; preferably human or humanised) reactive against at least two constituents of the immunogenic composition of the invention, which could be used to treat or prevent infection by Gram positive bacteria, preferably staphylococci, more preferably *S. aureus* or *S. epidermidis*. Such pharmaceutical compositions comprise monoclonal antibodies that can be whole immunoglobulins of any class, chimeric antibodies, or hybrid antibodies with specificity to

two or more antigens of the invention. They may also be fragments (e.g., F(ab')₂, Fab', Fab, Fv and the like) including hybrid fragments.

Methods of making monoclonal antibodies are well known in the art and can include the fusion of splenocytes with myeloma cells (Kohler and Milstein, 1975; Harlow and Lane, 1988). Alternatively, monoclonal Fv fragments can be obtained by screening a suitable phage display library (Vaughan et al., 1998). Monoclonal antibodies may be humanized or part humanized by known methods.

VII. EXAMPLES

The following examples are given for the purpose of illustrating various embodiments of the invention and are not meant to limit the present invention in any fashion. One skilled in the art will appreciate readily that the present invention is well adapted to carry out the objects and obtain the ends and advantages mentioned, as well as those objects, ends and advantages inherent herein. The present examples, along with the methods described herein are presently representative of preferred embodiments, are exemplary, and are not intended as limitations on the scope of the invention. Changes therein and other uses which are encompassed within the spirit of the invention as defined by the scope of the claims will occur to those skilled in the art.

Example 1

Non-Toxicigenic Protein A Variants as Subunit Vaccines to Prevent *Staphylococcus aureus* Infections

An Animal Model for *S. aureus* Infection

BALB/c mice were infected by intravenous injection with 1×10^7 CFU of the human clinical isolate *S. aureus* Newman (Baba et al., 2007). Within 6 hours following infection, 99.999% of staphylococci disappeared from the blood stream and were distributed via the vasculature. Staphylococcal dissemination to peripheral tissues occurred rapidly, as the bacterial load in kidney and other peripheral organ tissues reached 1×10^5 CFU g⁻¹ within the first three hours. The staphylococcal load in kidney tissues increased by 1.5 log CFU within twenty-four hours. Forty-eight hours following infection, mice developed disseminated abscesses in multiple organs, detectable by light microscopy of hematoxylin-eosin stained, thin-sectioned kidney tissue. The initial abscess diameter was 524 μm (±65 μm); lesions were initially marked by an influx of polymorphonuclear leukocytes (PMNs) and harbored no discernable organization of staphylococci, most of which appeared to reside within PMNs. On day 5 of infection, abscesses increased in size and enclosed a central population of staphylococci, surrounded by a layer of eosinophilic, amorphous material and a large cuff of PMNs. Histopathology revealed massive necrosis of PMNs in proximity to the staphylococcal nidus at the center of abscess lesions as well as a mantle of healthy phagocytes. A rim of necrotic PMNs were observed at the periphery of abscess lesions, bordering eosinophilic, amorphous material that separates healthy renal tissue from lesions. Abscesses eventually reached a diameter of 1,524 μm on day 15 or 36. At later time intervals, the staphylococcal load was increased to 10^4 - 10^6 CFU g⁻¹ and growing abscess lesions migrated towards the organ capsule. Peripheral lesions were prone to rupture, thereby releasing necrotic material and staphylococci into the peritoneal cavity or the retroperito-

renal space. These events resulted in bacteremia as well as a secondary wave of abscesses, eventually precipitating a lethal outcome.

To enumerate staphylococcal load in renal tissue, animals were killed, their kidneys excised and tissue homogenate spread on agar media for colony formation. On day 5 of infection, a mean of 1×10^6 CFU g^{-1} renal tissue for *S. aureus* Newman was observed. To quantify abscess formation, kidneys were visually inspected, and each individual organ was given a score of one or zero. The final sum was divided by the total number of kidneys to calculate percent surface abscesses (Table 3). In addition, randomly chosen kidneys were fixed in formalin, embedded, thin sectioned, and stained with hematoxylin-eosin. For each kidney, four sagittal sections at 200 μ m intervals were viewed by microscopy. The numbers of lesions were counted for each section and averaged to quantify the number of abscesses within the kidneys. *S. aureus* Newman caused 4.364 ± 0.889 abscesses per kidney, and surface abscesses were observed on 14 out of 20 kidneys (70%) (Table 3).

When examined by scanning electron microscopy, *S. aureus* Newman was located in tightly associated lawns at the center of abscesses. Staphylococci were contained by an amorphous pseudocapsule that separated bacteria from the cuff of abscesses leukocytes. No immune cells were observed in these central nests of staphylococci, however occasional red blood cells were located among the bacteria. Bacterial populations at the abscess center, designated staphylococcal abscess communities (SAC), appeared homogenous and coated by an electron-dense, granular material. The kinetics of the appearance of infectious lesions and the morphological attributes of abscesses formed by *S. aureus* Newman were similar to those observed following mouse infection with *S. aureus* USA300 (LAC), the current epidemic community-acquired methicillin-resistant *S. aureus* (CA-MRSA) clone in the United States (Diep et al., 2006).

2002; Weiss et al., 2004). Compared to the wild-type parent (Baba et al., 2007), an isogenic *srtA* variant (*AsrtA*) failed to form abscess lesions on either macroscopic or histopathology examination on days 2, 5, or 15. In mice infected with the *strA* mutant, only 1×10^4 CFU g^{-1} was recovered from kidney tissue on day 5 of infection, which is a $2.046 \log_{10}$ CFU g^{-1} reduction compared to the wild-type parent strain ($P=6.73 \times 10^{-6}$). A similar defect was observed for the *srtA* mutant of MRSA strain USA300 (data not shown). Scanning electron microscopy showed that *srtA* mutants were highly dispersed and often associated with leukocytes in otherwise healthy renal tissue. On day fifteen following infection, *srtA* mutants were cleared from renal tissues, a $\geq 3.5 \log_{10}$ CFU g^{-1} reduction compared to the wild-type (Table 3). Thus, sortase A anchored surface proteins enable the formation of abscess lesions and the persistence of bacteria in host tissues, wherein staphylococci replicate as communities embedded in an extracellular matrix and shielded from surrounding leukocytes by an amorphous pseudocapsule.

Sortase A anchors a large spectrum of proteins with LPXTG motif sorting signals to the cell wall envelope, thereby providing for the surface display of many virulence factors (Mazmanian et al., 2002). To identify surface proteins required for staphylococcal abscess formation, bursa aurealis insertions were introduced in 5' coding sequences of genes that encode polypeptides with LPXTG motif proteins (Bae et al., 2004) and these mutations were transduced into *S. aureus* Newman. Mutations in the structural gene for Protein A (*spa*) reduced the staphylococcal load in infected mouse kidney tissues by $1.004 \log_{10}$ ($P=0.0144$). When analyzed for their ability to form abscesses in kidney tissues by histopathology, we observed that the *spa* mutants were unable to form abscesses as compared with the wild-type parent strain *S. aureus* Newman (wild-type *S. aureus* New-

TABLE 3

Genotype	Genetic requirements for <i>S. aureus</i> Newman abscess formation in mice					
	Staphylococcal load in kidney tissue			Abscess formation in kidney tissue		
	^a \log_{10} CFU g^{-1} tissue	^b Significance (P-value)	^c Reduction (\log_{10} CFU g^{-1})	^d Surface abscesses (%)	^e Number of abscesses per kidney	^f Significance (P-value)
wild-type	6.141 \pm 0.192	—	—	70	4.364 \pm 0.889	—
<i>AsrtA</i>	4.095 \pm 0.347	6.7×10^{-6}	2.046	0	0.000 \pm 0.000	0.0216
<i>spa</i>	5.137 \pm 0.374	0.0144	1.004	13	0.375 \pm 0.374	0.0356

^aMeans of staphylococcal load calculated as \log_{10} CFU g^{-1} in homogenized renal tissues 5 days following infection in cohorts of fifteen BALB/c mice per challenge strain. Standard error of the means (\pm SEM) is indicated.

^bStatistical significance was calculated with the Students t-test and P-values recorded; P-values < 0.05 were deemed significant.

^cReduction in bacterial load calculated as \log_{10} CFU g^{-1} .

^dAbscess formation in kidney tissues five days following infection was measured by macroscopic inspection (% positive)

^eHistopathology of hematoxylin-eosin stained, thin sectioned kidneys from eight to ten animals; the average number of abscesses per kidney was recorded and averaged again for the final mean (\pm SEM).

^fStatistical significance was calculated with the Students t-test and P-values recorded; P-values < 0.05 were deemed significant.

S. aureus Protein A (Spa) Mutants are Avirulent and Cannot Form Abscesses

Sortase A is a transpeptidase that immobilizes nineteen surface proteins in the envelope of *S. aureus* strain Newman (Mazmanian et al., 1999; Mazmanian et al., 2000). Earlier work identified sortase A as a virulence factor in multiple animal model systems, however the contributions of this enzyme and its anchored surface proteins to abscess formation or persistence have not yet been revealed (Jonsson et al.,

man 4.364 ± 0.889 abscesses per kidney vs. the isogenic *spa* mutant with 0.375 ± 0.374 lesions; $P=0.0356$).

Protein A Blocks Innate and Adaptive Immune Responses.

Studies identified Protein A as a critical virulence factor during the pathogenesis of *S. aureus* infections. Earlier work demonstrated that Protein A impedes phagocytosis of staphylococci by binding the Fc component of immunoglobulin (Jensen 1958; Uhlén et al., 1984), activates platelet aggregation via the von Willebrand factor (Hartleib et al.,

2000), functions as a B cell superantigen by capturing the F(ab)₂ region of VH3 bearing IgM (Roben et al., 1995), and, through its activation of TNFR1, can initiate staphylococcal pneumonia (Gomez et al., 2004). Due to the fact that Protein A captures immunoglobulin and displays toxic attributes, the possibility that this surface molecule may function as a vaccine in humans has not been rigorously pursued. The inventors demonstrate for the first time that Protein A variants no longer able to bind to immunoglobulins, vWF and TNFR-1 are removed of their toxigenic potential and are able to stimulate humoral immune responses that protect against staphylococcal disease.

Molecular Basis of Protein A Surface Display and Function.

Protein A is synthesized as a precursor in the bacterial cytoplasm and secreted via its YSIRK (SEQ ID NO.: 72) signal peptide at the cross wall, i.e., the cell division septum of staphylococci (FIG. 1A). (DeDent et al., 2007; DeDent et al., 2008). Following cleavage of the C-terminal LPXTG (SEQ ID NO.: 71) sorting signal, Protein A is anchored to bacterial peptidoglycan crossbridges by sortase A (Schneewind et al., 1995; Mazmanian et al., 1999; Mazmanian et al., 2000). Protein A is the most abundant surface protein of staphylococci; the molecule is expressed by virtually all *S. aureus* strains (Saïd-Salim et al., 2003; Cespedes et al., 2005; Kennedy et al., 2008). Staphylococci turn over 15-20% of their cell wall per division cycle (Navarre and Schneewind 1999). Murine hydrolases cleave the glycan strands and wall peptides of peptidoglycan, thereby releasing Protein A with its attached C-terminal cell wall disaccharide tetrapeptide into the extracellular medium (Ton-That et al., 1999). Thus, by physiological design, Protein A is both anchored to the cell wall and displayed on the bacterial surface but also released into surrounding tissues during host infection (Marraffini et al., 2006).

Protein A captures immunoglobulins on the bacterial surface and this biochemical activity enables staphylococcal escape from host innate and acquired immune responses (Jensen 1958; Goodyear and Silverman 2004). Interestingly, region X of Protein A (Guss et al., 1984), a repeat domain that tethers the IgG binding domains to the LPXTG sorting signal/cell wall anchor, is perhaps the most variable portion of the staphylococcal genome (Schneewind et al., 1992; Saïd-Salim et al., 2003). Each of the five immunoglobulin binding domains of Protein A (SpA), formed from three helix bundles and designated E, D, A, B, and C, exerts similar structural and functional properties (Sjödahl 1977; Jansson et al., 1998). The solution and crystal structure of domain D has been solved both with and without the Fc and V_{H3} (Fab) ligands, which bind Protein A in a non-competitive manner at distinct sites (Graille et al., 2000).

In the crystal structure complex, the Fab interacts with helix II and helix III of domain D via a surface composed of four VH region β -strands (Graille et al., 2000). The major axis of helix II of domain D is approximately 50° to the orientation of the strands, and the interhelical portion of domain D is most proximal to the CO strand. The site of interaction on Fab is remote from the Ig light chain and the heavy chain constant region. The interaction involves the following domain D residues: Asp-36 of helix II as well as Asp-37 and Gln-40 in the loop between helix II and helix III, in addition to several other residues with SpA-D (Graille et al., 2000). Both interacting surfaces are composed predominantly of polar side chains, with three negatively charged residues on domain D and two positively charged residues on the 2A2 Fab buried by the interaction, providing an overall electrostatic attraction between the two molecules.

Of the five polar interactions identified between Fab and domain D, three are between side chains. A salt bridge is formed between Arg-H19 and Asp-36 and two hydrogen bonds are made between Tyr-H59 and Asp-37 and between Asn-H82a and Ser-33. Because of the conservation of Asp-36 and Asp-37 in all five IgG binding domains of Protein A, these residues were selected for mutagenesis.

The SpA-D sites responsible for Fab binding are structurally separate from the domain surface that mediates Fc γ binding. The interaction of Fc γ with domain B primarily involves residues in helix I with lesser involvement of helix II (Deisenhofer 1981; Gouda et al., 1992). With the exception of the Gln-32, a minor contact in both complexes, none of the residues that mediate the Fc γ interaction are involved in Fab binding. To examine the spatial relationship between these different Ig-binding sites, the SpA domains in these complexes have been superimposed to construct a model of a complex between Fab, the SpA-domain D, and the Fc γ molecule. In this ternary model, Fab and Fc γ form a sandwich about opposite faces of the helix II without evidence of steric hindrance of either interaction. These findings illustrate how, despite its small size (i.e., 56-61 aa), a SpA domain can simultaneously display both activities, explaining experimental evidence that the interactions of Fab with an individual domain are noncompetitive. Residues for the interaction between SpA-D and Fc γ are Gln-9 and Gln-10.

In contrast, occupancy of the Fc portion of IgG on the domain D blocks its interaction with vWF A1 and probably also TNFR1 (O'Seaghdha et al., 2006). Mutations in residues essential for IgG Fc binding (F5, Q9, Q10, S11, F13, Y14, L17, N28, I31 and K35) are also required for vWF A1 and TNFR1 binding (Cedergren et al., 1993; Gómez et al., 2006; O'Seaghdha et al. 2006), whereas residues critical for the V_{H3} interaction (Q26, G29, F30, S33, D36, D37, Q40, N43, E47) have no impact on the binding activities of IgG Fc, vWF A1 or TNFR1 (Jansson et al., 1998; Graille et al., 2000). The Protein A immunoglobulin Fab binding activity targets a subset of B cells that express VH3 family related IgM on their surface, i.e. these molecules function as VH3 type B cell receptors (Roben et al., 1995). Upon interaction with SpA, these B cells rapidly proliferate and then commit to apoptosis, leading to preferential and prolonged deletion of innate-like B lymphocytes (i.e. marginal zone B cells and follicular B2 cells) (Goodyear and Silverman 2003; Goodyear and Silverman 2004). It is important to note that more than 40% of circulating B cells are targeted by the Protein A interaction and the VH3 family represents the largest family of human B cell receptors to impart protective humoral responses against pathogens (Goodyear and Silverman 2003; Goodyear and Silverman 2004). Thus, Protein A functions analogously to staphylococcal superantigens (Roben et al., 1995), albeit that the latter class of molecules, for example SEB, TSST-1, TSST-2, form complexes with the T cell receptor to inappropriately stimulate host immune responses and thereby precipitating characteristic disease features of staphylococcal infections (Roben et al., 1995; Tiedemann et al., 1995). Together these findings document the contributions of Protein A in establishing staphylococcal infections and in modulating host immune responses.

Non-Toxicogenic Variant of Protein A.

The inventors have developed a non-toxicogenic variant of staphylococcal Protein A and, with this reagent in hand, aimed for the first time to measure the immune response of animals to Protein A immunization. Further, the inventors address whether immunization of animals with a non-toxi-

genic variant of Protein A could generate immune responses that raise protective immunity against staphylococcal infection.

To perturb the IgG Fc, vWF A1 and TNFR1 binding activities of Protein A, glutamine (Q) residues 9 and 10 [the numbering here is derived from that established for the SpA domain D] were modified generating lysine or glycine substitutions for both glutamines with the expectation that these substitutions abolish the ion bonds formed between wild-type Protein A and its ligands. The added effect of the dual lysine substitutions may be that these positively charged residues institute a repellent charge for immunoglobulins. To perturb IgM Fab VH3 binding, the inventors selected the aspartate (D) residues 36 and 37 of SpA-D, each of which is required for the association of Protein A with the B cell receptor. D36 and D37 were both substituted with alanine. The Q9,10K and D36,37A mutations were combined in the recombinant molecule SpA-D_{Q9,10K;D36,37A} and examined for the binding attributes of Protein A.

In brief, the Protein A (spa) genomic sequence of *Staphylococcus aureus* N315 was PCR amplified with the primers (GCTGCACATATGGCGCAACACGATGAAGCTCAAC [5' primer] (SEQ ID NO:35) and AGTGGATCCTTATGCTTTGTAGCATCTGC [3' primer] (SEQ ID NO:36)), cloned into the pET15b vector (pYSJ1, codons 48-486) (Stranger-Jones, et al., 2006) and recombinant plasmid transformed into *E. coli* BL21(DE3) (Studier et al., 1990). The Protein A product derived from pYSJ1 harbors SpA residues 36-265 fused to the N-terminal His₆ tag (MGSSH-HHHHHSSGLVPRGS (SEQ ID NO:37)). Following IPTG inducible expression, recombinant N-terminal His₆-tagged SpA was purified by affinity chromatography on Ni-NTA resin (Stranger-Jones et al., 2006). The domain D of SpA (SpA-D) was PCR amplified with a pair of specific primers (AACATATGTTCAACAAAGATCAACAAAGC [5' primer] (SEQ ID NO:38) and AAGGATCCAGATTCGTTTAATTTTTAGC [3' primer] (SEQ ID NO:39)), subcloned into the pET15b vector (pHAN1, spa codons 212-261) and recombinant plasmid transformed into *E. coli* BL21(DE3) to express and purify recombinant N-terminal His₆-tagged protein. To generate mutations in the SpA-D coding sequence, sets of two pairs of primers were synthesized (for D to A substitutions: CTTCATCAAAGCTCTAAAGCCGCCCAAGCCAAAGCACTAAC [5' primer] (SEQ ID NO:40) and GTTAGTGCTTTGGCTTGGGGCGCGCTTTAAGACTTTGAATGAAG [3' primer] (SEQ ID NO:41); for Q to K substitutions CATATGTCAACAAAGATAAAAAAGCGCCTTCTATGAAATC [5' primer] (SEQ ID NO:42) and GATTCATAGAAGGCGCTTTTTTATCTTTGTTGAACATATG [3' primer] (SEQ ID NO:43); for Q to G substitutions CATATGTTCAACAAAGATGGAGGAAGCGCCTTCTATGAAATC [5' primer] (SEQ ID NO:44) and GATTCATAGAAGGCGCTTCCATCTTTGTTGAACATATG [3' primer] (SEQ ID NO:45). Primers were used for quick-change mutagenesis protocols. Following mutagenesis, DNA sequences were confirmed for each of the recombinant proteins: SpA, SpA-D and SpA-D_{Q9,10K;D36,37A} and SpA-D_{Q9,10K;D36,37A}. All proteins were purified from lysates of recombinant *E. coli* using Ni-NTA chromatography and subsequently dialyzed against PBS and stored at 4° C.

To measure binding of immunoglobulin to Protein A and its variants, 200 µg of purified protein was diluted into a 1 ml volume using column buffer (50 mM Tris-HCl, 150 mM NaCl, pH7.5) and then loaded onto a pre-equilibrated Ni-NTA column (1 ml bed volume). Columns were washed with 10 ml of column buffer. 200 µg of purified human IgG was

diluted in a total volume of 1 ml column buffer and then applied to each of the columns charged with Protein A and its variants. The columns were subsequently washed with 5 ml wash buffer (10 mM imidazole in column buffer) and 5 ml column buffer. Protein samples were eluted with 2 ml elution buffer (500 mM imidazole in column buffer), fractions collected and aliquots subjected to SDS-PAGE gel electrophoresis, followed by COOMASSIE-BLUE staining. As shown in FIG. 1C, wild-type Protein A (SpA) and its SpA-domain D both retained immunoglobulin during chromatography. In contrast, the SpA-D_{Q9,10K;D36,37A} variant did not bind to immunoglobulin.

To quantify the binding of Protein A and its variants to the Fc portion of immunoglobulin and the VH3 domain of Fab, HRP conjugated human immunoglobulin G [hIgG], the Fc portion of human IgG [hFc] and the F(ab)₂ portion of human IgG [hF(ab)₂] as well as ELISA assays were used to quantify the relative amount binding to Protein A and its variants. The data in FIG. 1D demonstrate the binding of SpA and SpA-D to hIgG and hFc, whereas SpA-D_{Q9,10K;D36,37A} and SpA-D_{Q9,10K;D36,37A} displayed only background binding activities. SpA bound similar amounts of hFc and hF(ab)₂, however the binding of SpA-D to hF(ab)₂ was reduced compared to full length SpA. This result suggests that the presence of multiple IgG binding domains may cooperatively increase the ability of Protein A to bind to the B cell receptor. When compared with the reduced binding power of SpA-D for hF(ab)₂, of the two variants only SpA-D_{Q9,10K;D36,37A} displayed a significant reduction in the ability to bind the VH3 domain of immunoglobulin. To examine the toxigenic attributes of SpA-D and its variants, purified proteins were injected into mice, which were sacrificed after 4 hours to remove their spleens. Organ tissue was homogenized, capsular material removed and B cells stained with fluorescent CD19 antibodies. Following FACS analysis to quantify the abundance of B cells in splenic tissues, it was observed that SpA-D caused a 5% drop in the B cell count compared to a mock (PBS) control (FIG. 1E). In contrast, SpA-D_{Q9,10K;D36,37A} did not cause a reduction in B-cell counts, indicating that the mutant molecule had lost its toxigenic attributes of stimulating B cell proliferation and death (FIG. 1E). In summary, amino acid substitutions in the SpA-D residues Q9, Q10, D36, and D37 abolished the ability of Protein A domains to bind immunoglobulins or exert toxigenic functions in human and animal tissues.

Non-toxicogenic Protein A variants elicit vaccine protection. To test whether or not Protein A and its variants can function as vaccine antigens, SpA, SpA-D, SpA-D_{Q9,10K;D36,37A} and SpA-D_{Q9,10K;D36,37A} were emulsified with complete or incomplete Freund's adjuvant and immunized 4 week old BALB/c mice on day 1 and day 11 with 50 µg of purified protein. Cohort of animals (n=5) were analyzed for humoral immune responses to immunization by bleeding the animals before (day 0) and after the immunization schedule (day 21). Table 4 indicates that immunized mice generated only a modest humoral immune response directed at wild-type Protein A or its SpA-D module, whereas the amount of antibody raised following immunization with SpA-D_{Q9,10K;D36,37A} or SpA-D_{Q9,10K;D36,37A} was increased four to five fold. Following intravenous challenge with 1×10⁷ CFU *S. aureus* Newman, animals were killed on day 4, their kidneys removed and either analyzed for staphylococcal load (by plating tissue homogenate on agar plates and enumerating colony forming units, CFU) or histopathology. As expected, mock (PBS) immunized mice (n=19) harbored 6.46 log₁₀ (±0.25) CFU in kidney tissue and infectious lesions were organized into 3.7 (±1.2) abscesses per organ (n=10)(Table

4). Immunization of animals with SpA led to a 2.51 log₁₀ CFU reduction on day 5 (P=0.0003) with 2.1 (±1.2) abscesses per organ. The latter data indicate that there was no significant reduction in abscess formation (P=0.35). Immunization with SpA-D generated similar results: a 2.03 log₁₀ CFU reduction on day 5 (P=0.0001) with 1.5 (±0.8) abscesses per organ (P=0.15). In contrast, immunization with SpA-D_{Q9,10K;D36,37A} or SpA-D_{Q9,10G;D36,37A} created increased protection, with 3.07 log₁₀ and 3.03 log₁₀ CFU reduction on day 4, respectively (statistical significance P<0.0001 for both observations). Further, immunization with both SpA-D_{Q9,10K;D36,37A} and SpA-D_{Q9,10G;D36,37A} generated significant protection from staphylococcal abscess formation, as only 0.5 (±0.4) and 0.8 (±0.5) infectious lesions per organ (P=0.02 and P=0.04) were identified. Thus, immunization with non-toxic Protein A variants generates increased humoral immune responses for Protein A and provides protective immunity against staphylococcal challenge. These data indicate that Protein A is an ideal candidate for a human vaccine that prevents *S. aureus* disease.

These exciting results have several implications for the design of a human vaccine. First, the generation of substitution mutations that affect the ability of the immunoglobulin binding domains of Protein A, either alone or in combination of two or more domains, can generate non-toxic variants suitable for vaccine development. It seems likely that a combination of mutant IgG binding domains closely resembling the structure of Protein A can generate even better humoral immune responses as is reported here for the SpA-domain D alone. Further, a likely attribute of Protein A specific antibodies may be that the interaction of antigen binding sites with the microbial surface can neutralize the ability of staphylococci to capture immunoglobulins via their Fc portion or to stimulate the B cell receptor via the VH3 binding activities.

Materials and Methods

Murine Abscess—

BALB/c mice (24-day-old female, 8-10 mice per group, Charles River Laboratories, Wilmington, Mass.) are immunized by intramuscular injection into the hind leg with purified protein (Chang et al., 2003; Schneewind et al., 1992). Purified SpA, SpA-D or SpA-D_{Q9,10K;D36,37A} (50 µg protein) is administered on days 0 (emulsified 1:1 with complete Freund's adjuvant) and 11 (emulsified 1:1 with incomplete Freund's adjuvant). Blood samples are drawn by retroorbital bleeding on days 0, 11, and 20. Sera are examined by ELISA for IgG titers for specific SpA-D and SpA-D_{Q9,10K;D36,37A} binding activity. Immunized animals are challenged on day 21 by retroorbital injection of 100 µl of *S. aureus* Newman or *S. aureus* USA300 suspension (1×10⁷ cfu). For this, overnight cultures of *S. aureus* Newman are diluted 1:100 into fresh tryptic soy broth and grown for 3 h at 37° C. Staphylococci are centrifuged, washed twice, and diluted in PBS to yield an A600 of 0.4 (1×10⁸ cfu per ml). Dilutions are verified experimentally by agar plating and colony formation. Mice are anesthetized by intraperitoneal injection of 80-120 mg of ketamine and 3-6 mg of xylazine per kilogram of body weight and infected by retroorbital injection. On day 5 or 15 following challenge, mice are euthanized by compressed CO₂ inhalation. Kidneys are removed and homogenized in 1% TRITON X-100. Aliquots are diluted and plated on agar medium for triplicate determination of cfu. For histology, kidney tissue is incubated at room temperature in 10% formalin for h. Tissues are embedded in paraffin, thin-sectioned, stained with hematoxylin-eosin, and examined by microscopy.

Murine Lethal Infection—

BALB/c mice (24-day-old female, 8-10 mice per group, Charles River Laboratories, Wilmington, Mass.) are immunized by intramuscular injection into the hind leg with

TABLE 4

Antigen	Bacterial load in kidney (n = number of mice)			Abscess formation in mice (n = number of mice)					
	^a log ₁₀ CFU g ⁻¹	^b Reduction	^c P value	IgG titer	^d Surface abscess	Reduction	^e Histopathology	Reduction	^f p value
Mock	6.46 ± 0.25 (n = 19)	—	—	<100	14/19 (70%)	—	3.7 ± 1.2 (n = 10)	—	—
SpA	3.95 ± 0.56 (n = 20)	2.51	0.0003	1706 ± 370	10/20 (50%)	32%	2.1 ± 1.2 (n = 10)	2.2	0.35
SpA-D	4.43 ± 0.41 (n = 18)	2.03	0.0001	381 ± 27	10/18 (55%)	25%	1.5 ± 0.8 (n = 10)	2.2	0.15
SpA-D1	3.39 ± 0.50 (n = 19)	3.07	<0.0001	5600 ± 801	6/20 (30%)	59%	0.5 ± 0.4 (n = 10)	3.2	0.02
SpA-D2	3.43 ± 0.46 (n = 19)	3.03	<0.0001	3980 ± 676	6/19 (32%)	57%	0.8 ± 0.5 (n = 10)	2.9	0.04

^aMeans of staphylococcal load calculated as log₁₀ CFU g⁻¹ in homogenized renal tissues 4 days following infection in cohorts of 18 to 20 BALB/c mice. Standard error of the means (±SEM) is indicated.

^cStatistical significance was calculated with the Students t-test and P-values recorded; P-values < 0.05 were deemed significant.

^bReduction in bacterial load calculated as log₁₀ CFU g⁻¹.

^dAbscess formation in kidney tissues four days following infection was measured by macroscopic inspection (% positive)

^eHistopathology of hematoxylin-eosin stained, thin sectioned kidneys from ten animals; the number of abscesses per kidney was recorded and averaged for the final mean (±SEM).

^fStatistical significance was calculated with the Students t-test and P-values recorded; P-values < 0.05 were deemed significant.

SpA-D1 and SpA-D2 represent SpA-D_{Q9, 10K; D36, 37A} and SpA-D_{Q9, 10G; D36, 37A}, respectively.

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Vaccine Protection in Murine Abscess, Murine Lethal Infection, and Murine Pneumonia Models.

Three animal models have been established for the study of *S. aureus* infectious disease. These models are used here to examine the level of protective immunity provided via the generation of Protein A specific antibodies.

65 purified SpA, SpA-D or SpA-D_{Q9,10K;D36,37A} (50 µg protein). Vaccine is administered on days 0 (emulsified 1:1 with complete Freund's adjuvant) and 11 (emulsified 1:1 with incomplete Freund's adjuvant). Blood samples are drawn by retroorbital bleeding on days 0, 11, and 20. Sera are examined by ELISA for IgG titers with specific SpA-D and

SpA-D_{Q9,10K;D36,37A} binding activity. Immunized animals are challenged on day 21 by retroorbital injection of 100 μ l of *S. aureus* Newman or *S. aureus* USA300 suspension (15×10^7 cfu) (34). For this, overnight cultures of *S. aureus* Newman are diluted 1:100 into fresh tryptic soy broth and grown for 3 h at 37° C. Staphylococci are centrifuged, washed twice, diluted in PBS to yield an A₆₀₀ of 0.4 (1×10^8 cfu per ml) and concentrated. Dilutions are verified experimentally by agar plating and colony formation. Mice are anesthetized by intraperitoneal injection of 80-120 mg of ketamine and 3-6 mg of xylazine per kilogram of body weight. Immunized animals are challenged on day 21 by intraperitoneal inject with 2×10^{10} cfu of *S. aureus* Newman or $3-10 \times 10^9$ cfu of clinical *S. aureus* isolates. Animals are monitored for 14 days, and lethal disease is recorded.

Murine Pneumonia Model—

S. aureus strains Newman or USA300 (LAC) are grown at 37° C. in tryptic soy broth/agar to OD₆₆₀ 0.5. 50-ml culture aliquots are centrifuged, washed in PBS, and suspended in 750 μ l PBS for mortality studies ($3-4 \times 10^8$ CFU per 30 μ l volume), or 1,250 μ l PBS (2×10^8 CFU per 30 μ l volume) for bacterial load and histopathology experiments (2, 3). For lung infection, 7-wk-old C57BL/6J mice (The Jackson Laboratory) are anesthetized before inoculation of 30 μ l of *S. aureus* suspension into the left nare. Animals are placed into the cage in a supine position for recovery and observed for 14 days. For active immunization, 4-wk-old mice receive 20 μ g SpA-D or SpA-D_{Q9,10K;D36,37A} in CFA on day 0 via the i.m. route, followed by a boost with 20 μ g SpA-D or SpA-D_{Q9,10K;D36,37A} in incomplete Freund's adjuvant (IFA) on day 10. Animals are challenged with *S. aureus* on day 21. Sera are collected before immunization and on day 20 to assess specific antibody production. For passive immunization studies, 7-wk-old mice receive 100 μ l of either NRS (normal rabbit serum) or SpA-D-specific rabbit antisera via i.p. injection 24 h before challenge. To assess the pathological correlates of pneumonia, infected animals are killed via forced CO₂ inhalation before removal of both lungs. The right lung is homogenized for enumeration of lung bacterial load. The left lung is placed in 1% formalin and paraffin embedded, thin sectioned, stained with hematoxylin-eosin, and analyzed by microscopy.

Rabbit Antibodies—

Purified 200 μ g SpA-D or SpA-D_{Q9,10K;D36,37A} is used as an immunogen for the production of rabbit antisera. 200 μ g protein is emulsified with CFA for injection at day 0, followed by booster injections with 200 μ g protein emulsified with IFA on days 21 and 42. Rabbit antibody titers are determined by ELISA. Purified antibodies are obtained by affinity chromatography of rabbit serum on SpA-D or SpA-D_{Q9,10K;D36,37A} sepharose. The concentration of eluted antibodies is measured by absorbance at A₂₈₀ and specific antibody titers are determined by ELISA.

Active Immunization with SpA-Domain D Variants.—

To determine vaccine efficacy, animals are actively immunized with purified SpA-D or SpAD_{Q9,10K;D36,37A}. As a control, animals are immunized with adjuvant alone. Antibody titers against Protein A preparations are determined using SpA-D or SpA-D_{Q9,10K;D36,37A} as antigens; note that the SpA-D_{Q9,10K;D36,37A} variant cannot bind the Fc or Fab portion of IgG. Using infectious disease models described above, any reduction in bacterial load (murine abscess and pneumonia), histopathology evidence of staphylococcal disease (murine abscess and pneumonia) and protection from lethal disease (murine lethal challenge and pneumonia) is measured.

Passive Immunization with Affinity Purified Rabbit Polyclonal Antibodies Generated Against SpA-Domain D Variants.

To determine protective immunity of Protein A specific rabbit antibodies, mice are passively immunized with 5 mg/kg of purified SpA-D or SpA-D_{Q9,10K;D36,37A} derived rabbit antibodies. Both of these antibody preparations are purified by affinity chromatography using immobilized SpA-D or SpA-D_{Q9,10K;D36,37A}. As a control, animals are passively immunized with rV10 antibodies (a plague protective antigen that has no impact on the outcome of staphylococcal infections). Antibody titers against all Protein A preparations are determined using SpA-D_{Q9,10K;D36,37A} as an antigen, as this variant cannot bind the Fc or Fab portion of IgG. Using the infectious disease models described above, the reduction in bacterial load (murine abscess and pneumonia), histopathology evidence of staphylococcal disease (murine abscess and pneumonia), and the protection from lethal disease (murine lethal challenge and pneumonia) is measured.

Example 2

Non-Toxicogenic Protein A Vaccine for Methicillin-Resistant *Staphylococcus aureus* Infection

Clinical isolates of *S. aureus* express protein A (Shopsin et al., 1999, whose primary translational product is comprised of an N-terminal signal peptide (DeDent et al., 2008), five Ig-BDs (designated E, D, A, B and C) (Sjodahl, 1977), region X with variable repeats of an eight residue peptide (Guss et al., 1984), and C-terminal sorting signal for the cell wall anchoring of SpA (Schneewind et al., 1992; Schneewind et al., 1995) (FIG. 1A-1B). Guided by amino acid homology (Uhlen et al., 1984), the triple α -helical bundle structure of IgBDs (Deisenhofer et al., 1978; Deisenhofer et al., 1981) and their atomic interactions with Fab V_H3 (Graille et al., 2000) or Fc γ (Gouda et al., 1998), glutamine 9 and 10 were selected as well as aspartate 36 and 37 as critical for the association of SpA with antibodies or B cell receptor, respectively. Substitutions Gln9Lys, Gln10Lys, Asp36Ala and Asp37Ala were introduced into the D domain to generate SpA-D_{KKAA} (FIG. 1B) (SEQ ID NO.: 69). The ability of isolated SpA-D or SpA-D_{KKAA} (SEQ ID NO.: 69) to bind human IgG was analyzed by affinity chromatography (FIG. 1D). Polyhistidine tagged SpA-D as well as full-length SpA retained human IgG on Ni-NTA, whereas SpA-D_{KKAA} (SEQ ID NO.: 69) and a negative control (SrtA) did not (FIG. 1C). A similar result was observed with von Willebrand factor (Hartleib et al., 2000), which, along with tumor necrosis factor receptor 1 (TNFR1) (Gomez et al., 2004), can also bind protein A via glutamine 9 and 10 (FIG. 1D). Human immunoglobulin encompasses 60-70% V_H3-type IgG. The inventors distinguish between Fc domain and B cell receptor activation of Igs and measured association of human Fc γ and F(ab)₂ fragments, both of which bound to full-length SpA or SpA-D, but not to SpA-D_{KKAA} (FIG. 1D) (SEQ ID NO.: 69). Injection of SpA-D into the peritoneal cavity of mice resulted in B cell expansion followed by apoptotic collapse of CD19+ lymphocytes in spleen tissue of BALB/c mice (Goodyear and Silverman, 2003) (FIG. 1E). B cell superantigen activity was not observed following injection with SpA-D_{KKAA} (SEQ ID NO.: 69), and TUNEL-staining of splenic tissue failed to detect the increase in apoptotic cells that follows injection of SpA or SpA-D (FIG. 1E).

