

THE UNIVERSITY OF CHICAGO

PALLADIUM CATALYZED CYCLIZATION AND C–O BOND FORMATION CASCADE,
AND ITS APPLICATION IN TOTAL SYNTHESIS

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ABSTRACT

Palladium catalyzed cyclization and C–O bond formation cascade, and its application in total synthesis

Chapter 1. Total synthesis inspired cascade reactions

Total synthesis of complex natural products has long inspired many creative methodologies. Cascade reactions have shown their ability to construct complex structures in total synthesis. In our group's former work on *(-)-presilphiperfolan-8-ol*, a cascade cyclization was used to construct the highly strained molecule. Inspired by our former results and synthesis of terpenes possessing a 1,3-*trans* structure, such as *botrydial* family, we designed a cascade reaction.

Chapter 2. Palladium catalyzed cyclization and C–O bond formation cascade

Heck type cyclization and nucleophilic attack cascades matched the stereochemistry demand. However, a C–O bond formation process, which was required in those terpene syntheses, was not preceded. To find a solution of the challenge, we developed a cyclization and C–O bond formation cascade. The methodology worked for a wide scope of substrates, resulting in moderate to high yields. We investigated the mechanism and found that a S_N2 process through a cyclopropane intermediate would be the most possible, deferent from former proposals. The methodology was used in total synthesis of *botrydial* and *epi-10-oxodehydro-dihydrobotrydial*, indicating the potential power of this methodology in organic synthesis.

Chapter 3. Synthetic study towards *paraliane* family of natural products

Paralianes are terpenes with complex tetracyclic structures. They are bioactive in an anti-inflammatory capacity. Though first isolated in 1998, none of the members in this family have been synthesized. We synthesized the east fragment with our palladium catalyzed methodology. The west fragment was synthesized from a commercial chiral starting material. The two fragments were coupled together by the Nozaki–Hiyama–Kishi reaction. In model study we achieved the

unprecedented Mukaiyama radical coupling of alkene and aldehyde for ring closure. Further study is still going on in real system to synthesize the target molecule.

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LIST OF ABBREVIATIONS

Ac	acetyl
acac	acetylacetonate
Ad	adamantyl
BDSB	bromodiethylsulfonium bromopentachloroantimate
Bn	benzyl
DBU	1,8-diazabicyclo[5.4.0]undec-7-ene
DCE	1,2-dichloroethane
DDQ	2,3-dicyano-5,6-dichlorobenzoquinone
DIBAL-H	diisobutylaluminum hydride
DIPA	diisopropylamine
DMF	<i>N,N</i> -dimethylformamide
DMP	Dess-Martin periodinane
DMSO	dimethylsulfoxide
dpm	2,2,6,6-tetramethyl-3,5-heptanedionate
HMPA	hexamethylphosphoramide
HOBt	1-hydroxybenzotriazole

IBX	2-iodoxybenzoic acid
KHMDS	potassium bis(trimethylsilyl)amide
LAH	lithium aluminum hydride
LDA	lithium diisopropylamide
LHMDS	lithium bis(trimethylsilyl)amide
<i>m</i>CPBA	<i>meta</i> -chloroperoxybenzoic acid
MPO	4-methoxypyridine <i>N</i> -oxide
MVK	methyl vinyl ketone
NBS	<i>N</i> -bromosuccinimide
NMO	4-methylmorpholine <i>N</i> -oxide
PCC	pyridinium chlorochromate
PDC	pyridinium dichromate
Phth	phthalimide
Piv	pivaloyl
PMB	<i>para</i> -methoxybenzyl
PMP	1,2,2,6,6-pentamethylpiperidine
<i>t</i>-AmylOH	2-methylbutan-2-ol
TBAF	tetrabutylammonium fluoride

TBAI	tetrabutylammonium iodide
TBHP	<i>tert</i> -butyl hydroperoxide
TBS	<i>tert</i> -butyldimethylsilyl
TES	triethylsilyl
Tf	trifluoromethanesulfonate
THF	tetrahydrofuran
TIPS	triisopropylsilyl
TMG	1,1,3,3-tetramethylguanidine
TMS	trimethylsilyl
Ts	tosyl

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CHAPTER 1

TOTAL SYNTHESIS INSPIRED CASCADE REACTIONS

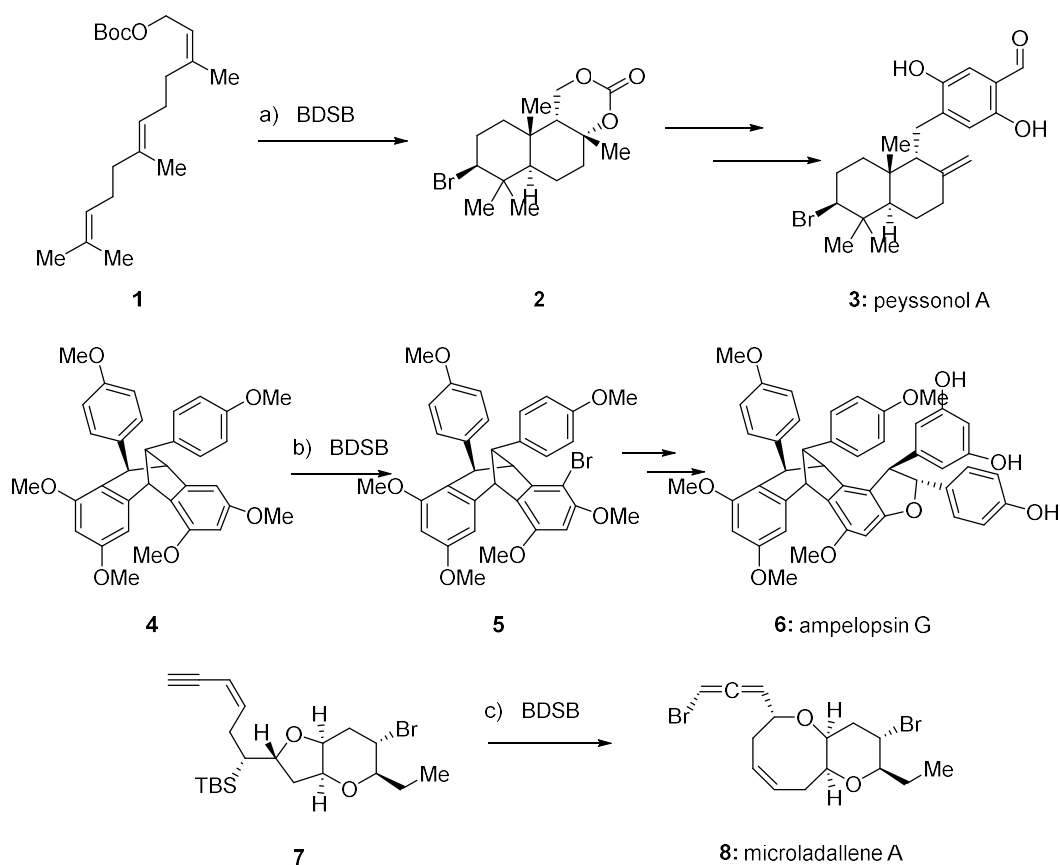
1.1 Total synthesis inspired methodology

Total synthesis of complex natural products is an arena of organic chemistry. The application in total synthesis often illustrated the power of one method. The challenging problems that are unable to draw upon previous methods are often the driving force for the discovery of new methodologies. Since the very start of total synthesis about 200 years ago,^[1] chemists have been developing many new approaches for total synthesis. Stories laid on how new methodologies were inspired from difficulties met in total synthesis. The useful DCC esterification^[2] was a fruit of long struggle with the β -lactam structure in *penicillin* synthesis and peptide formation. The demand for enantio-selective method to synthesize *prostaglandin* led to Corey's borane based asymmetric reduction of ketones, Corey-Bakshi-Shibata reaction.^[3] In recent research, Lee's diazo radical tandem cyclization was closely related to their terpene total synthesis.^[4]

Those examples are just the tip of a huge iceberg in the research history of organic synthesis. Our group has been working on total synthesis and inspired methodologies for a long time. For selective bromination and polyene cyclization in halogen-containing natural products, our group developed BDSB as a new and convenient reagent as a bromonium resource.^[5] This reagent played an important role in the synthesis of *resveratrol* oligomers (**6**) as a selective bromination reagent of aromatic rings.^[6] The bromonium introduced polyene cyclization also led to the synthesis of *peyssonol* (**3**) and *laurencia* (**8**) family natural products.^[7] Those synthetic applications showed the power of BDSB as a bromination reagent. Our recent pyrone Diels-Alder reaction^[8] and asymmetric Pictet-Spengler reaction^[9] are also related to total synthesis. (Scheme 1-1)

Also inspired by oxidized terpene synthesis, herein we will introduce our development of a cyclization methodology and its application in total synthesis.

Scheme 1-1: BDSB in total synthesis

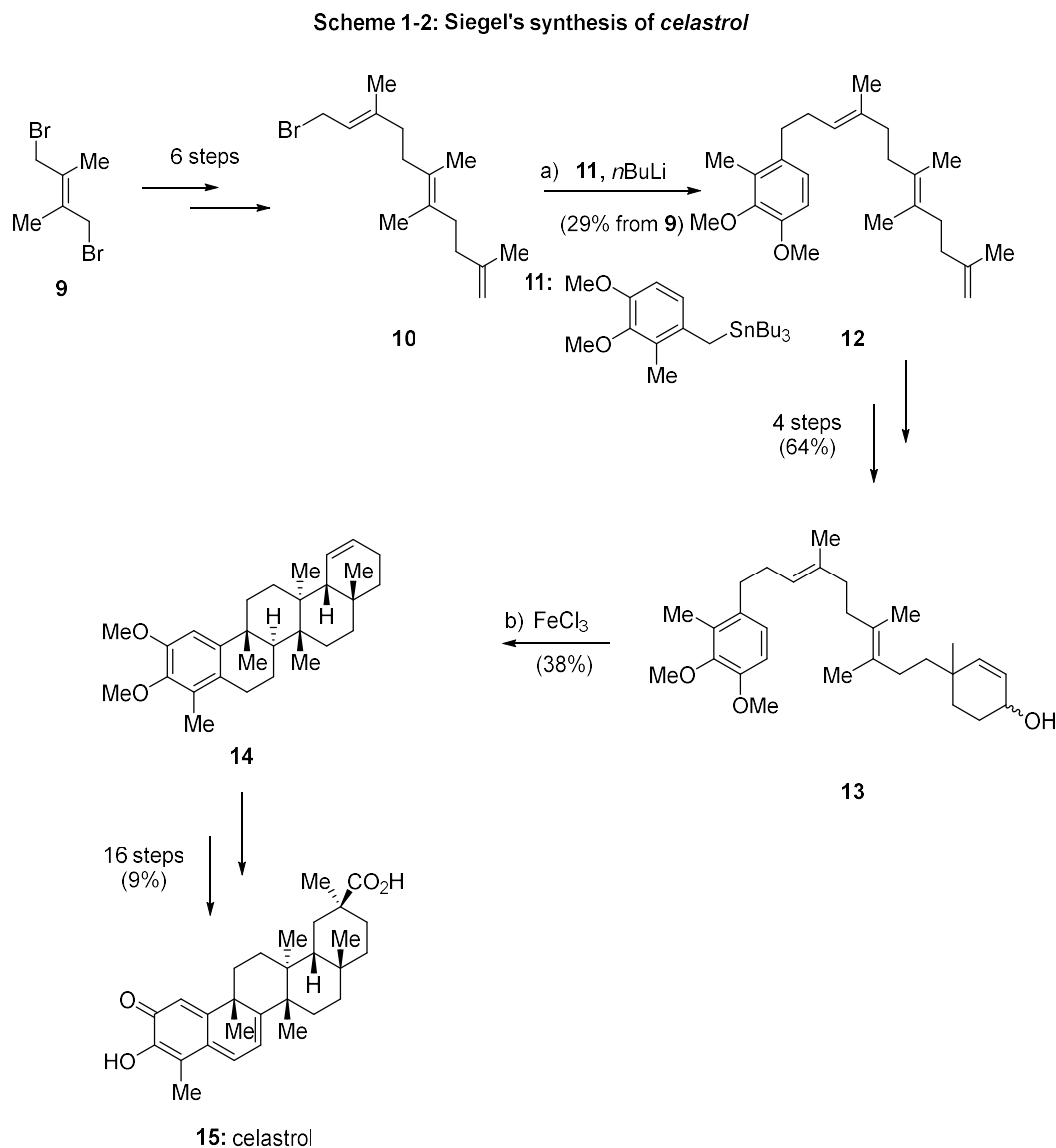


1.2 Cascade reaction in total synthesis

Cascade reactions can form complex ring structures, such as continuous stereocenters, in one step. Thus, they are very useful tools in the total synthesis of natural products. Robinson's famous synthesis of tropinone in 1917 was the first light of cascade cyclization.^[10] From then on, many cascade reactions have been developed.

Polyene cyclization is typical of cascade reactions. They played a significant role in the natural synthesis of terpenes and steroids, and in total synthesis since the middle of the 20th century.^[11] The BDSB research in our group described above was also closely related to polyene cyclization.^[5,7] One more recent example would be Siegel's synthesis of *celastrol* (**15**) in 2015 from simple alkene **9**. **9** was elongated to triene **10**, which was coupled with organotin **11** to

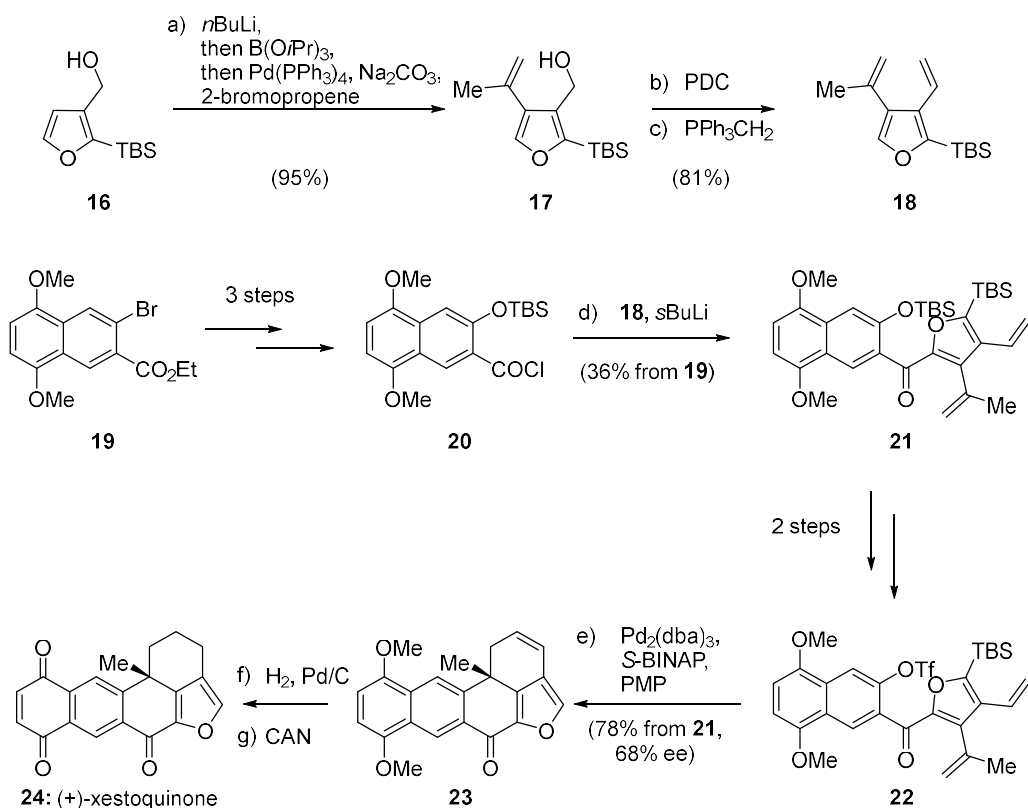
provide intermediate **12**. Then, **12** was transformed into cascade precursor **13** in 4 steps. Lewis acid FeCl_3 catalyzed polyene cyclization provided polycyclic structure **14** in moderate yield with all rings formed and all stereocenters stored correctly. Further transformations led to *celastrol* (**15**) in a total of 28 steps.^[12] (Scheme 1-2)



Transition metal-catalyzed cascade reactions are also useful tools in total synthesis. Early in 1996, Keay et al. reported a short synthesis of (+)-*xestoquinone* (**24**), with a Pd-catalyzed cascade Heck reaction as the key step. They first synthesized fragment **18** in 3 steps from **16** in

high yield. Another fragment, **20**, was also prepared in 3 steps from **19**. Then, **18** and **20** were coupled by a nucleophilic attack to form **21**, which was transformed to cascade precursor **22**. Palladium catalyzed cascade Heck reaction, with *S*-(+)-BINAP as chiral ligand, provided **23** asymmetrically, forming two rings in one step and constructing all carbon skeleton. **23** could produce **24** after hydrogenation and dearomatization.^[13] (Scheme 1-3)