Antibodies Against SpA-D_{KKAA} (SEQ ID NO.: 69) Protect Against MSSA and MRSA Infections.

Naive six week old BALB/c mice were injected with 50 µg of each of purified SpA, SpA-D or SpA-D_{KKAA} (SEQ ID NO.: 69) emulsified in CFA and boosted with the same antigen emulsified in IFA. In agreement with the hypothesis that SpA-D promotes the apoptotic collapse of activated clonal B cell populations, the inventors observed a ten-fold higher titer of SpA-D_{KKAA} (SEQ ID NO.: 69) specific antibodies following immunization of mice with the non-toxicogenic variant as compared to the B cell superantigen (SpA-D vs. SpA-D_{KKAA} (SEQ ID NO.: 69) $P < 0.0001$, Table 5). Antibody titers raised by immunization with full-length SpA were higher than those elicited by SpA-D ($P = 0.0022$), which is likely due to the larger size and reiterative domain structure of this antigen (Table 5). Nevertheless, even SpA elicited lower antibody titers than SpA-D_{KKAA} (SEQ ID NO.: 69) ($P = 0.0003$), which encompasses only 50 amino acids of protein A (520 residues, SEQ ID NO.: 33). Immunized mice were challenged by intravenous inoculation with *S. aureus* Newman and the ability of staphylococci to seed abscesses in renal tissues was examined by necropsy four days after challenge. In homogenized renal tissue of mock (PBS/adjuvant) immunized mice, an average staphylococcal load of $6.46 \log_{10}$ CFU was enumerated (Table 5). Immunization of mice with SpA or SpA-D led to a reduction in staphylococcal load, however SpA-D_{KKAA} (SEQ ID NO.: 69) vaccinated animals displayed an even greater, $3.07 \log_{10}$ CFU g^{-1} reduction of *S. aureus* Newman in renal tissues ($P < 0.0001$, Table 5). Abscess formation in kidneys was analyzed by histopathology (FIG. 2). Mock immunized animals harbored an average of $3.7 (\pm 1.2)$ abscesses per kidney (Table 5). Vaccination with SpA-D_{KKAA} (SEQ ID NO.: 69) reduced the average number of abscesses to $0.5 (\pm 0.4)$ ($P = 0.0204$), whereas immunization with SpA or SpA-D did not cause a significant reduction in the number of abscess lesions (Table 5). Lesions from SpA-D_{KKAA} (SEQ ID NO.: 69) vaccinated animals were smaller in size, with fewer infiltrating PMNs and characteristically lacked staphylococcal abscess communities (Cheng et al., 2009) (FIG. 2). Abscesses in animals that had been immunized with SpA or SpA-D displayed the same overall structure of lesions in mock immunized animals (FIG. 2).

The inventors examined whether SpA-D_{KKAA} (SEQ ID NO.: 69) immunization can protect mice against MRSA strains and selected the USA300 LAC isolate for animal challenge (Diep et al., 2006). This highly virulent CA-MRSA strain spread rapidly throughout the United States, causing significant human morbidity and mortality (Kennedy et al., 2008). Compared to adjuvant control mice, SpA-D_{KKAA} (SEQ ID NO.: 69) immunized animals harbored a $1.07 \log_{10}$ CFU g^{-1} reduction in bacterial load of infected kidney tissues. Histopathology examination of renal tissue following *S. aureus* USA300 challenge revealed that the average number of abscesses was reduced from $4.04 (\pm 0.8)$ to $1.6 (\pm 0.6)$ ($P = 0.02774$). In contrast, SpA or SpA-D immunization did not cause a significant reduction in bacterial load or abscess formation (Table 5).

SpA-D_{KKAA} (SEQ ID NO.: 69) Antibodies Prevent Immunoglobulin-Protein A Interaction.

Rabbits were immunized with SpA-D_{KKAA} (SEQ ID NO.: 69) and specific antibodies were purified on SpA-D_{KKAA} (SEQ ID NO.: 69) affinity column followed by SDS-PAGE (FIG. 3). SpA-D_{KKAA} (SEQ ID NO.: 69) specific IgG was cleaved with pepsin to generate Fcγ and F(ab)₂ fragments, the latter of which were purified by chromatography on SpA-D_{KKAA} (SEQ ID NO.: 69) column (FIG. 3). Binding of

human IgG or vWF to SpA or SpA-D was perturbed by SpA-D_{KKAA} (SEQ ID NO.: 69) specific F(ab)₂, indicating that SpA-D_{KKAA} (SEQ ID NO.: 69) derived antibodies neutralize the B cell superantigen function of protein A as well as its interactions with Ig (FIG. 3).

SpA_{KKAA} (SEQ ID NO.: 34) generates improved protective immune responses. To further improve the vaccine properties for non-toxicogenic protein A, the inventors generated SpA_{KKAA} (SEQ ID NO.: 34) which includes all five IgBDs with four amino acid substitutions—substitutions corresponding to Gln9Lys, Gln10Lys, Asp36Ala and Asp37Ala of domain D—in each of its five domains (E, D, A, B and C). Polyhistidine tagged SpA_{KKAA} (SEQ ID NO.: 34) was purified by affinity chromatography and analyzed by COOMASSIE-BLUE-stained SDS-PAGE (FIG. 4). Unlike full-length SpA, SpA_{KKAA} (SEQ ID NO.: 34) did not bind human IgG, Fc and F(ab)₂ or vWF (FIG. 4). SpA_{KKAA} (SEQ ID NO.: 34) failed to display B cell superantigen activity, as injection of the variant into BALB/c mice did not cause a depletion of CD19+ B cells in splenic tissue (FIG. 4). SpA_{KKAA} (SEQ ID NO.: 34) vaccination generated higher specific antibody titers than SpA-D_{KKAA} (SEQ ID NO.: 69) immunization and provided mice with elevated protection against *S. aureus* USA300 challenge (Table 5). Four days following challenge, SpA_{KKAA} (SEQ ID NO.: 34) vaccinated animals harbored $3.54 \log_{10}$ CFU g^{-1} fewer staphylococci in renal tissues ($P = 0.0001$) and also caused a greater reduction in the number of abscess lesions ($P = 0.0109$) (Table 5). As a test whether protein A vaccines impact other MRSA strains, mice were challenged with the Japanese vancomycin-resistant MRSA isolate Mu50 (Hiramatsu et al., 1997). Similar to the data observed with the MRSA isolate USA300, SpA_{KKAA} (SEQ ID NO.: 34) vaccinated animals harbored fewer Mu50 staphylococci in renal tissues than mock immunized animals ($P = 0.0248$, FIG. 7).

Passive Transfer of SpA-Specific Antibodies Prevents Staphylococcal Disease.

SpA_{KKAA} (SEQ ID NO.: 34) was used to immunize rabbits. Rabbit antibodies specific for SpA-D_{KKAA} (SEQ ID NO.: 69) or SpA_{KKAA} (SEQ ID NO.: 34) were affinity purified on matrices with immobilized cognate antigen and injected at a concentration of 5 mg kg^{-1} body weight into the peritoneal cavity of BALB/c mice (Table 6). Twenty-four hours later, specific antibody titers were determined in serum and animals challenged by intravenous inoculation with *S. aureus* Newman. Passive transfer reduced the staphylococcal load in kidney tissues for SpA-D_{KKAA} (SEQ ID NO.: 69) ($P = 0.0016$) or SpA_{KKAA} (SEQ ID NO.: 34) ($P = 0.0005$) specific antibodies. On histopathology examination, both antibodies reduced the abundance of abscess lesions in the kidneys of mice challenged with *S. aureus* Newman (Table 6). Together these data reveal that vaccine protection following immunization with SpA-D_{KKAA} (SEQ ID NO.: 69) or SpA_{KKAA} (SEQ ID NO.: 34) is conferred by antibodies that neutralize protein A.

The inventors also sought to ascertain whether protein A-specific antibodies can protect animals against lethal challenge. BALB/c mice were actively or passively immunized to raise antibodies against SpA_{KKAA} (SEQ ID NO.: 34) and then challenged by intraperitoneal injection with lethal doses of *S. aureus* Newman (FIG. 6). Antibodies against SpA_{KKAA} (SEQ ID NO.: 34), whether raised by active ($P = 0.0475$, SpA_{KKAA} (SEQ ID NO.: 34) vs. mock) or passive immunization ($P = 0.0493$, SpA_{KKAA} (SEQ ID NO.: 34) vs. mock), conferred protection against lethal challenge with *S. aureus* Newman (FIG. 6).

TABLE 5

Active immunization of mice with protein A vaccines.						
Staphylococcal load and abscess formation in renal tissue						
Antigen	^a log ₁₀ CFU g ⁻¹	^b P-value	^c Reduction (log ₁₀ CFU g ⁻¹)	^d IgG Titer	^e Number of abscesses	^f P-value
<i>S. aureus</i> Newman challenge						
Mock	6.46 ± 0.25	—	—	<100	3.7 ± 1.2	—
SpA	3.95 ± 0.56	0.0003	2.51	1706 ± 370	2.1 ± 1.2	0.3581
SpA-D	4.43 ± 0.41	0.0001	2.03	381 ± 27	1.5 ± 0.8	0.1480
SpA-D _{KKAA} (SEQ ID NO.: 69)	3.39 ± 0.50	<0.000	3.07	5600 ± 801	0.5 ± 0.4	0.0204
<i>S. aureus</i> USA300 (LAC) challenge						
Mock	7.20 ± 0.24	—	—	<100	4.0 ± 0.8	—
SpA	6.81 ± 0.26	0.2819	0.39	476 ± 60	3.3 ± 1.0	0.5969
SpA-D	6.34 ± 0.52	0.1249	0.86	358 ± 19	2.2 ± 0.6	0.0912
SpA-D _{KKAA} (SEQ ID NO.: 69)	6.00 ± 0.42	0.0189	1.20	3710 ± 1147	1.6 ± 0.6	0.0277
SpA _{KKAA} (SEQ ID NO.: 34)	3.66 ± 0.76	0.0001	3.54	10200 ± 2476	1.2 ± 0.5	0.0109

^aMeans of staphylococcal load calculated as log₁₀ CFU g⁻¹ in homogenized renal tissues 4 days following infection in cohorts of fifteen to twenty BALB/c mice per immunization. A representative of three independent and reproducible animal experiments is shown. Standard error of the means (±SEM) is indicated.

^bStatistical significance was calculated with the unpaired two-tailed Students t-test and P-values recorded; P-values < 0.05 were deemed significant.

^cReduction in bacterial load calculated as log₁₀ CFU g⁻¹.

^dMeans of five randomly chosen serum IgG titers were measured prior to staphylococcal infection by ELISA.

^eHistopathology of hematoxylin-eosin stained, thin sectioned kidneys from ten animals; the average number of abscesses per kidney was recorded and averaged again for the final mean (±SEM).

TABLE 6

Passive immunization of mice with antibodies against protein A.						
Staphylococcal load and abscess formation in renal tissue						
^a Antibody	^b log ₁₀ CFU g ⁻¹	^c P-value	^d Reduction (log ₁₀ CFU g ⁻¹)	^e IgG Titer	^f Number of abscesses	^g P-value
Mock	7.10 ± 0.14	—	—	<100	4.5 ± 0.8	—
α-SpA-D _{KKAA} (SEQ ID NO.: 69)	5.53 ± 0.43	0.0016	1.57	466 ± 114	1.9 ± 0.7	0.0235
α-SpA _{KKAA} (SEQ ID NO.: 34)	5.69 ± 0.34	0.0005	1.41	1575 ± 152	1.6 ± 0.5	0.0062

^aAffinity purified antibodies were injected into the peritoneal cavity of BALB/c mice at a concentration of 5 mg · kg⁻¹ twenty-four hours prior to intravenous challenge with 1 × 10⁷ CFU *S. aureus* Newman.

^bMeans of staphylococcal load calculated as log₁₀ CFU g⁻¹ in homogenized renal tissues 4 days following infection in cohorts of fifteen BALB/c mice per immunization. A representative of two independent and reproducible animal experiments is shown. Standard error of the means (±SEM) is indicated.

^cStatistical significance was calculated with the unpaired two-tailed Students t-test and P-values recorded; P-values < 0.05 were deemed significant.

^dReduction in bacterial load calculated as log₁₀ CFU g⁻¹.

^eMeans of five randomly chosen serum IgG titers were measured prior to staphylococcal infection by ELISA.

^fHistopathology of hematoxylin-eosin stained, thin sectioned kidneys from ten animals; the average number of abscesses per kidney was recorded and averaged again for the final mean (±SEM).

Immune Response to Protein A Following Staphylococcal Infection or SpA_{KKAA} (SEQ ID NO.: 34) Immunization.

Following infection with virulent *S. aureus*, mice do not develop protective immunity against subsequent infection with the same strain (Burts et al., 2008) (FIG. 8). The average abundance of SpA-D_{KKAA} (SEQ ID NO.: 69) specific IgG in these animals was determined by dot blot as 0.20 μg ml⁻¹ (±0.04) and 0.14 μg ml⁻¹ (±0.01) for strains Newman and USA300 LAC, respectively (FIG. 4). The minimal concentration of protein A-specific IgG required for disease protection in SpA_{KKAA} (SEQ ID NO.: 34) or SpA-D_{KKAA} (SEQ ID NO.: 69) vaccinated animals (P 0.005 log₁₀

reduction in staphylococcal CFU g⁻¹ renal tissue) was calculated as 4.05 μg ml⁻¹ (±0.88). Average serum concentration of SpA-specific IgG in adult healthy human volunteers (n=16) was 0.21 μg ml⁻¹ (±0.02). Thus, *S. aureus* infections in mice or humans are not associated with immune responses that raise significant levels of neutralizing antibodies directed against protein A, which is likely due to the B cell superantigen attributes of this molecule. In contrast, the average serum concentration of IgG specific for diphtheria toxin in human volunteers, 0.068 μg ml⁻¹ (±0.20), was within range for protective immunity against diphtheria (Behring, 1890; Lagergard et al., 1992).

Clinical *S. aureus* isolates express protein A, an essential virulence factor whose B cell superantigen activity and evasive attributes towards opsonophagocytic clearance are absolutely required for staphylococcal abscess formation (Palmqvist et al., 2005; Cheng et al., 2009; Silverman and Goodyear, 2006). Protein A can thus be thought of as a toxin, essential for pathogenesis, whose molecular attributes must be neutralized in order to achieve protective immunity. By generating non-toxic variants unable to bind Igs via Fc γ or VH $_3$ -Fab domains, the inventors measure here for the first time protein A neutralizing immune responses as a correlate for protective immunity against *S. aureus* infection. In contrast to many methicillin-sensitive strains, CA-MRSA isolate USA300 LAC is significantly more virulent (Cheng et al., 2009). For example, immunization of experimental animals with the surface protein IsdB (Kuklin et al., 2006; Stranger-Jones et al., 2006) raises antibodies that confer protection against *S. aureus* Newman (Stranger-Jones et al., 2009) but not against USA300 challenge.

Material and Methods

Bacterial Strains and Growth.

Staphylococcus aureus strains Newman and USA300 were grown in tryptic soy broth (TSB) at 37° C. *Escherichia coli* strains DH5a and BL21 (DE3) were grown in Luria-Bertani (LB) broth with 100 $\mu\text{g ml}^{-1}$ ampicillin at 37° C.

Rabbit Antibodies.

The coding sequence for SpA was PCR-amplified with two primers, gctgcacatgatggcgcaacacgatgaagctcaac (SEQ ID NO:35) and agtggatccttatgcttgagctttagcatctgc (SEQ ID NO:36) using *S. aureus* Newman template DNA. SpA-D was PCR-amplified with two primers, aacatattgtcaacaaagatcaacaaagc (SEQ ID NO:38) and aagatccagattcgttaatttttagc (SEQ ID NO:39). The sequence for SpA-D_{KKAA} (SEQ ID NO.: 69) was mutagenized with two sets of primers catatgttcaacaaagataaaaaagcgcttctatgaaatc (SEQ ID NO:42) and gattcatagaagggcgtttttatcttggtaacatag (SEQ ID NO:43) for Q9K, Q10K as well as ctccattcaaagtcttaagccgcccacgcaaaagcactaac (SEQ ID NO:40) and gttagtcttggcttggggcgcgctttaagactttgaaagaag (SEQ ID NO:41) for D36A,D37A. The sequence of SpA_{KKAA} (SEQ ID NO.: 34) was synthesized by Integrated DNA Technologies, Inc. PCR products were cloned into pET-15b generating N-terminal His $_6$ tagged recombinant protein. Plasmids were transformed into BL21(DE3). Overnight cultures of transformants were diluted 1:100 into fresh media and grown at 37° C. to an OD $_{600}$ 0.5, at which point cultures were induced with 1 mM isopropyl β -D-1-thiogalactopyranoside (IPTG) and grown for an additional three hours. Bacterial cells were sedimented by centrifugation, suspended in column buffer (50 mM Tris-HCl, pH 7.5, 150 mM NaCl) and disrupted with a French pressure cell at 14,000 psi. Lysates were cleared of membrane and insoluble components by ultracentrifugation at 40,000 \times g. Proteins in the soluble lysate were subjected to nickel-nitrilotriacetic acid (Ni-NTA, Qiagen) affinity chromatography. Proteins were eluted in column buffer containing successively higher concentrations of imidazole (100-500 mM). Protein concentrations were determined by bicinchoninic acid (BCA) assay (Thermo Scientific). For antibody generation, rabbits (6 month old New-Zealand white, female, Charles River Laboratories) were immunized with 500 μg protein emulsified in Complete Freund's Adjuvant (Difco) by subcapsular injection. For booster immunizations, proteins emulsified in Incomplete Freund's Adjuvant and injected 24 or 48 days following the initial immunization. On day 60, rabbits were bled and serum recovered.

Antibody Isolation.

Purified antigen (5 mg protein) was covalently linked to HiTrap NETS-activated HP columns (GE Healthcare). Anti-gen-matrix was used for affinity chromatography of 10-20 ml of rabbit serum at 4° C. Charged matrix was washed with 50 column volumes of PBS, antibodies eluted with elution buffer (1 M glycine, pH 2.5, 0.5 M NaCl) and immediately neutralized with 1M Tris-HCl, pH 8.5. Purified antibodies were dialyzed overnight against PBS at 4° C.

F(ab) $_2$ Fragments.

Affinity purified antibodies were mixed with 3 mg of pepsin at 37 ° C. for 30 minutes. The reaction was quenched with 1 M Tris-HCl, pH 8.5 and F(ab) $_2$ fragments were affinity purified with specific antigen-conjugated HiTrap NETS-activated HP columns. Purified antibodies were dialyzed overnight against PBS at 4° C., loaded onto SDS-PAGE gel and visualized with COOMASSIE-BLUE staining.

Active and Passive Immunization.

BALB/c mice (3 week old, female, Charles River Laboratories) were immunized with 50 μg protein emulsified in Complete Freund's Adjuvant (Difco) by intramuscular injection. For booster immunizations, proteins were emulsified in Incomplete Freund's Adjuvant and injected 11 days following the initial immunization. On day 20 following immunization, 5 mice were bled to obtain sera for specific antibody titers by enzyme-linked immunosorbent assay (ELISA).

BALB/c mice were immunized by intramuscular injection and boosted with the same antigen in Alum after 11 and 25 days. On day 34, mice were bled to obtain serum for specific antibody titers. Affinity purified antibodies were injected into the peritoneal cavity of BALB/c mice either 24 hours or 4 hours prior to sub-lethal or lethal challenge, respectively. Animal blood was collected via periorbital vein puncture and antigen specific serum antibody titers measured by ELISA.

Mouse Renal Abscess.

Overnight cultures of *S. aureus* Newman or USA300 (LAC) were diluted 1:100 into fresh TSB and grown for 2 hours at 37° C. Staphylococci were sedimented, washed and suspended PBS at OD $_{600}$ of 0.4 ($\sim 1 \times 10^8$ CFU ml $^{-1}$). Inocula were quantified by spreading sample aliquots on TSA and enumerating colonies formed. BALB/c mice (6 week old, female, Charles River Laboratories) were anesthetized via intraperitoneal injection with 100 mg ml $^{-1}$ ketamine and 20 mg ml $^{-1}$ xylazine per kilogram of body weight. Mice were infected by retroorbital injection with 1×10^7 CFU of *S. aureus* Newman or 5×10^6 CFU of *S. aureus* USA300. On day 4 following challenge, mice were killed by CO $_2$ inhalation. Both kidneys were removed, and the staphylococcal load in one organ was analyzed by homogenizing renal tissue with PBS, 1% TRITON X-100. Serial dilutions of homogenate were spread on TSA and incubated for colony formation. The remaining organ was examined by histopathology. Briefly, kidneys were fixed in 10% formalin for 24 hours at room temperature. Tissues were embedded in paraffin, thin-sectioned, stained with hematoxylin-eosin, and inspected by light microscopy to enumerate abscess lesions. All mouse experiments were performed in accordance with the institutional guidelines following experimental protocol review and approval by the Institutional Biosafety Committee (IBC) and the Institutional Animal Care and Use Committee (IACUC) at the University of Chicago.

Mouse Infection.

Staphylococci were used to infect anesthetized mice by retro-orbital injection (1×10^7 CFU of *S. aureus* Newman,

5×10⁶ CFU of *S. aureus* USA300 or 3×10⁷ CFU of *S. aureus* Mu50). On day 4, 15 or 30, mice were killed, kidneys removed, and homogenized tissue spread on agar for colony formation. Organ tissue was also thin-sectioned, stained with hematoxylin-eosin, and viewed by microscopy. Animal experiments were performed in accordance with the institutional guidelines following experimental protocol review and approval by the Institutional Biosafety Committee (IBC) and the Institutional Animal Care and Use Committee (IACUC) at the University of Chicago.

For lethal challenge experiments, BALB/c mice (cohorts of 8-10 animals per experiment) were injected with a suspension of 2-6×10⁸ CFU of *S. aureus* Newman or its isogenic *Aspa* variant into the peritoneal cavity. Animal survival was monitored over a period of 15 days and statistical significance of survival data analyzed with the log-rank test.

Protein A Binding.

For human IgG binding, Ni-NTA affinity columns were pre-charged with 200 µg of purified proteins (SpA, SpA-D, SpA-D_{KKAA} (SEQ ID NO.: 69), and SrtA) in column buffer. After washing, 200 µg of human IgG (Sigma) was loaded onto the column. Protein samples were collected from washes and elutions and subjected to SDS-PAGE gel electrophoresis, followed by COOMASSIE-BLUE staining. Purified proteins (SpA, SpA_{KKAA} (SEQ ID NO.: 34), SpA-D and SpA-D_{KKAA} (SEQ ID NO.: 69)) were coated onto MaxiSorp ELISA plates (NUNC) in 0.1M carbonate buffer (pH 9.5) at 1 µg ml⁻¹ concentration overnight at 4° C. Plates were next blocked with 5% whole milk followed by incubation with serial dilutions of peroxidase-conjugated human IgG, Fc or F(ab)₂ fragments for one hour. Plates were washed and developed using OptEIA ELISA reagents (BD). Reactions were quenched with 1 M phosphoric acid and A₄₅₀ readings were used to calculate half maximal titer and percent binding.

von Willebrand Factor (vWF) Binding Assays.

Purified proteins (SpA, SpA_{KKAA} (SEQ ID NO.: 34), SpA D and SpA-D_{KKAA} (SEQ ID NO.: 69)) were coated and blocked as described above. Plates were incubated with human vWF at 1 µg ml⁻¹ concentration for two hours, then washed and blocked with human IgG for another hour. After washing, plates were incubated with serial dilution of peroxidase-conjugated antibody directed against human vWF for one hour. Plates were washed and developed using OptEIA ELISA reagents (BD). Reactions were quenched with 1 M phosphoric acid and A₄₅₀ readings were used to calculate half maximal titer and percent binding. For inhibition assays, plates were incubated with affinity purified F(ab)₂ fragments specific for SpA-D_{KKAA} (SEQ ID NO.: 69) at 10 µg ml⁻¹ concentration for one hour prior to ligand binding assays.

Splenocyte Apoptosis.

Affinity purified proteins (150 µg of SpA, SpA-D, SpA_{KKAA} (SEQ ID NO.: 34), and SpA-D_{KKAA} (SEQ ID NO.: 69)) were injected into the peritoneal cavity of BALB/c mice (6 week old, female, Charles River Laboratories). Four hours following injection, animals were killed by CO₂ inhalation. Their spleens were removed and homogenized. Cell debris were removed using cell strainer and suspended cells were transferred to ACK lysis buffer (0.15 M NH₄Cl, 10 mM KHCO₃, 0.1 mM EDTA) to lyse red blood cells. White blood cells were sedimented by centrifugation, suspended in PBS and stained with 1:250 diluted R-PE conjugated anti-CD19 monoclonal antibody (Invitrogen) on ice and in the dark for one hour. Cells were washed with 1% FBS and fixed with 4% formalin overnight at 4° C. The following day, cells

were diluted in PBS and analyzed by flow cytometry. The remaining organ was examined for histopathology. Briefly, spleens were fixed in 10% formalin for 24 hours at room temperature. Tissues were embedded in paraffin, thin-sectioned, stained with the Apoptosis detection kit (Millipore), and inspected by light microscopy.

Antibody Quantification.

Sera were collected from healthy human volunteers or BALB/c mice that had been either infected with *S. aureus* Newman or USA300 for 30 days or that had been immunized with SpA-D_{KKAA} (SEQ ID NO.: 69)/SpA_{KKAA} (SEQ ID NO.: 34) as described above. Human/mouse IgG (Jackson Immunochemistry Laboratory), SpA_{KKAA} (SEQ ID NO.: 34), and CRM₁₉₇ were blotted onto nitrocellulose membrane. Membranes were blocked with 5% whole milk, followed by incubation with either human or mouse sera. IRDye 700DX conjugated affinity purified anti-human/mouse IgG (Rockland) was used to quantify signal intensities using the Odyssey™ infrared imaging system (Li-cor). Experiments with blood from human volunteers involved protocols that were reviewed, approved and performed under regulatory supervision of The University of Chicago's Institutional Review Board (IRB).

Statistical Analysis.

Two tailed Student's t tests were performed to analyze the statistical significance of renal abscess, ELISA, and B cell superantigen data. Animal survival data were analyzed with the log-rank test (Prism).

Example 3

Coagulases of *Staphylococcus aureus* Contribute to Abscesses Formation and Function as Protective Antigens

All clinical *S. aureus* isolates display coagulase activity—the clotting of blood or plasma through non-proteolytic activation of prothrombin to cleave fibrinogen. The inventors identified prothrombin, fibrinogen, coagulase (Coa) and von Willebrand-factor binding protein (vWbp) in staphylococcal abscess lesions of infected mice. Secreted Coa and vWbp both contributed to *S. aureus* Newman coagulase activity, thereby enabling abscess formation as well as lethal disease in mice. Antibodies raised against purified Coa or vWbp specifically block association of the corresponding polypeptide with prothrombin and fibrinogen. Coa- and vWbp-specific antibodies, whether raised by active or passive immunization, prevented abscess formation and mortality of mice infected with staphylococci.

VIII. RESULTS

Localization of Coagulase and Coagulation Factors in Staphylococcal Abscesses.

Previous work established the mouse renal abscess model, whereby 1×10⁷ CFU of the human clinical isolate *S. aureus* Newman (Baba et al., 2007) are injected into the blood stream of BALB/c mice (Albus et al., 1991). Forty-eight hours following infection, mice develop disseminated abscesses in multiple organs, detectable by light microscopy of hematoxylin-eosin stained, thin-sectioned kidney tissue initially as an accumulation of polymorphonuclear leukocytes (PMNs) with few bacteria (Cheng et al., 2009). By day 5 of infection, abscesses increase in size and enclose a central population of staphylococci (staphylococcal abscess community—SAC), surrounded by a layer of eosinophilic,

amorphous material (the pseudocapsule) and a large cuff of PMNs (Cheng et al., 2009). Histopathology reveals massive necrosis of PMNs in proximity to the staphylococcal nidus at the center of abscess lesions as well as a mantle of healthy phagocytes. At later time intervals, SACs increase and abscesses rupture, releasing necrotic material and staphylococci into the bloodstream. A new round of abscess formation is initiated, eventually precipitating a lethal outcome of infections (Cheng et al., 2009).

To localize coagulases in abscess lesions, kidneys of mice that had been infected for 5 days with *S. aureus* Newman were thin-sectioned and stained by immuno-histochemistry with affinity purified Coa- or vWbp-specific rabbit antibodies (FIG. 10). The inventors observed intense Coa staining in the pseudocapsule surrounding SACs and in the periphery of abscess lesions, i.e., the fibrin capsule bordering uninfected tissue. vWbp staining occurred throughout abscess lesions, but also with accumulation at the periphery. Prothrombin specific antibodies revealed staining of the zymogen in the pseudocapsule and in the periphery, whereas fibrinogen/fibrin specific staining occurred throughout abscess lesions. Together these data indicate that the eosinophilic pseudocapsule of staphylococcal abscesses harbors prothrombin and fibrinogen, which co-localize with Coa. At the periphery of abscess lesions, Coa, vWbp, prothrombin and fibrinogen/fibrin are co-localized. These observations prompted further investigation in to whether Coa and vWbp are crucial contributors to the establishment of abscesses by triggering prothrombin-mediated conversion of fibrinogen to fibrin.

Staphylococcus aureus Coa and vWbp Contribute to the Clotting of Mouse Blood.

The coa and/or vWbp genes on the chromosome of *S. aureus* Newman were deleted by allelic replacement using pKOR1 technology (Bae and Schneewind, 2005). Two complementing plasmids, pcoa and pvWbp, were generated by cloning coa or vWbp structural genes as well as their upstream promoter sequences into p051 (Schneewind et al., 1993). Plasmids were electroporated into staphylococci and their continued replication selected on media supplemented with chloramphenicol (Schneewind et al., 1992). When probed for coagulases with specific antibodies, the inventors observed Coa secretion by the wild-type as well as the Δ vWbp strain, but not by Δ coa or Δ coa/ Δ vWbp variants (FIG. 11). The phenotypic defect of Δ coa and Δ coa/ Δ vWbp mutants was restored by electroporation with pcoa but not by pvWbp (FIG. 11). Similarly, secretion of vWbp was observed in *S. aureus* Newman (wild-type) as well as Δ coa mutant cultures, but not in Δ vWbp or Δ coa/ Δ vWbp variants (FIG. 11). This defect was restored by electroporation with pvWbp, but not by pcoa.

Clotting of blood is effectively inhibited by hirudin (lepirudin) (Harvey et al., 1986), a 65 residue peptide from leech that forms a 1:1 complex with thrombin, thereby blocking proteolytic conversion of fibrinogen to fibrin (Markwardt, 1955). Inoculation of fresh lepirudin-treated mouse blood

with *S. aureus* Newman triggered clotting in less than 12 hours, whereas mock infected blood remained without clots for more than 48 hours (FIG. 11C). Using this assay, it was observed that staphylococcal variants lacking coagulase activity displayed delays in clotting time, Δ coa 36 hours and Δ vWbp 24 hours (FIG. 11C). The double mutant, Δ coa/ Δ vWbp, was unable to clot mouse blood. These defects were complemented by electroporation with plasmids pvWbp as well as pcoa. Taken together, these data indicate that the two coagulases, Coa and vWbp, contribute to the ability of *S. aureus* Newman to clot mouse blood (FIG. 11C).

Coa and vWbp are Required for Staphylococcal Survival in Blood, Abscess Formation and Lethal Bacteremia in Mice.

To analyze the virulence contributions of coagulases, the inventors first examined staphylococcal survival in lepirudin-treated blood. Wild-type strain *S. aureus* Newman was not killed in mouse blood, however isogenic variants lacking Coa, i.e. Δ coa and Δ coa/ Δ vWbp, each displayed a significant reduction in CFU after 30 min incubation. This defect in survival was restored by pcoa, but not by pvWbp, suggesting that only Coa is required for staphylococcal survival in mouse blood.

Staphylococcal bacteremia is a frequent cause of human mortality in hospital settings (Klebens et al., 2007). The inventors sought to ascertain whether coagulases are required for lethal challenge of BALB/c mice, following intravenous injection of 1×10^8 CFU *S. aureus* Newman. All animals infected with the wild-type parent strain Newman succumbed to infection within 24 hours (FIG. 12B). Animals infected with single mutants, Δ coa or Δ vWbp, each displayed a short but statistically significant delay in time-to-death (FIG. 12B). The double mutant strain was significantly more impaired than mutants with single deletions and animals infected with the Δ coa/ Δ vWbp strain displayed the largest reduction in virulence as compared to the wild-type (FIG. 12B).

The inventors next analyzed abscess formation in renal tissues of infected mice and observed that Δ coa variants were impaired in their ability to form abscesses by day 5 and 15 of infection (Table 7, FIG. 12G, 12I). The Δ vWbp mutant continued to form abscesses, although the bacterial load, the overall size of staphylococcal abscess communities and the amount of immune cell infiltrates were reduced in these variants (Table 7, FIG. 12D, 12F). Mutants in coagulase are slightly more attenuated in virulence than those in vWbp, as Δ coa has lower abscess formation and bacterial load by day 15. However, the Δ coa/ Δ vWbp double mutants markedly incapacitated in their ability to form abscesses and persist in infected tissues (Table 7, FIG. 12H, 12K). Thus, both coagulase and von Willebrand factor binding protein are important for staphylococcal survival in the host, whether in the bloodstream or end organ tissues.

TABLE 7

Virulence of <i>S. aureus</i> Newman coa, vWbp, and coa/vWbp mutants						
Strain	Abscess formation in kidney tissue*					
	Staphylococcal load in kidney tissue*			Number of		
	^a log ₁₀ CFU g ⁻¹ of kidney tissue	^b Significance (P-value)	^c Reduction in log ₁₀ CFU g ⁻¹	^d Surface abscesses (%)	abscesses per kidney	^e Significance (P-value)
Day 5 analysis of staphylococcal load and abscess formation						
PBS	6.034 ± 0.899	—	—	75	2.333 ± 0.623	—
Coa	5.538 ± 0.560	0.3750	0.492	38	1.111 ± 0.389	0.1635
vWbp	5.247 ± 0.311	0.0859	0.783	56	1.750 ± 0.650	0.6085
coa/vWbp	4.908 ± 0.251	0.0044	1.395	25	0.750 ± 0.342	0.0786
Day 15 analysis of staphylococcal load and abscess formation						
PBS	5.380 ± 0.294	—	—	81	3.000 ± 1.234	—
Coa	4.023 ± 0.324	0.0077	1.357	44	1.400 ± 0.452	0.1862
vWbp	5.140 ± 0.689	0.0688	0.240	50	1.625 ± 0.298	0.2974
coa/vWbp	3.300 ± 0.552	0.0056	2.080	20	0.556 ± 0.154	0.0341

*BALB/c mice (n = 18-20) were injected into the peritoneum with 100 µl each of affinity purified rabbit antibodies against vWbp (α-vWbp), Coa (α-Coa) or vWbp and Coa (α-vWbp/Coa) on day 0. Twenty four hours later, animals were examined for IgG antibody titers in serum and were challenged by intravenous inoculation with 1 × 10⁷ colony forming units (CFU) *S. aureus* Newman or mutants thereof. Five or fifteen days later, animals were killed and both kidneys removed. One kidney was fixed in formaldehyde, embedded in paraffin, thin sectioned, hematoxylin-eosin stained and four sagittal sections per kidney were analyzed for abscess formation. The other kidney was homogenized in PBS buffer, homogenate spread on agar medium for colony formation, and staphylococcal load enumerated as CFU.