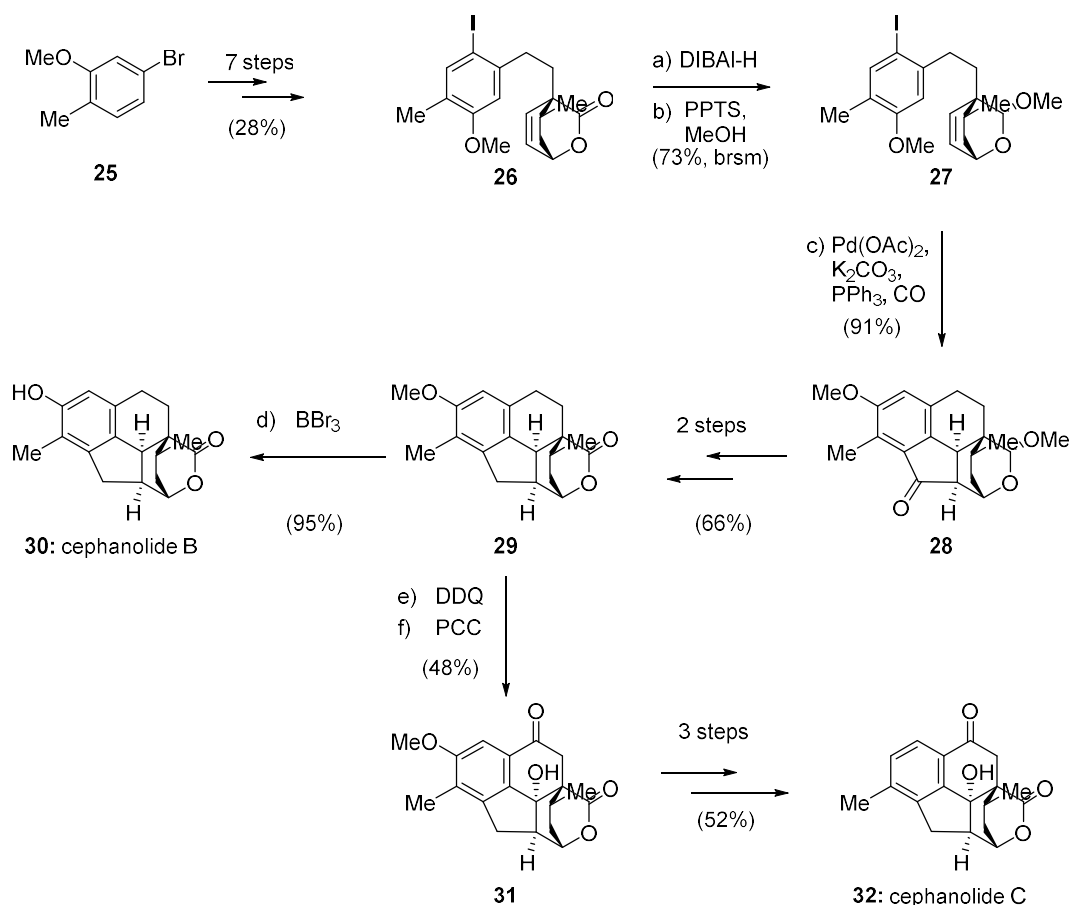
Scheme 1-3: Keay's synthesis of (+)-xestoquinone



Zhao et al. synthesized *cephanolides*, providing another example of a Pd-catalyzed cascade reaction. They synthesized intermediate **26** from **25**, but **26** would give undesired diastereomer in the cascade Heck coupling and carbonylation, so, they transformed **26** into **27** which smoothly produced the desired **28**. **29** could be synthesized from **28** by oxidation and reduction, the deprotection would provide *cephanolide B* (**30**). *Cephanolide C* (**32**) could be synthesized by a

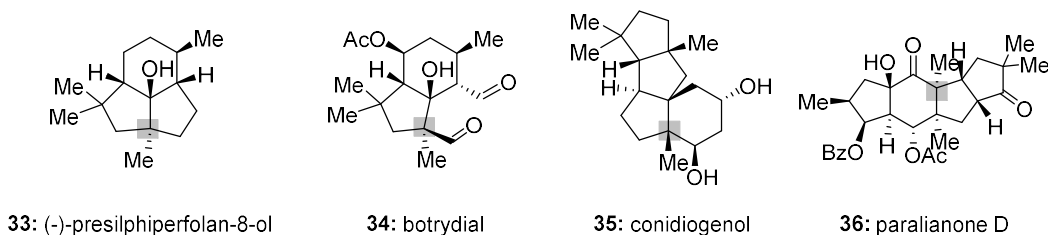
further oxidation/reduction sequence. They also expanded the scope of their cascade reaction to other substrates and showed its potential use in organic synthesis.^[14] (Scheme 1-4)

Scheme 1-4: Zhao's synthesis of *cephanolides*



In our group's former work of *(-)-presilphiperfolan-8-ol* synthesis, to solve the challenge of synthesizing a highly strained trans-5,5 bicyclic structure, a Pd-catalyzed cascade reaction was developed.^[15] We noticed that the stereo-specific Heck reaction would result in an 1,3-*trans* structure, which also presents in other natural products.^[16] (Figure 1-1) For **34-36**, structures of those terpenes also share a quaternary center and one C–O bond next to it. These common features inspired our development of a common solution of syntheses of such structures: a cascade reaction.

Figure 1-1: 1,3-*trans* structure in natural products



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CHAPTER 2

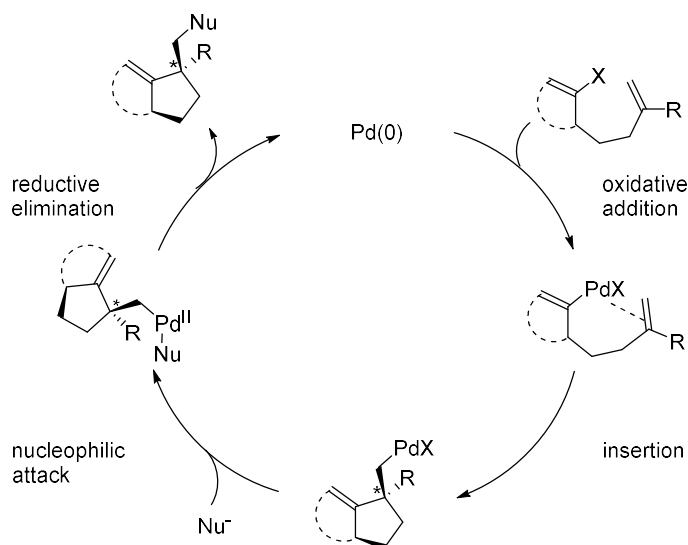
**PALLADIUM CATALYZED CYCLIZATION AND C–O BOND
FORMATION CASCADE**

2.1 Introduction

2.1.1 Heck-type cyclization and nucleophile trapping cascade

Since its original development,^[1] the Heck reaction has proven a powerful tool in organic synthesis.^[2] The catalytic cycle of the Heck reaction ends with β -H elimination of the palladium intermediate and release of the product. When no β -H is available, Grigg et al. reported that in a 5-exo intramolecular cyclization, the addition of extra nucleophile could trap the intermediate and introduce a new C–X bond.^[3] Their proposed cascade mechanism included a nucleophilic attack to the alkyl palladium next to the labeled quaternary center which prevented β -H elimination, followed by reductive elimination (Figure 2-1).

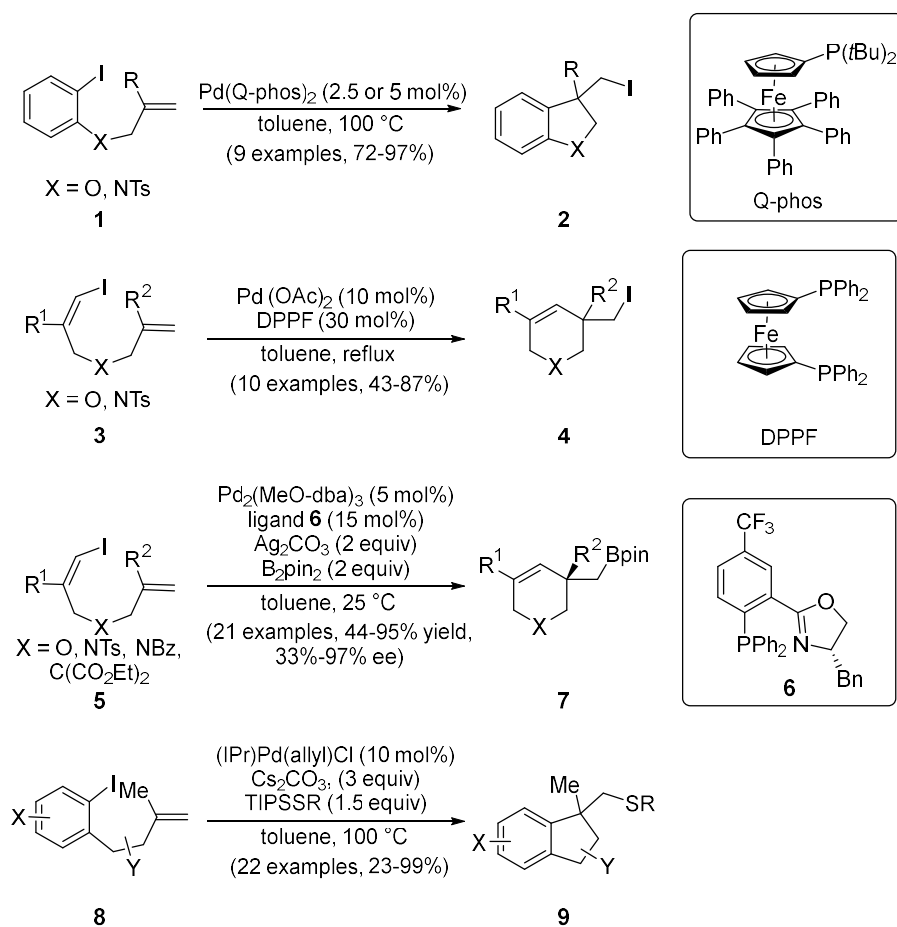
Figure 2-1: Proposed mechanism of nucleophile trapping



This process was widely studied and proved to have various applications.^[4] The reaction generated a quaternary center and was stereospecific. As shown in Figure 2-1, the *cis* insertion guaranteed 1,3-*trans* stereochemistry, which perfectly matched the proposed molecules in **Chapter 1**, thus being a potential general solution of such structures in natural product synthesis.

A large scope of heteroatom nucleophiles has been investigated since Grigg's original discovery with carbon and hydride. Lautens et al. in 2011 reported that, starting from phenyl iodide **1**, with very bulky ligand Q-phos, they could achieve a cyclization and iodination cascade with high yields.^[5a] In the same year, Tong et al. reported a similar process using vinyl iodide **3** instead, with the bident ligand DPPF.^[5b] Later in 2017, Tong et al. succeeded in cyclization and C–B bond formation asymmetrically with chiral ligand **6**, resulting both high yields and enantiomeric excess in most examples.^[6] Recently in 2019, Nakada et al. used the TIPS thioether as nucleophile to form C–S bond in this cascade.^[7] (Scheme 2-1)

Scheme 2-1: Heteroatom nucleophiles in cyclization cascade



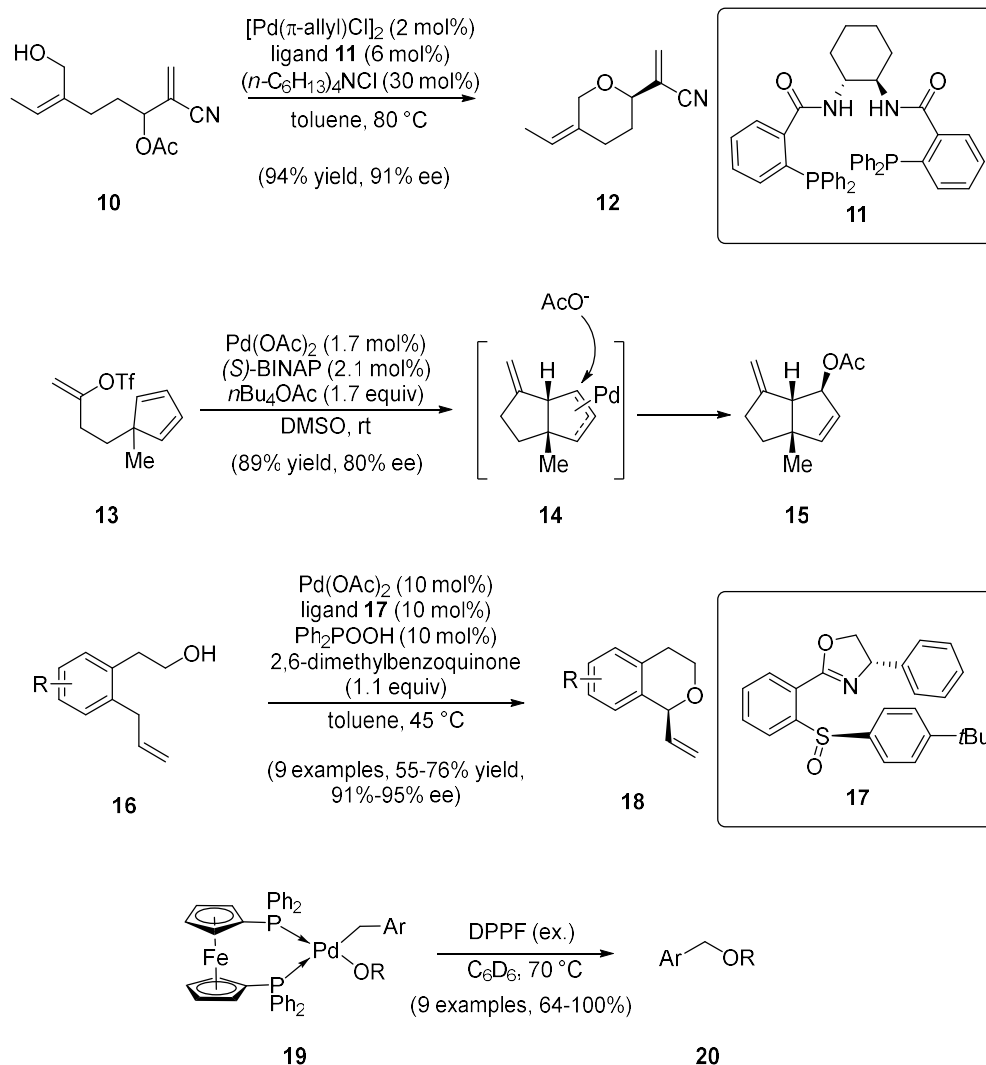
However, C–O bond formation, which is of interest to construct our proposed molecules, was still unprecedented to the best of our knowledge in this cascade. (One close example will be discussed in 2.3.2)

2.1.2 Palladium catalyzed C–O bond formation

Many different C–O bond formation process catalyzed by palladium, with different mechanisms, have been reported.

In contrast to the unactivated alkyl palladium(II) intermediate in cyclization cascade above, activated palladium(II), such as allyl palladium(II), was readily available for a nucleophilic attack to form a C–O bond. The Tsuji-Trost reaction, generating an allyl palladium(II) intermediate from allyl acetate, was a famous example. Trost et al. reported that using an intramolecular hydroxyl group as nucleophile, with chiral ligands, the asymmetric cyclic ether **12** could be synthesized from allyl acetate **10** via dynamic kinetic transformation.^[8] In a process by Shibasaki et al. similar with Heck-type cyclization but via an allyl palladium intermediate **14**, a nucleophilic attack by *n*Bu₄NOAc could form an enantioenriched allylic acetate **15**.^[9] The White group did a series of work on allylic oxidation with palladium(II) and sulfoxide ligands (known as the White catalyst), including the synthesis of asymmetric cyclic ether **18** through intramolecular allylic oxidation.^[10] Recently this sulfoxide ligand catalysis was applied in the synthesis of artemisininic alcohols.^[11] Other than allyl palladium(II), Hartwig et al. heated a pre-synthesized benzyl palladium(II) DPPF complex **19** with excess DPPF to trap Pd(0), and also observed C–O bond formation.^[12] (Scheme 2-2)

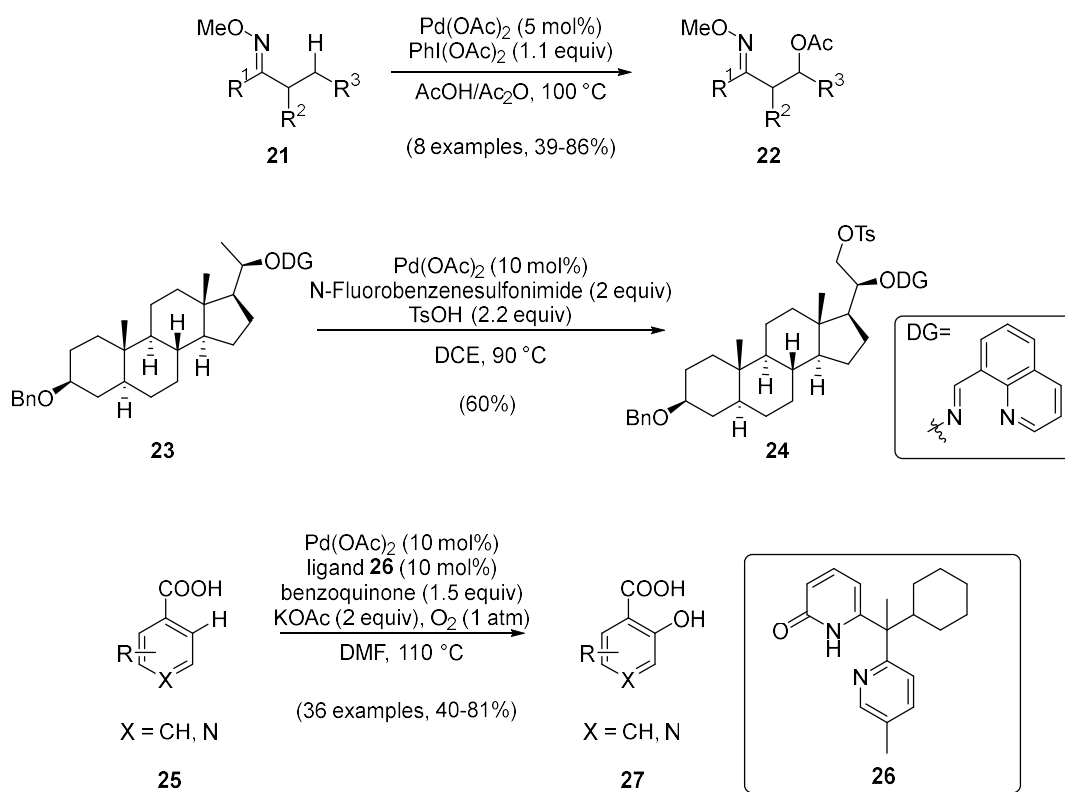
Scheme 2-2: C-O bond formation via activated Pd(II) intermediate



In addition to C–O bond formation via the palladium(II) intermediate, C–O bond formation via palladium(IV) species is also well studied. The formation of these high valence intermediate generally requires the use of an extra oxidant, such as $\text{PhI}(\text{OAc})_2$ or O_2 . Sanford et al. found that oximes were a good directing group, activating its β -position, helping palladium insertion into C–H bond, and resulting in C–H bond oxidation.^[13] Dong et al. expanded this chemistry. They established a methodology to introduce a tosyl group by oxime directed C–H bond oxidation on the α -position of the oxime oxygen. This would work with the pregnanediol derivative **23** and thus

potentially work in late-stage oxidation in total synthesis.^[14] The Yu group did intensive research on C–H bond oxidation, and recently reported an interesting methodology of sp^2 C–H bond oxidation using molecular oxygen. The tautomeric ligand **26** was essential, as its tautomerization between pyridone and hydroxyl pyridine accelerated proton extraction from the substrate and palladium(IV) intermediate formation.^[15] (Scheme 2-3)

Scheme 2-3: C–O bond formation via Pd(IV) intermediate



2.2 Development of methodology

2.2.1 Original design

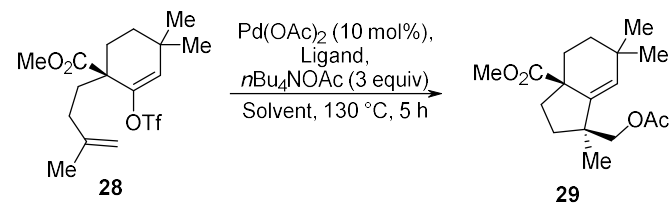
While oxidation via palladium(IV) intermediates are not compatible with Pd(0)/Pd(II) catalytic cycle of Heck-type cyclizations, we proposed a new methodology to achieve the cyclization and C–O bond formation cascade. In terms of oxygen nucleophiles, only nucleophilic attacks on activated allyl or benzyl palladium(II) were known. We deduced that a good nucleophile

would help in the case of our unactivated alkyl palladium(II) intermediate. Thus, we chose the nucleophile $n\text{Bu}_4\text{NOAc}$ used by Shibasaki et al.^[9] as our acetate source and Buchwald's ligands were used^[16] to achieve this transformation.

2.2.2 Optimization of conditions

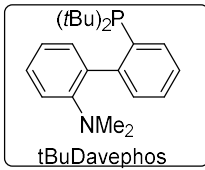
With Dr. Pengfei Hu, we started our investigation on Grigg's vinyl triflate **28**.^[3b] To our delight, with Buchwald's bulky ligand Trixiephos^[16a] and $n\text{Bu}_4\text{NOAc}$ we observed a considerable amount of the desired cascade product, acetate **29** as a single diastereomer (entry 1). In contrast, KOAc as the nucleophile produced only trace amounts of **29** (entry 2), indicating that a suitable nucleophile was essential. When the amount of ligand was reduced to 15 mol%, the yield increased to 83% (entry 3). A decreased equivalence of nucleophiles, however, resulted in yield loss (entry 4). Then we screened other ligands, of which tBuMephos^[16b] proved to be the best, providing a 95% yield (entries 5-8). A decrease in the loading of tBuMephos decreased the yield, so we insisted with 15 mol% (entry 9). Solvent screening showed that product was formed in comparable yields in both EtOAc and xylene, while DMF resulted in worse yields and no desired product was observed in DCE (entries 10-13). It was then found that the temperature could be lowered to 90 °C without a deduction of yield, providing the optimized reaction conditions (entry 14). This set of conditions would serve as our optimized reaction conditions. (Table 2-1)

Table 2-1: Reaction condition screening^a

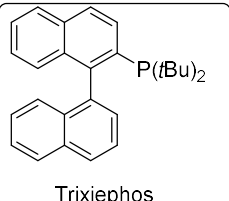


Pd(OAc)₂ (10 mol%),
Ligand,
*n*Bu₄NOAc (3 equiv)
Solvent, 130 °C, 5 h

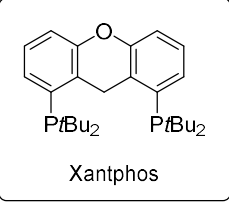
Entry	Ligand	Solvent	NMR Yield(%)
1	Trixiephos (30 mol%)	toluene	61
2 ^b	Trixiephos (30 mol%)	toluene	trace
3	Trixiephos (15 mol%)	toluene	83
4 ^c	Trixiephos (15 mol%)	toluene	61
5	tBuDavephos (15 mol%)	toluene	82
6	Johnphos (15 mol%)	toluene	76
7	Xantphos (15 mol%)	toluene	52
8	tBuMephos (15 mol%)	toluene	95
9	tBuMephos (11 mol%)	toluene	86
10	tBuMephos (15 mol%)	EtOAc	93
11	tBuMephos (15 mol%)	xylene	91
12	tBuMephos (15 mol%)	DMF	73
13	tBuMephos (15 mol%)	DCE	0
14^d	tBuMephos (15 mol%)	toluene	96



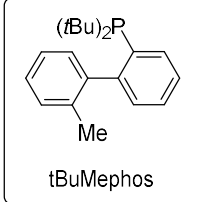
tBuDavephos



Trixiephos



Xantphos



tBuMephos

a. reaction conditions: **28** (0.1 mmol), Pd(OAc)₂ (0.01 mmol), solvent (1 ml), ligands and acetate salt (0.3 mmol) as indicated.

b. KOAc was used instead of *n*Bu₄NOAc;

c. *n*Bu₄NOAc was 1.2 equiv;

d. reaction temperature was 90 °C

2.2.3 Substrate scope

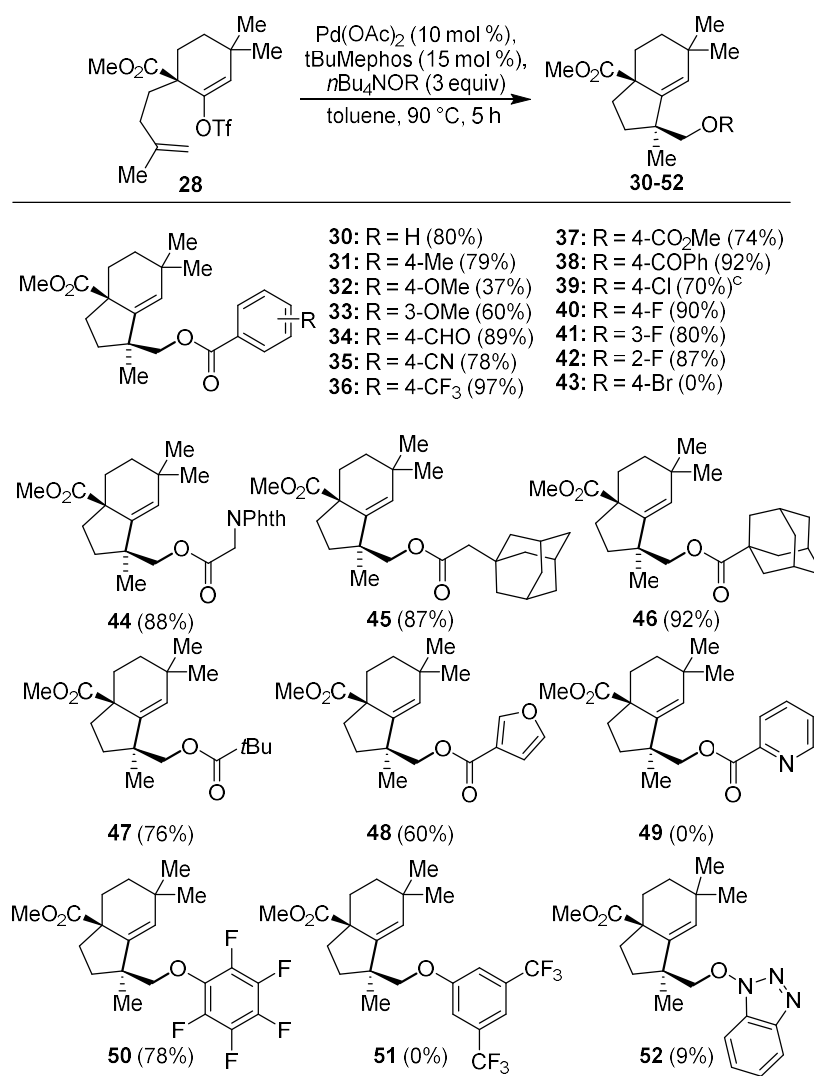
With the optimized condition in hand, we expanded the oxygen nucleophile scope to various tetrabutylammonium salts. While these salts are highly hydroscopic, this reaction calls for anhydrous conditions. Thus, they have to be prepared and crystalized under anhydrous conditions carefully and stored in a dry box (see experiment section for more details). Improper storage of these salts resulted in a dramatic yield loss. The *n*Bu₄NOAc powder, which was exposed to open air for 10 min before setting up a reaction, resulted in a 45-50% yield of **29** with all other conditions unchanged. However, when kept free from moisture, they could be stored for months without a

loss in reactivity. Phenolate salts (see table 2-2, **50, 51**), however, are not stable and should be used freshly.

Many benzoic acid nucleophiles (**30-42**), both electron-rich and electron-poor ones, are compatible with our methodology. Substitutions such as carbonyls (**34, 38**), esters (**37**), cyanide (**35**), fluoride (**40-42**) and chloride (**39**) on the benzene ring are tolerated in this reaction. With a bromide substitution (**43**), however, we observed no desired product. The proposed reason was that bromide was also prone to palladium oxidative addition. Those salts, in most cases, provided good yields of the desired product, higher than 70%. However, the two most electron-rich examples (**32, 33**) only gave moderate yields. -OMe groups at para position (**32**), which were the most electron donating, resulted in the lowest yield. This trend indicated that an electron-poor nucleophile was better for this reaction.

We then investigated other types of nucleophiles. Alkyl acid nucleophiles like protected glycine (**44**), hindered adamantyl acids (**45, 46**), and pivalate (**47**) all reacted smoothly to get high yields. 3-furoic acid (**48**) worked in 60% yield, while picolinic acid (**49**) was not compatible. The proposed reason was that picolinic acid itself was also a strong ligand to palladium^[17] and poisoned the phosphine coordinating catalyst. When phenol was used instead of acid (**50, 51**), only those highly electron deficient, with an acidity in the range of normal carboxyl acid, like pentafluorophenol (**50**) ($pK_a = 5.5$ in H_2O)^[18a], could provide the desired product. No product was observed with 3,5-ditrifluoromethylphenol (**51**) which was not as electron poor, $pK_a = 8.26$ in H_2O .^[18b] This proved consistent with the trend shown with electron poor benzoic acids serving as better nucleophiles. Additionally, acidic *N*-oxide HOBt could also act as nucleophile, though with a very low 9% yield (**52**). (Table 2-2)

Table 2-2: Oxygen nucleophile scope of cyclization cascade.^{a,b}



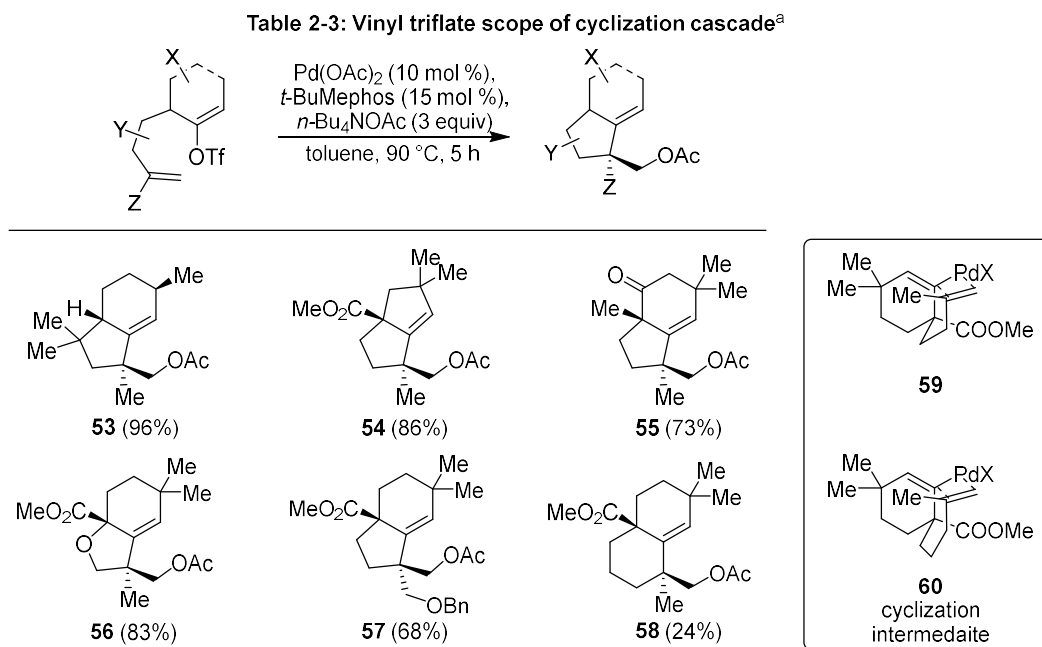
a. reaction conditions unless otherwise noted: **28** (0.1 mmol), Pd(OAc)₂ (0.01 mmol), tBuMephos (0.015 mol), ammonium salt (0.3 mmol), toluene (1 mL), 90 °C, 5 h;

b. isolated yields;

c. reaction temperature was 130 °C.

With the success of the nucleophile scope, we investigated other vinyl triflate substrates. A chiral substrate resulted in a single enantiomer product **53**. Besides 6,5-bicycle, a 5,5-bicyclic structure **54** could also be constructed in high yields. Carbonyl substitution in **55** did not influence the reactivity. A tetrahydrofuran structure **56** could be synthesized from triflate with an O-linked side chain. In **57**, two hydroxymethyl substitutions on the same quaternary center could be differentiated by benzyl and acetate protecting groups. All examples above were 5-exo cyclizations,

but the 6-exo cyclization product **58** could also be synthesized, though with much lower yield, because of a more strained cyclization intermediate **60**.^[19] (Table 2-3)



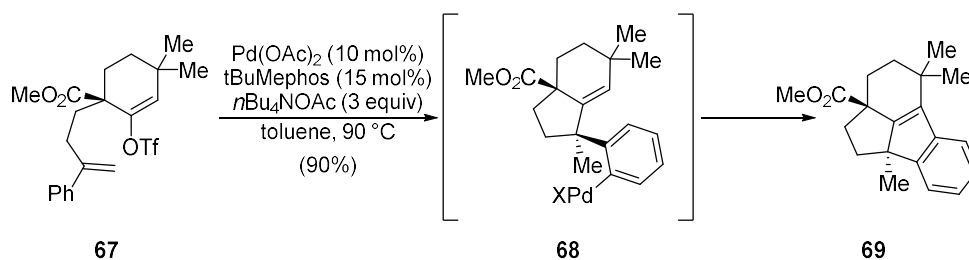
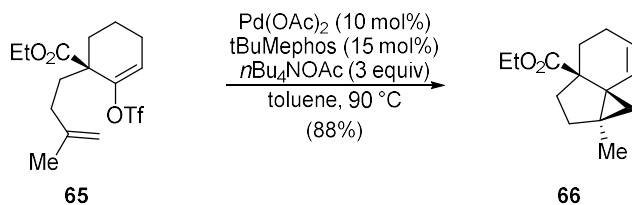
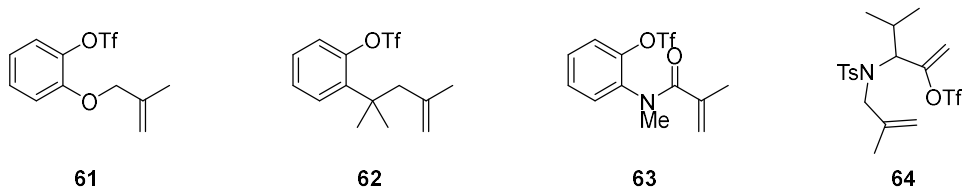
a. reaction conditions: substrate (0.1 mmol), Pd(OAc)₂ (0.01 mmol), tBuMephos (0.015 mol), nBu₄NOAc (0.3 mmol), toluene (1 mL), 90 °C, 5 h.

2.2.4 Substrate limitations

Though we showed the power of this reaction, the methodology also had its limitations. In addition to **43**, **49**, and **51** mentioned above, some other substrates failed in our system. All phenyl triflates and acyclic vinyl triflates we tried, such as **61-64**, failed to form any of the desired product. With **61-63**, only *ipso* substitution of triflate by acetate was observed. **64** resulted in decomposition.