^aMeans of staphylococcal load calculated as log₁₀ CFU g⁻¹ in homogenized renal tissues 4 days following infection in cohorts of eighteen to twenty BALB/c mice per immunization. Standard error of the means (±SEM) is indicated.

^bStatistical significance was calculated with the unpaired two-tailed Students t-test and P-values recorded; P-values < 0.05 were deemed significant.

^cReduction in bacterial load calculated as log₁₀ CFU g⁻¹.

^dAbscess formation in kidney tissues four days following infection was measured by macroscopic inspection (% positive)

^eHistopathology of hematoxyline-eosin stained, thin sectioned kidneys from ten animals; the average number of abscesses per kidney was recorded and averaged again for the final mean (±SEM).

^fStatistical significance was calculated with the unpaired two-tailed Students t-test and P-values recorded; P-values < 0.05 were deemed significant.

Antibodies Against Coagulases and their Effect on Blood Clotting Induced During Staphylococcal Infection.

Recombinant His₆-Coa and His₆-vWbp were purified by affinity chromatography on Ni-NTA (FIG. 13A), emulsified in adjuvant and injected into rabbits to raise specific antibodies that were purified on affinity matrices harboring recombinant protein. Antibodies directed against Coa preferentially bound to Coa, not to vWbp (FIG. 13B). The reciprocal was true for antibodies directed against vWbp (FIG. 13B). When added to lepirudin-treated mouse blood infected with *S. aureus* Newman, the inventors observed that antibodies directed against Coa, vWbp or Coa and vWbp each blocked the coagulation of blood (FIG. 13C). As controls, mock treated samples or the irrelevant V10 antibody (which provides protection against *Yersinia pestis* type III injection (DeBord et al., 2006)) had no effect (FIG. 13C).

To examine the role of antibodies on isolated Coa or vWbp, the inventors purified recombinant, functionally active proteins (Friedrich et al., 2003) that were then added to lepirudin treated mouse blood. Coa or vWbp treated mouse blood coagulated in less than 30 minutes (FIG. 13D). As a control, mock (PBS) or treatment with irrelevant V10 antibody did not affect clotting. Antibodies directed against Coa or vWbp delayed clotting of mouse blood treated with recombinant proteins and this occurred even for the cross-reacting homologous factor (FIG. 13D). Minimal cross reactivity of the antibodies was observed by ELISA and western blot, yet there is cross inhibition of function.

Antibodies that block association between coagulases and prothrombin or fibrinogen. Surface plasmon resonance (SPR) was used to investigate how αCoa and αvWbp antibodies interfere with the physiological functions of coagulases. Prothrombin was immobilized on a CM5 chip. Flowing purified Coa over the sample, a dissociation constant K_D 28 nM was calculated, a measurement that is

commensurate with other reports in the literature (Friedrich et al., 2003). The addition of αCoa led to a concentration-dependent decrease in response signal for the formation of prothrombin•Coa, indicating that these antibodies block association of Coa with prothrombin (FIG. 14A). SPR further confirmed association between coagulase and fibrinogen (K_D 93.1 nM, FIG. 14B). Upon pre-incubation with αCoa, the inventors observed a dramatic decrease in the binding of Coa to fibrinogen (FIG. 14B). Taken together, these results indicate that antibodies directed against Coa block the association of this molecule with blood coagulation factors.

Purified vWbp displayed strong affinity for prothrombin (K_D 38.4 nM, FIG. 13C) and fibrinogen (484 nM, FIG. 13D), the latter of which had hitherto not been appreciated (Kroh et al., 2009). Further, pre-incubation with antibodies raised against vWbp blocked the association between vWbp and prothrombin or fibrinogen in a dose-dependent manner (FIG. 13C, 13D). These findings support results from the blood coagulation assays, demonstrating that specific polyclonal antibodies can block the interaction between Coa or vWbp and specific components of the coagulation cascade (FIG. 12).

To test whether antibodies specific for coagulases block the conversion of fibrinogen to fibrin, the ability of prothrombin•coagulase complexes to cleave S-2238 was measured, a surrogate for the cleavage of fibrinogen to fibrin (FIG. 14E, 14F). Addition of specific antibodies to prothrombin•Coa or prothrombin•vWbp reduced the ability of these complexes to convert substrate to product. Further, cross-inhibition of coagulase-specific antibodies was observed, where the addition of cross-reacting antibodies caused a reduction in activity of the prothrombin•vWbp complex. These data suggest that specific antibodies

directed against Coa or vWbp neutralize the pathophysiological effect of the secreted product to which they bind.

Antibodies Against Coagulases Provide Protection Against Staphylococcal Disease.

IgG type antibodies specific for Coa or vWbp were isolated from rabbit serum by chromatography over an affinity column, generated by covalent crosslinking of the antigen to CNBr SEPHAROSE. The inventors attempted to perturb staphylococcal pathogenesis by administration of neutralizing antibodies, directed against Coa and/or vWbp. Mice were administered rabbit antibodies and challenged with a lethal dose of *S. aureus* strain Newman. Injection of Coa or vWbp specific antibodies significantly prolonged murine survival (FIG. 15).

lesions (P=0.039); abscess lesions with staphylococcal communities at the nidus of large PMN infiltrates were not detected (FIG. 16 and Table 8). Animals that received both antibodies, α Wbp and α Coa, displayed an even greater reduction in staphylococcal load (P=0.013) and a reduction in the abundance of inflammatory lesions (P=0.0078) (Table 8). Together, these data indicate that antibodies against coagulases, administered by passive immunization, protect mice against abscess formation and enable clearance of the invading pathogen from host tissues. Antibodies against vWbp contribute relatively little to vaccine protection, in agreement with the finding that vWbp does not play the same critical role as Coa during the pathogenesis of *S. aureus* infections in mice (Table 8).

TABLE 8

Purified Rabbit Antibody	Staphylococcal load in kidney tissue*				Abscess formation in kidney tissue*		
	^a log ₁₀ CFU g ⁻¹ of kidney tissue	^b Significance (P-value)	^c Reduction in log ₁₀ CFU g ⁻¹	^d IgG Titer	^e Surface abscesses (%)	^f Number of abscesses per kidney	^g Significance (P-value)
Mock	5.86 ± 0.29	—	—	<100	75	4.6 ± 1.4	—
α -vWbp	5.25 ± 0.36	0.3554	0.60	1,100 ± 200	39	1.4 ± 0.5	0.0592
α -Coa	4.68 ± 0.47	0.0420	1.18	1,300 ± 250	20	1.2 ± 0.7	0.0396
α -vWbp/Coa	4.29 ± 0.52	0.0130	1.53	1,000 ± 300	25	0.3 ± 0.2	0.0078

*BALB/c mice (n = 18-20) were injected into the peritoneum with 100 μ l each of affinity purified rabbit antibodies against vWbp (α -vWbp), Coa (α -Coa) or vWbp and Coa (α -vWbp/Coa) on day 0. Twenty four hours later, animals were examined for IgG antibody titers in serum and were challenged by intravenous inoculation with 1×10^7 colony forming units (CFU) *S. aureus* Newman. Five days later, animals were killed and both kidneys removed. One kidney was fixed in formaldehyde, embedded in paraffin, thin sectioned, hematoxylin-eosin stained and four sagittal sections per kidney were analyzed for abscess formation. The other kidney was homogenized in PBS buffer, homogenate spread on agar medium for colony formation, and staphylococcal load enumerated as CFU.

^aMeans of staphylococcal load calculated as log₁₀ CFU g⁻¹ in homogenized renal tissues 4 days following infection in cohorts of eighteen to twenty BALB/c mice per immunization. Standard error of the means (\pm SEM) is indicated.

^bStatistical significance was calculated with the unpaired two-tailed Students t-test and P-values recorded; P-values < 0.05 were deemed significant.

^cReduction in bacterial load calculated as log₁₀ CFU g⁻¹.

^dAbscess formation in kidney tissues four days following infection was measured by macroscopic inspection (% positive)

^eHistopathology of hematoxyline-eosin stained, thin sectioned kidneys from ten animals; the average number of abscesses per kidney was recorded and averaged again for the final mean (\pm SEM).

^gStatistical significance was calculated with the unpaired two-tailed Students t-test and P-values recorded; P-values < 0.05 were deemed significant.

To test antibody reagents for possible vaccine protection against lethal bacteremia, affinity purified IgG (5 mg kg⁻¹ body weight) were injected into the peritoneal cavity of mice. Twenty-four hours later, animals were injected with a suspension of 1×10^8 CFU *S. aureus* Newman in PBS into the retro-orbital plexus. Monitoring animals over time, the inventors observed that antibodies directed against vWbp (α vWbp) led to increased time-to-death and to 10% survival, as compared to animals that had received irrelevant α V10 antibodies and died within 12-48 hours (FIG. 15). Antibodies against Coa (α Coa) further increased the time-to-death of passively immunized mice (FIG. 15). A mixture of both antibodies (α Coa/ α vWbp) did not generate a statistically significant improvement in survival or time-to-death over α Coa antibodies.

To examine the passive immunization for protection against staphylococcal abscess formation, purified antibodies (5 mg kg⁻¹ body weight) were injected into the peritoneal cavity of mice and abscess formation was monitored for five days after intravenous challenge with 1×10^7 CFU *S. aureus* Newman. Antibodies against vWbp did not lead to a significant reduction in staphylococcal load or in the number of inflammatory lesions (Table 8), although the observed lesions harbored smaller abscess communities and reduced PMN infiltrates as compared to mock immunized mice (FIG. 16). Antibodies against coagulase reduced the staphylococcal load (P=0.042) as well as the number of inflammatory

Coagulases Function as Protective Antigens for Staphylococcal Infections.

Poly-histidine tagged CoA and vWbp were purified from *E. coli* and used as subunit vaccine antigens. Proteins (100 μ g emulsified in CFA or IFA) were injected into naïve BALB/c mice on day 0 (CFA) or 11 (IFA). Animals were challenged on day 21 by intravenous inoculation of *S. aureus* Newman. Five control animals were bled at the time of challenge and serum antibody titers against vaccine antigens were determined by ELISA (Table 9). Animals were killed five or fifteen days following challenge staphylococcal load and histopathology of abscess lesions were analyzed. Immunization with Coa reduced the bacterial load by day 5 (P=0.03, PBS mock vs. Coa) and day 15 (P=4.286 $\times 10^{-5}$, PBS mock vs. Coa, see Table 9). Coa vaccination also diminished the number of infectious lesions that formed in kidney tissues, mock vs. Coa, P=0.03 (day 5) and P=0.0522 (day 15) (Table 9). Of note, none of the Coa-immunized mice developed typical abscess lesions (FIG. 17). On occasion small accumulations of PMNs that were not associated with staphylococcal abscess communities were observed (FIG. 17). Immunization with vWbp did not significantly reduce staphylococcal load on day 5 (P=0.39, PBS mock vs. vWbp) or on day 15 (P=0.09, PBS mock vs. vWbp). The total number of inflammatory lesions was not reduced. Nevertheless, the architecture of abscesses had changed following immunization with vWbp. Staphylococcal communities were not detected at the center of abscesses and

instead PMN infiltrations were observed (FIG. 17). The combination vaccine, vWbp-Coa, further reduced the number of inflammatory cells in kidney tissues and infected animals did not display abscess lesions on day 5 or 15 (Table 9).

samples of each culture centrifuged at 13,000×g for 10 min in a table top centrifuge and the supernatant was recovered. Trichloroacetic acid, 75 µl of 100% w/v solution, was added and samples were incubated on ice for 10 min, followed by centrifugation and wash with 1 ml ice-cold 100% acetone.

TABLE 9

		Active immunization of mice with Coa and/or vWbp				Abscess formation in kidney tissue*		
		Staphylococcal load in kidney tissue*			Abscess formation in kidney tissue*			
Purified Vaccine Antigen	^a log ₁₀ CFU g ⁻¹ of kidney tissue	^b Significance (P-value)	^c Reduction in ^a log ₁₀ CFU g ⁻¹	^d IgG Titer	^e Surface abscesses (%)	^f Number of abscesses per kidney	^g Significance (P-value)	
Day 5 ^h	Mock	5.75 ± 0.42	—	—	<100	56	1.3 ± 0.3	—
	vWbp	4.94 ± 0.46	0.1413	0.81	14,000 ± 5,000	45	1.8 ± 0.5	0.39
	Coa	4.86 ± 0.50	0.1417	0.88	19,000 ± 4,000	25	0.3 ± 0.3	0.03
	vWbp/Coa	4.84 ± 0.38	0.1195	0.90	7,000 ± 1,500	25	0.3 ± 0.3	0.03
Day 15 ⁱ	Mock	6.68 ± 0.22	—	—	<100	75	6.0 ± 1.9	—
	vWbp	3.41 ± 0.47	0.4503	3.27	14,000 ± 5,000	20	1.8 ± 1.1	0.09
	Coa	3.43 ± 0.54	0.1681	3.25	19,000 ± 4,000	20	1.2 ± 0.8	0.05
	vWbp/Coa	3.79 ± 0.37	0.0263	2.89	7,000 ± 1,500	30	0.7 ± 0.5	0.01

*BALB/c mice (n = 18-20) were injected with 100 µg each of purified vWbp, Coa or vWbp and Coa emulsified in CFA on day 0 and boosted with the same antigen emulsified in IFA on day 11. On day 20, animals were examined for IgG antibody titers and on day 21 animals were challenged by intravenous inoculation with either 1 × 10⁷ colony forming units (CFU) *S. aureus* Newman. On day 25, animals were killed and both kidneys removed. One kidney was fixed in formaldehyde, embedded in paraffin, thin sectioned, hematoxylin-eosin stained and four sagittal sections per kidney were analyzed for abscess formation. The other kidney was homogenized in PBS buffer, homogenate spread on agar medium for colony formation, and staphylococcal load enumerated as CFU.

^aMeans of staphylococcal load calculated as log₁₀ CFU g⁻¹ in homogenized renal tissues 4 days following infection in cohorts of eighteen to twenty BALB/c mice per immunization. Standard error of the means (±SEM) is indicated.

^bStatistical significance was calculated with the unpaired two-tailed Students t-test and P-values recorded; P-values < 0.05 were deemed significant.

^cReduction in bacterial load calculated as log₁₀ CFU g⁻¹.

^dMeans of five randomly chosen serum IgG titers were measured prior to staphylococcal infection by ELISA with SpA-D_{KKAA} (SEQ ID NO.: 69) antigen

^eAbscess formation in kidney tissues four days following infection was measured by macroscopic inspection (% positive)

^fHistopathology of hematoxyline-eosin stained, thin sectioned kidneys from ten animals; the average number of abscesses per kidney was recorded and averaged again for the final mean (±SEM).

^gStatistical significance was calculated with the unpaired two-tailed Students t-test and P-values recorded; P-values < 0.05 were deemed significant.

^hAnalysis of mice 5 days following infection with *S. aureus* Newman.

ⁱAnalysis of mice 15 days following infection with *S. aureus* Newman.

IX. MATERIALS AND METHODS

Bacterial Strains and Growth of Cultures.

Staphylococci were cultured on tryptic soy agar or broth at 37° C. *E. coli* strains DH5a and BL21(DE3) (Studier et al., 1990) were cultured on Luria agar or broth at 37° C. Ampicillin (100 µg/ml) and chloramphenicol (10 µg/ml)

were used for pET15b (Studier et al., 1990) and pOS1 (Schneewind et al., 1993) plasmid selection, respectively.

Generation of Mutants.

DNA sequences 1 kb upstream and downstream of *coa* and *vWbp* were PCR amplified using the primers attB1_Coa, Coa1_BamHI, Coa2_BamHI, attbB2_Coa and attB1_vWF, vWF1_BamHI, vWF2_BamHI, attbB2_vWF (Table 10). The fragments were exchanged onto pKOR1 using the BP clonase II kit (Invitrogen) (Bae and Schneewind, 2005). These vectors were electroporated into *S. aureus* Newman and subjected to temperature shift induced allelic exchange to generate the corresponding deletion (Bae and Schneewind, 2005). Mutants were verified by PCR amplification of the gene locus, DNA sequencing, and immunoblot analysis.

To generate complementing plasmids, the primers Coa_promoter_BamHI_F, Coa_out_PstI_R, vWbp_promoter_BamHI_F, vWbp_out_PstI_R (Table 10) were designed to include the upstream promoter region of *vWbp* or *coa* and the amplified regions were cloned into pOS1. These plasmids were verified by sequencing and then electroporated into staphylococcal strains. For immunoblot analysis, overnight cultures of staphylococci grown in tryptic soy broth (Difco) were refreshed 1:100 and grown with shaking at 37° C. until they reached OD₆₀₀ of 0.4. One ml

35 Samples were air dried overnight and solubilized in 50 µl sample buffer (4% SDS, 50 mM Tris-HCl, pH8, 10% glycerol, and bromophenol blue).

Blood Survival Assay and Blood Coagulation.

Overnight cultures of staphylococcal strains were diluted 1:100 into fresh TSB and grown at 37° C. until they reached an OD₆₀₀ 0.4. One ml of culture was centrifuged, and staphylococci washed and suspended in 10 ml of sterile PBS to generate a suspension of 1×10⁷ CFU/ml. Whole blood from naïve 6 week old Balb/c mice was collected and REFLUDAN™ (lepirudin, Bayer) was added to a final concentration 50 µg/ml. 450 µL blood was aliquoted into a 1 ml eppendorf tube and mixed with 50 µl bacterial sample (1×10⁵ CFU/ml). Samples were incubated at 37° C. with slow rotation. 100 µl aliquots were removed at times 0 min and 30 min, mixed 1:1 with 2% saponin/PBS and incubated on ice for 30 minutes. Five 1:10 serial dilutions were prepared and 10 µl aliquots spread on TSA agar for colony formation and enumeration.

To assess bacterial blood coagulating activity, 10 µl of the above stock bacterial culture was added to 100 µl of anti-coagulated mouse blood in a sterile plastic test tube (BD falcon) to achieve an end concentration of 1×10⁵ CFU/ml. For antibody perturbation, an additional 10 ul of PBS containing 3×10⁻⁵ Mol of antibody was added to the mixture. To assess recombinant proteins, 10 µl of protein in PBS buffer added to an end concentration of 50 µM. Test tubes were incubated at room temperature and blood liquidity or coagulation was verified by tipping the tubes to 45° angles in timed intervals.

Protein purification. For vaccination studies, full-length coding sequence of mature Coa or vWbp was cloned into

pET15b vector using the primers Coa_foward_XhoI, Coa_reverse_BamHI, vWbp_forward_XhoI, vWbp_reverse_BamHI (Table 10) to obtain His6-Coa and His6-vWbp. *E. coli* BL21(DE3) harboring expression vectors were grown at 37° C. and induced with 1 mM IPTG after two hours. Four hours after induction, cells were centrifuged at 6,000×g, suspended in 1× column buffer (0.1 M Tris-HCl pH 7.5, 0.5 M NaCl) and lysed in a French press at 14,000 lb/in². Lysates were subjected to ultracentrifugation at 40,000×g for 30 min and the supernatant was subjected to Ni-NTA chromatography, washed with column buffer containing 25 μM imidazole, followed by elution with 500 μM imidazole. Eluate was dialyzed with 1×PBS. To remove endotoxin, 1:1,000 TRITON-X114 was added and the solution was chilled for 5 min, incubated at 37° C. for 10 min, and centrifuged at 13,000×g. Supernatant was loaded onto a HiTrap desalting column to remove any remnant of TRITON-X114.

loaded onto a Superdex 75 (GE Healthcare) column for final purification. All eluted proteins were stored in 1×PBS.

Rabbit Antibodies.

Protein concentration was determined using a BCA kit (Thermo Scientific). Purity was verified by SDS page gel analysis and COOMASSIE BRILLIANT BLUE staining. Six month old New-Zealand white female rabbits (Charles River Laboratories) were immunized with 500 μg protein emulsified in CFA (Difco) for initial immunization or IFA for booster immunizations on day 24 and 48. On day 60, rabbits were bled and serum recovered for immunoblotting or passive transfer experiments. For antibody purification, recombinant His6-Coa or His6-vWbp (5 mg) was covalently linked to HiTrap NETS-activated HP columns (GE Healthcare). This antigen-matrix was then used for affinity chromatography of 10-20 ml of rabbit serum at 4° C. Charged matrix was washed with 50 column volumes of PBS, antibodies eluted with elution buffer (1 M glycine pH 2.5, 0.5 M

TABLE 10

Primers used in this study	
Primer name	Sequence
attB1_Coa	GGGGACAAGTTTGTACAAAAAAGCAGGCTgatgactaagttgaaaaagaag (SEQ ID NO: 46)
Coa1_BamHI	aaGGATCCcctccaaaatgtaattgccc (SEQ ID NO: 47)
Coa2_BamHI	aaGGATCCgtttgtaactctatccaagac (SEQ ID NO: 48)
attbB2_Coa	GGGGACCACTTTGTACAAGAAAGCTGGGTgacacctattgcacgattcgc (SEQ ID NO: 49)
attB1_vWF	GGGGACAAGTTTGTACAAAAAAGCAGGCTcagatagcgattcagattcag (SEQ ID NO: 50)
vWF1_BamHI	aaGGATCCctgtatctttctccttaattttcc (SEQ ID NO: 51)
vWF2_BamHI	aaGGATCCcatggctgcaagcaataatg (SEQ ID NO: 52)
attbB2_vWF	GGGGACCACTTTGTACAAGAAAGCTGGGTgcctctgtgtaacaaatttatg (SEQ ID NO: 53)
Coa_promoter_BamHI_F	gaaGGATCCgtttattctagttaatatagttaatg (SEQ ID NO: 54)
Coa_out_PstI_R	gaaCTGCAGctgtatgtctttggatagagttac (SEQ ID NO: 55)
vWbp_promoter_BamHI_F	gaaGGATCCggtggcttttttacttggattttc (SEQ ID NO: 56)
vWbp_out_PstI_R	gaaCTGCAGcgacaaactcattatttcttgc (SEQ ID NO: 57)
Coa_foward_XhoI	GAACTCGAGTCTAGCTTATTACATGG (SEQ ID NO: 58)
Coa_Xho_factorXa_F	GAACTCGAGatagaaggcagaatagtaacaaaggattatagtggg (SEQ ID NO: 59)
Coa_reverse_BamHI	GTAGGATCCTGGGATAGAGTTACAAAC (SEQ ID NO: 60)
vWbp_forward_XhoI	GAACTCGAGgcattatgtgtatcacaatttggg (SEQ ID NO: 61)
vWbp_Xho_factorXa_F	GAACTCGAGatagaaggcagagtggtttctggggagaagaatc (SEQ ID NO: 62)
vWbp_reverse_BamHI	GAACTCGAGgcagccatgcattaattatttggc (SEQ ID NO: 63)

For enzymatic studies, ELISA, and SPR, full-length coding sequence of mature Coa or vWbp was cloned into pET15b with primers Coa_Xho_factorXa_F, Coa_reverse_BamHI, vWbp_Xho_factorXa_F, vWbp_reverse_BamHI (Table 10) which contain a Factor Xa site preceding the initial Ile-Val-Thr-Lys (SEQ ID NO.: 73) of coagulase and Val-Val-Ser-Gly (SEQ ID NO.: 74) of vWbp. These proteins were expressed and purified using the above protocol, then cleaved with 10 units Factor Xa/1 ml for 1 hour at 25° C. to remove the His₆ tag from the N-terminus. Proteins were then

NaCl) and immediately neutralized with 1M Tris-HCl, pH 8.5. Purified antibodies were dialyzed overnight against PBS at 4° C.

Surface Plasmon Resonance.

Affinity and rates of association and dissociation were measured on a BIAcore 3000. Buffers were sterile filtered and degassed. A CM5 chip was prepared for amine linkage by injection of human prothrombin (500 nM, pH 4.0) (Innovative Research) and human fibrinogen (200 nM, pH 4.5) (Innovative Research) in presence of 0.2 M EDC and 0.05 M NHS. To measure the interaction of coagulase with

prothrombin and fibrinogen, Coa was diluted into HBS-P buffer (20 mM HEPES [pH 7.4], 150 mM NaCl, 0.005% [vol/vol] surfactant P20) at concentrations 0-75 nM with successive injections of coagulase for 300 seconds followed by 300 seconds for dissociation followed by regeneration with NaOH (50 μ L, 30 seconds). K_D and χ^2 were determined using the BiaEvaluation software and best fit was determined with a 1:1 binding model with drifting baseline and local R_{max} . The interaction of von Willebrand factor with prothrombin and fibrinogen was measured in the same way. All experiments were repeated in triplicate. Inhibition experiments with polyclonal antibodies were conducted by successive injections of coagulase (25 nM) incubated with α Coa at 0 nM-200 nM under the same injection conditions described above. vWF (50 nM) was similarly incubated with α vWF at 0 nM-400 nM. Response difference was measured as the change in response units from before the injection to the end of the injection.

Measurements of Coagulase Activity.

1×10^{-16} M prothrombin (Innovative Research) was pre-incubated for 20 min with an equimolar amount of functional coagulase or vWbp at room temperature, followed by addition of S-2238 (a chromogenic substrate) to an end concentration of 1 mM in a total reaction buffer of 100 μ l 1xPBS. The change in absorbance was measured at 450 nm for 10 minutes in a spectrophotometer, plotted as a function of time and fitted to a linear curve. The slope of the curve (dA/dt) was interpreted to be the rate of S-2238 hydrolysis, and thus reflective of enzymatic function (% coagulase-prothrombin or vWbp-prothrombin complex activity). The assay was repeated in presence of specific or cross antibodies added in 3M excess (3×10^{-16} M) and the data was normalized to the % average activity without inhibition.

Renal Abscess Model and Lethal Challenge.

Overnight cultures of staphylococcal strains were diluted 1:100 into fresh TSB and grown until they reached an OD₆₀₀ of 0.4. 10 ml of bacteria were centrifuged at 7,500xg, washed, and suspended in 10 ml of 1xPBS. Six week old female BALB/c mice (Charles River) were injected retro-orbitally with 1×10^7 CFU staphylococcal suspension in 100 μ l of PBS. Cohorts of 10 mice were used. On the fifth day post infection, these mice were killed by CO₂ asphyxiation and their kidneys were excised. All organs were examined for surface lesions and 8-10 right kidneys were sent for histopathology sectioning and hematoxylin-eosin staining. These slides were examined by light microscopy for internal abscesses. For the lethal challenge model, all experimental conditions remain the same except that 1×10^8 CFU staphylococci were administered and that the mice were monitored for 10 days post infection for survival.

Immunohistochemistry Staining of Renal Sections.

Sectioned kidneys were deparaffinized and rehydrated through xylene and serial dilutions of EtOH to distilled water. They were incubated in antigen retrieval buffer (DAKO, pH 6.0) and heated in steamer at over 96° C. for 20 minutes. After rinsing, the slides were incubated in 3% hydrogen peroxide for 5 minutes and then 10% normal serum in 0.025% TRITON X-100-PBS for 30 minutes. 10% human IgG was used as blocking reagent for 30 minutes incubation (Sigma-Aldrich). Primary antibody was applied on the slides for over night incubation at 4° C. degree in a humidity chamber. The primary antibodies used were 1:500 rat anti-mouse Prothrombin (Innovative Research), 1:500 rabbit anti-mouse fibrinogen (Innovative Research), 1:250 rabbit anti-staphylocoagulase, or 1:250 rabbit anti-staphylococcus vwbp. Following TBS wash, the slides were incubated with biotinylated secondary antibody (1:50 dilution of

biotinylated anti-rat IgG, BA-4001 from Vector Laboratories; or 1:200 dilution of biotinylated anti-rabbit IgG, BA-1000 from Vector), and then ABC reagents (Vector Laboratories). The antigen-antibody binding was detected by DAB substrate chromogen system. The slide were briefly immersed in hematoxylin for counterstaining and evaluated under light microscope.

Active Immunization.

Three week old BALB/c mice were injected with 50 μ g protein each, emulsified in 100 μ l CFA. Cohorts of 15 mice were used, with 5 mice reserved for bleeding and antibody titers. Eleven days post vaccination, these mice were boosted with 50 μ g protein each, emulsified in 100 μ l IFA. On day 21, mice were injected with 1×10^7 CFU of staphylococci for the renal abscess model or 1×10^8 CFU for lethal challenge. At the time of infection 5 mice were bled to obtain antibody titers.

Passive Transfer of Antibodies.

Twenty four hours prior to infection, six week old BALB/c mice were injected with purified antibodies against Coa and/or vWbp at a dose of 5 mg/kg body weight. Cohorts of 10 mice were used. These mice were challenged by retro-orbital injection with 1×10^7 CFU (renal abscess model) or 1×10^8 CFU staphylococci (lethal bacteremia).

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Ala Ala Asn Ala Ala Gln His Asp Glu Ala Gln Gln Asn Ala Phe Tyr
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Gln Val Leu Asn Met Pro Asn Leu Asn Ala Asp Gln Arg Asn Gly Phe
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 385 390 395 400
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 405 410 415
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Ala Ala Asn Ala Ala Gln His Asp Glu Ala Gln Gln Asn Ala Phe Tyr	35	40	45
Gln Val Leu Asn Met Pro Asn Leu Asn Ala Asp Gln Arg Asn Gly Phe	50	55	60
Ile Gln Ser Leu Lys Asp Asp Pro Ser Gln Ser Ala Asn Val Leu Gly	65	70	80
Glu Ala Gln Lys Leu Asn Asp Ser Gln Ala Pro Lys Ala Asp Ala Gln	85	90	95
Gln Asn Asn Phe Asn Lys Asp Gln Gln Ser Ala Phe Tyr Glu Ile Leu	100	105	110
Asn Met Pro Asn Leu Asn Glu Ala Gln Arg Asn Gly Phe Ile Gln Ser	115	120	125
Leu Lys Asp Asp Pro Ser Gln Ser Thr Asn Val Leu Gly Glu Ala Lys	130	135	140
Lys Leu Asn Glu Ser Gln Ala Pro Lys Ala Asp Asn Asn Phe Asn Lys	145	150	160
Glu Gln Gln Asn Ala Phe Tyr Glu Ile Leu Asn Met Pro Asn Leu Asn	165	170	175
Glu Glu Gln Arg Asn Gly Phe Ile Gln Ser Leu Lys Asp Asp Pro Ser	180	185	190
Gln Ser Ala Asn Leu Leu Ser Glu Ala Lys Lys Leu Asn Glu Ser Gln	195	200	205
Ala Pro Lys Ala Asp Asn Lys Phe Asn Lys Glu Gln Gln Asn Ala Phe	210	215	220
Tyr Glu Ile Leu His Leu Pro Asn Leu Asn Glu Glu Gln Arg Asn Gly	225	230	240
Phe Ile Gln Ser Leu Lys Asp Asp Pro Ser Val Ser Lys Glu Ile Leu	245	250	255
Ala Glu Ala Lys Lys Leu Asn Asp Ala Gln Ala Pro Lys Glu Glu Asp	260	265	270
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Gly Asn Lys Pro Gly Lys Glu Asp Asn Asn Lys Pro Gly Lys Glu Asp	305	310	315
Gly Asn Lys Pro Gly Lys Glu Asp Asn Asn Lys Pro Gly Lys Glu Asp	325	330	335
Gly Asn Lys Pro Gly Lys Glu Asp Gly Asn Lys Pro Gly Lys Glu Asp	340	345	350
Gly Asn Gly Val His Val Val Lys Pro Gly Asp Thr Val Asn Asp Ile	355	360	365
Ala Lys Ala Asn Gly Thr Thr Ala Asp Lys Ile Ala Ala Asp Asn Lys	370	375	380
Leu Ala Asp Lys Asn Met Ile Lys Pro Gly Gln Glu Leu Val Val Asp	385	390	395
Lys Lys Gln Pro Ala Asn His Ala Asp Ala Asn Lys Ala Gln Ala Leu	405	410	415
Pro Glu Thr Gly Glu Glu Asn Pro Phe Ile Gly Thr Thr Val Phe Gly	420	425	430

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Glu Leu
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Thr Arg Ala Gln Gly Glu Ile Ala Ala Asn Trp Glu Gly Gln Ala Phe
 35 40 45

Ser Arg Phe Glu Glu Gln Phe Gln Gln Leu Ser Pro Lys Val Glu Lys
 50 55 60

Phe Ala Gln Leu Leu Glu Glu Ile Lys Gln Gln Leu Asn Ser Thr Ala
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Asp Ala Val Gln Glu Gln Asp Gln Gln Leu Ser Asn Asn Phe Gly Leu
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Gln

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 20 25 30

Lys Gln Thr Gln Gln Leu Ala Glu Tyr Ile Glu Gly Ser Asp Trp Glu
 35 40 45

Gly Gln Phe Ala Asn Lys Val Lys Asp Val Leu Leu Ile Met Ala Lys
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Phe Gln Glu Glu Leu Val Gln Pro Met Ala Asp His Gln Lys Ala Ile
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Gln Gly Leu Asp Arg Val
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Ala Ser Ile Leu Val Gly Thr Thr Leu Ile Phe Gly Leu Gly Asn Gln

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 Lys Met Thr Tyr Gly Ser Asn Gln Ser Val Asn Leu Asp Phe Gly Asp
 500 505 510
 Ile Thr Ser Ala Tyr Val Val Met Val Asn Thr Lys Phe Gln Tyr Thr
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 Asn Ser Glu Ser Pro Thr Leu Val Gln Met Ala Thr Leu Ser Ser Thr
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 Gly Asn Lys Ser Val Ser Thr Gly Asn Ala Leu Gly Phe Thr Asn Asn
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 Trp Glu Asp Thr Asn Lys Asn Gly Val Gln Glu Leu Gly Glu Lys Gly
 580 585 590
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 595 600 605
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 Asn Gly Leu Ser Ser Val Ile Thr Val Asn Gly Lys Asp Asn Leu Ser
 660 665 670
 Ala Asp Leu Gly Ile Tyr Lys Pro Lys Tyr Asn Leu Gly Asp Tyr Val
 675 680 685
 Trp Glu Asp Thr Asn Lys Asn Gly Ile Gln Asp Gln Asp Glu Lys Gly
 690 695 700
 Ile Ser Gly Val Thr Val Thr Leu Lys Asp Glu Asn Gly Asn Val Leu
 705 710 715 720
 Lys Thr Val Thr Thr Asp Ala Asp Gly Lys Tyr Lys Phe Thr Asp Leu
 725 730 735
 Asp Asn Gly Asn Tyr Lys Val Glu Phe Thr Thr Pro Glu Gly Tyr Thr
 740 745 750
 Pro Thr Thr Val Thr Ser Gly Ser Asp Ile Glu Lys Asp Ser Asn Gly
 755 760 765
 Leu Thr Thr Thr Gly Val Ile Asn Gly Ala Asp Asn Met Thr Leu Asp
 770 775 780
 Ser Gly Phe Tyr Lys Thr Pro Lys Tyr Asn Leu Gly Asn Tyr Val Trp
 785 790 795 800
 Glu Asp Thr Asn Lys Asp Gly Lys Gln Asp Ser Thr Glu Lys Gly Ile
 805 810 815
 Ser Gly Val Thr Val Thr Leu Lys Asn Glu Asn Gly Glu Val Leu Gln
 820 825 830
 Thr Thr Lys Thr Asp Lys Asp Gly Lys Tyr Gln Phe Thr Gly Leu Glu
 835 840 845
 Asn Gly Thr Tyr Lys Val Glu Phe Glu Thr Pro Ser Gly Tyr Thr Pro
 850 855 860
 Thr Gln Val Gly Ser Gly Thr Asp Glu Gly Ile Asp Ser Asn Gly Thr
 865 870 875 880

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1280	1285	1290
Ser Asp Ser Asp Ser Asp Ser Asp Ser Asp Ser Asp Ser Asp Ser		
1295	1300	1305
Asp Ser Asp Ser Asp Ser Asp Ser Asp Ser Asp Ser Asp Ser Asp		
1310	1315	1320
Ser Asp Ala Gly Lys His Thr Pro Val Lys Pro Met Ser Thr Thr		
1325	1330	1335
Lys Asp His His Asn Lys Ala Lys Ala Leu Pro Glu Thr Gly Asn		
1340	1345	1350
Glu Asn Ser Gly Ser Asn Asn Ala Thr Leu Phe Gly Gly Leu Phe		
1355	1360	1365
Ala Ala Leu Gly Ser Leu Leu Leu Phe Gly Arg Arg Lys Lys Gln		
1370	1375	1380
Asn Lys		
1385		

<210> SEQ ID NO 14

<211> LENGTH: 1141

<212> TYPE: PRT

<213> ORGANISM: Staphylococcus sp.