Some substrates formed undesired products. When the gem-dimethyl structure was absent, we did not observe the desired product, rather, a 3-membered ring structure **66**, which was observed by Grigg et al. in a similar system.^[20] Palladium insertion into styrene in **67** resulted in a polycyclic structure **69** instead of our desired product. Activation of benzene C–H bonds by the alkyl palladium(II) intermediate followed by an intramolecular Heck reaction should be responsible. Zhu et al. reported a similar example.^[21] (Table 2-4)

Table 2-4: Failed substrates

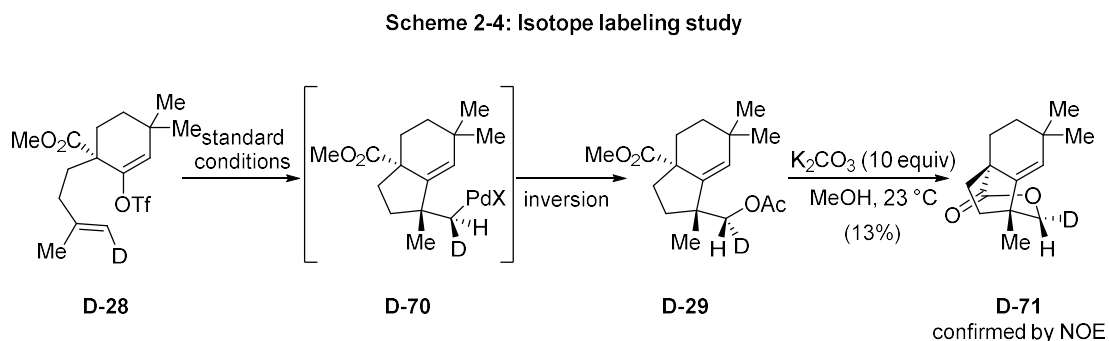


2.3 Mechanism study

2.3.1 Isotope labeling experiment

To illustrate mechanism of the reaction, we first did an isotope labeling experiment. We synthesized a D-labeled vinyl triflate **D-28**.^[22] Under our standard condition, the labeled cyclization product **D-29** was formed smoothly. As we observed a single product, a radical process did not tend to happen. For more stereochemical information, we tried several modifications and found that simply treating **D-29** with excess base could form a lactone **D-71**, though only in low yield. NOE NMR analysis of **D-71** revealed that the remained hydrogen had a NOE interaction with the angular methyl group, so the deuterium atom was found to be *trans* to that methyl, which indicated an inversion from cyclized intermediate **D-70**, where the deuterium atom was *cis* to the

methyl, to cascade product **D-29**. This provided evidence for an S_N2 process in the mechanism. (Scheme 2-4)



2.3.2 Cyclopropane intermediate

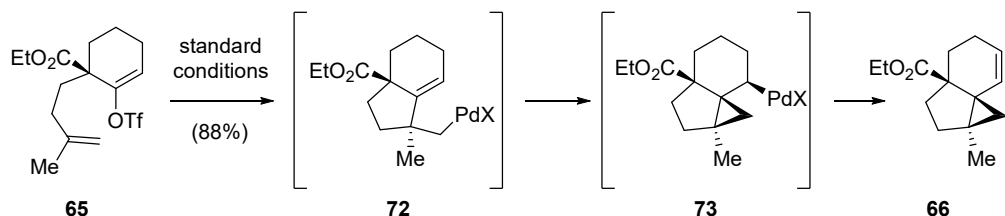
As we confirmed a S_N2 mechanism of this reaction, the detailed mechanism was still of interest. The incompatibility of phenyl triflates in our reaction, while reported possible with halogen and sulfur nucleophiles,^[5a,7] was of particular interest. Grigg et al. proposed a direct nucleophilic attack mechanism (Figure 2-1).^[3] However in this case, C–O reductive elimination with inactivated alkyl palladium(II) would be difficult. As mentioned in 2.1.1, Mulzer et al. observed a similar cyclization reaction on vinyl bromide substrate in the synthetic study of *bielschowskysin*, proposing an acetoxy-palladation mechanism.^[23] However, this was also difficult when oxidative addition was possible, and as shown on **43** (Table 2-2), oxidative addition of bromide might interfere with this cascade. Thus, those proposals were not satisfying.

One hint of the mechanism came from **66** with the 3-membered ring. An intramolecular Heck-type insertion to the double bond in alkyl palladium(II) intermediate **72** formed cyclopropane intermediate **73**. It was followed by β-H elimination, which is the termination of ‘normal’ Heck reactions, to provide **66**.^[20] It was reasonable to propose that the cyclopropane intermediate was general for this type of reaction. This could also explain the necessity of the gem-dimethyl blocking group (or a blocking group *cis* to the palladium, like **53**) to prevent β-H

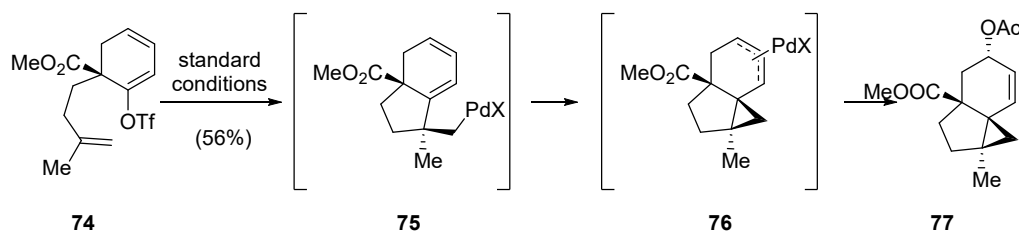
elimination and allow nucleophilic attack. We also noted that the strain of the cyclopropane structure would enhance the reactivity and make it ready to nucleophilic attack and ring opening.^[24]

Scheme 2-5: Cyclopropane intermediate mechanism

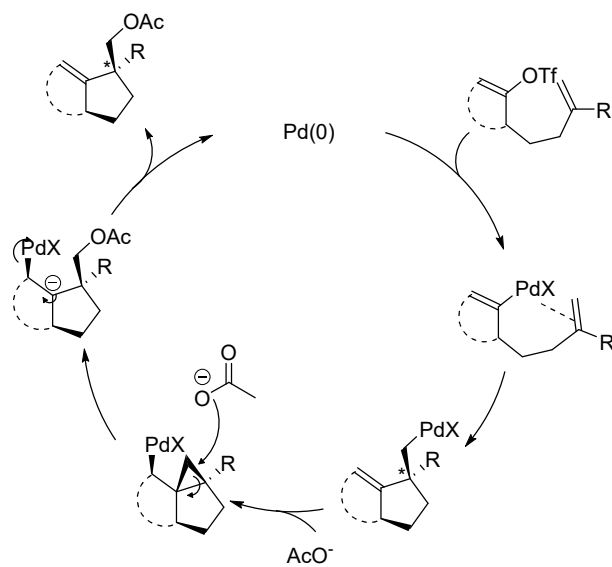
formation of cyclopropane structure:



conjugated vinyl triflate experiment:



proposed catalytic cycle:



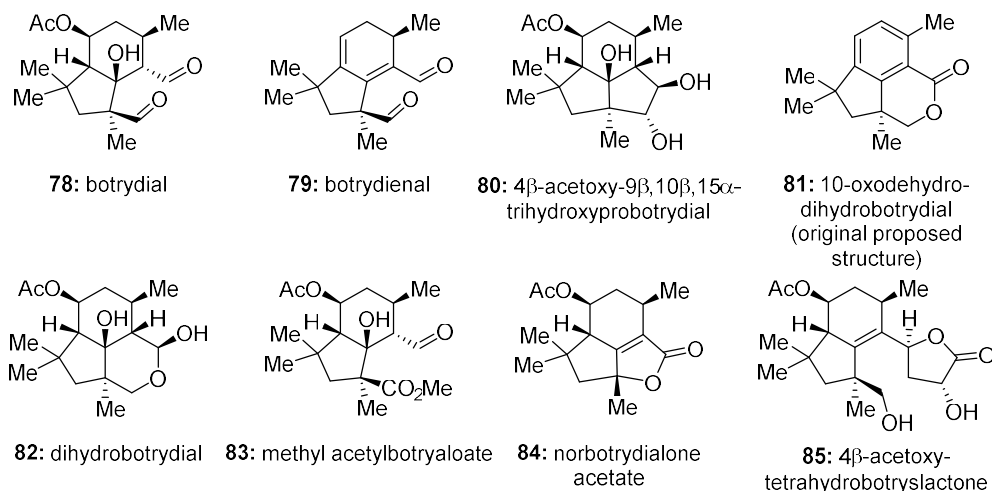
We proposed that the cyclopropane structure was the actual active intermediate. To provide more sound evidence, we synthesized conjugated vinyl triflate **74**. After intramolecular insertion, due to the additional double bond, the cyclopropane intermediate **76** would contain an activated allyl palladium(II) instead of alkyl. If a nucleophilic attack happened on **76**, the allyl palladium(II)

species would be more reactive than cyclopropane ring, which was confirmed by the observed product **77** and the lack of the cascade product. Based on these results, we proposed the revised catalytic cycle of this reaction, with one extra cyclopropane intermediate. The strain of 3-membered ring was essential to allow the oxygen nucleophilic attack. As the elimination did not follow antiperiplanar manner, it was possibly a stepwise process, first ring-opening of the cyclopropane then cleavage of C–Pd bond. The relationship between acidity and reactivity (see 2.2.3) was possibly because that a too strong nucleophile might attack palladium to replace the ligand, rather than the carbon on cyclopropane ring, thus making the palladium a bad leaving group and terminating this process. (Scheme 2-5)

2.4 Synthetic application: botrydial

2.4.1 Introduction

Figure 2-2: Structures of selected members of *botrydial* family



The *botrydial* family were first isolated in 1974 from *Botrytis cinerea*, a useful fungus in wine industry.^[25] They are phytotoxic metabolites, causing ‘noble rot’ in grapes and have been made use of in production of trockenbeerenauslese, an expensive sweet wine, since the 18th century.

Botrydial **79**, which was isolated in 1985,^[26] proved to be highly bioactive, with $ID_{50} = 0.1-1 \mu\text{g/ml}$.^[27]

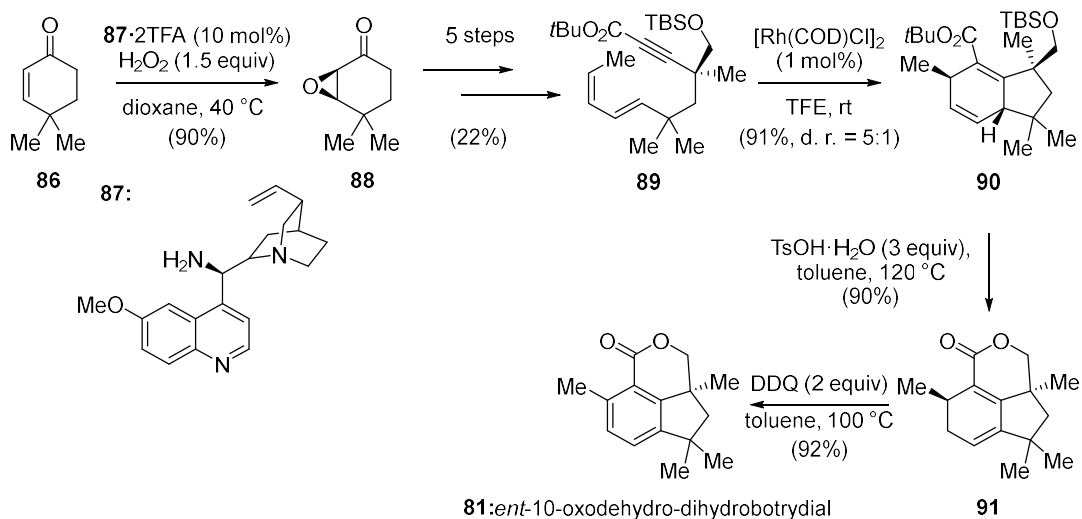
This terpene family, having more than 15 members, share a 6,5-bicyclic skeleton, with different oxidation statuses. Some of them have an extra lactone or acetal ring (**81**, **84**), or a highly strained *trans*-5,5-bicyclic structure (**80**). Many members have continuous stereocenters with oxygen (**78**, **80**, **82**, **83**, **85**) and are challenging targets of synthesis. (Figure 2-2)

Though first isolated decades ago, not many synthetic studies of the *botrydial* family were reported. Collado et al. reported some transformations between family members.^[27] The Li group made significant contributions in this field with their Diels-Alder chemistry. In 2016 they reported the first asymmetric synthesis of proposed structure of *10-oxodehydro-dihydrobotrydial* in 9 steps.^[28] They achieved asymmetric epoxidation of ketone **86** by quinine catalyst **87**. The enantio-rich epoxide **88** was then transferred to compound **89**. Then a Diels-Alder reaction catalyzed by rhodium provided bicyclic **90** stereo-selectively. Then lactonization and double bond migration by acid catalysis followed by oxidative aromatization provide natural product **81**. However, the product they synthesized had reversed optical rotation of the natural product, effectively proving to be an enantiomer of the target. In 2017 in a following study the transferred intermediate **91** to *botrydial* **79** in 2 steps.^[29]

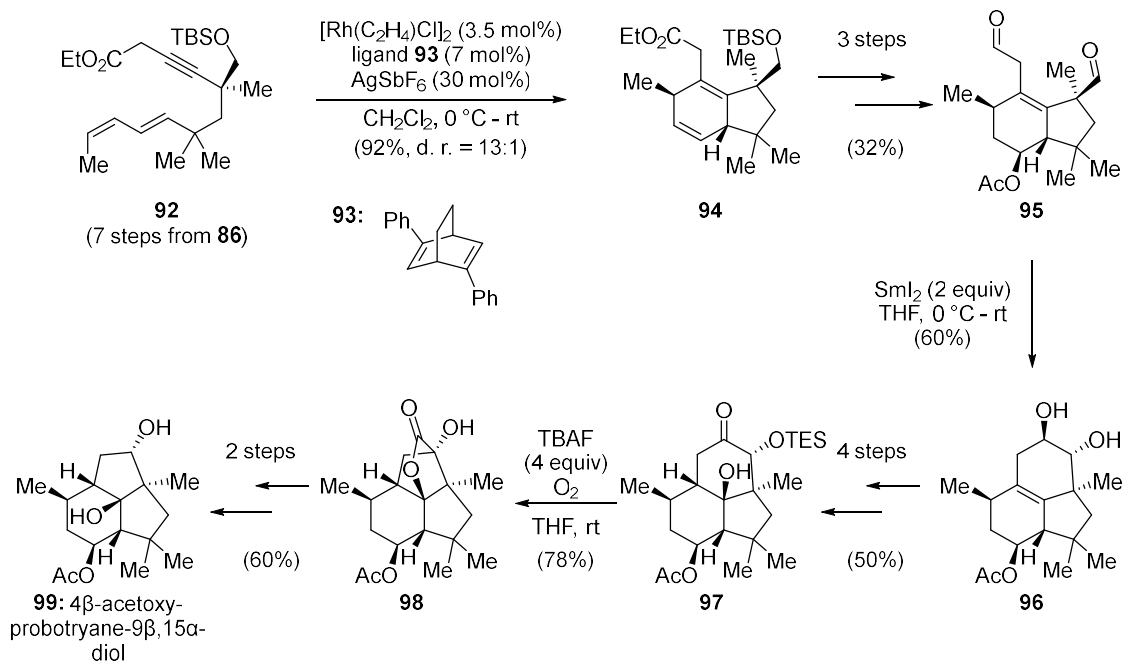
A more strained and challenging product *4 β -acetoxyprobotryane-9 β ,15 α -diol* **99** was also achieved by Li et al.^[30] They synthesized Diels-Alder product **94** in a similar route. This was transferred to dial **95**, followed by SmI_2 reductive coupling to construct the third ring. After further oxidation and protection, oxidative ring contraction led to strained *trans*-5,5-bicyclic structure **98**. Finally oxidative cleavage of the lactone and reduction produced natural product **99**. (Scheme 2-6)

Scheme 2-6: Li's synthesis of *botrydial* family

asymmetric total synthesis of proposed structure of *10-oxodehydro-dihydrobotrydial*:



asymmetric total synthesis of *4β-acetoxyprobotryane-9β,15α-diol*:

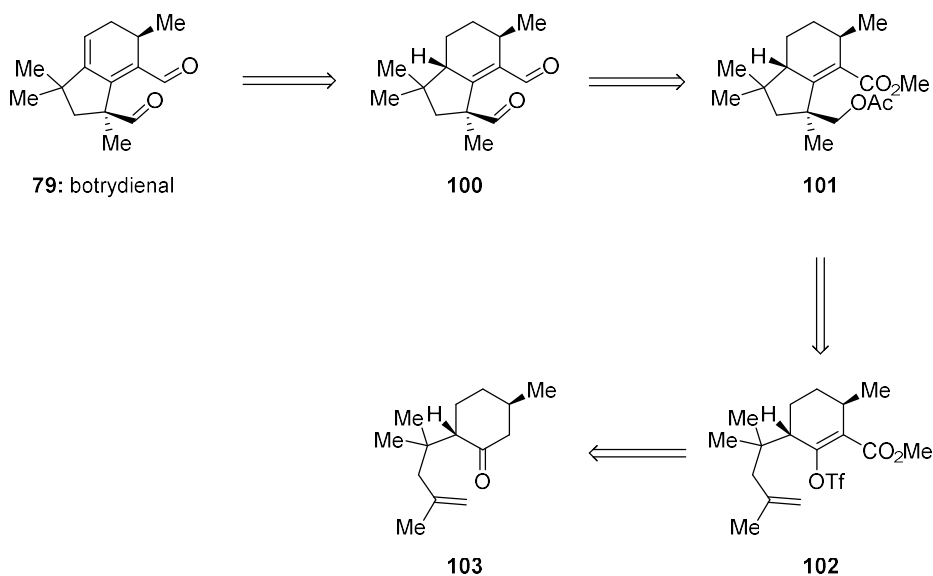


Li's total synthesis based on intramolecular Diels-Alder reactions and ring contraction was impressive. However, the synthesis of Diels-Alder precursor was not concise. We proposed that our cascade reaction would be an alternative and effective method to build up the bicyclic structure.

2.4.2 Overall design of synthetic route

To demonstrate the power of our cascade methodology we planned to achieve the synthesis of *botrydial* (**79**). We proposed that the 5-membered ring and C–O bond could be built with our cascade chemistry in one step stereo-selectively. As shown in scheme 2-7, we proposed that the conjugated diene in our target **79** could be introduced in a later stage. The intermediate **100** can be traced back to **101** after oxidation status management. **101** could be synthesized by our cascade reaction from triflate **102**, which could be easily prepared from known chiral compound **103**.^[31]

Scheme 2-7: Proposed synthetic route of *botrydial*



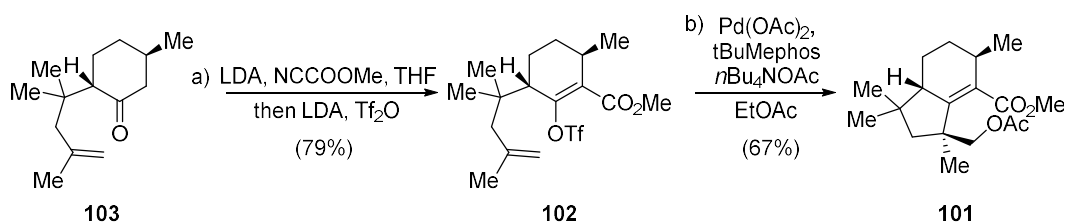
2.4.3 Route to cyclization cascade and aromatization problem

Our synthesis started with known compound **103**, which was prepared from commercial (+)-*pulegone* in one step by Sakurai reaction with 50% yield.^[31b] A one-pot α -acylation and triflation of **103** resulted in triflate **102**. This one-pot process produced better yields than the isolated two-step procedure, which exhibited a yield of less than 50%. Our optimized condition of methodology in toluene worked for this substrate to provide cascade product **101**, however with

only 35% yield. After additional screening we found EtOAc (also good for the methodology, see Table 2-1, entry 10) was the optimized solvent for this specific reaction, resulting in 67% yield.

(Scheme 2-8)

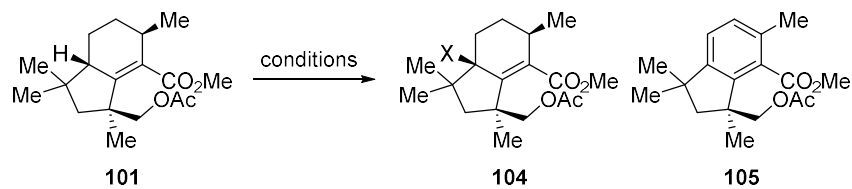
Scheme 2-8: Synthesis towards cyclization cascade^a



a. reaction conditions: (a) LDA (3 equiv), THF, -78 °C - 0 °C, 90 min, then NCCOOMe (3.3 equiv), HMPA (3 equiv), -78 °C, 1 h, then LDA (10 equiv), Tf₂O (10 equiv), -78 °C, 4 h; (b) Pd(OAc)₂ (10 mol%), tBuMephos (15 mol%), nBu₄NOAc (3 equiv), EtOAc, 90 °C, 12 h.

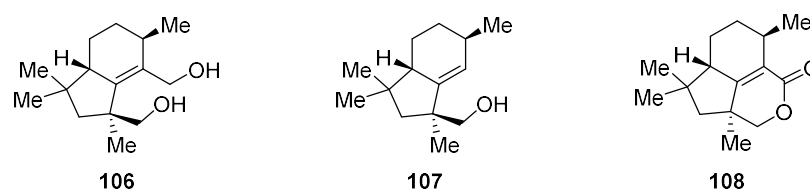
Though the cyclization worked smoothly, the following step proved to be challenging. Li et al. showed that their intermediate **91** was readily susceptible to aromatization,^[28a] which was also true of intermediate **101**. We tried various conditions, including radical substitution, allylic oxidation, and epoxidation, but the desired allylic functionalization was not successful. Only aromatization product was observed. We also tried other substrates, such as free diol **106**, mono alcohol **107** and lactone **108**, but all these reactants led to undesired aromatization. However, we made use of this aromatization of **108** to synthesis *10-oxodehydro-dihydrobotrydial*. Treatment of **101** with excess base formed lactone **108**, followed by aromatization, providing **81**. Optical data of our sample was consistent with Li's^[28a] leading us to believe that this was the enantiomer of the natural product isolated by Collado et al..^[28b] (Scheme 2-9)

Scheme 2-9: Aromatization and synthesis of 10-oxodehydro-dihydrobotrydial

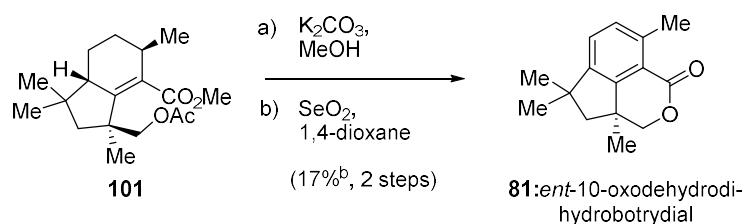


Entry	Conditions	Results
1	NBS, CCl ₄	105 only
2	Br ₂ , <i>hν</i> , CCl ₄	105 only
3	SeO ₂ , 1,4-dioxane	105 only
4	TBHP, <i>n</i> BuLi, THF	no reaction
5	<i>m</i> CPBA, K ₂ CO ₃ , CH ₂ Cl ₂	no reaction

Other failed substrates:



synthesis of 10-oxodehydro-dihydrobotrydial:^a



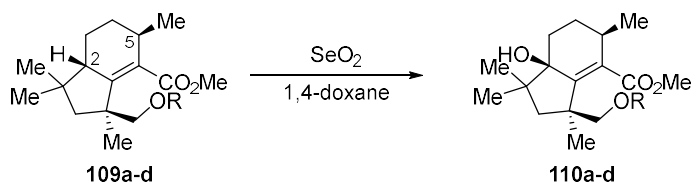
a. reaction conditions: (a) K₂CO₃ (10 equiv), MeOH, 23 °C, 10 h; (b) SeO₂ (4 equiv), 1,4-dioxane, reflux, 12 h.
b. 32% **108** was recovered.

2.4.4 Final steps towards botrydial

After extensive screening, we finally found that a bulky protection group of the hydroxyl, instead of acetate, could prevent aromatization. TBS protected **109c** (easily prepared from **101** by basic alcoholysis and TBS protection, see Scheme 2-10) was found to be the best, resulting in 49% yield with SeO₂ oxidation.^[32] (Table 2-5) We proposed that aromatization was a 2-step process, consisting of oxidation of both allylic positions (C-2 and C-5 in **109**) and double elimination. The

hindered TBS group, though remote, could mask the undesired allylic position at C-5 position and thus provided the desired mono oxidized product.

Table 2-5: Protection group and allylic oxidation^a



Substrate	R group	isolated yield of 110
109a	piv	0% ^b
109b	TES	0% ^b
109c	TBS	49% ^c
109d	TIPS	37% ^c

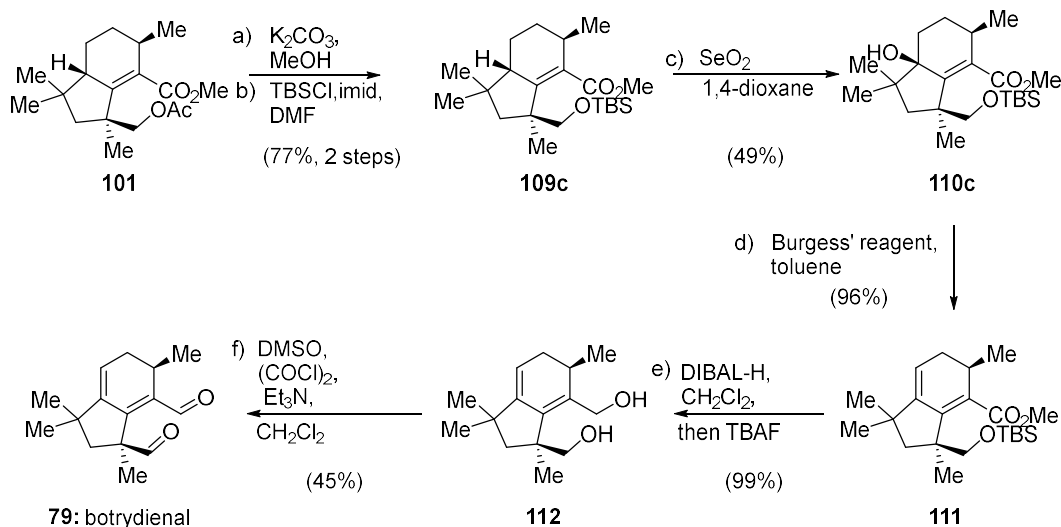
a. reaction conditions: SeO₂ (4 equiv), 1,4-dioxane, reflux, 12 h.

b. only aromatized product was observed.

c. with some aromatized product.

With the allylic oxidation succeeded, we continued the synthesis toward *botrydial*. Allylic hydroxyl in **110c** could be eliminated by Burgess' reagent^[33] to produce the conjugated diene structure in **111**. Then a one-pot DIBAL-H reduction and deprotection of the TBS group provided diol **112** in nearly quantitative yield. DMP oxidation of **112** only resulted in trace amounts of the desired natural product **79**, but Swern oxidation with a high excess of the amount of reagents provided 45% yield of **79**. We finished the total synthesis of *botrydial* in 9 steps from commercial (+)-*pulegone*, with 4.3% yield overall. (Scheme 2-10)

Scheme 2-10: Finish of synthesis of *botrydiena*^a



a. reaction conditions: (a) K_2CO_3 (1.05 equiv), MeOH, 23 °C, 10 h; (b) TBSCl (1.5 equiv), imidazole (1.6 equiv), DMF, 23 °C, 12 h; (c) SeO_2 (4 equiv), 1,4-dioxane, reflux, 12 h; (d) Burgess reagent (3 equiv), toluene, 80 °C, 12 h; (e) DIBAL-H (3 equiv), CH_2Cl_2 , 0 °C, 1 h, then TBAF (3 equiv), 0 °C, 3 h; (f) DMSO (20 equiv), $(\text{COCl})_2$ (10 equiv), CH_2Cl_2 , -78 °C, 3 h, then Et_3N (30 equiv), 30 min.

2.5 Summary and outlook

Palladium catalyzed cyclization and nucleophile capture cascades have been widely investigated. However, C–O formation in such cascades were not well studied, as this via an inactivated alkyl palladium(II) intermediate was challenging. C–O bonds are very useful in organic synthesis for various further transformations. Such a stereospecific C–O bond formation cascade would be a potential solution to a series of natural products with 1,3-*trans* structures. Herein we developed an unprecedented cascade methodology of cyclization and C–O bond formation, via a Pd(0)/Pd(II) catalytic cycle. This reaction was compatible with many substrates with different substitutions, mostly resulting in high yields, and could construct various ring structures.

Mechanism study indicated that the reaction proceeded via a cyclopropane key intermediate which was proposed to be the actual intermediate of such a nucleophile capture cascade, different from former proposals. We also showed the power of our methodology by short

asymmetric synthesis of *botrydial* and *epi-10-oxodehydro-dihydrobotrydial*, in 9 and 5 steps respectively, more concise than former syntheses.

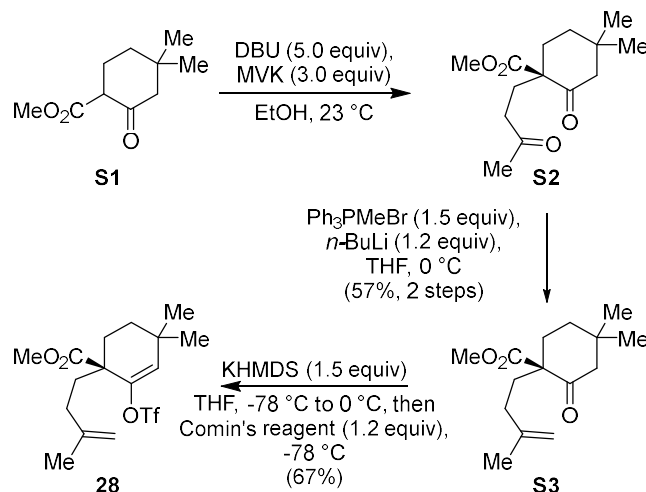
This methodology was also applied in our group's synthesis of *conidiogenones* to construct a 5,5-bicyclic moiety and a C–O bond.^[34] Further studies, such as C–N bond formation cascades with azide salts are now under investigation in our group. We have found that azide nucleophiles could react smoothly to provide C–N bond formation product with the same substrate **28**.

2.6 Experiment section

General Procedures. All reactions were conducted under an argon atmosphere with dry solvents under anhydrous conditions, unless otherwise noted. Dry tetrahydrofuran (THF), toluene, dimethylformamide (DMF), diethyl ether (Et₂O) and dichloromethane (CH₂Cl₂) were obtained by passing commercially available pre-dried, oxygen-free formulations through activated alumina columns. Yields refer to chromatographically and spectroscopically (¹H and ¹³C NMR) homogeneous materials, unless otherwise stated. Reagents were purchased at the highest commercial quality and used without further purification, unless otherwise stated. Reactions were magnetically stirred and monitored by thin-layer chromatography (TLC) carried out on 0.25 mm E. SiliCycle silica gel plates silica gel plates (60F-254) using UV light as visualizing agent, and an ethanolic solution of phosphomolybdic acid and cerium sulfate, and heat as developing agents. SiliCycle silica gel (60, academic grade, particle size 0.040–0.063 mm) was used for flash column chromatography. Preparative thin-layer chromatography separations were carried out on 0.50 mm E. Merck silica gel plates (60F-254). NMR spectra were recorded on Bruker 500 MHz and 400 MHz instruments and calibrated using residual undeuterated solvent as an internal reference ($\delta(\text{CDCl}_3) = 7.26$, $\delta(\text{DMSO-d}_6) = 2.50$ for ¹H NMR and $\delta(\text{CDCl}_3) = 77.0$, $\delta(\text{DMSO-d}_6) = 39.5$ for ¹³C NMR). The following abbreviations were used to explain the multiplicities: s = singlet, d

= doublet, t = triplet, q = quartet, sx = sextet, m = multiplet. IR spectra were recorded on a Perkin-Elmer 1000 series FT-IR spectrometer. High-resolution mass spectra (HRMS) were recorded on an Agilent 6244 ToF-MS using ESI (Electrospray Ionization) at the University of Chicago Mass Spectroscopy Core Facility.

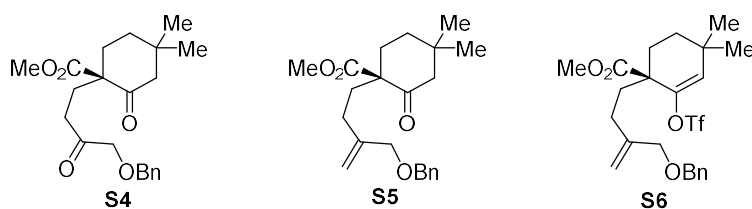
Methodology



Triflate 28. To a solution of **methyl 4,4-dimethyl-2-oxocyclohexane-1-carboxylate (S1)** (1.84 g, 10.0 mmol, 1.0 equiv) in EtOH (50 mL) at 23 °C was added DBU (7.60 g, 50.0 mmol, 5.0 equiv) and methyl vinyl ketone (2.10 g, 30.0 mmol, 3.0 equiv). The resultant solution was then stirred at 23 °C for 3 h. Upon completion, the reaction contents were diluted with EtOAc (30 mL) and quenched by the addition of saturated aqueous NH₄Cl (30 mL). The resultant mixture was poured into a separatory funnel and the layers were separated. The aqueous layer was then extracted with EtOAc (2 × 30 mL). The combined organic layers were washed with H₂O (30 mL) and brine (30 mL) before being dried (Na₂SO₄), filtered, and concentrated. The resultant residue was purified by flash column chromatography (silica gel, hexanes:EtOAc, 5:1) to provide the desired Michael adduct **S2** (2.41 g, 95% yield) as a pale-yellow oil. **S2**: R_f = 0.23 (silica gel, hexanes: EtOAc = 5:1); IR (film) ν_{\max} 3628, 3414, 2956, 1713, 1435, 1369, 1102, 1075, 940, 853