<400> SEQUENCE: 14

Met	Ile	Asn	Arg	Asp	Asn	Lys	Lys	Ala	Ile	Thr	Lys	Lys	Gly	Met	Ile
1				5					10					15	
Ser	Asn	Arg	Leu	Asn	Lys	Phe	Ser	Ile	Arg	Lys	Tyr	Thr	Val	Gly	Thr
		20						25					30		
Ala	Ser	Ile	Leu	Val	Gly	Thr	Thr	Leu	Ile	Phe	Gly	Leu	Gly	Asn	Gln
		35				40						45			
Glu	Ala	Lys	Ala	Ala	Glu	Asn	Thr	Ser	Thr	Glu	Asn	Ala	Lys	Gln	Asp
		50				55					60				
Asp	Ala	Thr	Thr	Ser	Asp	Asn	Lys	Glu	Val	Val	Ser	Glu	Thr	Glu	Asn
65					70					75					80
Asn	Ser	Thr	Thr	Glu	Asn	Asp	Ser	Thr	Asn	Pro	Ile	Lys	Lys	Glu	Thr
			85						90					95	
Asn	Thr	Asp	Ser	Gln	Pro	Glu	Ala	Lys	Glu	Glu	Ser	Thr	Thr	Ser	Ser
			100					105						110	
Thr	Gln	Gln	Gln	Gln	Asn	Asn	Val	Thr	Ala	Thr	Thr	Glu	Thr	Lys	Pro
			115				120						125		
Gln	Asn	Ile	Glu	Lys	Glu	Asn	Val	Lys	Pro	Ser	Thr	Asp	Lys	Thr	Ala
			130				135					140			
Thr	Glu	Asp	Thr	Ser	Val	Ile	Leu	Glu	Glu	Lys	Lys	Ala	Pro	Asn	Tyr
145					150					155					160
Thr	Asn	Asn	Asp	Val	Thr	Thr	Lys	Pro	Ser	Thr	Ser	Glu	Ile	Gln	Thr
			165						170					175	
Lys	Pro	Thr	Thr	Pro	Gln	Glu	Ser	Thr	Asn	Ile	Glu	Asn	Ser	Gln	Pro
			180						185					190	
Gln	Pro	Thr	Pro	Ser	Lys	Val	Asp	Asn	Gln	Val	Thr	Asp	Ala	Thr	Asn
			195				200						205		
Pro	Lys	Glu	Pro	Val	Asn	Val	Ser	Lys	Glu	Glu	Leu	Lys	Asn	Asn	Pro
			210				215						220		
Glu	Lys	Leu	Lys	Glu	Leu	Val	Arg	Asn	Asp	Asn	Asn	Thr	Asp	Arg	Ser
225					230					235					240
Thr	Lys	Pro	Val	Ala	Thr	Ala	Pro	Thr	Ser	Val	Ala	Pro	Lys	Arg	Leu
				245					250						255

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675				680				685							
Ser	Asn	Gly	Ser	Ser	Ile	Thr	Val	Lys	Ile	Asn	Gly	Lys	Asp	Asp	Met
690						695					700				
Ser	Leu	Asp	Thr	Gly	Phe	Tyr	Lys	Glu	Pro	Lys	Tyr	Asn	Leu	Gly	Asp
705					710					715					720
Tyr	Val	Trp	Glu	Asp	Thr	Asn	Lys	Asp	Gly	Ile	Gln	Asp	Ala	Asn	Glu
				725					730					735	
Pro	Gly	Ile	Lys	Asp	Val	Lys	Val	Thr	Leu	Lys	Asp	Ser	Thr	Gly	Lys
			740					745					750		
Val	Ile	Gly	Thr	Thr	Thr	Thr	Asp	Ala	Ser	Gly	Lys	Tyr	Lys	Phe	Thr
		755					760					765			
Asp	Leu	Asp	Asn	Gly	Asn	Tyr	Thr	Val	Glu	Phe	Glu	Thr	Pro	Ala	Gly
770						775					780				
Tyr	Thr	Pro	Thr	Val	Lys	Asn	Thr	Thr	Ala	Glu	Asp	Lys	Asp	Ser	Asn
785					790					795					800
Gly	Leu	Thr	Thr	Thr	Gly	Val	Ile	Lys	Asp	Ala	Asp	Asn	Met	Thr	Leu
				805					810					815	
Asp	Ser	Gly	Phe	Tyr	Lys	Thr	Pro	Lys	Tyr	Ser	Leu	Gly	Asp	Tyr	Val
			820					825					830		
Trp	Tyr	Asp	Ser	Asn	Lys	Asp	Gly	Lys	Gln	Asp	Ser	Thr	Glu	Lys	Gly
		835					840					845			
Ile	Lys	Asp	Val	Lys	Val	Thr	Leu	Leu	Asn	Glu	Lys	Gly	Glu	Val	Ile
850						855					860				
Gly	Thr	Thr	Lys	Thr	Asp	Glu	Asn	Gly	Lys	Tyr	Arg	Phe	Asp	Asn	Leu
865					870					875					880
Asp	Ser	Gly	Lys	Tyr	Lys	Val	Ile	Phe	Glu	Lys	Pro	Ala	Gly	Leu	Thr
				885					890					895	
Gln	Thr	Val	Thr	Asn	Thr	Thr	Glu	Asp	Asp	Lys	Asp	Ala	Asp	Gly	Gly
			900					905					910		
Glu	Val	Asp	Val	Thr	Ile	Thr	Asp	His	Asp	Asp	Phe	Thr	Leu	Asp	Asn
		915					920					925			
Gly	Tyr	Phe	Glu	Glu	Asp	Thr	Ser	Asp	Ser	Asp	Ser	Asp	Ser	Asp	Ser
930						935					940				
Asp	Ser	Asp	Ser	Asp	Ser	Asp	Ser	Asp	Ser	Asp	Ser	Asp	Ser	Asp	Ser
945					950					955					960
Asp	Ser	Asp	Ser	Asp	Ser	Asp	Ser	Asp	Ser	Asp	Ser	Asp	Ser	Asp	Ser
				965					970					975	
Asp	Ser	Asp	Ser	Asp	Ser	Asp	Ser	Asp	Ser	Asp	Ser	Asp	Ser	Asp	Ser
				980				985					990		
Asp	Ser	Asp	Ser	Asp	Ser	Asp	Ser	Asp	Ser	Asp	Ser	Asp	Ser	Asp	Ser
				995				1000				1005			
Asp	Ser	Asp	Ser	Asp	Ser	Asp	Ser	Asp	Ser	Asp	Ser	Asp	Ser	Asp	Ser
1010						1015						1020			
Ser	Asp	Ser	Asp	Ser	Asp	Ser	Asp	Ser	Asp	Ser	Asp	Ser	Asp	Ser	Ser
1025						1030						1035			
Asp	Ser	Asp	Ser	Asp	Ser	Asp	Ser	Asp	Ser	Asp	Ser	Asp	Ser	Asp	Ser
1040						1045						1050			
Ser	Asp	Ser	Asp	Ser	Asp	Ser	Asp	Ser	Asp	Ser	Asp	Ser	Asp	Ser	Ser
1055						1060						1065			
Asp	Ser	Asp	Ser	Asp	Ser	Asp	Ser	Asp	Ser	Asp	Ser	Asp	Ala	Gly	
1070						1075						1080			
Lys	His	Thr	Pro	Val	Lys	Pro	Met	Ser	Thr	Thr	Lys	Asp	His	His	
1085						1090						1095			

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Asn Lys Ala Lys Ala Leu Pro Glu Thr Gly Ser Glu Asn Asn Gly
1100 1105 1110

Ser Asn Asn Ala Thr Leu Phe Gly Gly Leu Phe Ala Ala Leu Gly
1115 1120 1125

Ser Leu Leu Leu Phe Gly Arg Arg Lys Lys Gln Asn Lys
1130 1135 1140

<210> SEQ ID NO 15

<211> LENGTH: 350

<212> TYPE: PRT

<213> ORGANISM: Staphylococcus sp.

<400> SEQUENCE: 15

Met Thr Lys His Tyr Leu Asn Ser Lys Tyr Gln Ser Glu Gln Arg Ser
1 5 10 15

Ser Ala Met Lys Lys Ile Thr Met Gly Thr Ala Ser Ile Ile Leu Gly
20 25 30

Ser Leu Val Tyr Ile Gly Ala Asp Ser Gln Gln Val Asn Ala Ala Thr
35 40 45

Glu Ala Thr Asn Ala Thr Asn Asn Gln Ser Thr Gln Val Ser Gln Ala
50 55 60

Thr Ser Gln Pro Ile Asn Phe Gln Val Gln Lys Asp Gly Ser Ser Glu
65 70 75 80

Lys Ser His Met Asp Asp Tyr Met Gln His Pro Gly Lys Val Ile Lys
85 90 95

Gln Asn Asn Lys Tyr Tyr Phe Gln Thr Val Leu Asn Asn Ala Ser Phe
100 105 110

Trp Lys Glu Tyr Lys Phe Tyr Asn Ala Asn Asn Gln Glu Leu Ala Thr
115 120 125

Thr Val Val Asn Asp Asn Lys Lys Ala Asp Thr Arg Thr Ile Asn Val
130 135 140

Ala Val Glu Pro Gly Tyr Lys Ser Leu Thr Thr Lys Val His Ile Val
145 150 155 160

Val Pro Gln Ile Asn Tyr Asn His Arg Tyr Thr Thr His Leu Glu Phe
165 170 175

Glu Lys Ala Ile Pro Thr Leu Ala Asp Ala Ala Lys Pro Asn Asn Val
180 185 190

Lys Pro Val Gln Pro Lys Pro Ala Gln Pro Lys Thr Pro Thr Glu Gln
195 200 205

Thr Lys Pro Val Gln Pro Lys Val Glu Lys Val Lys Pro Thr Val Thr
210 215 220

Thr Thr Ser Lys Val Glu Asp Asn His Ser Thr Lys Val Val Ser Thr
225 230 235 240

Asp Thr Thr Lys Asp Gln Thr Lys Thr Gln Thr Ala His Thr Val Lys
245 250 255

Thr Ala Gln Thr Ala Gln Glu Gln Asn Lys Val Gln Thr Pro Val Lys
260 265 270

Asp Val Ala Thr Ala Lys Ser Glu Ser Asn Asn Gln Ala Val Ser Asp
275 280 285

Asn Lys Ser Gln Gln Thr Asn Lys Val Thr Lys His Asn Glu Thr Pro
290 295 300

Lys Gln Ala Ser Lys Ala Lys Glu Leu Pro Lys Thr Gly Leu Thr Ser
305 310 315 320

Val Asp Asn Phe Ile Ser Thr Val Ala Phe Ala Thr Leu Ala Leu Leu

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Tyr Glu Ser Val Glu Asn Asn Glu Ser Met Met Asp Thr Phe Val Lys
 355 360 365
 His Pro Ile Lys Thr Gly Met Leu Asn Gly Lys Lys Tyr Met Val Met
 370 375 380
 Glu Thr Thr Asn Asp Asp Tyr Trp Lys Asp Phe Met Val Glu Gly Gln
 385 390 395 400
 Arg Val Arg Thr Ile Ser Lys Asp Ala Lys Asn Asn Thr Arg Thr Ile
 405 410 415
 Ile Phe Pro Tyr Val Glu Gly Lys Thr Leu Tyr Asp Ala Ile Val Lys
 420 425 430
 Val His Val Lys Thr Ile Asp Tyr Asp Gly Gln Tyr His Val Arg Ile
 435 440 445
 Val Asp Lys Glu Ala Phe Thr Lys Ala Asn Thr Asp Lys Ser Asn Lys
 450 455 460
 Lys Glu Gln Gln Asp Asn Ser Ala Lys Lys Glu Ala Thr Pro Ala Thr
 465 470 475 480
 Pro Ser Lys Pro Thr Pro Ser Pro Val Glu Lys Glu Ser Gln Lys Gln
 485 490 495
 Asp Ser Gln Lys Asp Asp Asn Lys Gln Leu Pro Ser Val Glu Lys Glu
 500 505 510
 Asn Asp Ala Ser Ser Glu Ser Gly Lys Asp Lys Thr Pro Ala Thr Lys
 515 520 525
 Pro Thr Lys Gly Glu Val Glu Ser Ser Ser Thr Thr Pro Thr Lys Val
 530 535 540
 Val Ser Thr Thr Gln Asn Val Ala Lys Pro Thr Thr Ala Ser Ser Lys
 545 550 555 560
 Thr Thr Lys Asp Val Val Gln Thr Ser Ala Gly Ser Ser Glu Ala Lys
 565 570 575
 Asp Ser Ala Pro Leu Gln Lys Ala Asn Ile Lys Asn Thr Asn Asp Gly
 580 585 590
 His Thr Gln Ser Gln Asn Asn Lys Asn Thr Gln Glu Asn Lys Ala Lys
 595 600 605
 Ser Leu Pro Gln Thr Gly Glu Glu Ser Asn Lys Asp Met Thr Leu Pro
 610 615 620
 Leu Met Ala Leu Leu Ala Leu Ser Ser Ile Val Ala Phe Val Leu Pro
 625 630 635 640
 Arg Lys Arg Lys Asn
 645

<210> SEQ ID NO 17

<211> LENGTH: 80

<212> TYPE: PRT

<213> ORGANISM: Staphylococcus sp.

<400> SEQUENCE: 17

Met Asn Gln His Val Lys Val Thr Phe Asp Phe Thr Asn Tyr Asn Tyr
 1 5 10 15
 Gly Thr Tyr Asp Leu Ala Val Pro Ala Tyr Leu Pro Ile Lys Asn Leu
 20 25 30
 Ile Ala Leu Val Leu Asp Ser Leu Asp Ile Ser Ile Phe Asp Val Asn
 35 40 45
 Thr Gln Ile Lys Val Met Thr Lys Gly Gln Leu Leu Val Glu Asn Asp
 50 55 60
 Arg Leu Ile Asp Tyr Gln Ile Ala Asp Gly Asp Ile Leu Lys Leu Leu
 65 70 75 80

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<210> SEQ ID NO 18
 <211> LENGTH: 877
 <212> TYPE: PRT
 <213> ORGANISM: Staphylococcus sp.

<400> SEQUENCE: 18

Met Lys Lys Arg Ile Asp Tyr Leu Ser Asn Lys Gln Asn Lys Tyr Ser
 1 5 10 15
 Ile Arg Arg Phe Thr Val Gly Thr Thr Ser Val Ile Val Gly Ala Thr
 20 25 30
 Ile Leu Phe Gly Ile Gly Asn His Gln Ala Gln Ala Ser Glu Gln Ser
 35 40 45
 Asn Asp Thr Thr Gln Ser Ser Lys Asn Asn Ala Ser Ala Asp Ser Glu
 50 55 60
 Lys Asn Asn Met Ile Glu Thr Pro Gln Leu Asn Thr Thr Ala Asn Asp
 65 70 75 80
 Thr Ser Asp Ile Ser Ala Asn Thr Asn Ser Ala Asn Val Asp Ser Thr
 85 90 95
 Thr Lys Pro Met Ser Thr Gln Thr Ser Asn Thr Thr Thr Thr Glu Pro
 100 105 110
 Ala Ser Thr Asn Glu Thr Pro Gln Pro Thr Ala Ile Lys Asn Gln Ala
 115 120 125
 Thr Ala Ala Lys Met Gln Asp Gln Thr Val Pro Gln Glu Ala Asn Ser
 130 135 140
 Gln Val Asp Asn Lys Thr Thr Asn Asp Ala Asn Ser Ile Ala Thr Asn
 145 150 155 160
 Ser Glu Leu Lys Asn Ser Gln Thr Leu Asp Leu Pro Gln Ser Ser Pro
 165 170 175
 Gln Thr Ile Ser Asn Ala Gln Gly Thr Ser Lys Pro Ser Val Arg Thr
 180 185 190
 Arg Ala Val Arg Ser Leu Ala Val Ala Glu Pro Val Val Asn Ala Ala
 195 200 205
 Asp Ala Lys Gly Thr Asn Val Asn Asp Lys Val Thr Ala Ser Asn Phe
 210 215 220
 Lys Leu Glu Lys Thr Thr Phe Asp Pro Asn Gln Ser Gly Asn Thr Phe
 225 230 235 240
 Met Ala Ala Asn Phe Thr Val Thr Asp Lys Val Lys Ser Gly Asp Tyr
 245 250 255
 Phe Thr Ala Lys Leu Pro Asp Ser Leu Thr Gly Asn Gly Asp Val Asp
 260 265 270
 Tyr Ser Asn Ser Asn Asn Thr Met Pro Ile Ala Asp Ile Lys Ser Thr
 275 280 285
 Asn Gly Asp Val Val Ala Lys Ala Thr Tyr Asp Ile Leu Thr Lys Thr
 290 295 300
 Tyr Thr Phe Val Phe Thr Asp Tyr Val Asn Asn Lys Glu Asn Ile Asn
 305 310 315 320
 Gly Gln Phe Ser Leu Pro Leu Phe Thr Asp Arg Ala Lys Ala Pro Lys
 325 330 335
 Ser Gly Thr Tyr Asp Ala Asn Ile Asn Ile Ala Asp Glu Met Phe Asn
 340 345 350
 Asn Lys Ile Thr Tyr Asn Tyr Ser Ser Pro Ile Ala Gly Ile Asp Lys
 355 360 365
 Pro Asn Gly Ala Asn Ile Ser Ser Gln Ile Ile Gly Val Asp Thr Ala

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370					375					380					
Ser	Gly	Gln	Asn	Thr	Tyr	Lys	Gln	Thr	Val	Phe	Val	Asn	Pro	Lys	Gln
385					390					395					400
Arg	Val	Leu	Gly	Asn	Thr	Trp	Val	Tyr	Ile	Lys	Gly	Tyr	Gln	Asp	Lys
				405					410					415	
Ile	Glu	Glu	Ser	Ser	Gly	Lys	Val	Ser	Ala	Thr	Asp	Thr	Lys	Leu	Arg
			420					425						430	
Ile	Phe	Glu	Val	Asn	Asp	Thr	Ser	Lys	Leu	Ser	Asp	Ser	Tyr	Tyr	Ala
		435					440							445	
Asp	Pro	Asn	Asp	Ser	Asn	Leu	Lys	Glu	Val	Thr	Asp	Gln	Phe	Lys	Asn
	450						455							460	
Arg	Ile	Tyr	Tyr	Glu	His	Pro	Asn	Val	Ala	Ser	Ile	Lys	Phe	Gly	Asp
	465				470						475				480
Ile	Thr	Lys	Thr	Tyr	Val	Val	Leu	Val	Glu	Gly	His	Tyr	Asp	Asn	Thr
				485					490						495
Gly	Lys	Asn	Leu	Lys	Thr	Gln	Val	Ile	Gln	Glu	Asn	Val	Asp	Pro	Val
			500						505						510
Thr	Asn	Arg	Asp	Tyr	Ser	Ile	Phe	Gly	Trp	Asn	Asn	Glu	Asn	Val	Val
		515					520								525
Arg	Tyr	Gly	Gly	Gly	Ser	Ala	Asp	Gly	Asp	Ser	Ala	Val	Asn	Pro	Lys
	530						535								540
Asp	Pro	Thr	Pro	Gly	Pro	Pro	Val	Asp	Pro	Glu	Pro	Ser	Pro	Asp	Pro
	545				550						555				560
Glu	Pro	Glu	Pro	Thr	Pro	Asp	Pro	Glu	Pro	Ser	Pro	Asp	Pro	Glu	Pro
				565					570						575
Glu	Pro	Ser	Pro	Asp	Pro	Asp	Pro	Asp	Ser	Asp	Ser	Asp	Ser	Asp	Ser
			580						585						590
Gly	Ser	Asp	Ser	Asp	Ser	Gly	Ser	Asp	Ser	Asp	Ser	Glu	Ser	Asp	Ser
		595					600						605		
Asp	Ser	Asp	Ser	Asp	Ser	Asp	Ser	Asp	Ser	Asp	Ser	Asp	Ser	Glu	Ser
	610						615								620
Asp	Ser	Asp	Ser	Glu	Ser	Asp	Ser	Asp	Ser	Asp	Ser	Asp	Ser	Asp	Ser
	625				630						635				640
Asp	Ser	Asp	Ser	Asp	Ser	Glu	Ser	Asp	Ser	Asp	Ser	Asp	Ser	Asp	Ser
				645					650						655
Asp	Ser	Asp	Ser	Asp	Ser	Asp	Ser	Glu	Ser	Asp	Ser	Asp	Ser	Asp	Ser
			660						665						670
Asp	Ser	Glu	Ser	Asp	Ser	Asp	Ser	Asp	Ser	Asp	Ser	Asp	Ser	Asp	Ser
		675							680						685
Asp	Ser	Asp	Ser	Asp	Ser	Asp	Ser	Asp	Ser	Asp	Ser	Asp	Ser	Asp	Ser
									695						700
Asp	Ser	Asp	Ser	Asp	Ser	Asp	Ser	Glu	Ser	Asp	Ser	Asp	Ser	Asp	Ser
									710						720
Asp	Ser	Asp	Ser	Asp	Ser	Asp	Ser	Asp	Ser	Asp	Ser	Asp	Ser	Asp	Ser
				725					730						735
Asp	Ser	Asp	Ser	Asp	Ser	Asp	Ser	Asp	Ser	Asp	Ser	Asp	Ser	Asp	Ser
				740					745						750
Asp	Ser	Asp	Ser	Asp	Ser	Asp	Ser	Asp	Ser	Asp	Ser	Asp	Ser	Asp	Ser
				755					760						765
Asp	Ser	Asp	Ser	Asp	Ser	Asp	Ser	Asp	Ser	Asp	Ser	Asp	Ser	Asp	Ser
				770					775						780
Asp	Ser	Asp	Ser	Asp	Ser	Asp	Ser	Asp	Ser	Asp	Ser	Asp	Ser	Asp	Ser
				785					790						800

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Lys Lys His Phe Ala Ser Thr Gly Asp Thr Ser Ser Asp Asp Ile Leu
 435 440 445
 Lys Ala Ile Leu Asn Asn Ala Lys Asp Lys Lys Gln Ala Ile Glu Thr
 450 455 460
 Ile Leu Ala Thr Arg Ile Glu Arg Gln Lys Ala Lys Leu Leu Ala Asp
 465 470 475 480
 Leu Ile Thr Lys Ile Glu Thr Asp Gln Asn Lys Ile Phe Asn Leu Val
 485 490 495
 Lys Ser Ala Leu Asn Gly Lys Ala Asp Asp Leu Leu Asn Leu Gln Lys
 500 505 510
 Arg Leu Asn Gln Thr Lys Lys Asp Ile Asp Tyr Ile Leu Ser Pro Ile
 515 520 525
 Val Asn Arg Pro Ser Leu Leu Asp Arg Leu Asn Lys Asn Gly Lys Thr
 530 535 540
 Thr Asp Leu Asn Lys Leu Ala Asn Leu Met Asn Gln Gly Ser Asn Leu
 545 550 555 560
 Leu Asp Ser Ile Pro Asp Ile Pro Thr Pro Lys Pro Glu Lys Thr Leu
 565 570 575
 Thr Leu Gly Lys Gly Asn Gly Leu Leu Ser Gly Leu Leu Asn Ala Asp
 580 585 590
 Gly Asn Val Ser Leu Pro Lys Ala Gly Glu Thr Ile Lys Glu His Trp
 595 600 605
 Leu Pro Ile Ser Val Ile Val Gly Ala Met Gly Val Leu Met Ile Trp
 610 615 620
 Leu Ser Arg Arg Asn Lys Leu Lys Asn Lys Ala
 625 630 635

<210> SEQ ID NO 21

<211> LENGTH: 953

<212> TYPE: PRT

<213> ORGANISM: Staphylococcus sp.

<400> SEQUENCE: 21

Met Asn Asn Lys Lys Thr Ala Thr Asn Arg Lys Gly Met Ile Pro Asn
 1 5 10 15
 Arg Leu Asn Lys Phe Ser Ile Arg Lys Tyr Ser Val Gly Thr Ala Ser
 20 25 30
 Ile Leu Val Gly Thr Thr Leu Ile Phe Gly Leu Ser Gly His Glu Ala
 35 40 45
 Lys Ala Ala Glu His Thr Asn Gly Glu Leu Asn Gln Ser Lys Asn Glu
 50 55 60
 Thr Thr Ala Pro Ser Glu Asn Lys Thr Thr Glu Lys Val Asp Ser Arg
 65 70 75 80
 Gln Leu Lys Asp Asn Thr Gln Thr Ala Thr Ala Asp Gln Pro Lys Val
 85 90 95
 Thr Met Ser Asp Ser Ala Thr Val Lys Glu Thr Ser Ser Asn Met Gln
 100 105 110
 Ser Pro Gln Asn Ala Thr Ala Ser Gln Ser Thr Thr Gln Thr Ser Asn
 115 120 125
 Val Thr Thr Asn Asp Lys Ser Ser Thr Thr Tyr Ser Asn Glu Thr Asp
 130 135 140
 Lys Ser Asn Leu Thr Gln Ala Lys Asn Val Ser Thr Thr Pro Lys Thr
 145 150 155 160
 Thr Thr Ile Lys Gln Arg Ala Leu Asn Arg Met Ala Val Asn Thr Val
 165 170 175

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Ala Ala Pro Gln Gln Gly Thr Asn Val Asn Asp Lys Val His Phe Thr
 180 185 190

Asn Ile Asp Ile Ala Ile Asp Lys Gly His Val Asn Lys Thr Thr Gly
 195 200 205

Asn Thr Glu Phe Trp Ala Thr Ser Ser Asp Val Leu Lys Leu Lys Ala
 210 215 220

Asn Tyr Thr Ile Asp Asp Ser Val Lys Glu Gly Asp Thr Phe Thr Phe
 225 230 235 240

Lys Tyr Gly Gln Tyr Phe Arg Pro Gly Ser Val Arg Leu Pro Ser Gln
 245 250 255

Thr Gln Asn Leu Tyr Asn Ala Gln Gly Asn Ile Ile Ala Lys Gly Ile
 260 265 270

Tyr Asp Ser Lys Thr Asn Thr Thr Thr Tyr Thr Phe Thr Asn Tyr Val
 275 280 285

Asp Gln Tyr Thr Asn Val Ser Gly Ser Phe Glu Gln Val Ala Phe Ala
 290 295 300

Lys Arg Glu Asn Ala Thr Thr Asp Lys Thr Ala Tyr Lys Met Glu Val
 305 310 315 320

Thr Leu Gly Asn Asp Thr Tyr Ser Lys Asp Val Ile Val Asp Tyr Gly
 325 330 335

Asn Gln Lys Gly Gln Gln Leu Ile Ser Ser Thr Asn Tyr Ile Asn Asn
 340 345 350

Glu Asp Leu Ser Arg Asn Met Thr Val Tyr Val Asn Gln Pro Lys Lys
 355 360 365

Thr Tyr Thr Lys Glu Thr Phe Val Thr Asn Leu Thr Gly Tyr Lys Phe
 370 375 380

Asn Pro Asp Ala Lys Asn Phe Lys Ile Tyr Glu Val Thr Asp Gln Asn
 385 390 395 400

Gln Phe Val Asp Ser Phe Thr Pro Asp Thr Ser Lys Leu Lys Asp Val
 405 410 415

Thr Gly Gln Phe Asp Val Ile Tyr Ser Asn Asp Asn Lys Thr Ala Thr
 420 425 430

Val Asp Leu Leu Asn Gly Gln Ser Ser Ser Asp Lys Gln Tyr Ile Ile
 435 440 445

Gln Gln Val Ala Tyr Pro Asp Asn Ser Ser Thr Asp Asn Gly Lys Ile
 450 455 460

Asp Tyr Thr Leu Glu Thr Gln Asn Gly Lys Ser Ser Trp Ser Asn Ser
 465 470 475 480

Tyr Ser Asn Val Asn Gly Ser Ser Thr Ala Asn Gly Asp Gln Lys Lys
 485 490 495

Tyr Asn Leu Gly Asp Tyr Val Trp Glu Asp Thr Asn Lys Asp Gly Lys
 500 505 510

Gln Asp Ala Asn Glu Lys Gly Ile Lys Gly Val Tyr Val Ile Leu Lys
 515 520 525

Asp Ser Asn Gly Lys Glu Leu Asp Arg Thr Thr Thr Asp Glu Asn Gly
 530 535 540

Lys Tyr Gln Phe Thr Gly Leu Ser Asn Gly Thr Tyr Ser Val Glu Phe
 545 550 555 560

Ser Thr Pro Ala Gly Tyr Thr Pro Thr Thr Ala Asn Ala Gly Thr Asp
 565 570 575

Asp Ala Val Asp Ser Asp Gly Leu Thr Thr Thr Gly Val Ile Lys Asp
 580 585 590

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Ala Asp Asn Met Thr Leu Asp Ser Gly Phe Tyr Lys Thr Pro Lys Tyr
595 600 605

Ser Leu Gly Asp Tyr Val Trp Tyr Asp Ser Asn Lys Asp Gly Lys Gln
610 615 620

Asp Ser Thr Glu Lys Gly Ile Lys Gly Val Lys Val Thr Leu Gln Asn
625 630 635 640

Glu Lys Gly Glu Val Ile Gly Thr Thr Glu Thr Asp Glu Asn Gly Lys
645 650 655

Tyr Arg Phe Asp Asn Leu Asp Ser Gly Lys Tyr Lys Val Ile Phe Glu
660 665 670

Lys Pro Ala Gly Leu Thr Gln Thr Gly Thr Asn Thr Thr Glu Asp Asp
675 680 685

Lys Asp Ala Asp Gly Gly Glu Val Asp Val Thr Ile Thr Asp His Asp
690 695 700

Asp Phe Thr Leu Asp Asn Gly Tyr Tyr Glu Glu Glu Thr Ser Asp Ser
705 710 715 720

Asp Ser Asp Ser Asp Ser Asp Ser Asp Ser Asp Ser Asp Ser Asp Ser
725 730 735

Asp Ser Asp Ser Asp Ser Asp Ser Asp Ser Asp Ser Asp Ser Asp Ser
740 745 750

Asp Ser Asp Ser Asp Ser Asp Ser Asp Ser Asp Ser Asp Ser Asp Ser
755 760 765

Asp Ser Asp Ser Glu Ser Asp Ser Asp Ser Asp Ser Asp Ser Asp Ser
770 775 780

Asp Ser Asp Ser Asp Ser Asp Ser Asp Ser Asp Ser Asp Ser Asp Ser
785 790 795 800

Asp Ser Asp Ser Asp Ser Asp Ser Asp Ser Asp Ser Asp Ser Asp Ser
805 810 815

Asp Ser Asp Ser Asp Ser Asp Asn Asp Ser Asp Ser Asp Ser Asp Ser
820 825 830

Asp Ser Asp Ser Asp Ser Asp Ser Asp Ser Asp Ser Asp Ser Asp Ser
835 840 845

Asp Ser Asp Ser Asp Ser Asp Ser Asp Ser Asp Ser Asp Ser Asp Ser
850 855 860

Asp Ser Asp Ser Asp Ser Asp Ser Asp Ser Asp Ser Asp Ser Asp Ser
865 870 875 880

Asp Ser Asp Ser Asp Ser Asp Ser Asp Ser Asp Ser Asp Ala Gly Lys
885 890 895

His Thr Pro Thr Lys Pro Met Ser Thr Val Lys Asp Gln His Lys Thr
900 905 910

Ala Lys Ala Leu Pro Glu Thr Gly Ser Glu Asn Asn Asn Ser Asn Asn
915 920 925

Gly Thr Leu Phe Gly Gly Leu Phe Ala Ala Leu Gly Ser Leu Leu Leu
930 935 940

Phe Gly Arg Arg Lys Lys Gln Asn Lys
945 950

<210> SEQ ID NO 22

<211> LENGTH: 989

<212> TYPE: PRT

<213> ORGANISM: Staphylococcus sp.

<400> SEQUENCE: 22

Met Asn Met Lys Lys Lys Glu Lys His Ala Ile Arg Lys Lys Ser Ile
1 5 10 15

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Ile	Lys	Val	Tyr	Arg	Val	Asp	Asn	Ala	Asn	Asp	Leu	Ser	Glu	Ser	Tyr
		435					440					445			
Tyr	Val	Asn	Pro	Ser	Asp	Phe	Glu	Asp	Val	Thr	Asn	Gln	Val	Arg	Ile
	450					455					460				
Ser	Phe	Pro	Asn	Ala	Asn	Gln	Tyr	Lys	Val	Glu	Phe	Pro	Thr	Asp	Asp
465					470					475					480
Asp	Gln	Ile	Thr	Thr	Pro	Tyr	Ile	Val	Val	Val	Asn	Gly	His	Ile	Asp
				485					490					495	
Pro	Ala	Ser	Thr	Gly	Asp	Leu	Ala	Leu	Arg	Ser	Thr	Phe	Tyr	Gly	Tyr
			500					505					510		
Asp	Ser	Asn	Phe	Ile	Trp	Arg	Ser	Met	Ser	Trp	Asp	Asn	Glu	Val	Ala
		515					520					525			
Phe	Asn	Asn	Gly	Ser	Gly	Ser	Gly	Asp	Gly	Ile	Asp	Lys	Pro	Val	Val
	530					535					540				
Pro	Glu	Gln	Pro	Asp	Glu	Pro	Gly	Glu	Ile	Glu	Pro	Ile	Pro	Glu	Asp
545					550					555					560
Ser	Asp	Ser	Asp	Pro	Gly	Ser	Asp	Ser	Gly	Ser	Asp	Ser	Asn	Ser	Asp
				565					570					575	
Ser	Gly	Ser	Asp	Ser	Gly	Ser	Asp	Ser	Thr	Ser	Asp	Ser	Gly	Ser	Asp
			580					585					590		
Ser	Ala	Ser	Asp	Ser	Asp	Ser	Ala	Ser	Asp	Ser	Asp	Ser	Ala	Ser	Asp
		595					600					605			
Ser	Asp	Ser	Ala	Ser	Asp	Ser	Asp	Ser	Ala	Ser	Asp	Ser	Asp	Ser	Ala
	610					615					620				
Ser	Asp	Ser	Asp	Ser	Ala	Ser	Asp	Ser	Asp	Ser	Ala	Ser	Asp	Ser	Asp
625					630					635					640
Ser	Ala	Ser	Asp	Ser	Asp	Ser	Ala	Ser	Asp	Ser	Asp	Ser	Ala	Ser	Asp
				645					650					655	
Ser	Asp	Ser	Ala	Ser	Asp	Ser	Asp	Ser	Ala	Ser	Asp	Ser	Asp	Ser	Asp
			660					665					670		
Ser	Asp	Ser	Asp	Ser	Asp	Ser	Asp	Ser	Asp	Ser	Asp	Ser	Asp	Ser	Asp
		675					680					685			
Ser	Asp	Ser	Asp	Ser	Asp	Ser	Asp	Ser	Asp	Ser	Asp	Ser	Asp	Ser	Asp
	690					695				700					
Ser	Asp	Ser	Asp	Ser	Asp	Ser	Asp	Ser	Asp	Ser	Asp	Ser	Asp	Ser	Asp
705					710					715					720
Ser	Asp	Ser	Asp	Ser	Asp	Ser	Asp	Ser	Asp	Ser	Asp	Ser	Asp	Ser	Asp
				725					730					735	
Ser	Asp	Ser	Asp	Ser	Asp	Ser	Asp	Ser	Asp	Ser	Asp	Ser	Asp	Ser	Asp
			740					745					750		
Ser	Asp	Ser	Asp	Ser	Asp	Ser	Asp	Ser	Asp	Ser	Asp	Ser	Asp	Ser	Asp
		755					760					765			
Ser	Asp	Ser	Asp	Ser	Asp	Ser	Asp	Ser	Asp	Ser	Asp	Ser	Asp	Ser	Asp
	770					775				780					
Ser	Asp	Ser	Asp	Ser	Asp	Ser	Asp	Ser	Asp	Ser	Asp	Ser	Asp	Ser	Asp
785					790					795					800
Ser	Asp	Ser	Asp	Ser	Asp	Ser	Asp	Ser	Asp	Ser	Asp	Ser	Asp	Ser	Asp
				805					810					815	
Ser	Asp	Ser	Asp	Ser	Ala	Ser	Asp	Ser	Asp	Ser	Asp	Ser	Asp	Ser	Glu
				820				825					830		
Ser	Asp	Ser	Asp	Ser	Asp	Ser	Asp	Ser	Asp	Ser	Asp	Ser	Asp	Ser	Asp
	835						840					845			
Ser	Asp	Ser	Asp	Ser	Asp	Ser	Glu	Ser	Asp	Ser	Asp	Ser	Asp	Ser	Asp

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850	855	860
Ser Asp Ser Glu Ser Asp Ser Asp Ser Asp Ser Asp Ser Asp Ser Asp		
865	870	875 880
Ser Ala Ser Asp Ser Asp Ser Gly Ser Asp Ser Asp Ser Ser Ser Asp		
	885	890 895
Ser Asp Ser Asp Ser Thr Ser Asp Thr Gly Ser Asp Asn Asp Ser Asp		
	900	905 910
Ser Asp Ser Asn Ser Asp Ser Glu Ser Gly Ser Asn Asn Asn Val Val		
	915	920 925
Pro Pro Asn Ser Pro Lys Asn Gly Thr Asn Ala Ser Asn Lys Asn Glu		
	930	935 940
Ala Lys Asp Ser Lys Glu Pro Leu Pro Asp Thr Gly Ser Glu Asp Glu		
	945	950 955 960
Ala Asn Thr Ser Leu Ile Trp Gly Leu Leu Ala Ser Leu Gly Ser Leu		
	965	970 975
Leu Leu Phe Arg Arg Lys Lys Glu Asn Lys Asp Lys Lys		
	980	985

<210> SEQ ID NO 23

<211> LENGTH: 584

<212> TYPE: PRT

<213> ORGANISM: Staphylococcus sp.