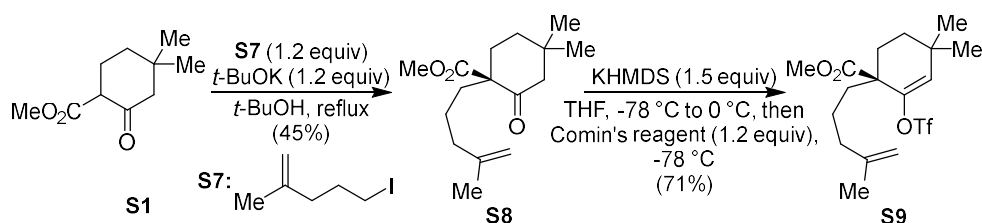
cm⁻¹; ¹H NMR (500 MHz, CDCl₃) δ 3.72 (s, 3 H), 2.56–2.49 (m, 1 H); 2.43–2.39 (m, 1 H), 2.34 (d, *J* = 13.3 Hz, 1 H), 2.18–2.13 (m, 1 H), 2.12 (s, 3 H), 2.11– 2.07 (m, 1 H), 1.89–1.83 (m, 1 H), 1.65–1.49 (m, 2 H), 1.51–1.49 (m, 1 H), 1.02 (s, 3 H), 0.87 (s, 3 H); ¹³C NMR (125 MHz, CDCl₃) δ 207.7, 207.4, 172.4, 58.8, 53.8, 52.5, 38.8, 36.7, 35.6, 31.9, 31.1, 30.0, 28.1, 25.7; HRMS (ESI) calcd for C₁₄H₂₃O₄⁺ [*M* + *H*⁺] 255.1591, found: 255.1593. Next, to a suspension of Ph₃PCH₃Br (2.68 g, 7.50 mmol, 1.5 equiv) in THF (30 mL) at 0 °C was added *n*-BuLi (2.40 mL, 2.5 M in THF, 6.00 mmol, 1.2 equiv). The resultant mixture was stirred at 0 °C for 15 min and then a solution of the intermediate from above (1.27 g, 5.00 mmol, 1.0 equiv) in THF (20 mL) was added. The resultant mixture was stirred at 0 °C for 2 h. Upon completion, the reaction contents were diluted with EtOAc (30 mL) and quenched by the addition of saturated aqueous NH₄Cl (30 mL). The mixture was warmed to 23 °C, poured into a separatory funnel, and the resultant layers were separated. The aqueous layer was then extracted with EtOAc (2 × 30 mL). The combined organic layers were washed with brine (30 mL) before being dried (Na₂SO₄), filtered, and concentrated. The resultant residue was purified by flash column chromatography (silica gel, hexanes:EtOAc, 50:1) to provide ketone **S3** (0.756 g, 60% yield) as a pale-yellow oil. **S3**: *R*_f = 0.68 (silica gel, hexanes:EtOAc, 5:1); IR (film) *v*_{max} 2956, 2857, 1714, 1650, 1454, 1369, 1252, 1199, 1100, 887 cm⁻¹; ¹H NMR (500 MHz, CDCl₃) δ 4.70 (s, 1 H), 4.68 (s, 1 H), 3.72 (s, 3 H), 2.46 (dt, *J* = 13.6, 3.7 Hz, 1 H), 2.31 (d, *J* = 13.1 Hz, 1 H), 2.15 (dd, *J* = 13.1, 2.4 Hz, 1 H), 2.06–2.00 (m, 1 H), 1.92–1.86 (m, 2 H), 1.72 (s, 3 H), 1.70–1.57 (m, 3 H), 1.51–1.47 (m, 1 H), 1.02 (s, 3 H), 0.87 (s, 3 H); ¹³C NMR (125 MHz, CDCl₃) δ 207.3, 172.3, 145.3, 110.0, 59.4, 53.9, 52.3, 36.7, 35.7, 32.7, 32.2, 31.4, 31.3, 25.5, 22.6; HRMS (ESI) calcd for C₁₅H₂₅O₃⁺ [*M* + *H*⁺] 253.1798, found: 253.1805. Finally, to a solution of ketone **S3** (1.26 g, 5.00 mmol) in THF (40 mL) at –78 °C was added KHMDS (7.50 mL, 1.0 M in THF, 7.50 mmol, 1.5 equiv). The resultant yellow solution was then

stirred at 0 °C for 90 min before being cooled to –78 °C. A solution of Comin’s reagent (2.35 g, 6.00 mmol, 1.2 equiv) in THF (10 mL) was then added. The resultant solution was stirred at –78 °C for 1 h before being diluted by EtOAc (30 mL) and quenched by the addition of saturated aqueous NH₄Cl (30 mL). The mixture was warmed to 0 °C, poured into a separatory funnel, and the resultant layers were separated. The aqueous layer was then extracted with EtOAc (2 × 30 mL). The combined organic layers were washed with brine (30 mL) before being dried (Na₂SO₄), filtered, and concentrated. The resultant residue was purified by flash column chromatography (Et₃N-buffered silica gel, hexanes) to provide triflate **17** (1.29 g, 67% yield) as a colorless oil. **17**: *R_f* = 0.69 (silica gel, hexanes: EtOAc, 10:1); IR (film) ν_{max} 2962, 2869, 1739, 1669, 1651, 1417, 1214, 1144, 1004, 950, 899, 816 cm⁻¹; ¹H NMR (500 MHz, CDCl₃) δ 5.63 (s, 1 H), 4.73 (s, 1 H), 4.67 (s, 1 H), 3.74 (s, 3 H), 2.23–2.18 (m, 1 H), 2.05–1.92 (m, 3 H), 1.87–1.76 (m, 2 H), 1.73 (s, 3 H), 1.50–1.48 (m, 2 H), 1.10 (s, 3 H), 1.06 (s, 3 H); ¹³C NMR (125 MHz, CDCl₃) δ 173.1, 146.8, 144.7, 129.9, 119.3, 117.0, 110.6, 52.3, 50.2, 33.48, 33.46, 33.3, 32.2, 29.9, 29.1, 27.8, 22.3; HRMS (ESI) calcd for C₁₆H₂₄O₅F₃S⁺ [M + H⁺] 385.1291, found: 385.1298.



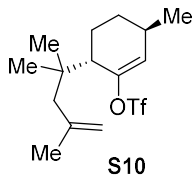
Triflate S6. Prepared as described above starting from **4,4-dimethyl-2-oxocyclohexane-1-carboxylate** (0.552 g, 3.00 mmol, 1.0 equiv) and **1-(benzyloxy)but-3-en-2-one**^[35] (1.58 g, 9.00 mmol, 3.0 equiv) to afford triflate **S6** (0.139 g, 9% overall yield) as a colorless oil. **S4**: *R_f* = 0.29 (silica gel, hexanes:EtOAc, 5:1); IR (film) ν_{max} 3510, 2954, 2866, 1712, 1454, 1248, 1201, 1101 cm⁻¹; ¹H NMR (500 MHz, CDCl₃) δ 7.42–7.30 (m, 5 H), 4.57 (s, 2 H), 4.04 (s, 2 H), 3.70 (s, 3 H), 2.59–2.52 (m, 1 H), 2.45–2.37 (m, 2 H), 2.34 (d, *J* = 13.2 Hz, 1 H), 2.16–2.11 (m, 2 H), 1.92–1.86 (m, 1 H), 1.62–1.49 (m, 3 H), 1.02 (s, 3 H), 0.87 (s, 3 H); ¹³C NMR (125 MHz, CDCl₃) δ 207.6,

207.2, 172.3, 137.2, 128.5, 128.0, 127.9, 74.8, 73.4, 58.8, 53.8, 52.4, 36.7, 35.6, 34.2, 31.8, 31.1, 27.6, 25.6; HRMS (ESI) calcd for $C_{21}H_{29}O_5^+$ [$M + H^+$] 361.2010, found: 361.2009. **S5**: $R_f = 0.50$ (silica gel, hexanes:EtOAc, 10:1); IR (film) ν_{max} 3443, 3030, 2954, 2863, 1713, 1652, 1496, 1368, 1199, 1098, 1028, 904, 698 cm^{-1} ; 1H NMR (500 MHz, $CDCl_3$) δ 7.37–7.28 (m, 5 H), 5.06 (s, 1 H), 4.96 (s, 1 H), 4.48 (s, 2 H), 3.96 (s, 2 H), 3.70 (s, 3 H), 2.58–2.52 (m, 1 H), 2.45–2.37 (m, 2 H), 2.33 (d, $J = 13.2$ Hz, 1 H), 2.16–2.09 (m, 2 H), 1.91–1.85 (m, 1 H), 1.64–1.56 (m, 2 H), 1.50–1.48 (m, 1 H), 1.01 (s, 3 H), 0.86 (s, 3 H); ^{13}C NMR (125 MHz, $CDCl_3$) δ 207.1, 172.2, 145.5, 138.4, 128.3, 128.1, 127.7, 127.5, 73.0, 72.0, 59.4, 53.9, 52.3, 36.7, 35.7, 35.1, 32.7, 31.31, 31.28; HRMS (ESI) calcd for $C_{22}H_{31}O_4^+$ [$M + H^+$] 359.2217, found: 359.2231. **S6**: $R_f = 0.62$ (silica gel, hexanes:EtOAc, 10:1); IR (film) ν_{max} 3032, 2959, 2866, 1738, 1669, 1454, 1416, 1365, 1213, 1144, 1093, 901, 817 cm^{-1} ; 1H NMR (500 MHz, $CDCl_3$) δ 7.37–7.28 (m, 5 H), 5.63 (s, 1 H), 5.08 (s, 1 H), 4.96 (s, 1 H), 4.51–4.46 (m, 2 H), 3.96 (dd, $J = 16.1, 12.5$ Hz, 1 H), 3.72 (s, 3 H), 2.22–2.18 (m, 1 H), 2.12–2.02 (m, 3 H), 1.89–1.83 (m, 1 H), 1.80–1.75 (m, 1 H), 1.51–1.48 (m, 2 H), 1.09 (s, 3 H), 1.05 (s, 3 H); ^{13}C NMR (125 MHz, $CDCl_3$) δ 173.1, 146.7, 145.0, 138.2, 130.1, 128.4, 127.7, 127.6, 122.1, 119.6, 117.0, 114.5, 112.6, 73.0, 72.0, 52.4, 50.2, 33.5, 33.4, 33.3, 29.9, 29.0, 27.8, 27.6; HRMS (ESI) calcd for $C_{23}H_{30}O_6F_3S^+$ [$M + H^+$] 491.1710, found: 491.1694.

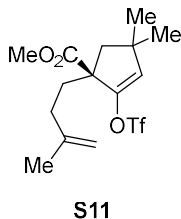


Triflate S9. To *t*-BuOK (2.4 mL, 1.0 M in *t*-BuOH, 2.40 mmol, 1.2 equiv) at 23 °C was added **methyl 4,4-dimethyl-2-oxocyclohexane-1-carboxylate** (0.368 g, 2.00 mmol, 1.0 equiv). The resultant mixture was stirred at 23 °C for 10 min before **5-iodo-2-methylpent-1-ene (S7)**^[36] (0.504 g, 2.40 mmol, 1.2 equiv) was added. The resultant mixture was heated to 85 °C and stirred

at that temperature for 18 h.^[37] Upon completion, the reaction contents were cooled to 23 °C, diluted with EtOAc (10 mL), and quenched by the addition of saturated aqueous NH₄Cl (10 mL). The mixture was poured into a separatory funnel and the layers were separated. The aqueous layer was then extracted with EtOAc (2 × 10 mL). The combined organic layers were washed with brine (10 mL) before being dried (Na₂SO₄), filtered, and concentrated. The resultant residue was purified by flash column chromatography (silica gel, hexanes:EtOAc, 100:1→50:1) to provide ketone **S8** (0.241 g, 45% yield) as a pale-yellow oil. **S8**: R_f = 0.45 (silica gel, hexanes:EtOAc, 10:1); IR (film) ν_{max} 3073, 2956, 2869, 1712, 1649, 1456, 1255, 1199, 1168, 1101, 888, 749 cm⁻¹; ¹H NMR (500 MHz, CDCl₃) δ 4.70 (s, 1 H), 4.66 (s, 1 H), 3.72 (s, 3 H), 2.44 (dt, *J* = 13.7, 3.6 Hz, 1 H), 2.30 (d, *J* = 13.1 Hz, 1 H), 2.14 (dd, *J* = 13.1, 2.3 Hz, 1 H), 2.00 (t, *J* = 7.4 Hz, 1 H), 1.88–1.82 (m, 1 H), 1.69 (s, 3 H), 1.65–1.46 (m, 4 H) 1.36–1.30 (m, 1 H), 1.01 (s, 3 H), 0.86 (s, 3 H); ¹³C NMR (125 MHz, CDCl₃) δ 207.3, 172.5, 145.2, 110.2, 59.6, 53.9, 52.2, 37.9, 36.7, 35.7, 34.0, 31.4, 31.3, 25.5, 22.2, 22.1; HRMS (ESI) calcd for C₁₆H₂₇O₃⁺ [M + H⁺] 267.1955, found: 267.1955. Pressing forward, triflate **S9** was prepared as described above starting from **S8** (0.084 g, 0.31 mmol, 1.0 equiv) to afford triflate **S9** (0.089 g, 71% yield) as a colorless oil. **S9**: R_f = 0.58 (silica gel, hexanes:EtOAc, 10:1); IR (film) ν_{max} 2960, 2890, 1739, 1670, 1651, 1417, 1212, 1144, 1005, 900, 816 cm⁻¹; ¹H NMR (500 MHz, CDCl₃) δ 5.60 (s, 1 H), 4.71 (s, 1 H), 4.66 (s, 1 H), 3.72 (s, 3 H), 2.22–2.17 (m, 1 H), 2.06–1.97 (m, 2 H), 1.86–1.64 (m, 3 H), 1.68 (s, 3 H), 1.49–1.38 (m, 4 H), 1.09 (s, 3 H), 1.05 (s, 3 H); ¹³C NMR (125 MHz, CDCl₃) δ 173.3, 147.0, 144.9, 129.7, 119.6, 117.0, 110.5, 52.3, 50.2, 37.8, 34.6, 33.5, 33.4, 29.9, 29.3, 27.8, 22.1, 22.0; HRMS (ESI) calcd for C₁₇H₂₆O₅F₃S⁺ [M + H⁺] 399.1448, found: 399.1454.

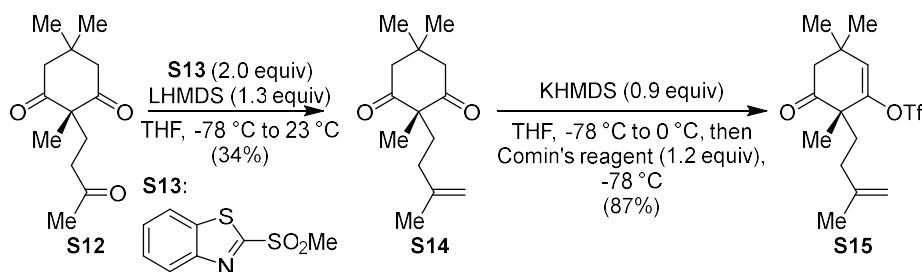


Triflate S10. Prepared as described above starting from **103** (0.694 g, 3.33 mmol, 1.0 equiv) to afford triflate **S10** (0.985 g, 87% yield) as a colorless oil. **S10**: $R_f = 0.48$ (silica gel, hexanes); $[\alpha]_D^{20} = +57.0^\circ$ ($c = 0.53$ in CHCl_3); IR (film) ν_{max} 2963, 2874, 1666, 1643, 1417, 1209, 1144, 969, 893 cm^{-1} ; ^1H NMR (500 MHz, CDCl_3) δ 5.73 (s, 1 H), 4.89–4.87 (m, 1 H), 4.68–4.67 (m, 1 H), 2.46–2.42 (m, 1 H), 2.32–2.26 (m, 1 H), 2.12 (d, $J = 13.2$ Hz, 1 H), 1.97 (d, $J = 13.3$ Hz, 1 H), 1.98–1.92 (m, 1 H), 1.82–1.77 (m, 1 H), 1.78 (s, 3 H), 1.59–1.52 (m, 1 H), 1.15–1.08 (m, 1 H), 1.05 (s, 3 H), 1.04 (d, $J = 7.1$ Hz, 3 H), 1.00 (s, 3 H); ^{13}C NMR (125 MHz, CDCl_3) δ 151.5, 142.9, 128.2, 121.5, 119.8, 117.2, 114.9, 47.6, 46.6, 37.3, 30.4, 30.1, 26.7, 26.5, 25.6, 25.5, 21.2; HRMS (CI) calcd for $\text{C}_{15}\text{H}_{24}\text{O}_3\text{F}_3\text{S}^+ [\text{M} + \text{H}^+]$ 341.1393, found: 341.1391.



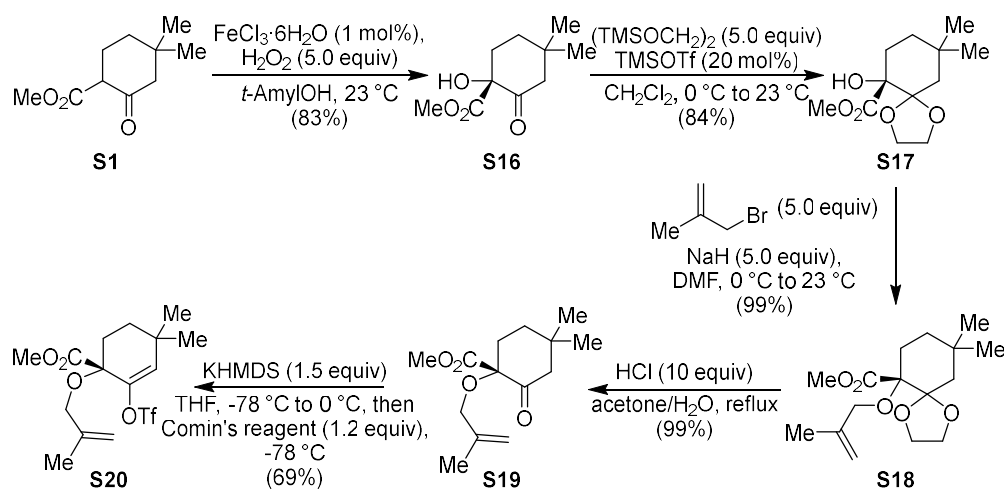
Triflate S11. Prepared as described above starting from **methyl 4,4-dimethyl-2-oxocyclopentane-1-carboxylate**^[38] (0.200 g, 1.17 mmol, 1.0 equiv) and **5-iodo-2-methylpent-1-ene** (0.275 g, 1.40 mmol, 1.2 equiv) to afford triflate **S11** (0.298 g, 69% yield) as a pale-yellow oil. **S11**: $R_f = 0.58$ (silica gel, hexanes:EtOAc, 10:1); IR (film) ν_{max} 2960, 2870, 1740, 1654, 1424, 1366, 1249, 1215, 1142, 1047, 995, 842, 805, 762, 602 cm^{-1} ; ^1H NMR (500 MHz, CDCl_3) δ 5.60 (s, 1 H), 4.73 (s, 1 H), 4.69 (s, 1 H), 3.74 (s, 3 H), 2.44 (d, $J = 13.5$ Hz, 1H), 2.10–2.04 (m, 1 H), 1.95–1.91 (m, 2 H), 1.78–1.73 (m, 1 H), 1.74 (d, $J = 13.5$ Hz, 1 H), 1.73 (s, 3 H), 1.17 (s, 3 H), 1.14 (s, 3 H); ^{13}C NMR (125 MHz, CDCl_3) δ 173.5, 145.9, 144.5, 127.2, 122.2, 119.7, 117.2, 114.6,

110.5, 57.9, 52.4, 45.9, 40.0, 34.1, 32.6, 29.9, 28.9, 22.4; HRMS (ESI) calcd for $C_{15}H_{22}O_5F_3S^+ [M + H^+]$ 371.1135, found: 371.1141.



Triflate S15. To a solution of **2-methanesulfonylbenzothiazole (S13)** (1.28 g, 6.00 mmol, 2.0 equiv) and **2,5,5-trimethyl-2-(3-oxobutyl)cyclohexane-1,3-dione (S12)**^[39] (0.672 g, 3.00 mmol, 1.0 equiv) in THF (30 mL) at $-78\text{ }^\circ\text{C}$ was added LHMDS (3.90 mL, 1.0 M in THF, 3.90 mmol, 1.3 equiv). The resultant mixture was stirred at $-78\text{ }^\circ\text{C}$ for 3 h, and then the mixture was warmed to $23\text{ }^\circ\text{C}$ and stirred for an additional 1 h. Upon completion, the reaction mixture was diluted with EtOAc (20 mL), cooled to $0\text{ }^\circ\text{C}$, quenched by the addition of saturated aqueous NH_4Cl (20 mL), and warmed to $23\text{ }^\circ\text{C}$. The resultant mixture was then poured into a separatory funnel and the layers were separated. The aqueous layer was extracted with EtOAc ($2 \times 30\text{ mL}$). The combined organic layers were washed with brine (30 mL) before being dried (Na_2SO_4), filtered, and concentrated. The resultant residue was purified by flash column chromatography (silica gel, hexanes:EtOAc, 20:1 \rightarrow 10:1) to provide ketone **S14** (0.226 g, 34% yield) as a yellow oil. [Note: **S12** was not stable under strong basic conditions, thus attempts at a Wittig reaction variant gave only trace product]. **S14**: $R_f = 0.47$ (silica gel, hexanes:EtOAc, 4:1); IR (film) ν_{max} 2955, 2871, 1727, 1693, 1650, 1456, 1373, 1331, 1254, 1070, 955, 889 cm^{-1} ; ^1H NMR (500 MHz, CDCl_3) δ 4.69 (s, 1 H), 4.63 (s, 1 H), 2.70 (d, $J = 14.5\text{ Hz}$, 2 H), 2.46 (d, $J = 14.5\text{ Hz}$, 2 H), 1.87–1.80 (m, 4 H), 1.67 (s, 3 H), 1.22 (s, 3 H), 1.06 (s, 3 H), 0.87 (s, 3 H); ^{13}C NMR (125 MHz, CDCl_3) δ 209.8, 144.6, 110.4, 64.4, 51.4, 35.9, 32.2, 30.6, 29.4, 27.6, 22.5, 17.9; HRMS (ESI) calcd for $C_{14}H_{23}O_2^+$

[M + H⁺] 233.1693, found: 233.1699. Pressing forward, triflate **S15** was prepared as described above starting from **S15** (0.044 g, 0.20 mmol, 1.0 equiv) and using KHMDS (0.18 mL, 1.0 M in THF, 0.9 equiv) to afford triflate **S15** (0.063 g, 87% yield) as a colorless oil. **S15**: R_f = 0.68 (silica gel, hexanes: EtOAc, 10:1); IR (film) ν_{max} 2966, 2937, 1724, 1673, 1416, 1245, 1214, 1142, 1001, 887, 797 cm⁻¹; ¹H NMR (500 MHz, CDCl₃) δ 5.88 (s, 1 H), 4.70 (s, 1 H), 4.66 (s, 1 H), 2.52 (d, *J* = 14.0 Hz, 2 H), 2.37 (d, *J* = 13.9 Hz, 2 H), 2.04–1.98 (m, 1 H), 1.82–1.78 (m, 2 H), 1.69 (s, 3 H), 1.66–1.61 (m, 1 H), 1.30 (s, 3 H), 1.16 (s, 3 H), 1.13 (s, 3 H); ¹³C NMR (125 MHz, CDCl₃) δ 208.2, 149.1, 144.5, 126.7, 119.6, 117.0, 110.5, 51.6, 50.9, 34.0, 33.1, 33.0, 29.8, 29.3, 23.4, 22.3; HRMS (ESI) calcd for C₁₅H₂₂O₄F₃S⁺ [M + H⁺] 355.1185, found: 355.1194.



Triflate S20. **S16** (0.423 g, 83% yield b.r.s.m.) was prepared as a pale yellow oil following the reported procedure starting from **S1** (0.736 g, 4.00 mmol, 1.0 equiv).^[40] **S16**: R_f = 0.26 (silica gel, hexanes:EtOAc, 5: 1); IR (film) ν_{max} 3459, 2958, 2870, 1720, 1456, 1436, 1370, 1289, 1258, 1200, 1164, 1123, 1099, 1039, 1006, 978, 796; ¹H NMR (500 MHz, CDCl₃) δ 4.31 (s, 1 H), 3.79 (s, 3 H), 2.60–2.56 (m, 1 H), 2.48–2.39 (m, 2 H), 1.83–1.79 (m, 1 H), 1.63–1.60 (m, 1H), 1.08 (s, 3 H), 0.93 (s, 3 H); ¹³C NMR (125 MHz, CDCl₃) δ 207.0, 170.5, 79.9, 53.0, 51.4, 36.9, 35.1, 33.4, 30.6, 26.1; HRMS (ESI) calcd for C₁₀H₁₅O₃⁺ [M + H⁺ – H₂O] 183.1016, found: 183.1022. Next,

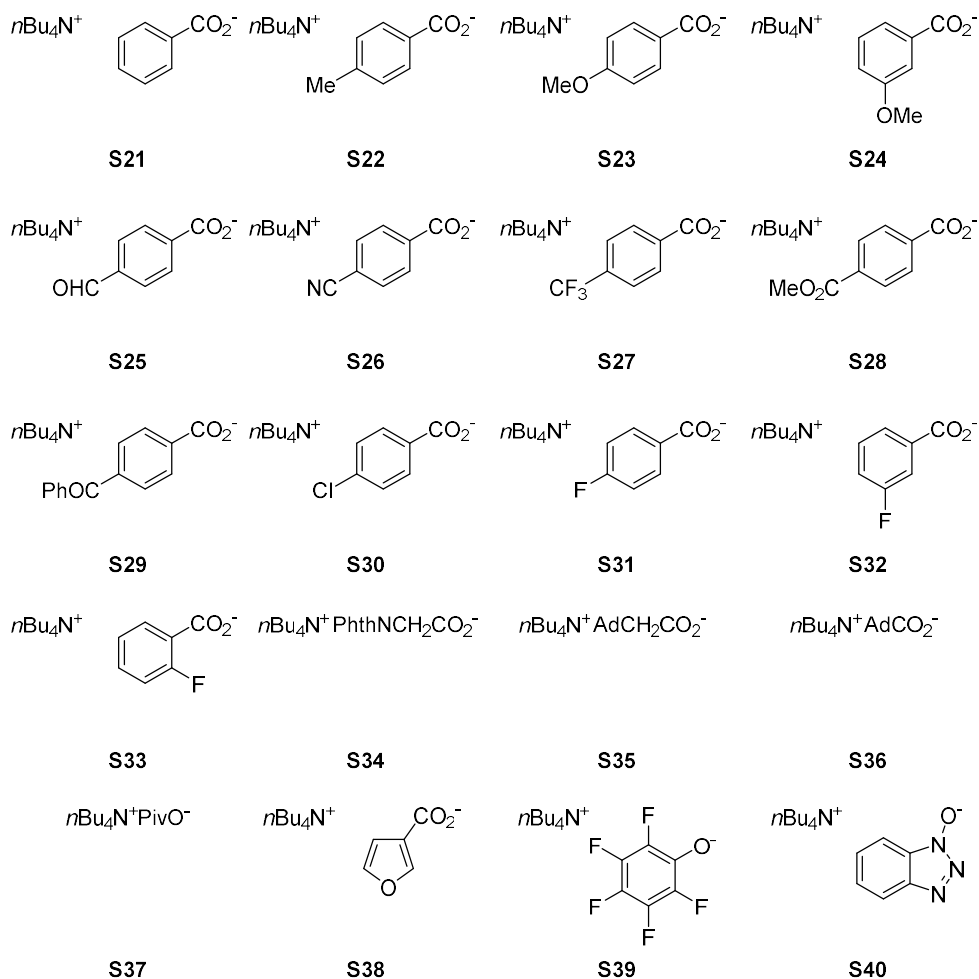
to a solution of **S16** (0.200 g, 1.00 mmol, 1.0 equiv) in CH₂Cl₂ (10 ml) at 0 °C was added 1,2-bis(trimethylsiloxy)ethane (0.618 g, 5.00 mmol, 5.0 equiv) and TMSOTf (44.0 mg, 0.20 mmol, 20 mol %). The mixture was then stirred at 23 °C for 20 h. Upon completion, the reaction contents were quenched by the addition of saturated aqueous NaHCO₃ (10 mL). The mixture was poured into a separatory funnel and the layers were separated. The aqueous layer was then extracted with CH₂Cl₂ (2 × 10 mL). The combined organic layers were dried (Na₂SO₄), filtered, and concentrated. The resultant residue was purified by flash column chromatography (hexanes:EtOAc, 10:1→5:1) to provide ketal **S17** (0.205 g, 84% yield) as a white solid. **S17**: R_f = 0.22 (silica gel, hexanes:EtOAc, 5:1); IR (film) ν_{max} 3505, 2952, 1732, 1449, 1388, 1264, 1217, 1168, 1074, 1043, 988, 826, 614; ¹H NMR (500 MHz, CDCl₃) δ 3.94–3.89 (m, 3 H), 3.78–3.74 (m, 1 H), 3.77 (s, 3 H), 3.38 (d, *J* = 1.7 Hz, 1 H), 2.29–2.22 (m, 1 H), 1.78–1.74 (m, 2 H), 1.58–1.52 (m, 1 H), 1.45 (dd, *J* = 13.8, 1.4 Hz, 1 H), 1.32 (dtd, *J* = 13.6, 4.8, 1.3 Hz, 1 H), 1.04 (s, 3 H), 0.96 (s, 3 H); ¹³C NMR (125 MHz, CDCl₃) δ 174.3, 110.1, 76.9, 65.8, 64.7, 52.8, 43.7, 32.9, 32.0, 31.2, 28.7, 26.7; HRMS (ESI) calcd for C₁₂H₁₉O₄⁺ [M + H⁺ – H₂O] 227.1278, found: 227.1281 Pressing forward, to a suspension of NaH (0.200 g, 60% dispersion in mineral oil, 5.00 mmol, 5.0 equiv) in DMF (5 ml) at 0 °C was added a solution of ketal **S17** (0.244 g, 1.00 mmol, 1.0 equiv) in DMF (5 ml). The resultant mixture was then stirred for 30 min at 0 °C before 3-bromo-2-methylpropene (0.675 g, 5.00 mmol, 5.0 equiv) was added. The reaction contents were then warmed to 23 °C and stirred at that temperature for 2 h. Upon completion, the reaction contents were cooled to 0 °C, diluted with Et₂O (10 ml), and quenched by the slow addition of H₂O. The resultant mixture was poured into a separatory funnel and the layers were separated. The aqueous layer was then extracted with Et₂O (2 × 10 mL). The combined organic layers were washed with H₂O (3 × 10 mL) and brine (10 mL) before being dried (Na₂SO₄), filtered, and concentrated. The resultant residue was purified by flash

column chromatography (silica gel, hexanes:EtOAc, 5:1) to provide ketal **S18** (0.298 g, 99% yield) as a colorless oil. **S18**: $R_f = 0.28$ (silica gel, hexanes: EtOAc = 5:1); IR (film) ν_{\max} 2951, 2913, 1735, 1453, 1254, 1214, 1165, 1083, 1057, 900; ^1H NMR (500 MHz, CDCl_3) δ 5.03 (s, 1 H), 4.85 (s, 1 H), 3.96–3.92 (m, 1 H), 3.89–3.85 (m, 2 H), 3.81–3.78 (m, 3 H), 3.75 (s, 3 H), 2.26 (ddd, $J = 15.1, 13.6, 3.8$ Hz, 1 H), 2.01 (dt, $J = 15.2, 3.8$ Hz, 1 H), 1.85 (d, $J = 13.8$ Hz, 1 H), 1.75 (s, 3 H), 1.40 (dd, $J = 13.8, 1.7$ Hz, 1 H), 1.37 (td, $J = 13.5, 3.5$ Hz, 1 H), 1.23 (ttt, $J = 13.5, 5.6, 1.8$ Hz, 1 H), 1.04 (s, 3 H), 0.93 (s, 3 H); ^{13}C NMR (125 MHz, CDCl_3) δ 172.0, 141.5, 111.4, 110.0, 82.4, 67.5, 65.0, 64.9, 52.2, 43.6, 32.9, 32.6, 31.3, 26.5, 24.7, 19.7; HRMS (ESI) calcd for $\text{C}_{16}\text{H}_{27}\text{O}_5^+$ [$\text{M} + \text{H}^+$] 299.1853, found: 299.1870. Next, to a solution of ketal **S18** (0.298 g, 1.00 mmol, 1.0 equiv) in acetone (15 ml) at 23 °C was added HCl (10.0 mL, 1.0 M in H_2O , 10.0 mmol, 10 equiv). The resultant solution was heated to 60 °C for 12 h. Upon completion, the reaction contents were cooled to 23 °C, poured into a separatory funnel, and the resultant layers were separated. The aqueous layer was then extracted with EtOAc (3 × 10 mL). The combined organic layers were washed with brine (10 mL) before being dried (Na_2SO_4), filtered, and concentrated. The resultant residue was purified by flash column chromatography (silica gel, hexanes:EtOAc, 10:1) to provide ketone **S19** (0.254 g, 99% yield) as a colorless oil. Ketone **S19**: $R_f = 0.30$ (silica gel, hexanes: EtOAc = 10:1); IR (film) ν_{\max} 2957, 2928, 2870, 1751, 1724, 1456, 1370, 1298, 1256, 1160, 1127, 1080, 1054, 899; ^1H NMR (500 MHz, CDCl_3) δ 5.03 (s, 1 H), 4.89 (s, 1 H), 4.15 (d, $J = 12.0$ Hz, 1 H), 3.79 (s, 3 H), 3.78 (d, $J = 12.0$ Hz, 1 H), 2.54 (d, $J = 13.2$ Hz, 1 H), 2.34 (ddd, $J = 14.8, 5.6, 4.2$ Hz, 1 H), 2.17 (dd, $J = 13.2, 1.6$ Hz, 1 H), 2.12 (ddd, $J = 14.8, 11.3, 4.3$ Hz, 1 H), 1.83 (ddd, $J = 13.9, 11.2, 4.3$ Hz, 1 H), 1.77 (s, 3 H), 1.36 (dddd, $J = 13.8, 5.5, 4.3, 1.6$ Hz, 1 H), 1.04 (s, 3 H), 0.98 (s, 3 H); ^{13}C NMR (125 MHz, CDCl_3) δ 205.8, 170.6, 141.8, 111.9, 85.4, 69.7, 52.3, 52.2, 36.5, 33.8, 31.3, 30.1, 26.7, 19.6; HRMS (ESI) calcd for $\text{C}_{14}\text{H}_{23}\text{O}_4^+$ [$\text{M} + \text{H}^+$] 255.1591, found:

255.1599. Pressing forward, triflate **S20** was prepared as described above starting from **S19** (0.079 g, 0.31 mmol, 1.0 equiv) to afford triflate **S20** (0.083 g, 69% yield) as a yellow oil. **S20**: $R_f = 0.32$ (silica gel, hexanes:EtOAc, 10:1); IR (film) ν_{\max} 2961, 2870, 1759, 1661, 1454, 1419, 1366, 1211, 1145, 1093, 1056, 1007, 952, 901, 811; $^1\text{H NMR}$ (500 MHz, CDCl_3) δ 5.76 (s, 1 H), 5.03 (s, 1 H), 4.88 (s, 1 H), 4.00 (dd, $J = 28.3, 11.6$ Hz, 2 H), 3.79 (s, 3 H), 2.25 (ddd, $J = 13.8, 11.4, 3.4$ Hz, 1 H), 2.10 (ddd, $J = 13.8, 6.9, 3.4$ Hz, 1 H), 1.78 (s, 3 H), 1.71 (ddd, $J = 13.8, 11.2, 3.4$ Hz, 1 H), 1.54 (ddd, $J = 13.8, 6.9, 3.4$ Hz, 1 H), 1.13 (s, 3 H), 1.12 (s, 3 H); $^{13}\text{C NMR}$ (125 MHz, CDCl_3) δ 170.7, 144.6, 141.9, 123.3, 119.6, 117.0, 112.0, 78.4, 69.6, 52.6, 33.6, 32.0, 30.0, 29.1, 27.6, 19.5; HRMS (ESI) calcd for $\text{C}_{15}\text{H}_{21}\text{O}_6\text{F}_3\text{SNa}^+ [\text{M} + \text{Na}^+]$ 409.0903, found: 409.0907.

Preparation of nucleophilic salts

To a solution of corresponding acid or phenol (5.00 mmol) in anhydrous MeOH (5 mL) was added *n*-Bu₄NOH (5.05 mL, 1.0 M in MeOH, 5.05 mmol, 1.01 equiv) at 23 °C, and the resultant solution was stirred at 23 °C for 1 h. Upon completion, the resultant mixture was co-evaporated with benzene under an Ar atmosphere and the residue was dried over P₂O₅ on vacuum overnight. If the dried residue was a liquid, it was cooled to 0 °C to form a solid (>95% yield). The solid was then recrystallized from minimal EtOAc under Ar atmosphere, then the mother liquid was removed through a cannula, and the residue was dried on vacuum again to provide the anhydrous salt crystal (30–60% yield) used for subsequent reaction. The salts can be kept stable in dry box for months. [Note: the pentafluorophenoate salt (**S39**) was unstable and should be used immediately].