<400> SEQUENCE: 23

Met Lys Phe Lys Ser Leu Ile Thr Thr Thr Leu Ala Leu Gly Val Leu		
1	5	10 15
Ala Ser Thr Gly Ala Asn Phe Asn Asn Asn Glu Ala Ser Ala Ala Ala		
	20	25 30
Lys Pro Leu Asp Lys Ser Ser Ser Ser Leu His His Gly Tyr Ser Lys		
	35	40 45
Val His Val Pro Tyr Ala Ile Thr Val Asn Gly Thr Ser Gln Asn Ile		
	50	55 60
Leu Ser Ser Leu Thr Phe Asn Lys Asn Gln Asn Ile Ser Tyr Lys Asp		
	65	70 75 80
Leu Glu Asp Arg Val Lys Ser Val Leu Lys Ser Asp Arg Gly Ile Ser		
	85	90 95
Asp Ile Asp Leu Arg Leu Ser Lys Gln Ala Lys Tyr Thr Val Tyr Phe		
	100	105 110
Lys Asn Gly Thr Lys Lys Val Ile Asp Leu Lys Ala Gly Ile Tyr Thr		
	115	120 125
Ala Asp Leu Ile Asn Thr Ser Glu Ile Lys Ala Ile Asn Ile Asn Val		
	130	135 140
Asp Thr Lys Lys Gln Val Glu Asp Lys Lys Lys Asp Lys Ala Asn Tyr		
	145	150 155 160
Gln Val Pro Tyr Thr Ile Thr Val Asn Gly Thr Ser Gln Asn Ile Leu		
	165	170 175
Ser Asn Leu Thr Phe Asn Lys Asn Gln Asn Ile Ser Tyr Lys Asp Leu		
	180	185 190
Glu Asp Lys Val Lys Ser Val Leu Glu Ser Asn Arg Gly Ile Thr Asp		
	195	200 205
Val Asp Leu Arg Leu Ser Lys Gln Ala Lys Tyr Thr Val Asn Phe Lys		
	210	215 220
Asn Gly Thr Lys Lys Val Ile Asp Leu Lys Ser Gly Ile Tyr Thr Ala		
	225	230 235 240

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Asn Leu Ile Asn Ser Ser Asp Ile Lys Ser Ile Asn Ile Asn Val Asp
      245      250      255

Thr Lys Lys His Ile Glu Asn Lys Ala Lys Arg Asn Tyr Gln Val Pro
      260      265      270

Tyr Ser Ile Asn Leu Asn Gly Thr Ser Thr Asn Ile Leu Ser Asn Leu
      275      280      285

Ser Phe Ser Asn Lys Pro Trp Thr Asn Tyr Lys Asn Leu Thr Ser Gln
      290      295      300

Ile Lys Ser Val Leu Lys His Asp Arg Gly Ile Ser Glu Gln Asp Leu
      305      310      315      320

Lys Tyr Ala Lys Lys Ala Tyr Tyr Thr Val Tyr Phe Lys Asn Gly Gly
      325      330      335

Lys Arg Ile Leu Gln Leu Asn Ser Lys Asn Tyr Thr Ala Asn Leu Val
      340      345      350

His Ala Lys Asp Val Lys Arg Ile Glu Ile Thr Val Lys Thr Gly Thr
      355      360      365

Lys Ala Lys Ala Asp Arg Tyr Val Pro Tyr Thr Ile Ala Val Asn Gly
      370      375      380

Thr Ser Thr Pro Ile Leu Ser Asp Leu Lys Phe Thr Gly Asp Pro Arg
      385      390      395      400

Val Gly Tyr Lys Asp Ile Ser Lys Lys Val Lys Ser Val Leu Lys His
      405      410      415

Asp Arg Gly Ile Gly Glu Arg Glu Leu Lys Tyr Ala Lys Lys Ala Thr
      420      425      430

Tyr Thr Val His Phe Lys Asn Gly Thr Lys Lys Val Ile Asn Ile Asn
      435      440      445

Ser Asn Ile Ser Gln Leu Asn Leu Leu Tyr Val Gln Asp Ile Lys Lys
      450      455      460

Ile Asp Ile Asp Val Lys Thr Gly Thr Lys Ala Lys Ala Asp Ser Tyr
      465      470      475      480

Val Pro Tyr Thr Ile Ala Val Asn Gly Thr Ser Thr Pro Ile Leu Ser
      485      490      495

Lys Leu Lys Ile Ser Asn Lys Gln Leu Ile Ser Tyr Lys Tyr Leu Asn
      500      505      510

Asp Lys Val Lys Ser Val Leu Lys Ser Glu Arg Gly Ile Ser Asp Leu
      515      520      525

Asp Leu Lys Phe Ala Lys Gln Ala Lys Tyr Thr Val Tyr Phe Lys Asn
      530      535      540

Gly Lys Lys Gln Val Val Asn Leu Lys Ser Asp Ile Phe Thr Pro Asn
      545      550      555      560

Leu Phe Ser Ala Lys Asp Ile Lys Lys Ile Asp Ile Asp Val Lys Gln
      565      570      575

Tyr Thr Lys Ser Lys Lys Asn Lys
      580

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<210> SEQ ID NO 24

<211> LENGTH: 10419

<212> TYPE: PRT

<213> ORGANISM: Staphylococcus sp.

<400> SEQUENCE: 24

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Met Asn Tyr Arg Asp Lys Ile Gln Lys Phe Ser Ile Arg Lys Tyr Thr
  1      5      10      15

Val Gly Thr Phe Ser Thr Val Ile Ala Thr Leu Val Phe Leu Gly Phe
      20      25      30

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Phe Ser Glu Ala Gly Ala Thr Pro Ala Thr Lys Ala Phe Gly Arg Gln
 450 455 460
 Asn Phe Glu Tyr Ile Asn Gly Gln Lys Pro Ala Glu Ser Pro Gly Val
 465 470 475 480
 Pro Lys Val Tyr Thr Phe Ile Gly Gln Gly Asp Ala Ser Tyr Thr Ile
 485 490 495
 Ser Phe Lys Thr Gln Gly Pro Thr Val Asn Lys Leu Tyr Tyr Ala Ala
 500 505 510
 Gly Gly Arg Ala Leu Glu Tyr Asn Gln Leu Phe Met Tyr Ser Gln Leu
 515 520 525
 Tyr Val Glu Ser Thr Gln Asp His Gln Gln Arg Leu Asn Gly Leu Arg
 530 535 540
 Gln Val Val Asn Arg Thr Tyr Arg Ile Gly Thr Thr Lys Arg Val Glu
 545 550 555 560
 Val Ser Gln Gly Asn Val Gln Thr Lys Lys Val Leu Glu Ser Thr Asn
 565 570 575
 Leu Asn Ile Asp Asp Phe Val Asp Asp Pro Leu Ser Tyr Val Lys Thr
 580 585 590
 Pro Ser Asn Lys Val Leu Gly Phe Tyr Ser Asn Ala Asn Thr Asn
 595 600 605
 Ala Phe Arg Pro Gly Gly Ala Gln Gln Leu Asn Glu Tyr Gln Leu Ser
 610 615 620
 Gln Leu Phe Thr Asp Gln Lys Leu Gln Glu Ala Ala Arg Thr Arg Asn
 625 630 635 640
 Pro Ile Arg Leu Met Ile Gly Phe Asp Tyr Pro Asp Ala Tyr Gly Asn
 645 650 655
 Ser Glu Thr Leu Val Pro Val Asn Leu Thr Val Leu Pro Glu Ile Gln
 660 665 670
 His Asn Ile Lys Phe Phe Lys Asn Asp Asp Thr Gln Asn Ile Ala Glu
 675 680 685
 Lys Pro Phe Ser Lys Gln Ala Gly His Pro Val Phe Tyr Val Tyr Ala
 690 695 700
 Gly Asn Gln Gly Asn Ala Ser Val Asn Leu Gly Gly Ser Val Thr Ser
 705 710 715 720
 Ile Gln Pro Leu Arg Ile Asn Leu Thr Ser Asn Glu Asn Phe Thr Asp
 725 730 735
 Lys Asp Trp Gln Ile Thr Gly Ile Pro Arg Thr Leu His Ile Glu Asn
 740 745 750
 Ser Thr Asn Arg Pro Asn Asn Ala Arg Glu Arg Asn Ile Glu Leu Val
 755 760 765
 Gly Asn Leu Leu Pro Gly Asp Tyr Phe Gly Thr Ile Arg Phe Gly Arg
 770 775 780
 Lys Glu Gln Leu Phe Glu Ile Arg Val Lys Pro His Thr Pro Thr Ile
 785 790 795 800
 Thr Thr Thr Ala Glu Gln Leu Arg Gly Thr Ala Leu Gln Lys Val Pro
 805 810 815
 Val Asn Ile Ser Gly Ile Pro Leu Asp Pro Ser Ala Leu Val Tyr Leu
 820 825 830
 Val Ala Pro Thr Asn Gln Thr Thr Asn Gly Gly Ser Glu Ala Asp Gln
 835 840 845
 Ile Pro Ser Gly Tyr Thr Ile Leu Ala Thr Gly Thr Pro Asp Gly Val
 850 855 860
 His Asn Thr Ile Thr Ile Arg Pro Gln Asp Tyr Val Val Phe Ile Pro

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865		870		875		880
Pro Val Gly Lys Gln Ile Arg Ala Val Val Tyr Tyr Asn Lys Val Val						
		885		890		895
Ala Ser Asn Met Ser Asn Ala Val Thr Ile Leu Pro Asp Asp Ile Pro		900		905		910
Pro Thr Ile Asn Asn Pro Val Gly Ile Asn Ala Lys Tyr Tyr Arg Gly		915		920		925
Asp Glu Val Asn Phe Thr Met Gly Val Ser Asp Arg His Ser Gly Ile		930		935		940
Lys Asn Thr Thr Ile Thr Thr Leu Pro Asn Gly Trp Thr Ser Asn Leu		945		950		955
				955		960
Thr Lys Ala Asp Lys Asn Asn Gly Ser Leu Ser Ile Thr Gly Arg Val		965		970		975
Ser Met Asn Gln Ala Phe Asn Ser Asp Ile Thr Phe Lys Val Ser Ala		980		985		990
Thr Asp Asn Val Asn Asn Thr Thr Asn Asp Ser Gln Ser Lys His Val		995		1000		1005
Ser Ile His Val Gly Lys Ile Ser Glu Asp Ala His Pro Ile Val		1010		1015		1020
Leu Gly Asn Thr Glu Lys Val Val Val Val Asn Pro Thr Ala Val		1025		1030		1035
Ser Asn Asp Glu Lys Gln Ser Ile Ile Thr Ala Phe Met Asn Lys		1040		1045		1050
Asn Gln Asn Ile Arg Gly Tyr Leu Ala Ser Thr Asp Pro Val Thr		1055		1060		1065
Val Asp Asn Asn Gly Asn Val Thr Leu His Tyr Arg Asp Gly Ser		1070		1075		1080
Ser Thr Thr Leu Asp Ala Thr Asn Val Met Thr Tyr Glu Pro Val		1085		1090		1095
Val Lys Pro Glu Tyr Gln Thr Val Asn Ala Ala Lys Thr Ala Thr		1100		1105		1110
Val Thr Ile Ala Lys Gly Gln Ser Phe Ser Ile Gly Asp Ile Lys		1115		1120		1125
Gln Tyr Phe Thr Leu Ser Asn Gly Gln Pro Ile Pro Ser Gly Thr		1130		1135		1140
Phe Thr Asn Ile Thr Ser Asp Arg Thr Ile Pro Thr Ala Gln Glu		1145		1150		1155
Val Ser Gln Met Asn Ala Gly Thr Gln Leu Tyr His Ile Thr Ala		1160		1165		1170
Thr Asn Ala Tyr His Lys Asp Ser Glu Asp Phe Tyr Ile Ser Leu		1175		1180		1185
Lys Ile Ile Asp Val Lys Gln Pro Glu Gly Asp Gln Arg Val Tyr		1190		1195		1200
Arg Thr Ser Thr Tyr Asp Leu Thr Thr Asp Glu Ile Ser Lys Val		1205		1210		1215
Lys Gln Ala Phe Ile Asn Ala Asn Arg Asp Val Ile Thr Leu Ala		1220		1225		1230
Glu Gly Asp Ile Ser Val Thr Asn Thr Pro Asn Gly Ala Asn Val		1235		1240		1245
Ser Thr Ile Thr Val Asn Ile Asn Lys Gly Arg Leu Thr Lys Ser		1250		1255		1260
Phe Ala Ser Asn Leu Ala Asn Met Asn Phe Leu Arg Trp Val Asn		1265		1270		1275

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Phe	Pro	Gln	Asp	Tyr	Thr	Val	Thr	Trp	Thr	Asn	Ala	Lys	Ile	Ala
1280						1285					1290			
Asn	Arg	Pro	Thr	Asp	Gly	Gly	Leu	Ser	Trp	Ser	Asp	Asp	His	Lys
1295						1300					1305			
Ser	Leu	Ile	Tyr	Arg	Tyr	Asp	Ala	Thr	Leu	Gly	Thr	Gln	Ile	Thr
1310						1315					1320			
Thr	Asn	Asp	Ile	Leu	Thr	Met	Leu	Lys	Ala	Thr	Thr	Thr	Val	Pro
1325						1330					1335			
Gly	Leu	Arg	Asn	Asn	Ile	Thr	Gly	Asn	Glu	Lys	Ser	Gln	Ala	Glu
1340						1345					1350			
Ala	Gly	Gly	Arg	Pro	Asn	Phe	Arg	Thr	Thr	Gly	Tyr	Ser	Gln	Ser
1355						1360					1365			
Asn	Ala	Thr	Thr	Asp	Gly	Gln	Arg	Gln	Phe	Thr	Leu	Asn	Gly	Gln
1370						1375					1380			
Val	Ile	Gln	Val	Leu	Asp	Ile	Ile	Asn	Pro	Ser	Asn	Gly	Tyr	Gly
1385						1390					1395			
Gly	Gln	Pro	Val	Thr	Asn	Ser	Asn	Thr	Arg	Ala	Asn	His	Ser	Asn
1400						1405					1410			
Ser	Thr	Val	Val	Asn	Val	Asn	Glu	Pro	Ala	Ala	Asn	Gly	Ala	Gly
1415						1420					1425			
Ala	Phe	Thr	Ile	Asp	His	Val	Val	Lys	Ser	Asn	Ser	Thr	His	Asn
1430						1435					1440			
Ala	Ser	Asp	Ala	Val	Tyr	Lys	Ala	Gln	Leu	Tyr	Leu	Thr	Pro	Tyr
1445						1450					1455			
Gly	Pro	Lys	Gln	Tyr	Val	Glu	His	Leu	Asn	Gln	Asn	Thr	Gly	Asn
1460						1465					1470			
Thr	Thr	Asp	Ala	Ile	Asn	Ile	Tyr	Phe	Val	Pro	Ser	Asp	Leu	Val
1475						1480					1485			
Asn	Pro	Thr	Ile	Ser	Val	Gly	Asn	Tyr	Thr	Asn	His	Gln	Val	Phe
1490						1495					1500			
Ser	Gly	Glu	Thr	Phe	Thr	Asn	Thr	Ile	Thr	Ala	Asn	Asp	Asn	Phe
1505						1510					1515			
Gly	Val	Gln	Ser	Val	Thr	Val	Pro	Asn	Thr	Ser	Gln	Ile	Thr	Gly
1520						1525					1530			
Thr	Val	Asp	Asn	Asn	His	Gln	His	Val	Ser	Ala	Thr	Ala	Pro	Asn
1535						1540					1545			
Val	Thr	Ser	Ala	Thr	Asn	Lys	Thr	Ile	Asn	Leu	Leu	Ala	Thr	Asp
1550						1555					1560			
Thr	Ser	Gly	Asn	Thr	Ala	Thr	Thr	Ser	Phe	Asn	Val	Thr	Val	Lys
1565						1570					1575			
Pro	Leu	Arg	Asp	Lys	Tyr	Arg	Val	Gly	Thr	Ser	Ser	Thr	Ala	Ala
1580						1585					1590			
Asn	Pro	Val	Arg	Ile	Ala	Asn	Ile	Ser	Asn	Asn	Ala	Thr	Val	Ser
1595						1600					1605			
Gln	Ala	Asp	Gln	Thr	Thr	Ile	Ile	Asn	Ser	Leu	Thr	Phe	Thr	Glu
1610						1615					1620			
Thr	Val	Pro	Asn	Arg	Ser	Tyr	Ala	Arg	Ala	Ser	Ala	Asn	Glu	Ile
1625						1630					1635			
Thr	Ser	Lys	Thr	Val	Ser	Asn	Val	Ser	Arg	Thr	Gly	Asn	Asn	Ala
1640						1645					1650			
Asn	Val	Thr	Val	Thr	Val	Thr	Tyr	Gln	Asp	Gly	Thr	Thr	Ser	Thr
1655						1660					1665			

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Val Thr 1670	Val Pro	Val Lys	His 1675	Val Ile	Pro Glu	Ile 1680	Val Ala	His			
Ser His 1685	Tyr Thr	Val Gln	Gly 1690	Gln Asp	Phe Pro	Ala 1695	Gly Asn	Gly			
Ser Ser 1700	Ala Ser	Asp Tyr	Phe 1705	Lys Leu	Ser Asn	Gly 1710	Ser Asp	Ile			
Ala Asp 1715	Ala Thr	Ile Thr	Trp 1720	Val Ser	Gly Gln	Ala 1725	Pro Asn	Lys			
Asp Asn 1730	Thr Arg	Ile Gly	Glu 1735	Asp Ile	Thr Val	Thr 1740	Ala His	Ile			
Leu Ile 1745	Asp Gly	Glu Thr	Thr 1750	Pro Ile	Thr Lys	Thr 1755	Ala Thr	Tyr			
Lys Val 1760	Val Arg	Thr Val	Pro 1765	Lys His	Val Phe	Glu 1770	Thr Ala	Arg			
Gly Val 1775	Leu Tyr	Pro Gly	Val 1780	Ser Asp	Met Tyr	Asp 1785	Ala Lys	Gln			
Tyr Val 1790	Lys Pro	Val Asn	Asn 1795	Ser Trp	Ser Thr	Asn 1800	Ala Gln	His			
Met Asn 1805	Phe Gln	Phe Val	Gly 1810	Thr Tyr	Gly Pro	Asn 1815	Lys Asp	Val			
Val Gly 1820	Ile Ser	Thr Arg	Leu 1825	Ile Arg	Val Thr	Tyr 1830	Asp Asn	Arg			
Gln Thr 1835	Glu Asp	Leu Thr	Ile 1840	Leu Ser	Lys Val	Lys 1845	Pro Asp	Pro			
Pro Arg 1850	Ile Asp	Ala Asn	Ser 1855	Val Thr	Tyr Lys	Ala 1860	Gly Leu	Thr			
Asn Gln 1865	Glu Ile	Lys Val	Asn 1870	Asn Val	Leu Asn	Asn 1875	Ser Ser	Val			
Lys Leu 1880	Phe Lys	Ala Asp	Asn 1885	Thr Pro	Leu Asn	Val 1890	Thr Asn	Ile			
Thr His 1895	Gly Ser	Gly Phe	Ser 1900	Ser Val	Val Thr	Val 1905	Ser Asp	Ala			
Leu Pro 1910	Asn Gly	Gly Ile	Lys 1915	Ala Lys	Ser Ser	Ile 1920	Ser Met	Asn			
Asn Val 1925	Thr Tyr	Thr Thr	Gln 1930	Asp Glu	His Gly	Gln 1935	Val Val	Thr			
Val Thr 1940	Arg Asn	Glu Ser	Val 1945	Asp Ser	Asn Asp	Ser 1950	Ala Thr	Val			
Thr Val 1955	Thr Pro	Gln Leu	Gln 1960	Ala Thr	Thr Glu	Gly 1965	Ala Val	Phe			
Ile Lys 1970	Gly Gly	Asp Gly	Phe 1975	Asp Phe	Gly His	Val 1980	Glu Arg	Phe			
Ile Gln 1985	Asn Pro	Pro His	Gly 1990	Ala Thr	Val Ala	Trp 1995	His Asp	Ser			
Pro Asp 2000	Thr Trp	Lys Asn	Thr 2005	Val Gly	Asn Thr	His 2010	Lys Thr	Ala			
Val Val 2015	Thr Leu	Pro Asn	Gly 2020	Gln Gly	Thr Arg	Asn 2025	Val Glu	Val			
Pro Val 2030	Lys Val	Tyr Pro	Val 2035	Ala Asn	Ala Lys	Ala 2040	Pro Ser	Arg			
Asp Val 2045	Lys Gly	Gln Asn	Leu 2050	Thr Asn	Gly Thr	Asp 2055	Ala Met	Asn			
Tyr Ile	Thr Phe	Asp Pro	Asn	Thr Asn	Thr Asn	Gly	Ile Thr	Ala			

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2060	2065	2070
Ala Trp Ala Asn Arg Gln Gln Pro Asn Asn Gln Gln Ala Gly Val 2075 2080 2085		
Gln His Leu Asn Val Asp Val Thr Tyr Pro Gly Ile Ser Ala Ala 2090 2095 2100		
Lys Arg Val Pro Val Thr Val Asn Val Tyr Gln Phe Glu Phe Pro 2105 2110 2115		
Gln Thr Thr Tyr Thr Thr Thr Val Gly Gly Thr Leu Ala Ser Gly 2120 2125 2130		
Thr Gln Ala Ser Gly Tyr Ala His Met Gln Asn Ala Thr Gly Leu 2135 2140 2145		
Pro Thr Asp Gly Phe Thr Tyr Lys Trp Asn Arg Asp Thr Thr Gly 2150 2155 2160		
Thr Asn Asp Ala Asn Trp Ser Ala Met Asn Lys Pro Asn Val Ala 2165 2170 2175		
Lys Val Val Asn Ala Lys Tyr Asp Val Ile Tyr Asn Gly His Thr 2180 2185 2190		
Phe Ala Thr Ser Leu Pro Ala Lys Phe Val Val Lys Asp Val Gln 2195 2200 2205		
Pro Ala Lys Pro Thr Val Thr Glu Thr Ala Ala Gly Ala Ile Thr 2210 2215 2220		
Ile Ala Pro Gly Ala Asn Gln Thr Val Asn Thr His Ala Gly Asn 2225 2230 2235		
Val Thr Thr Tyr Ala Asp Lys Leu Val Ile Lys Arg Asn Gly Asn 2240 2245 2250		
Val Val Thr Thr Phe Thr Arg Arg Asn Asn Thr Ser Pro Trp Val 2255 2260 2265		
Lys Glu Ala Ser Ala Ala Thr Val Ala Gly Ile Ala Gly Thr Asn 2270 2275 2280		
Asn Gly Ile Thr Val Ala Ala Gly Thr Phe Asn Pro Ala Asp Thr 2285 2290 2295		
Ile Gln Val Val Ala Thr Gln Gly Ser Gly Glu Thr Val Ser Asp 2300 2305 2310		
Glu Gln Arg Ser Asp Asp Phe Thr Val Val Ala Pro Gln Pro Asn 2315 2320 2325		
Gln Ala Thr Thr Lys Ile Trp Gln Asn Gly His Ile Asp Ile Thr 2330 2335 2340		
Pro Asn Asn Pro Ser Gly His Leu Ile Asn Pro Thr Gln Ala Met 2345 2350 2355		
Asp Ile Ala Tyr Thr Glu Lys Val Gly Asn Gly Ala Glu His Ser 2360 2365 2370		
Lys Thr Ile Asn Val Val Arg Gly Gln Asn Asn Gln Trp Thr Ile 2375 2380 2385		
Ala Asn Lys Pro Asp Tyr Val Thr Leu Asp Ala Gln Thr Gly Lys 2390 2395 2400		
Val Thr Phe Asn Ala Asn Thr Ile Lys Pro Asn Ser Ser Ile Thr 2405 2410 2415		
Ile Thr Pro Lys Ala Gly Thr Gly His Ser Val Ser Ser Asn Pro 2420 2425 2430		
Ser Thr Leu Thr Ala Pro Ala Ala His Thr Val Asn Thr Thr Glu 2435 2440 2445		
Ile Val Lys Asp Tyr Gly Ser Asn Val Thr Ala Ala Glu Ile Asn 2450 2455 2460		

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Asn	Ala	Val	Gln	Val	Ala	Asn	Lys	Arg	Thr	Ala	Thr	Ile	Lys	Asn
2465						2470					2475			
Gly	Thr	Ala	Met	Pro	Thr	Asn	Leu	Ala	Gly	Gly	Ser	Thr	Thr	Thr
2480						2485					2490			
Ile	Pro	Val	Thr	Val	Thr	Tyr	Asn	Asp	Gly	Ser	Thr	Glu	Glu	Val
2495						2500					2505			
Gln	Glu	Ser	Ile	Phe	Thr	Lys	Ala	Asp	Lys	Arg	Glu	Leu	Ile	Thr
2510						2515					2520			
Ala	Lys	Asn	His	Leu	Asp	Asp	Pro	Val	Ser	Thr	Glu	Gly	Lys	Lys
2525						2530					2535			
Pro	Gly	Thr	Ile	Thr	Gln	Tyr	Asn	Asn	Ala	Met	His	Asn	Ala	Gln
2540						2545					2550			
Gln	Gln	Ile	Asn	Thr	Ala	Lys	Thr	Glu	Ala	Gln	Gln	Val	Ile	Asn
2555						2560					2565			
Asn	Glu	Arg	Ala	Thr	Pro	Gln	Gln	Val	Ser	Asp	Ala	Leu	Thr	Lys
2570						2575					2580			
Val	Arg	Ala	Ala	Gln	Thr	Lys	Ile	Asp	Gln	Ala	Lys	Ala	Leu	Leu
2585						2590					2595			
Gln	Asn	Lys	Glu	Asp	Asn	Ser	Gln	Leu	Val	Thr	Ser	Lys	Asn	Asn
2600						2605					2610			
Leu	Gln	Ser	Ser	Val	Asn	Gln	Val	Pro	Ser	Thr	Ala	Gly	Met	Thr
2615						2620					2625			
Gln	Gln	Ser	Ile	Asp	Asn	Tyr	Asn	Ala	Lys	Lys	Arg	Glu	Ala	Glu
2630						2635					2640			
Thr	Glu	Ile	Thr	Ala	Ala	Gln	Arg	Val	Ile	Asp	Asn	Gly	Asp	Ala
2645						2650					2655			
Thr	Ala	Gln	Gln	Ile	Ser	Asp	Glu	Lys	His	Arg	Val	Asp	Asn	Ala
2660						2665					2670			
Leu	Thr	Ala	Leu	Asn	Gln	Ala	Lys	His	Asp	Leu	Thr	Ala	Asp	Thr
2675						2680					2685			
His	Ala	Leu	Glu	Gln	Ala	Val	Gln	Gln	Leu	Asn	Arg	Thr	Gly	Thr
2690						2695					2700			
Thr	Thr	Gly	Lys	Lys	Pro	Ala	Ser	Ile	Thr	Ala	Tyr	Asn	Asn	Ser
2705						2710					2715			
Ile	Arg	Ala	Leu	Gln	Ser	Asp	Leu	Thr	Ser	Ala	Lys	Asn	Ser	Ala
2720						2725					2730			
Asn	Ala	Ile	Ile	Gln	Lys	Pro	Ile	Arg	Thr	Val	Gln	Glu	Val	Gln
2735						2740					2745			
Ser	Ala	Leu	Thr	Asn	Val	Asn	Arg	Val	Asn	Glu	Arg	Leu	Thr	Gln
2750						2755					2760			
Ala	Ile	Asn	Gln	Leu	Val	Pro	Leu	Ala	Asp	Asn	Ser	Ala	Leu	Lys
2765						2770					2775			
Thr	Ala	Lys	Thr	Lys	Leu	Asp	Glu	Glu	Ile	Asn	Lys	Ser	Val	Thr
2780						2785					2790			
Thr	Asp	Gly	Met	Thr	Gln	Ser	Ser	Ile	Gln	Ala	Tyr	Glu	Asn	Ala
2795						2800					2805			
Lys	Arg	Ala	Gly	Gln	Thr	Glu	Ser	Thr	Asn	Ala	Gln	Asn	Val	Ile
2810						2815					2820			
Asn	Asn	Gly	Asp	Ala	Thr	Asp	Gln	Gln	Ile	Ala	Ala	Glu	Lys	Thr
2825						2830					2835			
Lys	Val	Glu	Glu	Lys	Tyr	Asn	Ser	Leu	Lys	Gln	Ala	Ile	Ala	Gly
2840						2845					2850			

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Leu Thr 2855	Pro Asp	Leu Ala	Pro 2860	Leu Gln Thr Ala	Lys Thr Gln Leu 2865
Gln Asn 2870	Asp Ile Asp	Gln Pro 2875	Thr Ser Thr Thr	Gly Met Thr Ser 2880	
Ala Ser 2885	Ile Ala Ala Phe	Asn 2890	Glu Lys Leu Ser	Ala Ala Arg Thr 2895	
Lys Ile 2900	Gln Glu Ile Asp	Arg 2905	Val Leu Ala Ser	His Pro Asp Val 2910	
Ala Thr 2915	Ile Arg Gln Asn	Val 2920	Thr Ala Ala Asn	Ala Ala Lys Ser 2925	
Ala Leu 2930	Asp Gln Ala Arg	Asn 2935	Gly Leu Thr Val	Asp Lys Ala Pro 2940	
Leu Glu 2945	Asn Ala Lys Asn	Gln 2950	Leu Gln His Ser	Ile Asp Thr Gln 2955	
Thr Ser 2960	Thr Thr Gly Met	Thr 2965	Gln Asp Ser Ile	Asn Ala Tyr Asn 2970	
Ala Lys 2975	Leu Thr Ala Ala	Arg 2980	Asn Lys Ile Gln	Gln Ile Asn Gln 2985	
Val Leu 2990	Ala Gly Ser Pro	Thr 2995	Val Glu Gln Ile	Asn Thr Asn Thr 3000	
Ser Thr 3005	Ala Asn Gln Ala	Lys 3010	Ser Asp Leu Asp	His Ala Arg Gln 3015	
Ala Leu 3020	Thr Pro Asp Lys	Ala 3025	Pro Leu Gln Thr	Ala Lys Thr Gln 3030	
Leu Glu 3035	Gln Ser Ile Asn	Gln 3040	Pro Thr Asp Thr	Thr Gly Met Thr 3045	
Thr Ala 3050	Ser Leu Asn Ala	Tyr 3055	Asn Gln Lys Leu	Gln Ala Ala Arg 3060	
Gln Lys 3065	Leu Thr Glu Ile	Asn 3070	Gln Val Leu Asn	Gly Asn Pro Thr 3075	
Val Gln 3080	Asn Ile Asn Asp	Lys 3085	Val Thr Glu Ala	Asn Gln Ala Lys 3090	
Asp Gln 3095	Leu Asn Thr Ala	Arg 3100	Gln Gly Leu Thr	Leu Asp Arg Gln 3105	
Pro Ala 3110	Leu Thr Thr Leu	His 3115	Gly Ala Ser Asn	Leu Asn Gln Ala 3120	
Gln Gln 3125	Asn Asn Phe Thr	Gln 3130	Gln Ile Asn Ala	Ala Gln Asn His 3135	
Ala Ala 3140	Leu Glu Thr Ile	Lys 3145	Ser Asn Ile Thr	Ala Leu Asn Thr 3150	
Ala Met 3155	Thr Lys Leu Lys	Asp 3160	Ser Val Ala Asp	Asn Asn Thr Ile 3165	
Lys Ser 3170	Asp Gln Asn Tyr	Thr 3175	Asp Ala Thr Pro	Ala Asn Lys Gln 3180	
Ala Tyr 3185	Asp Asn Ala Val	Asn 3190	Ala Ala Lys Gly	Val Ile Gly Glu 3195	
Thr Thr 3200	Asn Pro Thr Met	Asp 3205	Val Asn Thr Val	Asn Gln Lys Ala 3210	
Ala Ser 3215	Val Lys Ser Thr	Lys 3220	Asp Ala Leu Asp	Gly Gln Gln Asn 3225	
Leu Gln 3230	Arg Ala Lys Thr	Glu 3235	Ala Thr Asn Ala	Ile Thr His Ala 3240	
Ser Asp	Leu Asn Gln Ala	Gln	Lys Asn Ala Leu	Thr Gln Gln Val	

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3245		3250		3255
Asn Ser Ala Gln Asn Val Gln Ala Val Asn Asp Ile Lys Gln Thr				
3260		3265		3270
Thr Gln Ser Leu Asn Thr Ala Met Thr Gly Leu Lys Arg Gly Val				
3275		3280		3285
Ala Asn His Asn Gln Val Val Gln Ser Asp Asn Tyr Val Asn Ala				
3290		3295		3300
Asp Thr Asn Lys Lys Asn Asp Tyr Asn Asn Ala Tyr Asn His Ala				
3305		3310		3315
Asn Asp Ile Ile Asn Gly Asn Ala Gln His Pro Val Ile Thr Pro				
3320		3325		3330
Ser Asp Val Asn Asn Ala Leu Ser Asn Val Thr Ser Lys Glu His				
3335		3340		3345
Ala Leu Asn Gly Glu Ala Lys Leu Asn Ala Ala Lys Gln Glu Ala				
3350		3355		3360
Asn Thr Ala Leu Gly His Leu Asn Asn Leu Asn Asn Ala Gln Arg				
3365		3370		3375
Gln Asn Leu Gln Ser Gln Ile Asn Gly Ala His Gln Ile Asp Ala				
3380		3385		3390
Val Asn Thr Ile Lys Gln Asn Ala Thr Asn Leu Asn Ser Ala Met				
3395		3400		3405
Gly Asn Leu Arg Gln Ala Val Ala Asp Lys Asp Gln Val Lys Arg				
3410		3415		3420
Thr Glu Asp Tyr Ala Asp Ala Asp Thr Ala Lys Gln Asn Ala Tyr				
3425		3430		3435
Asn Ser Ala Val Ser Ser Ala Glu Thr Ile Ile Asn Gln Thr Thr				
3440		3445		3450
Asn Pro Thr Met Ser Val Asp Asp Val Asn Arg Ala Thr Ser Ala				
3455		3460		3465
Val Thr Ser Asn Lys Asn Ala Leu Asn Gly Tyr Glu Lys Leu Ala				
3470		3475		3480
Gln Ser Lys Thr Asp Ala Ala Arg Ala Ile Asp Ala Leu Pro His				
3485		3490		3495
Leu Asn Asn Ala Gln Lys Ala Asp Val Lys Ser Lys Ile Asn Ala				
3500		3505		3510
Ala Ser Asn Ile Ala Gly Val Asn Thr Val Lys Gln Gln Gly Thr				
3515		3520		3525
Asp Leu Asn Thr Ala Met Gly Asn Leu Gln Gly Ala Ile Asn Asp				
3530		3535		3540
Glu Gln Thr Thr Leu Asn Ser Gln Asn Tyr Gln Asp Ala Thr Pro				
3545		3550		3555
Ser Lys Lys Thr Ala Tyr Thr Asn Ala Val Gln Ala Ala Lys Asp				
3560		3565		3570
Ile Leu Asn Lys Ser Asn Gly Gln Asn Lys Thr Lys Asp Gln Val				
3575		3580		3585
Thr Glu Ala Met Asn Gln Val Asn Ser Ala Lys Asn Asn Leu Asp				
3590		3595		3600
Gly Thr Arg Leu Leu Asp Gln Ala Lys Gln Thr Ala Lys Gln Gln				
3605		3610		3615
Leu Asn Asn Met Thr His Leu Thr Thr Ala Gln Lys Thr Asn Leu				
3620		3625		3630
Thr Asn Gln Ile Asn Ser Gly Thr Thr Val Ala Gly Val Gln Thr				
3635		3640		3645

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Val	Gln	Ser	Asn	Ala	Asn	Thr	Leu	Asp	Gln	Ala	Met	Asn	Thr	Leu
3650						3655					3660			
Arg	Gln	Ser	Ile	Ala	Asn	Lys	Asp	Ala	Thr	Lys	Ala	Ser	Glu	Asp
3665						3670					3675			
Tyr	Val	Asp	Ala	Asn	Asn	Asp	Lys	Gln	Thr	Ala	Tyr	Asn	Asn	Ala
3680						3685					3690			
Val	Ala	Ala	Ala	Glu	Thr	Ile	Ile	Asn	Ala	Asn	Ser	Asn	Pro	Glu
3695						3700					3705			
Met	Asn	Pro	Ser	Thr	Ile	Thr	Gln	Lys	Ala	Glu	Gln	Val	Asn	Ser
3710						3715					3720			
Ser	Lys	Thr	Ala	Leu	Asn	Gly	Asp	Glu	Asn	Leu	Ala	Ala	Ala	Lys
3725						3730					3735			
Gln	Asn	Ala	Lys	Thr	Tyr	Leu	Asn	Thr	Leu	Thr	Ser	Ile	Thr	Asp
3740						3745					3750			
Ala	Gln	Lys	Asn	Asn	Leu	Ile	Ser	Gln	Ile	Thr	Ser	Ala	Thr	Arg
3755						3760					3765			
Val	Ser	Gly	Val	Asp	Thr	Val	Lys	Gln	Asn	Ala	Gln	His	Leu	Asp
3770						3775					3780			
Gln	Ala	Met	Ala	Ser	Leu	Gln	Asn	Gly	Ile	Asn	Asn	Glu	Ser	Gln
3785						3790					3795			
Val	Lys	Ser	Ser	Glu	Lys	Tyr	Arg	Asp	Ala	Asp	Thr	Asn	Lys	Gln
3800						3805					3810			
Gln	Glu	Tyr	Asp	Asn	Ala	Ile	Thr	Ala	Ala	Lys	Ala	Ile	Leu	Asn
3815						3820					3825			
Lys	Ser	Thr	Gly	Pro	Asn	Thr	Ala	Gln	Asn	Ala	Val	Glu	Ala	Ala
3830						3835					3840			
Leu	Gln	Arg	Val	Asn	Asn	Ala	Lys	Asp	Ala	Leu	Asn	Gly	Asp	Ala
3845						3850					3855			
Lys	Leu	Ile	Ala	Ala	Gln	Asn	Ala	Ala	Lys	Gln	His	Leu	Gly	Thr
3860						3865					3870			
Leu	Thr	His	Ile	Thr	Thr	Ala	Gln	Arg	Asn	Asp	Leu	Thr	Asn	Gln
3875						3880					3885			
Ile	Ser	Gln	Ala	Thr	Asn	Leu	Ala	Gly	Val	Glu	Ser	Val	Lys	Gln
3890						3895					3900			
Asn	Ala	Asn	Ser	Leu	Asp	Gly	Ala	Met	Gly	Asn	Leu	Gln	Thr	Ala
3905						3910					3915			
Ile	Asn	Asp	Lys	Ser	Gly	Thr	Leu	Ala	Ser	Gln	Asn	Phe	Leu	Asp
3920						3925					3930			
Ala	Asp	Glu	Gln	Lys	Arg	Asn	Ala	Tyr	Asn	Gln	Ala	Val	Ser	Ala
3935						3940					3945			
Ala	Glu	Thr	Ile	Leu	Asn	Lys	Gln	Thr	Gly	Pro	Asn	Thr	Ala	Lys
3950						3955					3960			
Thr	Ala	Val	Glu	Gln	Ala	Leu	Asn	Asn	Val	Asn	Asn	Ala	Lys	His
3965						3970					3975			
Ala	Leu	Asn	Gly	Thr	Gln	Asn	Leu	Asn	Asn	Ala	Lys	Gln	Ala	Ala
3980						3985					3990			
Ile	Thr	Ala	Ile	Asn	Gly	Ala	Ser	Asp	Leu	Asn	Gln	Lys	Gln	Lys
3995						4000					4005			
Asp	Ala	Leu	Lys	Ala	Gln	Ala	Asn	Gly	Ala	Gln	Arg	Val	Ser	Asn
4010						4015					4020			
Ala	Gln	Asp	Val	Gln	His	Asn	Ala	Thr	Glu	Leu	Asn	Thr	Ala	Met
4025						4030					4035			

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Gly	Thr	Leu	Lys	His	Ala	Ile	Ala	Asp	Lys	Thr	Asn	Thr	Leu	Ala
4040						4045					4050			
Ser	Ser	Lys	Tyr	Val	Asn	Ala	Asp	Ser	Thr	Lys	Gln	Asn	Ala	Tyr
4055						4060					4065			
Thr	Thr	Lys	Val	Thr	Asn	Ala	Glu	His	Ile	Ile	Ser	Gly	Thr	Pro
4070						4075					4080			
Thr	Val	Val	Thr	Thr	Pro	Ser	Glu	Val	Thr	Ala	Ala	Ala	Asn	Gln
4085						4090					4095			
Val	Asn	Ser	Ala	Lys	Gln	Glu	Leu	Asn	Gly	Asp	Glu	Arg	Leu	Arg
4100						4105					4110			
Glu	Ala	Lys	Gln	Asn	Ala	Asn	Thr	Ala	Ile	Asp	Ala	Leu	Thr	Gln
4115						4120					4125			
Leu	Asn	Thr	Pro	Gln	Lys	Ala	Lys	Leu	Lys	Glu	Gln	Val	Gly	Gln
4130						4135					4140			
Ala	Asn	Arg	Leu	Glu	Asp	Val	Gln	Thr	Val	Gln	Thr	Asn	Gly	Gln
4145						4150					4155			
Ala	Leu	Asn	Asn	Ala	Met	Lys	Gly	Leu	Arg	Asp	Ser	Ile	Ala	Asn
4160						4165					4170			
Glu	Thr	Thr	Val	Lys	Thr	Ser	Gln	Asn	Tyr	Thr	Asp	Ala	Ser	Pro
4175						4180					4185			
Asn	Asn	Gln	Ser	Thr	Tyr	Asn	Ser	Ala	Val	Ser	Asn	Ala	Lys	Gly
4190						4195					4200			
Ile	Ile	Asn	Gln	Thr	Asn	Asn	Pro	Thr	Met	Asp	Thr	Ser	Ala	Ile
4205						4210					4215			
Thr	Gln	Ala	Thr	Thr	Gln	Val	Asn	Asn	Ala	Lys	Asn	Gly	Leu	Asn
4220						4225					4230			
Gly	Ala	Glu	Asn	Leu	Arg	Asn	Ala	Gln	Asn	Thr	Ala	Lys	Gln	Asn
4235						4240					4245			
Leu	Asn	Thr	Leu	Ser	His	Leu	Thr	Asn	Asn	Gln	Lys	Ser	Ala	Ile
4250						4255					4260			
Ser	Ser	Gln	Ile	Asp	Arg	Ala	Gly	His	Val	Ser	Glu	Val	Thr	Ala
4265						4270					4275			
Thr	Lys	Asn	Ala	Ala	Thr	Glu	Leu	Asn	Thr	Gln	Met	Gly	Asn	Leu
4280						4285					4290			
Glu	Gln	Ala	Ile	His	Asp	Gln	Asn	Thr	Val	Lys	Gln	Ser	Val	Lys
4295						4300					4305			
Phe	Thr	Asp	Ala	Asp	Lys	Ala	Lys	Arg	Asp	Ala	Tyr	Thr	Asn	Ala
4310						4315					4320			
Val	Ser	Arg	Ala	Glu	Ala	Ile	Leu	Asn	Lys	Thr	Gln	Gly	Ala	Asn
4325						4330					4335			
Thr	Ser	Lys	Gln	Asp	Val	Glu	Ala	Ala	Ile	Gln	Asn	Val	Ser	Ser
4340						4345					4350			
Ala	Lys	Asn	Ala	Leu	Asn	Gly	Asp	Gln	Asn	Val	Thr	Asn	Ala	Lys
4355						4360					4365			
Asn	Ala	Ala	Lys	Asn	Ala	Leu	Asn	Asn	Leu	Thr	Ser	Ile	Asn	Asn
4370						4375					4380			
Ala	Gln	Lys	Arg	Asp	Leu	Thr	Thr	Lys	Ile	Asp	Gln	Ala	Thr	Thr
4385						4390					4395			
Val	Ala	Gly	Val	Glu	Ala	Val	Ser	Asn	Thr	Ser	Thr	Gln	Leu	Asn
4400						4405					4410			
Thr	Ala	Met	Ala	Asn	Leu	Gln	Asn	Gly	Ile	Asn	Asp	Lys	Thr	Asn
4415						4420					4425			
Thr	Leu	Ala	Ser	Glu	Asn	Tyr	His	Asp	Ala	Asp	Ser	Asp	Lys	Lys

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4430	4435	4440
Thr Ala Tyr Thr Gln Ala Val	Thr Asn Ala Glu Asn Ile Leu Asn	
4445	4450	4455
Lys Asn Ser Gly Ser Asn Leu	Asp Lys Thr Ala Val Glu Asn Ala	
4460	4465	4470
Leu Ser Gln Val Ala Asn Ala	Lys Gly Ala Leu Asn Gly Asn His	
4475	4480	4485
Asn Leu Glu Gln Ala Lys Ser	Asn Ala Asn Thr Thr Ile Asn Gly	
4490	4495	4500
Leu Gln His Leu Thr Thr Ala	Gln Lys Asp Lys Leu Lys Gln Gln	
4505	4510	4515
Val Gln Gln Ala Gln Asn Val	Ala Gly Val Asp Thr Val Lys Ser	
4520	4525	4530
Ser Ala Asn Thr Leu Asn Gly	Ala Met Gly Thr Leu Arg Asn Ser	
4535	4540	4545
Ile Gln Asp Asn Thr Ala Thr	Lys Asn Gly Gln Asn Tyr Leu Asp	
4550	4555	4560
Ala Thr Glu Arg Asn Lys Thr	Asn Tyr Asn Asn Ala Val Asp Ser	
4565	4570	4575
Ala Asn Gly Val Ile Asn Ala	Thr Ser Asn Pro Asn Met Asp Ala	
4580	4585	4590
Asn Ala Ile Asn Gln Ile Ala	Thr Gln Val Thr Ser Thr Lys Asn	
4595	4600	4605
Ala Leu Asp Gly Thr His Asn	Leu Thr Gln Ala Lys Gln Thr Ala	
4610	4615	4620
Thr Asn Ala Ile Asp Gly Ala	Thr Asn Leu Asn Lys Ala Gln Lys	
4625	4630	4635
Asp Ala Leu Lys Ala Gln Val	Thr Ser Ala Gln Arg Val Ala Asn	
4640	4645	4650
Val Thr Ser Ile Gln Gln Thr	Ala Asn Glu Leu Asn Thr Ala Met	
4655	4660	4665
Gly Gln Leu Gln His Gly Ile	Asp Asp Glu Asn Ala Thr Lys Gln	
4670	4675	4680
Thr Gln Lys Tyr Arg Asp Ala	Glu Gln Ser Lys Lys Thr Ala Tyr	
4685	4690	4695
Asp Gln Ala Val Ala Ala Ala	Lys Ala Ile Leu Asn Lys Gln Thr	
4700	4705	4710
Gly Ser Asn Ser Asp Lys Ala	Ala Val Asp Arg Ala Leu Gln Gln	
4715	4720	4725
Val Thr Ser Thr Lys Asp Ala	Leu Asn Gly Asp Ala Lys Leu Ala	
4730	4735	4740
Glu Ala Lys Ala Ala Ala Lys	Gln Asn Leu Gly Thr Leu Asn His	
4745	4750	4755
Ile Thr Asn Ala Gln Arg Thr	Asp Leu Glu Gly Gln Ile Asn Gln	
4760	4765	4770
Ala Thr Thr Val Asp Gly Val	Asn Thr Val Lys Thr Asn Ala Asn	
4775	4780	4785
Thr Leu Asp Gly Ala Met Asn	Ser Leu Gln Gly Ser Ile Asn Asp	
4790	4795	4800
Lys Asp Ala Thr Leu Arg Asn	Gln Asn Tyr Leu Asp Ala Asp Glu	
4805	4810	4815
Ser Lys Arg Asn Ala Tyr Thr	Gln Ala Val Thr Ala Ala Glu Gly	
4820	4825	4830

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Ile	Leu	Asn	Lys	Gln	Thr	Gly	Gly	Asn	Thr	Ser	Lys	Ala	Asp	Val
4835						4840					4845			
Asp	Asn	Ala	Leu	Asn	Ala	Val	Thr	Arg	Ala	Lys	Ala	Ala	Leu	Asn
4850						4855					4860			
Gly	Ala	Asp	Asn	Leu	Arg	Asn	Ala	Lys	Thr	Ser	Ala	Thr	Asn	Thr
4865						4870					4875			
Ile	Asp	Gly	Leu	Pro	Asn	Leu	Thr	Gln	Leu	Gln	Lys	Asp	Asn	Leu
4880						4885					4890			
Lys	His	Gln	Val	Glu	Gln	Ala	Gln	Asn	Val	Ala	Gly	Val	Asn	Gly
4895						4900					4905			
Val	Lys	Asp	Lys	Gly	Asn	Thr	Leu	Asn	Thr	Ala	Met	Gly	Ala	Leu
4910						4915					4920			
Arg	Thr	Ser	Ile	Gln	Asn	Asp	Asn	Thr	Thr	Lys	Thr	Ser	Gln	Asn
4925						4930					4935			
Tyr	Leu	Asp	Ala	Ser	Asp	Ser	Asn	Lys	Asn	Asn	Tyr	Asn	Thr	Ala
4940						4945					4950			
Val	Asn	Asn	Ala	Asn	Gly	Val	Ile	Asn	Ala	Thr	Asn	Asn	Pro	Asn
4955						4960					4965			
Met	Asp	Ala	Asn	Ala	Ile	Asn	Gly	Met	Ala	Asn	Gln	Val	Asn	Thr
4970						4975					4980			
Thr	Lys	Ala	Ala	Leu	Asn	Gly	Ala	Gln	Asn	Leu	Ala	Gln	Ala	Lys
4985						4990					4995			
Thr	Asn	Ala	Thr	Asn	Thr	Ile	Asn	Asn	Ala	His	Asp	Leu	Asn	Gln
5000						5005					5010			
Lys	Gln	Lys	Asp	Ala	Leu	Lys	Thr	Gln	Val	Asn	Asn	Ala	Gln	Arg
5015						5020					5025			
Val	Ser	Asp	Ala	Asn	Asn	Val	Gln	His	Thr	Ala	Thr	Glu	Leu	Asn
5030						5035					5040			
Ser	Ala	Met	Thr	Ala	Leu	Lys	Ala	Ala	Ile	Ala	Asp	Lys	Glu	Arg
5045						5050					5055			
Thr	Lys	Ala	Ser	Gly	Asn	Tyr	Val	Asn	Ala	Asp	Gln	Glu	Lys	Arg
5060						5065					5070			
Gln	Ala	Tyr	Asp	Ser	Lys	Val	Thr	Asn	Ala	Glu	Asn	Ile	Ile	Ser
5075						5080					5085			
Gly	Thr	Pro	Asn	Ala	Thr	Leu	Thr	Val	Asn	Asp	Val	Asn	Ser	Ala
5090						5095					5100			
Ala	Ser	Gln	Val	Asn	Ala	Ala	Lys	Thr	Ala	Leu	Asn	Gly	Asp	Asn
5105						5110					5115			
Asn	Leu	Arg	Val	Ala	Lys	Glu	His	Ala	Asn	Asn	Thr	Ile	Asp	Gly
5120						5125					5130			
Leu	Ala	Gln	Leu	Asn	Asn	Ala	Gln	Lys	Ala	Lys	Leu	Lys	Glu	Gln
5135						5140					5145			
Val	Gln	Ser	Ala	Thr	Thr	Leu	Asp	Gly	Val	Gln	Thr	Val	Lys	Asn
5150						5155					5160			
Ser	Ser	Gln	Thr	Leu	Asn	Thr	Ala	Met	Lys	Gly	Leu	Arg	Asp	Ser
5165						5170					5175			
Ile	Ala	Asn	Glu	Ala	Thr	Ile	Lys	Ala	Gly	Gln	Asn	Tyr	Thr	Asp
5180						5185					5190			
Ala	Ser	Pro	Asn	Asn	Arg	Asn	Glu	Tyr	Asp	Ser	Ala	Val	Thr	Ala
5195						5200					5205			
Ala	Lys	Ala	Ile	Ile	Asn	Gln	Thr	Ser	Asn	Pro	Thr	Met	Glu	Pro
5210						5215					5220			

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Asn Thr 5225	Ile Thr Gln Val Thr 5230	Ser Gln Val Thr Thr 5235	Lys Glu Gln
Ala Leu 5240	Asn Gly Ala Arg Asn 5245	Leu Ala Gln Ala Lys 5250	Thr Thr Ala
Lys Asn 5255	Asn Leu Asn Asn Leu 5260	Thr Ser Ile Asn Asn 5265	Ala Gln Lys
Asp Ala 5270	Leu Thr Arg Ser Ile 5275	Asp Gly Ala Thr Thr 5280	Val Ala Gly
Val Asn 5285	Gln Glu Thr Ala Lys 5290	Ala Thr Glu Leu Asn 5295	Asn Ala Met
His Ser 5300	Leu Gln Asn Gly Ile 5305	Asn Asp Glu Thr Gln 5310	Thr Lys Gln
Thr Gln 5315	Lys Tyr Leu Asp Ala 5320	Glu Pro Ser Lys Lys 5325	Ser Ala Tyr
Asp Gln 5330	Ala Val Asn Ala Ala 5335	Lys Ala Ile Leu Thr 5340	Lys Ala Ser
Gly Gln 5345	Asn Val Asp Lys Ala 5350	Ala Val Glu Gln Ala 5355	Leu Gln Asn
Val Asn 5360	Ser Thr Lys Thr Ala 5365	Leu Asn Gly Asp Ala 5370	Lys Leu Asn
Glu Ala 5375	Lys Ala Ala Ala Lys 5380	Gln Thr Leu Gly Thr 5385	Leu Thr His
Ile Asn 5390	Asn Ala Gln Arg Thr 5395	Ala Leu Asp Asn Glu 5400	Ile Thr Gln
Ala Thr 5405	Asn Val Glu Gly Val 5410	Asn Thr Val Lys Ala 5415	Lys Ala Gln
Gln Leu 5420	Asp Gly Ala Met Gly 5425	Gln Leu Glu Thr Ser 5430	Ile Arg Asp
Lys Asp 5435	Thr Thr Leu Gln Ser 5440	Gln Asn Tyr Gln Asp 5445	Ala Asp Asp
Ala Lys 5450	Arg Thr Ala Tyr Ser 5455	Gln Ala Val Asn Ala 5460	Ala Ala Thr
Ile Leu 5465	Asn Lys Thr Ala Gly 5470	Gly Asn Thr Pro Lys 5475	Ala Asp Val
Glu Arg 5480	Ala Met Gln Ala Val 5485	Thr Gln Ala Asn Thr 5490	Ala Leu Asn
Gly Ile 5495	Gln Asn Leu Asp Arg 5500	Ala Lys Gln Ala Ala 5505	Asn Thr Ala
Ile Thr 5510	Asn Ala Ser Asp Leu 5515	Asn Thr Lys Gln Lys 5520	Glu Ala Leu
Lys Ala 5525	Gln Val Thr Ser Ala 5530	Gly Arg Val Ser Ala 5535	Ala Asn Gly
Val Glu 5540	His Thr Ala Thr Glu 5545	Leu Asn Thr Ala Met 5550	Thr Ala Leu
Lys Arg 5555	Ala Ile Ala Asp Lys 5560	Ala Glu Thr Lys Ala 5565	Ser Gly Asn
Tyr Val 5570	Asn Ala Asp Ala Asn 5575	Lys Arg Gln Ala Tyr 5580	Asp Glu Lys
Val Thr 5585	Ala Ala Glu Asn Ile 5590	Val Ser Gly Thr Pro 5595	Thr Pro Thr
Leu Thr 5600	Pro Ala Asp Val Thr 5605	Asn Ala Ala Thr Gln 5610	Val Thr Asn
Ala Lys	Thr Gln Leu Asn Gly	Asn His Asn Leu Glu	Val Ala Lys

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5615	5620	5625
Gln Asn Ala Asn Thr Ala Ile Asp Gly Leu Thr Ser Leu Asn Gly 5630 5635 5640		
Pro Gln Lys Ala Lys Leu Lys Glu Gln Val Gly Gln Ala Thr Thr 5645 5650 5655		
Leu Pro Asn Val Gln Thr Val Arg Asp Asn Ala Gln Thr Leu Asn 5660 5665 5670		
Thr Ala Met Lys Gly Leu Arg Asp Ser Ile Ala Asn Glu Ala Thr 5675 5680 5685		
Ile Lys Ala Gly Gln Asn Tyr Thr Asp Ala Ser Gln Asn Lys Gln 5690 5695 5700		
Thr Asp Tyr Asn Ser Ala Val Thr Ala Ala Lys Ala Ile Ile Gly 5705 5710 5715		
Gln Thr Thr Ser Pro Ser Met Asn Ala Gln Glu Ile Asn Gln Ala 5720 5725 5730		
Lys Asp Gln Val Thr Ala Lys Glu Gln Ala Leu Asn Gly Gln Glu 5735 5740 5745		
Asn Leu Arg Thr Ala Gln Thr Asn Ala Lys Gln His Leu Asn Gly 5750 5755 5760		
Leu Ser Asp Leu Thr Asp Ala Gln Lys Asp Ala Val Lys Arg Gln 5765 5770 5775		
Ile Glu Gly Ala Thr His Val Asn Glu Val Thr Gln Ala Gln Asn 5780 5785 5790		
Asn Ala Asp Ala Leu Asn Thr Ala Met Thr Asn Leu Lys Asn Gly 5795 5800 5805		
Ile Gln Asp Gln Asn Thr Ile Lys Gln Gly Val Asn Phe Thr Asp 5810 5815 5820		
Ala Asp Glu Ala Lys Arg Asn Ala Tyr Thr Asn Ala Val Thr Gln 5825 5830 5835		
Ala Glu Gln Ile Leu Asn Lys Ala Gln Gly Pro Asn Thr Ser Lys 5840 5845 5850		
Asp Gly Val Glu Thr Ala Leu Glu Asn Val Gln Arg Ala Lys Asn 5855 5860 5865		
Glu Leu Asn Gly Asn Gln Asn Val Ala Asn Ala Lys Thr Thr Ala 5870 5875 5880		
Lys Asn Ala Leu Asn Asn Leu Thr Ser Ile Asn Asn Ala Gln Lys 5885 5890 5895		
Glu Ala Leu Lys Ser Gln Ile Glu Gly Ala Thr Thr Val Ala Gly 5900 5905 5910		
Val Asn Gln Val Ser Thr Thr Ala Ser Glu Leu Asn Thr Ala Met 5915 5920 5925		
Ser Asn Leu Gln Asn Gly Ile Asn Asp Glu Ala Ala Thr Lys Ala 5930 5935 5940		
Ala Gln Lys Tyr Thr Asp Ala Asp Arg Glu Lys Gln Thr Ala Tyr 5945 5950 5955		
Asn Asp Ala Val Thr Ala Ala Lys Thr Leu Leu Asp Lys Thr Ala 5960 5965 5970		
Gly Ser Asn Asp Asn Lys Ala Ala Val Glu Gln Ala Leu Gln Arg 5975 5980 5985		
Val Asn Thr Ala Lys Thr Ala Leu Asn Gly Asp Glu Arg Leu Asn 5990 5995 6000		
Glu Ala Lys Asn Thr Ala Lys Gln Gln Val Ala Thr Met Ser His 6005 6010 6015		

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Glu	Gly	Ala	Thr	Thr	Val	Asn	Gly	Val	Asn	Gly	Val	Lys	Thr	Lys
6410						6415					6420			
Ala	Gln	Asp	Leu	Asp	Gly	Ala	Met	Gln	Arg	Leu	Gln	Ser	Ala	Ile
6425						6430					6435			
Ala	Asn	Lys	Asp	Gln	Thr	Lys	Ala	Ser	Glu	Asn	Tyr	Ile	Asp	Ala
6440						6445					6450			
Asp	Pro	Thr	Lys	Lys	Thr	Ala	Phe	Asp	Asn	Ala	Ile	Thr	Gln	Ala
6455						6460					6465			
Glu	Ser	Tyr	Leu	Asn	Lys	Asp	His	Gly	Ala	Asn	Lys	Asp	Lys	Gln
6470						6475					6480			
Ala	Val	Glu	Gln	Ala	Ile	Gln	Ser	Val	Thr	Ser	Thr	Glu	Asn	Ala
6485						6490					6495			
Leu	Asn	Gly	Asp	Ala	Asn	Leu	Gln	Arg	Ala	Lys	Thr	Glu	Ala	Ile
6500						6505					6510			
Gln	Ala	Ile	Asp	Asn	Leu	Thr	His	Leu	Asn	Thr	Pro	Gln	Lys	Thr
6515						6520					6525			
Ala	Leu	Lys	Gln	Gln	Val	Asn	Ala	Ala	Gln	Arg	Val	Ser	Gly	Val
6530						6535					6540			
Thr	Asp	Leu	Lys	Asn	Ser	Ala	Thr	Ser	Leu	Asn	Asn	Ala	Met	Asp
6545						6550					6555			
Gln	Leu	Lys	Gln	Ala	Ile	Ala	Asp	His	Asp	Thr	Ile	Val	Ala	Ser
6560						6565					6570			
Gly	Asn	Tyr	Thr	Asn	Ala	Ser	Pro	Asp	Lys	Gln	Gly	Ala	Tyr	Thr
6575						6580					6585			
Asp	Ala	Tyr	Asn	Ala	Ala	Lys	Asn	Ile	Val	Asn	Gly	Ser	Pro	Asn
6590						6595					6600			
Val	Ile	Thr	Asn	Ala	Ala	Asp	Val	Thr	Ala	Ala	Thr	Gln	Arg	Val
6605						6610					6615			
Asn	Asn	Ala	Glu	Thr	Gly	Leu	Asn	Gly	Asp	Thr	Asn	Leu	Ala	Thr
6620						6625					6630			
Ala	Lys	Gln	Gln	Ala	Lys	Asp	Ala	Leu	Arg	Gln	Met	Thr	His	Leu
6635						6640					6645			
Ser	Asp	Ala	Gln	Lys	Gln	Ser	Ile	Thr	Gly	Gln	Ile	Asp	Ser	Ala
6650						6655					6660			
Thr	Gln	Val	Thr	Gly	Val	Gln	Ser	Val	Lys	Asp	Asn	Ala	Thr	Asn
6665						6670					6675			
Leu	Asp	Asn	Ala	Met	Asn	Gln	Leu	Arg	Asn	Ser	Ile	Ala	Asn	Lys
6680						6685					6690			
Asp	Asp	Val	Lys	Ala	Ser	Gln	Pro	Tyr	Val	Asp	Ala	Asp	Arg	Asp
6695						6700					6705			
Lys	Gln	Asn	Ala	Tyr	Asn	Thr	Ala	Val	Thr	Asn	Ala	Glu	Asn	Ile
6710						6715					6720			
Ile	Asn	Ala	Thr	Ser	Gln	Pro	Thr	Leu	Asp	Pro	Ser	Ala	Val	Thr
6725						6730					6735			
Gln	Ala	Ala	Asn	Gln	Val	Ser	Thr	Asn	Lys	Thr	Ala	Leu	Asn	Gly
6740						6745					6750			
Ala	Gln	Asn	Leu	Ala	Asn	Lys	Lys	Gln	Glu	Thr	Thr	Ala	Asn	Ile
6755						6760					6765			
Asn	Gln	Leu	Ser	His	Leu	Asn	Asn	Ala	Gln	Lys	Gln	Asp	Leu	Asn
6770						6775					6780			
Thr	Gln	Val	Thr	Asn	Ala	Pro	Asn	Ile	Ser	Thr	Val	Asn	Gln	Val
6785						6790					6795			
Lys	Thr	Lys	Ala	Glu	Gln	Leu	Asp	Gln	Ala	Met	Glu	Arg	Leu	Ile

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6800	6805	6810
Asn Gly Ile Gln Asp Lys Asp Gln Val Lys Gln Ser Val Asn Phe 6815 6820 6825		
Thr Asp Ala Asp Pro Glu Lys Gln Thr Ala Tyr Asn Asn Ala Val 6830 6835 6840		
Thr Ala Ala Glu Asn Ile Ile Asn Gln Ala Asn Gly Thr Asn Ala 6845 6850 6855		
Asn Gln Ser Gln Val Glu Ala Ala Leu Ser Thr Val Thr Thr Thr 6860 6865 6870		
Lys Gln Ala Leu Asn Gly Asp Arg Lys Val Thr Asp Ala Lys Asn 6875 6880 6885		
Asn Ala Asn Gln Thr Leu Ser Thr Leu Asp Asn Leu Asn Asn Ala 6890 6895 6900		
Gln Lys Gly Ala Val Thr Gly Asn Ile Asn Gln Ala His Thr Val 6905 6910 6915		
Ala Glu Val Thr Gln Ala Ile Gln Thr Ala Gln Glu Leu Asn Thr 6920 6925 6930		
Ala Met Gly Asn Leu Lys Asn Ser Leu Asn Asp Lys Asp Thr Thr 6935 6940 6945		
Leu Gly Ser Gln Asn Phe Ala Asp Ala Asp Pro Glu Lys Lys Asn 6950 6955 6960		
Ala Tyr Asn Glu Ala Val His Asn Ala Glu Asn Ile Leu Asn Lys 6965 6970 6975		
Ser Thr Gly Thr Asn Val Pro Lys Asp Gln Val Glu Ala Ala Met 6980 6985 6990		
Asn Gln Val Asn Ala Thr Lys Ala Ala Leu Asn Gly Thr Gln Asn 6995 7000 7005		
Leu Glu Lys Ala Lys Gln His Ala Asn Thr Ala Ile Asp Gly Leu 7010 7015 7020		
Ser His Leu Thr Asn Ala Gln Lys Glu Ala Leu Lys Gln Leu Val 7025 7030 7035		
Gln Gln Ser Thr Thr Val Ala Glu Ala Gln Gly Asn Glu Gln Lys 7040 7045 7050		
Ala Asn Asn Val Asp Ala Ala Met Asp Lys Leu Arg Gln Ser Ile 7055 7060 7065		
Ala Asp Asn Ala Thr Thr Lys Gln Asn Gln Asn Tyr Thr Asp Ala 7070 7075 7080		
Ser Gln Asn Lys Lys Asp Ala Tyr Asn Asn Ala Val Thr Thr Ala 7085 7090 7095		
Gln Gly Ile Ile Asp Gln Thr Thr Ser Pro Thr Leu Asp Pro Thr 7100 7105 7110		
Val Ile Asn Gln Ala Ala Gly Gln Val Ser Thr Thr Lys Asn Ala 7115 7120 7125		
Leu Asn Gly Asn Glu Asn Leu Glu Ala Ala Lys Gln Gln Ala Ser 7130 7135 7140		
Gln Ser Leu Gly Ser Leu Asp Asn Leu Asn Asn Ala Gln Lys Gln 7145 7150 7155		
Thr Val Thr Asp Gln Ile Asn Gly Ala His Thr Val Asp Glu Ala 7160 7165 7170		
Asn Gln Ile Lys Gln Asn Ala Gln Asn Leu Asn Thr Ala Met Gly 7175 7180 7185		
Asn Leu Lys Gln Ala Ile Ala Asp Lys Asp Ala Thr Lys Ala Thr 7190 7195 7200		

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Val Asn Phe Thr Asp Ala Asp Gln Ala Lys Gln Gln Ala Tyr Asn 7205 7210 7215
Thr Ala Val Thr Asn Ala Glu Asn Ile Ser Lys Ala Asn Gly Asn 7220 7225 7230
Ala Thr Gln Ala Glu Val Glu Gln Ala Ile Lys Gln Val Asn Ala 7235 7240 7245
Ala Lys Gln Ala Leu Asn Gly Asn Ala Asn Val Gln His Ala Lys 7250 7255 7260
Asp Glu Ala Thr Ala Leu Ile Asn Ser Ser Asn Asp Leu Asn Gln 7265 7270 7275
Ala Gln Lys Asp Ala Leu Lys Gln Gln Val Gln Asn Ala Thr Thr 7280 7285 7290
Val Ala Gly Val Asn Asn Val Lys Gln Thr Ala Gln Glu Leu Asn 7295 7300 7305
Asn Ala Met Thr Gln Leu Lys Gln Gly Ile Ala Asp Lys Glu Gln 7310 7315 7320
Thr Lys Ala Asp Gly Asn Phe Val Asn Ala Asp Pro Asp Lys Gln 7325 7330 7335
Asn Ala Tyr Asn Gln Ala Val Ala Lys Ala Glu Ala Leu Ile Ser 7340 7345 7350
Ala Thr Pro Asp Val Val Val Thr Pro Ser Glu Ile Thr Ala Ala 7355 7360 7365
Leu Asn Lys Val Thr Gln Ala Lys Asn Asp Leu Asn Gly Asn Thr 7370 7375 7380
Asn Leu Ala Thr Ala Lys Gln Asn Val Gln His Ala Ile Asp Gln 7385 7390 7395
Leu Pro Asn Leu Asn Gln Ala Gln Arg Asp Glu Tyr Ser Lys Gln 7400 7405 7410
Ile Thr Gln Ala Thr Leu Val Pro Asn Val Asn Ala Ile Gln Gln 7415 7420 7425
Ala Ala Thr Thr Leu Asn Asp Ala Met Thr Gln Leu Lys Gln Gly 7430 7435 7440
Ile Ala Asn Lys Ala Gln Ile Lys Gly Ser Glu Asn Tyr His Asp 7445 7450 7455
Ala Asp Thr Asp Lys Gln Thr Ala Tyr Asp Asn Ala Val Thr Lys 7460 7465 7470
Ala Glu Glu Leu Leu Lys Gln Thr Thr Asn Pro Thr Met Asp Pro 7475 7480 7485
Asn Thr Ile Gln Gln Ala Leu Thr Lys Val Asn Asp Thr Asn Gln 7490 7495 7500
Ala Leu Asn Gly Asn Gln Lys Leu Ala Asp Ala Lys Gln Asp Ala 7505 7510 7515
Lys Thr Thr Leu Gly Thr Leu Asp His Leu Asn Asp Ala Gln Lys 7520 7525 7530
Gln Ala Leu Thr Thr Gln Val Glu Gln Ala Pro Asp Ile Ala Thr 7535 7540 7545
Val Asn Asn Val Lys Gln Asn Ala Gln Asn Leu Asn Asn Ala Met 7550 7555 7560
Thr Asn Leu Asn Asn Ala Leu Gln Asp Lys Thr Glu Thr Leu Asn 7565 7570 7575
Ser Ile Asn Phe Thr Asp Ala Asp Gln Ala Lys Lys Asp Ala Tyr 7580 7585 7590

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Thr	Asn	Ala	Val	Ser	His	Ala	Glu	Gly	Ile	Leu	Ser	Lys	Ala	Asn
7595						7600					7605			
Gly	Ser	Asn	Ala	Ser	Gln	Thr	Glu	Val	Glu	Gln	Ala	Met	Gln	Arg
7610						7615					7620			
Val	Asn	Glu	Ala	Lys	Gln	Ala	Leu	Asn	Gly	Asn	Asp	Asn	Val	Gln
7625						7630					7635			
Arg	Ala	Lys	Asp	Ala	Ala	Lys	Gln	Val	Ile	Thr	Asn	Ala	Asn	Asp
7640						7645					7650			
Leu	Asn	Gln	Ala	Gln	Lys	Asp	Ala	Leu	Lys	Gln	Gln	Val	Asp	Ala
7655						7660					7665			
Ala	Gln	Thr	Val	Ala	Asn	Val	Asn	Thr	Ile	Lys	Gln	Thr	Ala	Gln
7670						7675					7680			
Asp	Leu	Asn	Gln	Ala	Met	Thr	Gln	Leu	Lys	Gln	Gly	Ile	Ala	Asp
7685						7690					7695			
Lys	Asp	Gln	Thr	Lys	Ala	Asn	Gly	Asn	Phe	Val	Asn	Ala	Asp	Thr
7700						7705					7710			
Asp	Lys	Gln	Asn	Ala	Tyr	Asn	Asn	Ala	Val	Ala	His	Ala	Glu	Gln
7715						7720					7725			
Ile	Ile	Ser	Gly	Thr	Pro	Asn	Ala	Asn	Val	Asp	Pro	Gln	Gln	Val
7730						7735					7740			
Ala	Gln	Ala	Leu	Gln	Gln	Val	Asn	Gln	Ala	Lys	Gly	Asp	Leu	Asn
7745						7750					7755			
Gly	Asn	His	Asn	Leu	Gln	Val	Ala	Lys	Asp	Asn	Ala	Asn	Thr	Ala
7760						7765					7770			
Ile	Asp	Gln	Leu	Pro	Asn	Leu	Asn	Gln	Pro	Gln	Lys	Thr	Ala	Leu
7775						7780					7785			
Lys	Asp	Gln	Val	Ser	His	Ala	Glu	Leu	Val	Thr	Gly	Val	Asn	Ala
7790						7795					7800			
Ile	Lys	Gln	Asn	Ala	Asp	Ala	Leu	Asn	Asn	Ala	Met	Gly	Thr	Leu
7805						7810					7815			
Lys	Gln	Gln	Ile	Gln	Ala	Asn	Ser	Gln	Val	Pro	Gln	Ser	Val	Asp
7820						7825					7830			
Phe	Thr	Gln	Ala	Asp	Gln	Asp	Lys	Gln	Gln	Ala	Tyr	Asn	Asn	Ala
7835						7840					7845			
Ala	Asn	Gln	Ala	Gln	Gln	Ile	Ala	Asn	Gly	Ile	Pro	Thr	Pro	Val
7850						7855					7860			
Leu	Thr	Pro	Asp	Thr	Val	Thr	Gln	Ala	Val	Thr	Thr	Met	Asn	Gln
7865						7870					7875			
Ala	Lys	Asp	Ala	Leu	Asn	Gly	Asp	Glu	Lys	Leu	Ala	Gln	Ala	Lys
7880						7885					7890			
Gln	Glu	Ala	Leu	Ala	Asn	Leu	Asp	Thr	Leu	Arg	Asp	Leu	Asn	Gln
7895						7900					7905			
Pro	Gln	Arg	Asp	Ala	Leu	Arg	Asn	Gln	Ile	Asn	Gln	Ala	Gln	Ala
7910						7915					7920			
Leu	Ala	Thr	Val	Glu	Gln	Thr	Lys	Gln	Asn	Ala	Gln	Asn	Val	Asn
7925						7930					7935			
Thr	Ala	Met	Ser	Asn	Leu	Lys	Gln	Gly	Ile	Ala	Asn	Lys	Asp	Thr
7940						7945					7950			
Val	Lys	Ala	Ser	Glu	Asn	Tyr	His	Asp	Ala	Asp	Ala	Asp	Lys	Gln
7955						7960					7965			
Thr	Ala	Tyr	Thr	Asn	Ala	Val	Ser	Gln	Ala	Glu	Gly	Ile	Ile	Asn
7970						7975					7980			
Gln	Thr	Thr	Asn	Pro	Thr	Leu	Asn	Pro	Asp	Glu	Ile	Thr	Arg	Ala

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7985		7990		7995										
Leu	Thr	Gln	Val	Thr	Asp	Ala	Lys	Asn	Gly	Leu	Asn	Gly	Glu	Ala
8000						8005					8010			
Lys	Leu	Ala	Thr	Glu	Lys	Gln	Asn	Ala	Lys	Asp	Ala	Val	Ser	Gly
8015						8020					8025			
Met	Thr	His	Leu	Asn	Asp	Ala	Gln	Lys	Gln	Ala	Leu	Lys	Gly	Gln
8030						8035					8040			
Ile	Asp	Gln	Ser	Pro	Glu	Ile	Ala	Thr	Val	Asn	Gln	Val	Lys	Gln
8045						8050					8055			
Thr	Ala	Thr	Ser	Leu	Asp	Gln	Ala	Met	Asp	Gln	Leu	Ser	Gln	Ala
8060						8065					8070			
Ile	Asn	Asp	Lys	Ala	Gln	Thr	Leu	Ala	Asp	Gly	Asn	Tyr	Leu	Asn
8075						8080					8085			
Ala	Asp	Pro	Asp	Lys	Gln	Asn	Ala	Tyr	Lys	Gln	Ala	Val	Ala	Lys
8090						8095					8100			
Ala	Glu	Ala	Leu	Leu	Asn	Lys	Gln	Ser	Gly	Thr	Asn	Glu	Val	Gln
8105						8110					8115			
Ala	Gln	Val	Glu	Ser	Ile	Thr	Asn	Glu	Val	Asn	Ala	Ala	Lys	Gln
8120						8125					8130			
Ala	Leu	Asn	Gly	Asn	Asp	Asn	Leu	Ala	Asn	Ala	Lys	Gln	Gln	Ala
8135						8140					8145			
Lys	Gln	Gln	Leu	Ala	Asn	Leu	Thr	His	Leu	Asn	Asp	Ala	Gln	Lys
8150						8155					8160			
Gln	Ser	Phe	Glu	Ser	Gln	Ile	Thr	Gln	Ala	Pro	Leu	Val	Thr	Asp
8165						8170					8175			
Val	Thr	Thr	Ile	Asn	Gln	Lys	Ala	Gln	Thr	Leu	Asp	His	Ala	Met
8180						8185					8190			
Glu	Leu	Leu	Arg	Asn	Ser	Val	Ala	Asp	Asn	Gln	Thr	Thr	Leu	Ala
8195						8200					8205			
Ser	Glu	Asp	Tyr	His	Asp	Ala	Thr	Ala	Gln	Arg	Gln	Asn	Asp	Tyr
8210						8215					8220			
Asn	Gln	Ala	Val	Thr	Ala	Ala	Asn	Asn	Ile	Ile	Asn	Gln	Thr	Thr
8225						8230					8235			
Ser	Pro	Thr	Met	Asn	Pro	Asp	Asp	Val	Asn	Gly	Ala	Thr	Thr	Gln
8240						8245					8250			
Val	Asn	Asn	Thr	Lys	Val	Ala	Leu	Asp	Gly	Asp	Glu	Asn	Leu	Ala
8255						8260					8265			
Ala	Ala	Lys	Gln	Gln	Ala	Asn	Asn	Arg	Leu	Asp	Gln	Leu	Asp	His
8270						8275					8280			
Leu	Asn	Asn	Ala	Gln	Lys	Gln	Gln	Leu	Gln	Ser	Gln	Ile	Thr	Gln
8285						8290					8295			
Ser	Ser	Asp	Ile	Ala	Ala	Val	Asn	Gly	His	Lys	Gln	Thr	Ala	Glu
8300						8305					8310			
Ser	Leu	Asn	Thr	Ala	Met	Gly	Asn	Leu	Ile	Asn	Ala	Ile	Ala	Asp
8315						8320					8325			
His	Gln	Ala	Val	Glu	Gln	Arg	Gly	Asn	Phe	Ile	Asn	Ala	Asp	Thr
8330						8335					8340			
Asp	Lys	Gln	Thr	Ala	Tyr	Asn	Thr	Ala	Val	Asn	Glu	Ala	Ala	Ala
8345						8350					8355			
Met	Ile	Asn	Lys	Gln	Thr	Gly	Gln	Asn	Ala	Asn	Gln	Thr	Glu	Val
8360						8365					8370			
Glu	Gln	Ala	Ile	Thr	Lys	Val	Gln	Thr	Thr	Leu	Gln	Ala	Leu	Asn
8375						8380					8385			

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Gly Asp	His Asn	Leu Gln	Val Ala	Lys Thr	Asn Ala	Thr Gln	Ala Ala	8390	8400
Ile Asp	Ala Leu	Thr Ser	Leu Asn	Asp Pro	Gln Lys	Thr Ala	Leu Leu	8405	8415
Lys Asp	Gln Val	Thr Ala	Ala Ala	Thr Leu	Val Thr	Ala Val	His Gln	8420	8430
Ile Glu	Gln Asn	Ala Asn	Thr Leu	Asn Gln	Ala Met	His Gly	Leu Leu	8435	8445
Arg Gln	Ser Ile	Gln Asp	Asn Ala	Ala Thr	Lys Ala	Asn Ser	Lys Lys	8450	8460
Tyr Ile	Asn Glu	Asp Gln	Pro Glu	Gln Gln	Asn Tyr	Asp Gln	Ala Ala	8465	8475
Val Gln	Ala Ala	Asn Asn	Ile Ile	Asn Glu	Gln Thr	Ala Thr	Leu Leu	8480	8490
Asp Asn	Asn Ala	Ile Asn	Gln Ala	Ala Thr	Thr Thr	Val Asn	Thr Thr	8495	8505
Lys Ala	Ala Leu	His Gly	Asp Val	Lys Leu	Gln Asn	Asp Lys	Asp Asp	8510	8520
His Ala	Lys Gln	Thr Val	Ser Gln	Leu Ala	His Leu	Asn Asn	Ala Ala	8525	8535
Gln Lys	His Met	Glu Asp	Thr Leu	Ile Asp	Ser Glu	Thr Thr	Arg Arg	8540	8550
Thr Ala	Val Lys	Gln Asp	Leu Thr	Glu Ala	Gln Ala	Leu Asp	Gln Gln	8555	8565
Leu Met	Asp Ala	Leu Gln	Gln Ser	Ile Ala	Asp Lys	Asp Ala	Thr Thr	8570	8580
Arg Ala	Ser Ser	Ala Tyr	Val Asn	Ala Glu	Pro Asn	Lys Lys	Gln Gln	8585	8595
Ser Tyr	Asp Glu	Ala Val	Gln Asn	Ala Glu	Ser Ile	Ile Ala	Gly Gly	8600	8610
Leu Asn	Asn Pro	Thr Ile	Asn Lys	Gly Asn	Val Ser	Ser Ala	Thr Thr	8615	8625
Gln Ala	Val Ile	Ser Ser	Lys Asn	Ala Leu	Asp Gly	Val Glu	Arg Arg	8630	8640
Leu Ala	Gln Asp	Lys Gln	Thr Ala	Gly Asn	Ser Leu	Asn His	Leu Leu	8645	8655
Asp Gln	Leu Thr	Pro Ala	Gln Gln	Gln Ala	Leu Glu	Asn Gln	Ile Ile	8660	8670
Asn Asn	Ala Thr	Thr Arg	Gly Glu	Val Ala	Gln Lys	Leu Thr	Glu Glu	8675	8685
Ala Gln	Ala Leu	Asn Gln	Ala Met	Glu Ala	Leu Arg	Asn Ser	Ile Ile	8690	8700
Gln Asp	Gln Gln	Gln Thr	Glu Ala	Gly Ser	Lys Phe	Ile Asn	Glu Glu	8705	8715
Asp Lys	Pro Gln	Lys Asp	Ala Tyr	Gln Ala	Ala Val	Gln Asn	Ala Ala	8720	8730
Lys Asp	Leu Ile	Asn Gln	Thr Asn	Asn Pro	Thr Leu	Asp Lys	Ala Ala	8735	8745
Gln Val	Glu Gln	Leu Thr	Gln Ala	Val Asn	Gln Ala	Lys Asp	Asn Asn	8750	8760
Leu His	Gly Asp	Gln Lys	Leu Ala	Asp Asp	Lys Gln	His Ala	Val Val	8765	8775

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Thr Asp 8780	Leu Asn	Gln Leu	Asn	Gly Leu	Asn Asn	Pro	Gln Arg	Gln		
Ala Leu 8795	Glu Ser	Gln Ile	Asn	Asn Ala	Ala Thr	Arg	Gly Glu	Val		
Ala Gln 8810	Lys Leu	Ala Glu	Ala	Lys Ala	Leu Asp	Gln	Ala Met	Gln		
Ala Leu 8825	Arg Asn	Ser Ile	Gln	Asp Gln	Gln Gln	Thr	Glu Ser	Gly		
Ser Lys 8840	Phe Ile	Asn Glu	Asp	Lys Pro	Gln Lys	Asp	Ala Tyr	Gln		
Ala Ala 8855	Val Gln	Asn Ala	Lys	Asp Leu	Ile Asn	Gln	Thr Gly	Asn		
Pro Thr 8870	Leu Asp	Lys Ser	Gln	Val Glu	Gln Leu	Thr	Gln Ala	Val		
Thr Thr 8885	Ala Lys	Asp Asn	Leu	His Gly	Asp Gln	Lys	Leu Ala	Arg		
Asp Gln 8900	Gln Gln	Ala Val	Thr	Thr Val	Asn Ala	Leu	Pro Asn	Leu		
Asn His 8915	Ala Gln	Gln Gln	Ala	Leu Thr	Asp Ala	Ile	Asn Ala	Ala		
Pro Thr 8930	Arg Thr	Glu Val	Ala	Gln His	Val Gln	Thr	Ala Thr	Glu		
Leu Asp 8945	His Ala	Met Glu	Thr	Leu Lys	Asn Lys	Val	Asp Gln	Val		
Asn Thr 8960	Asp Lys	Ala Gln	Pro	Asn Tyr	Thr Glu	Ala	Ser Thr	Asp		
Lys Lys 8975	Glu Ala	Val Asp	Gln	Ala Leu	Gln Ala	Ala	Glu Ser	Ile		
Thr Asp 8990	Pro Thr	Asn Gly	Ser	Asn Ala	Asn Lys	Asp	Ala Val	Asp		
Gln Val 9005	Leu Thr	Lys Leu	Gln	Glu Lys	Glu Asn	Glu	Leu Asn	Gly		
Asn Glu 9020	Arg Val	Ala Glu	Ala	Lys Thr	Gln Ala	Lys	Gln Thr	Ile		
Asp Gln 9035	Leu Thr	His Leu	Asn	Ala Asp	Gln Ile	Ala	Thr Ala	Lys		
Gln Asn 9050	Ile Asp	Gln Ala	Thr	Lys Leu	Gln Pro	Ile	Ala Glu	Leu		
Val Asp 9065	Gln Ala	Thr Gln	Leu	Asn Gln	Ser Met	Asp	Gln Leu	Gln		
Gln Ala 9080	Val Asn	Glu His	Ala	Asn Val	Glu Gln	Thr	Val Asp	Tyr		
Thr Gln 9095	Ala Asp	Ser Asp	Lys	Gln Asn	Ala Tyr	Lys	Gln Ala	Ile		
Ala Asp 9110	Ala Glu	Asn Val	Leu	Lys Gln	Asn Ala	Asn	Lys Gln	Gln		
Val Asp 9125	Gln Ala	Leu Gln	Asn	Ile Leu	Asn Ala	Lys	Gln Ala	Leu		
Asn Gly 9140	Asp Glu	Arg Val	Ala	Leu Ala	Lys Thr	Asn	Gly Lys	His		
Asp Ile 9155	Asp Gln	Leu Asn	Ala	Leu Asn	Asn Ala	Gln	Gln Asp	Gly		
Phe Lys	Gly Arg	Ile Asp	Gln	Ser Asn	Asp Leu	Asn	Gln Ile	Gln		

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9170	9175	9180
Gln Ile Val Asp Glu Ala Lys 9185	Ala Leu Asn Arg Ala Met Asp Gln 9190	9195
Leu Ser Gln Glu Ile Thr Asp 9200	Asn Glu Gly Arg Thr Lys Gly Ser 9205	9210
Thr Asn Tyr Val Asn Ala Asp 9215	Thr Gln Val Lys Gln Val Tyr Asp 9220	9225
Glu Thr Val Asp Lys Ala Lys 9230	Gln Ala Leu Asp Lys Ser Thr Gly 9235	9240
Gln Asn Leu Thr Ala Lys Gln 9245	Val Ile Lys Leu Asn Asp Ala Val 9250	9255
Thr Ala Ala Lys Lys Ala Leu 9260	Asn Gly Glu Glu Arg Leu Asn Asn 9265	9270
Arg Lys Ala Glu Ala Leu Gln 9275	Arg Leu Asp Gln Leu Thr His Leu 9280	9285
Asn Asn Ala Gln Arg Gln Leu 9290	Ala Ile Gln Gln Ile Asn Asn Ala 9295	9300
Glu Thr Leu Asn Lys Ala Ser 9305	Arg Ala Ile Asn Arg Ala Thr Lys 9310	9315
Leu Asp Asn Ala Met Gly Ala 9320	Val Gln Gln Tyr Ile Asp Glu Gln 9325	9330
His Leu Gly Val Ile Ser Ser 9335	Thr Asn Tyr Ile Asn Ala Asp Asp 9340	9345
Asn Leu Lys Ala Asn Tyr Asp 9350	Asn Ala Ile Ala Asn Ala Ala His 9355	9360
Glu Leu Asp Lys Val Gln Gly 9365	Asn Ala Ile Ala Lys Ala Glu Ala 9370	9375
Glu Gln Leu Lys Gln Asn Ile 9380	Ile Asp Ala Gln Asn Ala Leu Asn 9385	9390
Gly Asp Gln Asn Leu Ala Asn 9395	Ala Lys Asp Lys Ala Asn Ala Phe 9400	9405
Val Asn Ser Leu Asn Gly Leu 9410	Asn Gln Gln Gln Gln Asp Leu Ala 9415	9420
His Lys Ala Ile Asn Asn Ala 9425	Asp Thr Val Ser Asp Val Thr Asp 9430	9435
Ile Val Asn Asn Gln Ile Asp 9440	Leu Asn Asp Ala Met Glu Thr Leu 9445	9450
Lys His Leu Val Asp Asn Glu 9455	Ile Pro Asn Ala Glu Gln Thr Val 9460	9465
Asn Tyr Gln Asn Ala Asp Asp 9470	Asn Ala Lys Thr Asn Phe Asp Asp 9475	9480
Ala Lys Arg Leu Ala Asn Thr 9485	Leu Leu Asn Ser Asp Asn Thr Asn 9490	9495
Val Asn Asp Ile Asn Gly Ala 9500	Ile Gln Ala Val Asn Asp Ala Ile 9505	9510
His Asn Leu Asn Gly Asp Gln 9515	Arg Leu Gln Asp Ala Lys Asp Lys 9520	9525
Ala Ile Gln Ser Ile Asn Gln 9530	Ala Leu Ala Asn Lys Leu Lys Glu 9535	9540
Ile Glu Ala Ser Asn Ala Thr 9545	Asp Gln Asp Lys Leu Ile Ala Lys 9550	9555
Asn Lys Ala Glu Glu Leu Ala 9560	Asn Ser Ile Ile Asn Asn Ile Asn 9565	9570

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Lys	Ala	Thr	Ser	Asn	Gln	Ala	Val	Ser	Gln	Val	Gln	Thr	Ala	Gly
9575						9580					9585			
Asn	His	Ala	Ile	Glu	Gln	Val	His	Ala	Asn	Glu	Ile	Pro	Lys	Ala
9590						9595					9600			
Lys	Ile	Asp	Ala	Asn	Lys	Asp	Val	Asp	Lys	Gln	Val	Gln	Ala	Leu
9605						9610					9615			
Ile	Asp	Glu	Ile	Asp	Arg	Asn	Pro	Asn	Leu	Thr	Asp	Lys	Glu	Lys
9620						9625					9630			
Gln	Ala	Leu	Lys	Asp	Arg	Ile	Asn	Gln	Ile	Leu	Gln	Gln	Gly	His
9635						9640					9645			
Asn	Gly	Ile	Asn	Asn	Ala	Met	Thr	Lys	Glu	Glu	Ile	Glu	Gln	Ala
9650						9655					9660			
Lys	Ala	Gln	Leu	Ala	Gln	Ala	Leu	Gln	Asp	Ile	Lys	Asp	Leu	Val
9665						9670					9675			
Lys	Ala	Lys	Glu	Asp	Ala	Lys	Gln	Asp	Val	Asp	Lys	Gln	Val	Gln
9680						9685					9690			
Ala	Leu	Ile	Asp	Glu	Ile	Asp	Gln	Asn	Pro	Asn	Leu	Thr	Asp	Lys
9695						9700					9705			
Glu	Lys	Gln	Ala	Leu	Lys	Tyr	Arg	Ile	Asn	Gln	Ile	Leu	Gln	Gln
9710						9715					9720			
Gly	His	Asn	Asp	Ile	Asn	Asn	Ala	Leu	Thr	Lys	Glu	Glu	Ile	Glu
9725						9730					9735			
Gln	Ala	Lys	Ala	Gln	Leu	Ala	Gln	Ala	Leu	Gln	Asp	Ile	Lys	Asp
9740						9745					9750			
Leu	Val	Lys	Ala	Lys	Glu	Asp	Ala	Lys	Asn	Ala	Ile	Lys	Ala	Leu
9755						9760					9765			
Ala	Asn	Ala	Lys	Arg	Asp	Gln	Ile	Asn	Ser	Asn	Pro	Asp	Leu	Thr
9770						9775					9780			
Pro	Glu	Gln	Lys	Ala	Lys	Ala	Leu	Lys	Glu	Ile	Asp	Glu	Ala	Glu
9785						9790					9795			
Lys	Arg	Ala	Leu	Gln	Asn	Val	Glu	Asn	Ala	Gln	Thr	Ile	Asp	Gln
9800						9805					9810			
Leu	Asn	Arg	Gly	Leu	Asn	Leu	Gly	Leu	Asp	Asp	Ile	Arg	Asn	Thr
9815						9820					9825			
His	Val	Trp	Glu	Val	Asp	Glu	Gln	Pro	Ala	Val	Asn	Glu	Ile	Phe
9830						9835					9840			
Glu	Ala	Thr	Pro	Glu	Gln	Ile	Leu	Val	Asn	Gly	Glu	Leu	Ile	Val
9845						9850					9855			
His	Arg	Asp	Asp	Ile	Ile	Thr	Glu	Gln	Asp	Ile	Leu	Ala	His	Ile
9860						9865					9870			
Asn	Leu	Ile	Asp	Gln	Leu	Ser	Ala	Glu	Val	Ile	Asp	Thr	Pro	Ser
9875						9880					9885			
Thr	Ala	Thr	Ile	Ser	Asp	Ser	Leu	Thr	Ala	Lys	Val	Glu	Val	Thr
9890						9895					9900			
Leu	Leu	Asp	Gly	Ser	Lys	Val	Ile	Val	Asn	Val	Pro	Val	Lys	Val
9905						9910					9915			
Val	Glu	Lys	Glu	Leu	Ser	Val	Val	Lys	Gln	Gln	Ala	Ile	Glu	Ser
9920						9925					9930			
Ile	Glu	Asn	Ala	Ala	Gln	Gln	Lys	Ile	Asn	Glu	Ile	Asn	Asn	Ser
9935						9940					9945			
Val	Thr	Leu	Thr	Leu	Glu	Gln	Lys	Glu	Ala	Ala	Ile	Ala	Glu	Val
9950						9955					9960			

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10355		10360		10365
Ile Glu Ser Lys Asp Glu Asp Val Pro Leu Leu Leu Ala Lys Lys				
10370		10375		10380
Lys Asn Gln Lys Asp Asn Gln Ser Lys Asp Lys Lys Ser Ala Ser				
10385		10390		10395
Lys Asn Thr Ser Lys Lys Val Ala Ala Lys Lys Lys Lys Lys Lys				
10400		10405		10410
Ala Lys Lys Asn Lys Lys				
10415				

<210> SEQ ID NO 25

<211> LENGTH: 340

<212> TYPE: PRT

<213> ORGANISM: Staphylococcus sp.

<400> SEQUENCE: 25

Met Lys Lys Lys Leu Leu Val Leu Thr Met Ser Thr Leu Phe Ala Thr				
1	5	10	15	
Gln Ile Met Asn Ser Asn His Ala Lys Ala Ser Val Thr Glu Ser Val				
20	25	30		
Asp Lys Lys Phe Val Val Pro Glu Ser Gly Ile Asn Lys Ile Ile Pro				
35	40	45		
Ala Tyr Asp Glu Phe Lys Asn Ser Pro Lys Val Asn Val Ser Asn Leu				
50	55	60		
Thr Asp Asn Lys Asn Phe Val Ala Ser Glu Asp Lys Leu Asn Lys Ile				
65	70	75	80	
Ala Asp Ser Ser Ala Ala Ser Lys Ile Val Asp Lys Asn Phe Val Val				
85	90	95		
Pro Glu Ser Lys Leu Gly Asn Ile Val Pro Glu Tyr Lys Glu Ile Asn				
100	105	110		
Asn Arg Val Asn Val Ala Thr Asn Asn Pro Ala Ser Gln Gln Val Asp				
115	120	125		
Lys His Phe Val Ala Lys Gly Pro Glu Val Asn Arg Phe Ile Thr Gln				
130	135	140		
Asn Lys Val Asn His His Phe Ile Thr Thr Gln Thr His Tyr Lys Lys				
145	150	155	160	
Val Ile Thr Ser Tyr Lys Ser Thr His Val His Lys His Val Asn His				
165	170	175		
Ala Lys Asp Ser Ile Asn Lys His Phe Ile Val Lys Pro Ser Glu Ser				
180	185	190		
Pro Arg Tyr Thr His Pro Ser Gln Ser Leu Ile Ile Lys His His Phe				
195	200	205		
Ala Val Pro Gly Tyr His Ala His Lys Phe Val Thr Pro Gly His Ala				
210	215	220		
Ser Ile Lys Ile Asn His Phe Cys Val Val Pro Gln Ile Asn Ser Phe				
225	230	235	240	
Lys Val Ile Pro Pro Tyr Gly His Asn Ser His Arg Met His Val Pro				
245	250	255		
Ser Phe Gln Asn Asn Thr Thr Ala Thr His Gln Asn Ala Lys Val Asn				
260	265	270		
Lys Ala Tyr Asp Tyr Lys Tyr Phe Tyr Ser Tyr Lys Val Val Lys Gly				
275	280	285		
Val Lys Lys Tyr Phe Ser Phe Ser Gln Ser Asn Gly Tyr Lys Ile Gly				
290	295	300		

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Lys Pro Ser Leu Asn Ile Lys Asn Val Asn Tyr Gln Tyr Ala Val Pro
305 310 315 320

Ser Tyr Ser Pro Thr His Tyr Val Pro Glu Phe Lys Gly Ser Leu Pro
325 330 335

Ala Pro Arg Val
340

<210> SEQ ID NO 26
<211> LENGTH: 130
<212> TYPE: PRT
<213> ORGANISM: Staphylococcus sp.

<400> SEQUENCE: 26

Met Asn Phe Asn Asp Ile Glu Thr Met Val Lys Ser Lys Phe Lys Asp
1 5 10 15

Ile Lys Lys His Ala Glu Glu Ile Ala His Glu Ile Glu Val Arg Ser
20 25 30

Gly Tyr Leu Arg Lys Ala Glu Gln Tyr Lys Arg Leu Glu Phe Asn Leu
35 40 45

Ser Phe Ala Leu Asp Asp Ile Glu Ser Thr Ala Lys Asp Val Gln Thr
50 55 60

Ala Lys Ser Ser Ala Asn Lys Asp Ser Val Thr Val Lys Gly Lys Ala
65 70 75 80

Pro Asn Thr Leu Tyr Ile Glu Lys Arg Asn Leu Met Lys Gln Lys Leu
85 90 95

Glu Met Leu Gly Glu Asp Ile Asp Lys Asn Lys Glu Ser Leu Gln Lys
100 105 110

Ala Lys Glu Ile Ala Gly Glu Lys Ala Ser Glu Tyr Phe Asn Lys Ala
115 120 125

Met Asn
130

<210> SEQ ID NO 27
<211> LENGTH: 636
<212> TYPE: PRT
<213> ORGANISM: Staphylococcus sp.

<400> SEQUENCE: 27

Met Lys Lys Gln Ile Ile Ser Leu Gly Ala Leu Ala Val Ala Ser Ser
1 5 10 15

Leu Phe Thr Trp Asp Asn Lys Ala Asp Ala Ile Val Thr Lys Asp Tyr
20 25 30

Ser Gly Lys Ser Gln Val Asn Ala Gly Ser Lys Asn Gly Thr Leu Ile
35 40 45

Asp Ser Arg Tyr Leu Asn Ser Ala Leu Tyr Tyr Leu Glu Asp Tyr Ile
50 55 60

Ile Tyr Ala Ile Gly Leu Thr Asn Lys Tyr Glu Tyr Gly Asp Asn Ile
65 70 75 80

Tyr Lys Glu Ala Lys Asp Arg Leu Leu Glu Lys Val Leu Arg Glu Asp
85 90 95

Gln Tyr Leu Leu Glu Arg Lys Lys Ser Gln Tyr Glu Asp Tyr Lys Gln
100 105 110

Trp Tyr Ala Asn Tyr Lys Lys Glu Asn Pro Arg Thr Asp Leu Lys Met
115 120 125

Ala Asn Phe His Lys Tyr Asn Leu Glu Glu Leu Ser Met Lys Glu Tyr
130 135 140

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Asn Glu Leu Gln Asp Ala Leu Lys Arg Ala Leu Asp Asp Phe His Arg
 145 150 155 160

Glu Val Lys Asp Ile Lys Asp Lys Asn Ser Asp Leu Lys Thr Phe Asn
 165 170 175

Ala Ala Glu Glu Asp Lys Ala Thr Lys Glu Val Tyr Asp Leu Val Ser
 180 185 190

Glu Ile Asp Thr Leu Val Val Ser Tyr Tyr Gly Asp Lys Asp Tyr Gly
 195 200 205

Glu His Ala Lys Glu Leu Arg Ala Lys Leu Asp Leu Ile Leu Gly Asp
 210 215 220

Thr Asp Asn Pro His Lys Ile Thr Asn Glu Arg Ile Lys Lys Glu Met
 225 230 235 240

Ile Asp Asp Leu Asn Ser Ile Ile Asp Asp Phe Phe Met Glu Thr Lys
 245 250 255

Gln Asn Arg Pro Lys Ser Ile Thr Lys Tyr Asn Pro Thr Thr His Asn
 260 265 270

Tyr Lys Thr Asn Ser Asp Asn Lys Pro Asn Phe Asp Lys Leu Val Glu
 275 280 285

Glu Thr Lys Lys Ala Val Lys Glu Ala Asp Asp Ser Trp Lys Lys Lys
 290 295 300

Thr Val Lys Lys Tyr Gly Glu Thr Glu Thr Lys Ser Pro Val Val Lys
 305 310 315 320

Glu Glu Lys Lys Val Glu Glu Pro Gln Ala Pro Lys Val Asp Asn Gln
 325 330 335

Gln Glu Val Lys Thr Thr Ala Gly Lys Ala Glu Glu Thr Thr Gln Pro
 340 345 350

Val Ala Gln Pro Leu Val Lys Ile Pro Gln Gly Thr Ile Thr Gly Glu
 355 360 365

Ile Val Lys Gly Pro Glu Tyr Pro Thr Met Glu Asn Lys Thr Val Gln
 370 375 380

Gly Glu Ile Val Gln Gly Pro Asp Phe Leu Thr Met Glu Gln Ser Gly
 385 390 395 400

Pro Ser Leu Ser Asn Asn Tyr Thr Asn Pro Pro Leu Thr Asn Pro Ile
 405 410 415

Leu Glu Gly Leu Glu Gly Ser Ser Ser Lys Leu Glu Ile Lys Pro Gln
 420 425 430

Gly Thr Glu Ser Thr Leu Lys Gly Thr Gln Gly Glu Ser Ser Asp Ile
 435 440 445

Glu Val Lys Pro Gln Ala Thr Glu Thr Thr Glu Ala Ser Gln Tyr Gly
 450 455 460

Pro Arg Pro Gln Phe Asn Lys Thr Pro Lys Tyr Val Lys Tyr Arg Asp
 465 470 475 480

Ala Gly Thr Gly Ile Arg Glu Tyr Asn Asp Gly Thr Phe Gly Tyr Glu
 485 490 495

Ala Arg Pro Arg Phe Asn Lys Pro Ser Glu Thr Asn Ala Tyr Asn Val
 500 505 510

Thr Thr His Ala Asn Gly Gln Val Ser Tyr Gly Ala Arg Pro Thr Tyr
 515 520 525

Lys Lys Pro Ser Glu Thr Asn Ala Tyr Asn Val Thr Thr His Ala Asn
 530 535 540

Gly Gln Val Ser Tyr Gly Ala Arg Pro Thr Gln Asn Lys Pro Ser Lys
 545 550 555 560

Thr Asn Ala Tyr Asn Val Thr Thr His Gly Asn Gly Gln Val Ser Tyr

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Arg Gly Asp Tyr Leu Val Lys Gln Leu Ser Gln Asn Pro Gln Leu Lys
 305 310 315 320
 Lys Arg Ile Thr Val Ile Asp Gly His Ser His Thr Val Leu Gln Asn
 325 330 335
 Gly Gln Ile Tyr Asn Asn Asp Ala Leu Ala Gln Thr Gly Thr Ala Leu
 340 345 350
 Ala Asn Ile Gly Lys Ile Thr Phe Asn Tyr Arg Asn Gly Glu Val Ser
 355 360 365
 Asn Ile Lys Pro Ser Leu Ile Asn Val Lys Asp Val Glu Asn Val Thr
 370 375 380
 Pro Asn Lys Ala Leu Ala Glu Gln Ile Asn Gln Ala Asp Gln Thr Phe
 385 390 395 400
 Arg Ala Gln Thr Ala Glu Val Ile Ile Pro Asn Asn Thr Ile Asp Phe
 405 410 415
 Lys Gly Glu Arg Asp Asp Val Arg Thr Arg Glu Thr Asn Leu Gly Asn
 420 425 430
 Ala Ile Ala Asp Ala Met Glu Ala Tyr Gly Val Lys Asn Phe Ser Lys
 435 440 445
 Lys Thr Asp Phe Ala Val Thr Asn Gly Gly Gly Ile Arg Ala Ser Ile
 450 455 460
 Ala Lys Gly Lys Val Thr Arg Tyr Asp Leu Ile Ser Val Leu Pro Phe
 465 470 475 480
 Gly Asn Thr Ile Ala Gln Ile Asp Val Lys Gly Ser Asp Val Trp Thr
 485 490 495
 Ala Phe Glu His Ser Leu Gly Ala Pro Thr Thr Gln Lys Asp Gly Lys
 500 505 510
 Thr Val Leu Thr Ala Asn Gly Gly Leu Leu His Ile Ser Asp Ser Ile
 515 520 525
 Arg Val Tyr Tyr Asp Ile Asn Lys Pro Ser Gly Lys Arg Ile Asn Ala
 530 535 540
 Ile Gln Ile Leu Asn Lys Glu Thr Gly Lys Phe Glu Asn Ile Asp Leu
 545 550 555 560
 Lys Arg Val Tyr His Val Thr Met Asn Asp Phe Thr Ala Ser Gly Gly
 565 570 575
 Asp Gly Tyr Ser Met Phe Gly Gly Pro Arg Glu Glu Gly Ile Ser Leu
 580 585 590
 Asp Gln Val Leu Ala Ser Tyr Leu Lys Thr Ala Asn Leu Ala Lys Tyr
 595 600 605
 Asp Thr Thr Glu Pro Gln Arg Met Leu Leu Gly Lys Pro Ala Val Ser
 610 615 620
 Glu Gln Pro Ala Lys Gly Gln Gln Gly Ser Lys Gly Ser Lys Ser Gly
 625 630 635 640
 Lys Asp Thr Gln Pro Ile Gly Asp Asp Lys Val Met Asp Pro Ala Lys
 645 650 655
 Lys Pro Ala Pro Gly Lys Val Val Leu Leu Leu Ala His Arg Gly Thr
 660 665 670
 Val Ser Ser Gly Thr Glu Gly Ser Gly Arg Thr Ile Glu Gly Ala Thr
 675 680 685
 Val Ser Ser Lys Ser Gly Lys Gln Leu Ala Arg Met Ser Val Pro Lys
 690 695 700
 Gly Ser Ala His Glu Lys Gln Leu Pro Lys Thr Gly Thr Asn Gln Ser
 705 710 715 720
 Ser Ser Pro Glu Ala Met Phe Val Leu Leu Ala Gly Ile Gly Leu Ile

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          725          730          735
Ala Thr Val Arg Arg Arg Lys Ala Ser
          740          745

<210> SEQ ID NO 29
<211> LENGTH: 628
<212> TYPE: PRT
<213> ORGANISM: Staphylococcus sp.

<400> SEQUENCE: 29

Met Ser Asp Arg Phe Ile Lys Phe Asn Asp Glu Gln Leu Asp Ala Lys
 1          5          10          15
Gln Val Met Met Leu Gln Asp Leu Ala Arg Leu Leu Leu Lys Asn Glu
          20          25          30
Gln Thr Gln Val Lys Ile Gln Lys Phe Pro Tyr Tyr Asn Pro Val Gln
          35          40          45
Asn Val Leu Ile Thr Ser Trp Phe Trp Ser His Arg Pro Ser His Ile
          50          55          60
Glu Met Ala Gly Leu Lys Thr Asp Val Met Leu Ala Ala Tyr Gly Tyr
 65          70          75          80
His Met Met Asp Val Gln Ile Val Asn Glu Val Val Gln Asp Lys Thr
          85          90          95
Phe Lys His Pro Lys Phe Tyr Gln Gln Leu Phe Lys Leu Leu Glu Asp
          100          105          110
Met Arg Val Leu Asn Ser Ile Lys Val Glu Arg Pro Ser Thr Ala Lys
          115          120          125
Leu Ile Asp Leu Arg Leu Asp Thr Arg Ile Ser Tyr Thr Glu Ser Gln
          130          135          140
Ile Lys Val Tyr Arg Thr Lys Thr Gln Tyr Thr Asp Leu Leu Phe Leu
          145          150          155          160
Tyr Leu Glu His Ala Phe Leu Ser Gln Asp Phe Phe Asp Ile Pro Ser
          165          170          175
Ile His Ser Asp Leu Asp Asp Ile Leu Val Asn Met Phe Leu Tyr Leu
          180          185          190
Pro Asn Phe Phe Gln Asn Gln Asn Ser Glu Asp Asn Met Tyr Leu Ala
          195          200          205
Gln Arg Ile Met Tyr Gln Val Asp Asp Ile Leu Lys Glu Asp Met Leu
          210          215          220
Asn Glu Tyr Tyr Tyr Leu Pro Lys Thr Leu Tyr Asn Thr Leu Ala Ser
          225          230          235          240
Pro Glu Phe Asp Asp Leu Lys Arg Thr Asp Ala Ser Gln Val Asp Gly
          245          250          255
Gln Asp Asp Thr Ser Glu Asp Asp Asp Asn Glu Ser Glu Lys Ala Asp
          260          265          270
Ser Lys Ser Ala Asp Ser Glu Ser Lys Gly Gly Ala Tyr Leu Glu Met
          275          280          285
Glu Leu His Glu Gly Gln Asn Ser Glu Thr Leu Gly Asn Asp Glu Ala
          290          295          300
Arg Glu Gly Asp Ala Thr Asp Asp Met Thr Asp Met Met Thr Lys Lys
          305          310          315          320
Gly Lys Gly Ser Asn Asp Thr Leu Asn Arg Glu Glu Gly Asp Ala Val
          325          330          335
Gly Gln Ser Gln Ala Phe Gln Leu Asp Gly Val Asn Lys Asn Val Glu
          340          345          350

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Ile Lys Trp Gln Ile Pro Glu Ile Glu Pro Gln Tyr Val Leu Glu Tyr
 355 360 365

Gln Glu Ser Lys Gln Asp Val Gln Tyr Glu Ile Lys Asp Leu Ile Gln
 370 375 380

Ile Ile Lys Lys Thr Ile Glu Arg Glu Gln Arg Asp Ala Arg Phe Asn
 385 390 395 400

Leu Thr Lys Gly Arg Leu Gln Lys Asp Leu Ile Asn Trp Phe Ile Asp
 405 410 415

Asp Gln Tyr Lys Leu Phe Tyr Lys Lys Gln Asp Leu Ser Lys Ser Phe
 420 425 430

Asp Ala Thr Phe Thr Leu Leu Ile Asp Ala Ser Ala Ser Met His Asp
 435 440 445

Lys Met Ala Glu Thr Lys Lys Gly Val Val Leu Phe His Glu Thr Leu
 450 455 460

Lys Ala Leu Asn Ile Lys His Glu Ile Leu Ser Phe Ser Glu Asp Ala
 465 470 475 480

Phe Asp Ser Asp Glu His Ala Gln Pro Asn Ile Ile Asn Glu Ile Ile
 485 490 495

Asn Tyr Asp Tyr Ser Thr Phe Glu Lys Asp Gly Pro Arg Ile Met Ala
 500 505 510

Leu Glu Pro Gln Asp Asp Asn Arg Asp Gly Val Ala Ile Arg Val Ala
 515 520 525

Ser Glu Arg Leu Met Arg Arg Asn Gln His Gln Arg Phe Leu Ile Val
 530 535 540

Phe Ser Asp Gly Glu Pro Ser Ala Phe Asn Tyr Ser Gln Asp Gly Ile
 545 550 555 560

Ile Asp Thr Tyr Glu Ala Val Glu Met Ser Arg Lys Phe Gly Ile Glu
 565 570 575

Val Phe Asn Val Phe Leu Ser Gln Asp Pro Ile Thr Glu Asp Val Glu
 580 585 590

Gln Thr Ile His Asn Ile Tyr Gly Gln Tyr Ala Ile Phe Val Glu Gly
 595 600 605

Val Ala His Leu Pro Gly His Leu Ser Pro Leu Leu Lys Lys Leu Leu
 610 615 620

Leu Lys Ser Leu
 625

<210> SEQ ID NO 30

<211> LENGTH: 154

<212> TYPE: PRT

<213> ORGANISM: Staphylococcus sp.

<400> SEQUENCE: 30

Ala Glu Ile Asn Lys Gln Thr Thr Ser Gln Gly Val Thr Thr Glu Lys
 1 5 10 15

Asn Asn Gly Ile Ala Val Leu Glu Gln Asp Val Ile Thr Pro Thr Val
 20 25 30

Lys Pro Gln Ala Lys Gln Asp Ile Ile Gln Ala Val Thr Thr Arg Lys
 35 40 45

Gln Gln Ile Lys Lys Ser Asn Ala Ser Leu Gln Asp Glu Lys Asp Val
 50 55 60

Ala Asn Asp Lys Ile Gly Lys Ile Glu Thr Lys Ala Ile Lys Asp Ile
 65 70 75 80

Asp Ala Ala Thr Thr Asn Ala Gln Val Glu Ala Ile Lys Thr Lys Ala
 85 90 95

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Ile Asn Asp Ile Asn Gln Thr Thr Pro Ala Thr Thr Ala Lys Ala Ala
 100 105 110
 Ala Leu Glu Glu Phe Asp Glu Val Val Gln Ala Gln Ile Asp Gln Ala
 115 120 125
 Pro Leu Asn Pro Asp Thr Thr Asn Glu Glu Val Ala Glu Ala Ile Glu
 130 135 140
 Arg Ile Asn Ala Ala Lys Val Ser Gly Val
 145 150

<210> SEQ ID NO 31
 <211> LENGTH: 584
 <212> TYPE: PRT
 <213> ORGANISM: Staphylococcus sp.

<400> SEQUENCE: 31

Met Lys Phe Lys Ser Leu Ile Thr Thr Thr Leu Ala Leu Gly Val Leu
 1 5 10 15
 Ala Ser Thr Gly Ala Asn Phe Asn Asn Asn Glu Ala Ser Ala Ala Ala
 20 25 30
 Lys Pro Leu Asp Lys Ser Ser Ser Ser Leu His His Gly Tyr Ser Lys
 35 40 45
 Val His Val Pro Tyr Ala Ile Thr Val Asn Gly Thr Ser Gln Asn Ile
 50 55 60
 Leu Ser Ser Leu Thr Phe Asn Lys Asn Gln Asn Ile Ser Tyr Lys Asp
 65 70 75 80
 Leu Glu Asp Arg Val Lys Ser Val Leu Lys Ser Asp Arg Gly Ile Ser
 85 90 95
 Asp Ile Asp Leu Arg Leu Ser Lys Gln Ala Lys Tyr Thr Val Tyr Phe
 100 105 110
 Lys Asn Gly Thr Lys Lys Val Ile Asp Leu Lys Ala Gly Ile Tyr Thr
 115 120 125
 Ala Asp Leu Ile Asn Thr Ser Glu Ile Lys Ala Ile Asn Ile Asn Val
 130 135 140
 Asp Thr Lys Lys Gln Val Glu Asp Lys Lys Lys Asp Lys Ala Asn Tyr
 145 150 155 160
 Gln Val Pro Tyr Thr Ile Thr Val Asn Gly Thr Ser Gln Asn Ile Leu
 165 170 175
 Ser Asn Leu Thr Phe Asn Lys Asn Gln Asn Ile Ser Tyr Lys Asp Leu
 180 185 190
 Glu Asp Lys Val Lys Ser Val Leu Glu Ser Asn Arg Gly Ile Thr Asp
 195 200 205
 Val Asp Leu Arg Leu Ser Lys Gln Ala Lys Tyr Thr Val Asn Phe Lys
 210 215 220
 Asn Gly Thr Lys Lys Val Ile Asp Leu Lys Ser Gly Ile Tyr Thr Ala
 225 230 235 240
 Asn Leu Ile Asn Ser Ser Asp Ile Lys Ser Ile Asn Ile Asn Val Asp
 245 250 255
 Thr Lys Lys His Ile Glu Asn Lys Ala Lys Arg Asn Tyr Gln Val Pro
 260 265 270
 Tyr Ser Ile Asn Leu Asn Gly Thr Ser Thr Asn Ile Leu Ser Asn Leu
 275 280 285
 Ser Phe Ser Asn Lys Pro Trp Thr Asn Tyr Lys Asn Leu Thr Ser Gln
 290 295 300
 Ile Lys Ser Val Leu Lys His Asp Arg Gly Ile Ser Glu Gln Asp Leu

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305                310                315                320
Lys Tyr Ala Lys Lys Ala Tyr Tyr Thr Val Tyr Phe Lys Asn Gly Gly
                325                330                335
Lys Arg Ile Leu Gln Leu Asn Ser Lys Asn Tyr Thr Ala Asn Leu Val
                340                345                350
His Ala Lys Asp Val Lys Arg Ile Glu Ile Thr Val Lys Thr Gly Thr
                355                360                365
Lys Ala Lys Ala Asp Arg Tyr Val Pro Tyr Thr Ile Ala Val Asn Gly
                370                375                380
Thr Ser Thr Pro Ile Leu Ser Asp Leu Lys Phe Thr Gly Asp Pro Arg
385                390                395                400
Val Gly Tyr Lys Asp Ile Ser Lys Lys Val Lys Ser Val Leu Lys His
                405                410                415
Asp Arg Gly Ile Gly Glu Arg Glu Leu Lys Tyr Ala Lys Lys Ala Thr
                420                425                430
Tyr Thr Val His Phe Lys Asn Gly Thr Lys Lys Val Ile Asn Ile Asn
                435                440                445
Ser Asn Ile Ser Gln Leu Asn Leu Leu Tyr Val Gln Asp Ile Lys Lys
                450                455                460
Ile Asp Ile Asp Val Lys Thr Gly Thr Lys Ala Lys Ala Asp Ser Tyr
465                470                475                480
Val Pro Tyr Thr Ile Ala Val Asn Gly Thr Ser Thr Pro Ile Leu Ser
                485                490                495
Lys Leu Lys Ile Ser Asn Lys Gln Leu Ile Ser Tyr Lys Tyr Leu Asn
                500                505                510
Asp Lys Val Lys Ser Val Leu Lys Ser Glu Arg Gly Ile Ser Asp Leu
                515                520                525
Asp Leu Lys Phe Ala Lys Gln Ala Lys Tyr Thr Val Tyr Phe Lys Asn
                530                535                540
Gly Lys Lys Gln Val Val Asn Leu Lys Ser Asp Ile Phe Thr Pro Asn
545                550                555                560
Leu Phe Ser Ala Lys Asp Ile Lys Lys Ile Asp Ile Asp Val Lys Gln
                565                570                575
Tyr Thr Lys Ser Lys Lys Asn Lys
                580

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<210> SEQ ID NO 32

<211> LENGTH: 508

<212> TYPE: PRT

<213> ORGANISM: Staphylococcus sp.

<400> SEQUENCE: 32

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Met Lys Asn Lys Leu Leu Val Leu Ser Leu Gly Ala Leu Cys Val Ser
1                5                10                15
Gln Ile Trp Glu Ser Asn Arg Ala Ser Ala Val Val Ser Gly Glu Lys
                20                25                30
Asn Pro Tyr Val Ser Glu Ser Leu Lys Leu Thr Asn Asn Lys Asn Lys
                35                40                45
Ser Arg Thr Val Glu Glu Tyr Lys Lys Ser Leu Asp Asp Leu Ile Trp
                50                55                60
Ser Phe Pro Asn Leu Asp Asn Glu Arg Phe Asp Asn Pro Glu Tyr Lys
65                70                75                80
Glu Ala Met Lys Lys Tyr Gln Gln Arg Phe Met Ala Glu Asp Glu Ala
                85                90                95

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Leu Lys Lys Phe Phe Ser Glu Glu Lys Lys Ile Lys Asn Gly Asn Thr
 100 105 110
 Asp Asn Leu Asp Tyr Leu Gly Leu Ser His Glu Arg Tyr Glu Ser Val
 115 120 125
 Phe Asn Thr Leu Lys Lys Gln Ser Glu Glu Phe Leu Lys Glu Ile Glu
 130 135 140
 Asp Ile Lys Lys Asp Asn Pro Glu Leu Lys Asp Phe Asn Glu Glu Glu
 145 150 155 160
 Gln Leu Lys Cys Asp Leu Glu Leu Asn Lys Leu Glu Asn Gln Ile Leu
 165 170 175
 Met Leu Gly Lys Thr Phe Tyr Gln Asn Tyr Arg Asp Asp Val Glu Ser
 180 185 190
 Leu Tyr Ser Lys Leu Asp Leu Ile Met Gly Tyr Lys Asp Glu Glu Arg
 195 200 205
 Ala Asn Lys Lys Ala Val Asn Lys Arg Met Leu Glu Asn Lys Lys Glu
 210 215 220
 Asp Leu Glu Thr Ile Ile Asp Glu Phe Phe Ser Asp Ile Asp Lys Thr
 225 230 235 240
 Arg Pro Asn Asn Ile Pro Val Leu Glu Asp Glu Lys Gln Glu Glu Lys
 245 250 255
 Asn His Lys Asn Met Ala Gln Leu Lys Ser Asp Thr Glu Ala Ala Lys
 260 265 270
 Ser Asp Glu Ser Lys Arg Ser Lys Arg Ser Lys Arg Ser Leu Asn Thr
 275 280 285
 Gln Asn His Lys Pro Ala Ser Gln Glu Val Ser Glu Gln Gln Lys Ala
 290 295 300
 Glu Tyr Asp Lys Arg Ala Glu Glu Arg Lys Ala Arg Phe Leu Asp Asn
 305 310 315 320
 Gln Lys Ile Lys Lys Thr Pro Val Val Ser Leu Glu Tyr Asp Phe Glu
 325 330 335
 His Lys Gln Arg Ile Asp Asn Glu Asn Asp Lys Lys Leu Val Val Ser
 340 345 350
 Ala Pro Thr Lys Lys Pro Thr Ser Pro Thr Thr Tyr Thr Glu Thr Thr
 355 360 365
 Thr Gln Val Pro Met Pro Thr Val Glu Arg Gln Thr Gln Gln Gln Ile
 370 375 380
 Ile Tyr Asn Ala Pro Lys Gln Leu Ala Gly Leu Asn Gly Glu Ser His
 385 390 395 400
 Asp Phe Thr Thr Thr His Gln Ser Pro Thr Thr Ser Asn His Thr His
 405 410 415
 Asn Asn Val Val Glu Phe Glu Glu Thr Ser Ala Leu Pro Gly Arg Lys
 420 425 430
 Ser Gly Ser Leu Val Gly Ile Ser Gln Ile Asp Ser Ser His Leu Thr
 435 440 445
 Glu Arg Glu Lys Arg Val Ile Lys Arg Glu His Val Arg Glu Ala Gln
 450 455 460
 Lys Leu Val Asp Asn Tyr Lys Asp Thr His Ser Tyr Lys Asp Arg Ile
 465 470 475 480
 Asn Ala Gln Gln Lys Val Asn Thr Leu Ser Glu Gly His Gln Lys Arg
 485 490 495
 Phe Asn Lys Gln Ile Asn Lys Val Tyr Asn Gly Lys
 500 505

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<210> SEQ ID NO 33
<211> LENGTH: 520
<212> TYPE: PRT
<213> ORGANISM: Staphylococcus sp.

<400> SEQUENCE: 33

Met Leu Thr Leu Gln Ile His Thr Gly Gly Ile Asn Leu Lys Lys Lys
 1           5           10           15
Asn Ile Tyr Ser Ile Arg Lys Leu Gly Val Gly Ile Ala Ser Val Thr
 20           25           30
Leu Gly Thr Leu Leu Ile Ser Gly Gly Val Thr Pro Ala Ala Asn Ala
 35           40           45
Ala Gln His Asp Glu Ala Gln Gln Asn Ala Phe Tyr Gln Val Leu Asn
 50           55           60
Met Pro Asn Leu Asn Ala Asp Gln Arg Asn Gly Phe Ile Gln Ser Leu
 65           70           75           80
Lys Asp Asp Pro Ser Gln Ser Ala Asn Val Leu Gly Glu Ala Gln Lys
 85           90           95
Leu Asn Asp Ser Gln Ala Pro Lys Ala Asp Ala Gln Gln Asn Asn Phe
 100          105          110
Asn Lys Asp Gln Gln Ser Ala Phe Tyr Glu Ile Leu Asn Met Pro Asn
 115          120          125
Leu Asn Glu Ala Gln Arg Asn Gly Phe Ile Gln Ser Leu Lys Asp Asp
 130          135          140
Pro Ser Gln Ser Thr Asn Val Leu Gly Glu Ala Lys Lys Leu Asn Glu
 145          150          155          160
Ser Gln Ala Pro Lys Ala Asp Asn Asn Phe Asn Lys Glu Gln Gln Asn
 165          170          175
Ala Phe Tyr Glu Ile Leu Asn Met Pro Asn Leu Asn Glu Glu Gln Arg
 180          185          190
Asn Gly Phe Ile Gln Ser Leu Lys Asp Asp Pro Ser Gln Ser Ala Asn
 195          200          205
Leu Leu Ser Glu Ala Lys Lys Leu Asn Glu Ser Gln Ala Pro Lys Ala
 210          215          220
Asp Asn Lys Phe Asn Lys Glu Gln Gln Asn Ala Phe Tyr Glu Ile Leu
 225          230          235          240
His Leu Pro Asn Leu Asn Glu Glu Gln Arg Asn Gly Phe Ile Gln Ser
 245          250          255
Leu Lys Asp Asp Pro Ser Gln Ser Ala Asn Leu Leu Ala Glu Ala Lys
 260          265          270
Lys Leu Asn Asp Ala Gln Ala Pro Lys Ala Asp Asn Lys Phe Asn Lys
 275          280          285
Glu Gln Gln Asn Ala Phe Tyr Glu Ile Leu His Leu Pro Asn Leu Thr
 290          295          300
Glu Glu Gln Arg Asn Gly Phe Ile Gln Ser Leu Lys Asp Asp Pro Ser
 305          310          315          320
Val Ser Lys Glu Ile Leu Ala Glu Ala Lys Lys Leu Asn Asp Ala Gln
 325          330          335
Ala Pro Lys Glu Glu Asp Asn Asn Lys Pro Gly Lys Glu Asp Gly Asn
 340          345          350
Lys Pro Gly Lys Glu Asp Asn Asn Lys Pro Gly Lys Glu Asp Asn Lys
 355          360          365
Lys Pro Gly Lys Glu Asp Asn Asn Lys Pro Gly Lys Glu Asp Asn Asn
 370          375          380

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Lys Pro Gly Lys Glu Asp Gly Asn Lys Pro Gly Lys Glu Asp Asn Lys
 385 390 395 400
 Lys Pro Gly Lys Glu Asp Asn Asn Lys Pro Gly Lys Glu Asp Gly Asn
 405 410 415
 Lys Pro Gly Lys Glu Asp Gly Asn Gly Val His Val Val Lys Pro Gly
 420 425 430
 Asp Thr Val Asn Asp Ile Ala Lys Ala Asn Gly Thr Thr Ala Asp Lys
 435 440 445
 Ile Ala Ala Asp Asn Lys Leu Ala Asp Lys Asn Met Ile Lys Pro Gly
 450 455 460
 Gln Glu Leu Val Val Asp Lys Lys Gln Pro Ala Asn His Ala Asp Ala
 465 470 475 480
 Asn Lys Ala Gln Ala Leu Pro Glu Thr Gly Glu Glu Asn Pro Phe Ile
 485 490 495
 Gly Thr Thr Val Phe Gly Gly Leu Ser Leu Ala Leu Gly Ala Ala Leu
 500 505 510
 Leu Ala Gly Arg Arg Arg Glu Leu
 515 520

<210> SEQ ID NO 34
 <211> LENGTH: 291
 <212> TYPE: PRT
 <213> ORGANISM: Staphylococcus sp.

<400> SEQUENCE: 34

Ala Gln His Asp Glu Ala Lys Lys Asn Ala Phe Tyr Gln Val Leu Asn
 1 5 10 15
 Met Pro Asn Leu Asn Ala Asp Gln Arg Asn Gly Phe Ile Gln Ser Leu
 20 25 30
 Lys Ala Ala Pro Ser Gln Ser Ala Asn Val Leu Gly Glu Ala Gln Lys
 35 40 45
 Leu Asn Asp Ser Gln Ala Pro Lys Ala Asp Ala Gln Gln Asn Asn Phe
 50 55 60
 Asn Lys Asp Lys Lys Ser Ala Phe Tyr Glu Ile Leu Asn Met Pro Asn
 65 70 75 80
 Leu Asn Glu Ala Gln Arg Asn Gly Phe Ile Gln Ser Leu Lys Ala Ala
 85 90 95
 Pro Ser Gln Ser Thr Asn Val Leu Gly Glu Ala Lys Lys Leu Asn Glu
 100 105 110
 Ser Gln Ala Pro Lys Ala Asp Asn Asn Phe Asn Lys Glu Lys Lys Asn
 115 120 125
 Ala Phe Tyr Glu Ile Leu Asn Met Pro Asn Leu Asn Glu Glu Gln Arg
 130 135 140
 Asn Gly Phe Ile Gln Ser Leu Lys Ala Ala Pro Ser Gln Ser Ala Asn
 145 150 155 160
 Leu Leu Ser Glu Ala Lys Lys Leu Asn Glu Ser Gln Ala Pro Lys Ala
 165 170 175
 Asp Asn Lys Phe Asn Lys Glu Lys Lys Asn Ala Phe Tyr Glu Ile Leu
 180 185 190
 His Leu Pro Asn Leu Asn Glu Glu Gln Arg Asn Gly Phe Ile Gln Ser
 195 200 205
 Leu Lys Ala Ala Pro Ser Gln Ser Ala Asn Leu Leu Ala Glu Ala Lys
 210 215 220
 Lys Leu Asn Asp Ala Gln Ala Pro Lys Ala Asp Asn Lys Phe Asn Lys
 225 230 235 240

-continued

<212> TYPE: DNA
 <213> ORGANISM: Staphylococcus sp.
 <400> SEQUENCE: 41
 gttagtgcctt tggcttgggg cggctttaag actttgaatg aag 43

<210> SEQ ID NO 42
 <211> LENGTH: 42
 <212> TYPE: DNA
 <213> ORGANISM: Staphylococcus sp.
 <400> SEQUENCE: 42
 catatggttca acaaagataa aaaaagcgcc ttctatgaaa tc 42

<210> SEQ ID NO 43
 <211> LENGTH: 42
 <212> TYPE: DNA
 <213> ORGANISM: Staphylococcus sp.
 <400> SEQUENCE: 43
 gatttcatag aaggcgcttt ttttatcttt gttgaacata tg 42

<210> SEQ ID NO 44
 <211> LENGTH: 42
 <212> TYPE: DNA
 <213> ORGANISM: Staphylococcus sp.
 <400> SEQUENCE: 44
 catatggttca acaaagatgg aggaagcgcc ttctatgaaa tc 42

<210> SEQ ID NO 45
 <211> LENGTH: 42
 <212> TYPE: DNA
 <213> ORGANISM: Staphylococcus sp.
 <400> SEQUENCE: 45
 gatttcatag aaggcgcttc ctccatcttt gttgaacata tg 42

<210> SEQ ID NO 46
 <211> LENGTH: 52
 <212> TYPE: DNA
 <213> ORGANISM: Staphylococcus sp.
 <400> SEQUENCE: 46
 ggggacaagt ttgtacaaaa aagcaggctg atgactaagt tgaaaaaaga ag 52

<210> SEQ ID NO 47
 <211> LENGTH: 28
 <212> TYPE: DNA
 <213> ORGANISM: Staphylococcus sp.
 <400> SEQUENCE: 47
 aaggatcccc tccaaaatgt aattgccc 28

<210> SEQ ID NO 48
 <211> LENGTH: 30
 <212> TYPE: DNA
 <213> ORGANISM: Staphylococcus sp.
 <400> SEQUENCE: 48
 aaggatccgt ttgtaactct atccaaagac 30

<210> SEQ ID NO 49

-continued

<211> LENGTH: 49
 <212> TYPE: DNA
 <213> ORGANISM: Staphylococcus sp.
 <400> SEQUENCE: 49
 ggggaccact ttgtacaaga aagctgggtg acacctattg cacgattcg 49

<210> SEQ ID NO 50
 <211> LENGTH: 50
 <212> TYPE: DNA
 <213> ORGANISM: Staphylococcus sp.
 <400> SEQUENCE: 50
 ggggacaagt ttgtacaaaa aagcaggctc agatagcgat tcagattcag 50

<210> SEQ ID NO 51
 <211> LENGTH: 31
 <212> TYPE: DNA
 <213> ORGANISM: Staphylococcus sp.
 <400> SEQUENCE: 51
 aaggatccct gtattttctc cttaattttc c 31

<210> SEQ ID NO 52
 <211> LENGTH: 30
 <212> TYPE: DNA
 <213> ORGANISM: Staphylococcus sp.
 <400> SEQUENCE: 52
 aaggatccca tggctgcaaa gcaaataatg 30

<210> SEQ ID NO 53
 <211> LENGTH: 51
 <212> TYPE: DNA
 <213> ORGANISM: Staphylococcus sp.
 <400> SEQUENCE: 53
 ggggaccact ttgtacaaga aagctgggtg ccoctgggtgta acaaatttat g 51

<210> SEQ ID NO 54
 <211> LENGTH: 37
 <212> TYPE: DNA
 <213> ORGANISM: Staphylococcus sp.
 <400> SEQUENCE: 54
 gaaggatccg tttattctag ttaatatata gttaatg 37

<210> SEQ ID NO 55
 <211> LENGTH: 33
 <212> TYPE: DNA
 <213> ORGANISM: Staphylococcus sp.
 <400> SEQUENCE: 55
 gaactgcagc tgtatgtctt tggatagagt tac 33

<210> SEQ ID NO 56
 <211> LENGTH: 33
 <212> TYPE: DNA
 <213> ORGANISM: Staphylococcus sp.
 <400> SEQUENCE: 56
 gaaggatccg gtggcttttt tacttggatt ttc 33

-continued

<210> SEQ ID NO 57
 <211> LENGTH: 33
 <212> TYPE: DNA
 <213> ORGANISM: Staphylococcus sp.
 <400> SEQUENCE: 57
 gaactgcagc gacaaactca ttatttgctt tgc 33

<210> SEQ ID NO 58
 <211> LENGTH: 27
 <212> TYPE: DNA
 <213> ORGANISM: Staphylococcus sp.
 <400> SEQUENCE: 58
 gaactcgagt ctagcttatt tacatgg 27

<210> SEQ ID NO 59
 <211> LENGTH: 45
 <212> TYPE: DNA
 <213> ORGANISM: Staphylococcus sp.
 <400> SEQUENCE: 59
 gaactcgaga tagaaggcag aatagtaaca aaggattata gtggg 45

<210> SEQ ID NO 60
 <211> LENGTH: 27
 <212> TYPE: DNA
 <213> ORGANISM: Staphylococcus sp.
 <400> SEQUENCE: 60
 gtaggatcct gggatagagt tacaaaac 27

<210> SEQ ID NO 61
 <211> LENGTH: 34
 <212> TYPE: DNA
 <213> ORGANISM: Staphylococcus sp.
 <400> SEQUENCE: 61
 gaactcgagg cattatgtgt atcacaatt tggg 34

<210> SEQ ID NO 62
 <211> LENGTH: 43
 <212> TYPE: DNA
 <213> ORGANISM: Staphylococcus sp.
 <400> SEQUENCE: 62
 gaactcgaga tagaaggcag agtggtttct ggggagaaga atc 43

<210> SEQ ID NO 63
 <211> LENGTH: 33
 <212> TYPE: DNA
 <213> ORGANISM: Staphylococcus sp.
 <400> SEQUENCE: 63
 gaactcgagg cagccatgca ttaattattt gcc 33

<210> SEQ ID NO 64
 <211> LENGTH: 61
 <212> TYPE: PRT
 <213> ORGANISM: Staphylococcus sp. NT165
 <400> SEQUENCE: 64
 Ala Asp Ala Gln Gln Asn Gln His Asp Glu Ala Gln Gln Asn Ala Phe
 1 5 10 15

-continued

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1           5           10           15
Leu His Leu Pro Asn Leu Asn Ala Glu Gln Arg Asn Gly Phe Ile Gln
                20                25                30
Ser Leu Lys Asp Asp Pro Ser Gln Ser Ala Glu Leu Leu Ala Glu Ala
                35                40                45
Gln Lys Leu Asn Asp Ala Gln Ala Pro Lys
                50                55

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<210> SEQ ID NO 69
<211> LENGTH: 50
<212> TYPE: PRT
<213> ORGANISM: Staphylococcus sp. NT165

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<400> SEQUENCE: 69

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Phe Asn Lys Asp Lys Lys Ser Ala Phe Tyr Glu Ile Leu Asn Met Pro
1           5           10           15
Asn Leu Asn Glu Ala Gln Arg Asn Gly Phe Ile Gln Ser Leu Lys Ala
                20                25                30
Ala Pro Ser Gln Ser Thr Asn Val Leu Gly Glu Ala Lys Lys Leu Asn
                35                40                45
Glu Ser
                50

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<210> SEQ ID NO 70
<211> LENGTH: 6
<212> TYPE: PRT
<213> ORGANISM: Staphylococcus sp. NT165
<220> FEATURE:
<221> NAME/KEY: MISC_FEATURE
<222> LOCATION: (6)..(6)
<223> OTHER INFORMATION: X is either Gly or Ser

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<400> SEQUENCE: 70

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Tyr Ser Ile Arg Lys Xaa
1           5

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<210> SEQ ID NO 71
<211> LENGTH: 5
<212> TYPE: PRT
<213> ORGANISM: Staphylococcus sp. NT165
<220> FEATURE:
<221> NAME/KEY: MISC_FEATURE
<222> LOCATION: (3)..(3)
<223> OTHER INFORMATION: X is any amino acid

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<400> SEQUENCE: 71

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Leu Pro Xaa Thr Gly
1           5

```

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<210> SEQ ID NO 72
<211> LENGTH: 5
<212> TYPE: PRT
<213> ORGANISM: Staphylococcus sp. NT165

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<400> SEQUENCE: 72

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Tyr Ser Ile Arg Lys
1           5

```

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<210> SEQ ID NO 73
<211> LENGTH: 4
<212> TYPE: PRT
<213> ORGANISM: Staphylococcus sp. NT165

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<400> SEQUENCE: 73

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-continued

Ile Val Thr Lys
1

<210> SEQ ID NO 74
<211> LENGTH: 4
<212> TYPE: PRT
<213> ORGANISM: Staphylococcus sp. NT165

<400> SEQUENCE: 74

Val Val Ser Gly
1

The invention claimed is:

1. An isolated polypeptide comprising a Staphylococcal Protein A (SpA) variant domain D segment comprising amino acid substitutions at amino acids corresponding to positions 9, 10, 36 and 37 of SEQ ID NO: 2, wherein the substitutions at amino acids corresponding to positions 9 and 10 are with a lysine residue and the substitutions at amino acids corresponding to positions 36 and 37 are with an alanine residue, and wherein the amino acid sequence is at least 80% identical to SEQ ID NO:2.

2. The polypeptide of claim 1, wherein the amino acid sequence is at least 85% identical to SEQ ID NO: 2.

3. The isolated polypeptide of claim 1, further comprising one or more immunogenic fragments of SpA E domain, SpA A domain, SpA B domain, or SpA C domain.

4. The isolated polypeptide of claim 1, further comprising two or more of immunogenic SpA D domain fragments.

5. The isolated polypeptide of claim 1, further comprising an immunogenic fragment of a second staphylococcal antigen.

6. The isolated polypeptide of claim 5, wherein the second staphylococcal antigen is selected from the group consisting of Emp, EsxA, EsxB, EsaC, Eap, Ebh, EsaB, Coa, vWbp, vWh, Hla, SdrC, SdrD, SdrE, lsdA, lsdB, lsdC, ClfA, ClfB and SasF.

7. A composition comprising an isolated polypeptide comprising Staphylococcal Protein A (SpA) variant domain D segment comprising amino acid substitutions at amino acids corresponding to positions 9, 10, 36 and 37 of SEQ ID NO: 2, wherein the substitutions at amino acids corresponding to positions 9 and 10 are with a lysine residue and the substitutions at amino acids corresponding to positions 36 and 37 are with an alanine residue, and wherein the amino acid sequence is at least 80% identical to SEQ ID NO: 2, and optionally wherein the polypeptide further comprises one or more immunogenic fragments of SpA E domain, SpA A domain, SpA B domain, or SpA C domain.

8. The composition of claim 7, further comprising at least one other staphylococcal antigen selected from the group

consisting of Emp, EsxA, EsxB, EsaC, Eap, Ebh, EsaB, Coa, vWbp, vWh, Hla, SdrC, SdrD, SdrE, lsdA, lsdB, lsdC, ClfA, ClfB and SasF.

9. The composition of claim 7, wherein the composition contains less than 1% by weight of staphylococcal bacterial components other than the polypeptide comprising the SpA variant domain D segment.

10. The composition of claim 7, further comprising a staphylococcal PIA capsular polysaccharide or its oligosaccharide.

11. The composition of claim 7, further comprising type V or type VIII capsular polysaccharide of *S. aureus* or its oligosaccharide.

12. The composition of claim 7, further comprising a staphylococcal capsular polysaccharide conjugated to a protein carrier.

13. The isolated polypeptide of claim 1, wherein the polypeptide further comprises one or more immunogenic variants of SpA E domain, SpA A domain, SpA B domain, or SpA C domain that have an amino acid sequence that is at least 80% identical to SEQ ID NOS: 3, 4, 6, and 5, respectively, and wherein the one or more domains comprise amino acid substitutions at amino acids corresponding to positions 9, 10, 36 and 37 of SEQ ID NO: 2, wherein the amino acid substitutions corresponding to positions 9 and 10 are with a lysine residue and the amino acid substitutions corresponding to positions 36 and 37 are with an alanine residue.

14. The composition of claim 7, wherein the composition further comprises immunogenic fragments comprising one or more of SpA E domain, SpA A domain, SpA B domain, or SpA C domain that have an amino acid sequence that is at least 80% identical to SEQ ID NOS: 3, 4, 6, and 5, respectively, and wherein the one or more domains comprise amino acid substitutions at amino acids corresponding to positions 9, 10, 36 and 37 of SEQ ID NO: 2, wherein the amino acid substitutions corresponding to positions 9 and 10 are with a lysine residue and the amino acid substitutions corresponding to positions 36 and 37 are with an alanine residue.

* * * * *