Tetrabutylammonium benzoate (S21). Pale yellow solid; ^1H NMR (500 MHz, CDCl_3) δ 8.10–8.08 (m, 2 H), 7.30–7.28 (m, 3 H), 3.56–2.32 (m, 8 H), 1.66–1.59 (m, 8 H), 1.40 (sx, $J = 7.4$ Hz, 8 H), 0.96 (t, $J = 7.3$ Hz, 12 H); ^{13}C NMR (125 MHz, CDCl_3) δ 170.9, 141.4, 129.4, 128.2, 126.9, 58.5, 23.9, 19.6, 13.5.

Tetrabutylammonium 4-methylbenzoate (S22). Pale yellow solid; ^1H NMR (500 MHz, CDCl_3) δ 7.85 (d, $J = 8.0$ Hz, 2 H), 6.98 (d, $J = 7.8$ Hz, 2 H), 3.04–3.00 (m, 8 H), 2.23 (s, 3 H), 1.41–1.35 (m, 8 H), 1.27–1.19 (m, 8 H), 1.07 (sx, $J = 7.4$ Hz, 8 H), 0.83 (t, $J = 7.3$ Hz, 12 H); ^{13}C NMR (125 MHz, CDCl_3) δ 171.0, 138.3, 137.4, 129.4, 127.8, 58.0, 23.6, 21.1, 19.4, 13.5.

Tetrabutylammonium 4-methoxybenzoate (S23). Pale yellow solid; ^1H NMR (500 MHz, CDCl_3) δ 7.87–7.84 (m, 2 H), 6.61–6.58 (m, 2 H), 3.01–2.97 (m, 8 H), 1.36–1.29 (m, 8 H), 1.16 (sx, $J = 7.4$ Hz, 8 H), 0.74 (t, $J = 7.3$ Hz, 12 H); ^{13}C NMR (125 MHz, CDCl_3) δ 170.6, 160.0, 133.9, 130.9, 112.0, 58.3, 55.0, 23.8, 19.5, 13.5.

Tetrabutylammonium 3-methoxybenzoate (S24). White solid; ^1H NMR (500 MHz, CDCl_3) δ 7.56–7.55 (m, 2 H), 7.09–7.06 (m, 1 H), 6.75–6.73 (m, 1 H), 3.71 (s, 3 H), 3.09–3.06 (m, 8 H), 1.46–1.39 (m, 8 H), 1.25 (sx, $J = 7.3$ Hz, 8 H), 0.83 (t, $J = 7.2$ Hz, 12 H); ^{13}C NMR (125 MHz, CDCl_3) δ 170.9, 158.9, 142.3, 128.0, 122.0, 115.3, 113.9, 58.4, 55.2, 23.8, 19.6, 13.6.

Tetrabutylammonium 4-formylbenzoate (S25). White solid; ^1H NMR (500 MHz, CDCl_3) δ 9.94 (s, 1 H); 8.12 (d, $J = 8.0$ Hz, 2 H), 7.73 (d, $J = 8.0$ Hz, 2 H), 3.20–3.17 (m, 8 H), 1.55–1.49 (m, 8 H), 1.30 (sx, $J = 7.4$ Hz, 8 H), 0.86 (t, $J = 7.4$ Hz, 12 H); ^{13}C NMR (125 MHz, CDCl_3) δ 192.7, 170.1, 146.6, 136.3, 129.8, 129.0, 58.6, 23.9, 19.6, 13.6.

Tetrabutylammonium 4-cyanobenzoate (S26). Yellow solid; ^1H NMR (500 MHz, CDCl_3) δ 7.98 (d, $J = 8.4$ Hz, 2 H), 7.36 (d, $J = 8.4$ Hz, 2 H), 3.09–3.06 (m, 8 H), 1.44–1.37 (m, 8 H), 1.18 (sx, $J = 7.4$ Hz, 8 H), 0.74 (t, $J = 7.3$ Hz, 12 H); ^{13}C NMR (125 MHz, CDCl_3) δ 169.0, 145.6, 131.0, 129.8, 119.6, 111.2, 58.5, 23.8, 19.5, 13.5.

Tetrabutylammonium 4-(trifluoromethyl)benzoate (S27). Pale yellow solid; ^1H NMR (500 MHz, CDCl_3) δ 8.17 (d, $J = 7.9$ Hz, 2 H), 7.52 (d, $J = 8.1$ Hz, 2 H), 3.32–3.30 (m, 8 H), 1.64–1.58 (m, 8 H), 1.41–1.34 (m, 8 H), 1.26 (sx, $J = 7.4$ Hz, 8 H), 0.94 (t, $J = 7.3$ Hz, 12 H); ^{13}C NMR (125 MHz, CDCl_3) δ 169.5, 144.7, 130.0, 129.8, 129.5, 125.6, 124.0, 123.9, 58.4, 23.8, 19.5, 13.4.

Tetrabutylammonium 4-(methoxycarbonyl)benzoate (S28). White solid; ^1H NMR (500 MHz, CDCl_3) δ 8.08 (d, $J = 8.0$ Hz, 2 H), 7.92 (d, $J = 8.0$ Hz, 2 H), 3.85 (s, 3 H), 3.26–3.22

(m, 8 H), 1.58–1.52 (m, 8 H), 1.34 (sx, $J = 7.3$ Hz, 8 H), 0.90 (t, $J = 7.3$ Hz, 12 H); ^{13}C NMR (125 MHz, CDCl_3) δ 170.4, 167.5, 145.6, 129.7, 129.2, 128.6, 58.7, 51.8, 24.0, 19.7, 13.6.

Tetrabutylammonium 4-benzoylbenzoate (S29). Yellow solid; ^1H NMR (500 MHz, CDCl_3) δ 8.05–8.03 (m, 2 H), 7.63–7.61 (m, 2 H), 7.45–7.42 (m, 1 H), 7.34–7.31 (m, 2 H), 3.14–3.11 (m, 8 H), 1.49–1.42 (m, 8 H), 1.24 (sx, $J = 7.4$ Hz, 8 H), 0.80 (t, $J = 7.3$ Hz, 12 H); ^{13}C NMR (125 MHz, CDCl_3) δ 197.1, 170.2, 145.0, 137.8, 137.2, 132.2, 129.8, 129.2, 129.1, 128.1, 58.5, 23.8, 19.6, 13.6.

Tetrabutylammonium 4-chlorobenzoate (S30). Green solid; ^1H NMR (500 MHz, CDCl_3) δ 7.95–7.93 (m, 2 H), 7.16–7.13 (m, 2 H), 3.15–3.12 (m, 8 H), 1.51–1.45 (m, 8 H), 1.28 (sx, $J = 7.4$ Hz, 8 H), 0.86 (t, $J = 7.3$ Hz, 12 H); ^{13}C NMR (125 MHz, CDCl_3) δ 170.2, 139.2, 134.4, 130.9, 127.2, 58.5, 23.9, 19.6, 13.6.

Tetrabutylammonium 4-fluorobenzoate (S31). White solid; ^1H NMR (500 MHz, CDCl_3) δ 8.00–7.97 (m, 2 H), 6.86–6.82 (m, 2 H), 3.13–3.10 (m, 8 H), 1.49–1.42 (m, 8 H), 1.27 (sx, $J = 7.4$ Hz, 8 H), 0.85 (t, $J = 7.3$ Hz, 12 H); ^{13}C NMR (125 MHz, CDCl_3) δ 170.2, 164.3, 162.4, 131.4, 131.3, 125.2, 113.6, 113.5, 58.8, 24.1, 19.7, 13.6.

Tetrabutylammonium 3-fluorobenzoate (S32). White solid; ^1H NMR (500 MHz, CDCl_3) δ 7.75–7.73 (m, 1 H), 7.63–7.61 (m, 1 H), 7.13–7.09 (m, 1 H), 6.86–6.83 (m, 1 H), 3.14–3.10 (m, 8 H), 1.47–1.43 (m, 8 H), 1.26–1.21 (m, 8 H), 0.84–0.80 (m, 12 H); ^{13}C NMR (125 MHz, CDCl_3) δ 169.7, 163.4, 161.4, 143.9, 128.3, 125.0, 116.0, 115.8, 115.1, 115.0, 58.5, 23.9, 19.6, 13.6.

Tetrabutylammonium 2-fluorobenzoate (S33). White solid; ^1H NMR (500 MHz, CDCl_3) δ 7.65–7.62 (m, 1 H), 7.06–7.02 (m, 1 H), 6.91–6.88 (m, 1 H), 6.82–6.78 (m, 1 H), 3.17–3.13 (m, 8 H), 1.49–1.45 (m, 8 H), 1.25 (sx, $J = 7.4$ Hz, 8 H), 0.83 (t, $J = 7.4$ Hz, 12 H); ^{13}C NMR (125

MHz, CDCl₃) δ 169.0, 161.2, 159.2, 131.2, 131.1, 128.5, 123.0, 115.4, 115.2, 58.4, 23.9, 19.6, 13.6.

Tetrabutylammonium *N*-Phthaloylglycinate (S34). Pale yellow solid; ¹H NMR (500 MHz, CDCl₃) δ 7.77 (dd, $J = 5.4, 3.0$ Hz, 2 H), 7.62 (dd, $J = 5.4, 3.0$ Hz, 2 H), 4.21 (s, 2 H), 3.32–3.28 (m, 8 H), 1.66–1.60 (m, 8 H), 1.40 (sx, $J = 7.4$ Hz, 8 H), 0.97 (t, $J = 7.3$ Hz, 12 H); ¹³C NMR (125 MHz, CDCl₃) δ 169.7, 168.6, 133.1, 122.7, 58.6, 42.5, 24.0, 19.7, 13.6.

Tetrabutylammonium 2-(adamantan-1-yl)acetate (S35). Pale yellow solid; ¹H NMR (500 MHz, CDCl₃) δ 3.30–3.26 (m, 8 H), 1.87 (s, 2 H), 1.81 (s, 3 H), 1.63 (d, $J = 2.2$ Hz, 6 H), 1.59–1.53 (m, 14 H), 1.34 (sx, $J = 7.4$ Hz, 8 H), 0.90 (t, $J = 7.3$ Hz, 12 H); ¹³C NMR (125 MHz, CDCl₃) δ 176.9, 58.7, 55.0, 43.0, 37.3, 32.1, 29.0, 24.1, 19.7, 13.7.

Tetrabutylammonium adamantane-1-carboxylate (S36). Pale yellow solid; ¹H NMR (500 MHz, CDCl₃) δ 3.28–3.26 (m, 8 H), 1.87–1.84 (m, 9 H), 1.63–1.55 (m, 14 H), 1.37 (sx, $J = 7.4$ Hz, 8 H), 0.94 (t, $J = 7.3$ Hz, 12 H); ¹³C NMR (125 MHz, CDCl₃) δ 183.2, 8.7, 49.8, 41.8, 40.7, 37.4, 29.0, 24.1, 19.7, 13.7.

Tetrabutylammonium pivalate (S37). Pale yellow solid; ¹H NMR (500 MHz, CDCl₃) δ 3.24–3.20 (m, 8 H), 1.57–1.51 (m, 8 H), 1.33 (sx, $J = 7.4$ Hz, 8 H), 1.04 (s, 9 H), 0.89 (t, $J = 7.4$ Hz, 12 H); ¹³C NMR (125 MHz, CDCl₃) δ 183.4, 58.6, 39.5, 28.9, 24.0, 19.7, 13.7.

Tetrabutylammonium furan-3-carboxylate (S38). Brown solid; ¹H NMR (500 MHz, CDCl₃) δ 7.63–7.62 (m, 1 H), 7.10 (t, $J = 1.6$ Hz, 1 H), 6.53–6.52 (m, 1 H), 3.09–3.06 (m, 8 H), 1.45–1.38 (m, 8 H), 1.22 (sx, $J = 7.4$ Hz, 8 H), 0.79 (t, $J = 7.4$ Hz, 12 H); ¹³C NMR (125 MHz, CDCl₃) δ 179.8, 144.5, 141.5, 128.4, 111.6, 58.4, 23.8, 19.5, 13.5.

Tetrabutylammonium pentafluorophenonate (S39). Pale yellow solid, ^1H NMR (500 MHz, CDCl_3) δ 3.13–3.09 (m, 8 H), 1.53–1.46 (m, 8 H), 1.28 (sx, $J = 7.4$ Hz, 8 H), 0.87 (t, $J = 7.4$ Hz, 12 H); ^{13}C NMR (125 MHz, CDCl_3) δ 147.0, 142.1, 140.3, 139.7, 137.8, 125.1, 58.4, 23.6, 19.4, 13.3.

Tetrabutylammonium 1H-benzo[d][1,2,3]triazol-1-olate (S40). Brown solid, ^1H NMR (500 MHz, CDCl_3) δ 7.67–7.63 (m, 2 H), 7.07–7.01 (m, 2 H), 3.20–3.17 (m, 8 H), 1.55–1.49 (m, 8 H), 1.31 (sx, $J = 7.4$ Hz, 8 H), 0.89 (t, $J = 7.3$ Hz, 12 H); ^{13}C NMR (125 MHz, CDCl_3) δ 143.8, 127.8, 122.2, 121.1, 118.0, 112.4, 58.5, 23.8, 19.6, 13.6.

General procedure for the Pd-based cyclization

To a flame-dried seal tube was added $\text{Pd}(\text{OAc})_2$ (2.2 mg, 0.010 mmol, 0.1 equiv), *t*-BuMephos (4.7 mg, 0.015 mmol, 0.15 equiv), and the tetrabutylammonium salt (0.30 mmol, 3.0 equiv). Next, a solution of the desired vinyl triflate (0.10 mmol, 1.0 equiv) in toluene (1 mL) was added at 23 °C. The resultant mixture was then stirred at 90 °C until palladium black completely crashed out, typically for 5 h. Upon completion, the reaction mixture was cooled to 23 °C and filtered through a pad of silica gel (EtOAc). The filtrate was concentrated, and the resultant residue was purified by flash column chromatography (silica gel, hexanes:EtOAc) to provide the desired oxygenated product in the amount and form delineated below.

Product Acetate 29. Pale yellow oil, 28.2 mg, 96% yield; $R_f = 0.26$ (silica gel, hexanes:EtOAc, 10:1); IR (film) ν_{max} 2955, 2930, 2863, 1739, 1729, 1597, 1564, 1448, 1385, 1274, 1218, 1150, 1034 cm^{-1} ; ^1H NMR (500 MHz, CDCl_3) δ 5.27 (s, 1 H), 4.00 (dd, $J = 27.2, 10.5$ Hz, 2 H), 3.65 (s, 3 H), 2.26–2.22 (m, 1 H), 2.20–2.17 (m, 1 H), 2.06 (s, 3 H), 1.56–1.49 (m, 2 H), 1.47–1.40 (m, 2 H), 1.37–1.28 (m, 2 H), 1.09 (s, 3 H), 0.99 (s, 3 H), 0.92 (s, 3 H); ^{13}C NMR (125

MHz, CDCl₃) δ 176.7, 144.1, 133.2, 72.4, 55.5, 51.8, 44.0, 35.8, 35.4, 32.5, 31.8, 30.5, 29.7, 29.4, 28.2, 27.4, 26.1, 21.0; HRMS (ESI) calcd for C₁₇H₂₇O₄⁺ [M + H⁺] 295.1904, found: 295.1907.

Product benzolate 30. Colorless oil, 28.4 mg, 80% yield; R_f = 0.26 (silica gel, hexanes:EtOAc, 10:1); IR (film) ν_{\max} 2955, 2864, 1722, 1451, 1362, 1274, 1205, 1176, 1151, 1114, 1070, 1026, 961, 711; ¹H NMR (500 MHz, CDCl₃) δ 8.07–8.05 (m, 2 H), 7.56 (tt, J = 7.4, 1.3 Hz, 1 H), 7.46–7.43 (m, 2 H), 5.36 (s, 1 H), 4.25 (dd, J = 16.5, 10.5 Hz, 2 H), 3.65 (s, 3 H), 2.30–2.27 (m, 1 H), 2.23–2.20 (m, 1 H), 1.71–1.58 (m, 2 H), 1.51–1.45 (m, 2 H), 1.40–1.25 (m, 2 H), 1.20 (s, 3 H), 1.00 (s, 3 H), 0.94 (s, 3 H); ¹³C NMR (125 MHz, CDCl₃) δ 176.8, 166.7, 144.0, 133.4, 132.7, 130.7, 129.6, 128.3, 72.7, 55.5, 51.9, 44.4, 35.8, 35.6, 34.8, 32.6, 31.9, 30.5, 29.4, 26.2; HRMS (ESI) calcd for C₂₂H₂₈O₄Na⁺ [M + Na⁺] 379.1880, found: 379.1886.

Product 4-Me 31. White oil, 29.2 mg, 79% yield; R_f = 0.27 (silica gel, hexanes:EtOAc, 10:1); IR (film) ν_{\max} 2954, 2864, 1720, 1612, 1458, 1380, 1362, 1273, 1206, 1177, 1151, 1120, 1020, 961, 914, 841; ¹H NMR (500 MHz, CDCl₃) δ 7.94 (d, J = 8.2 Hz, 2 H), 7.24 (d, J = 8.1 Hz, 2 H), 5.34 (s, 1 H), 4.22 (dd, J = 18.6, 10.6 Hz, 2 H), 3.65 (s, 3 H), 2.41 (s, 3 H), 2.30–2.26 (m, 1 H), 2.22–2.19 (m, 1 H), 1.70–1.57 (m, 2 H), 1.49–1.44 (m, 2 H), 1.40–1.27 (m, 2 H), 1.19 (s, 3 H), 1.00 (s, 3 H), 0.94 (s, 3 H); ¹³C NMR (125 MHz, CDCl₃) δ 176.8, 166.8, 144.1, 143.3, 133.3, 129.6, 129.0, 128.0, 72.5, 55.5, 51.9, 44.4, 35.8, 35.6, 34.8, 32.6, 31.9, 30.5, 29.4, 26.2, 21.6; HRMS (ESI) calcd for C₂₃H₃₁O₄⁺ [M + H⁺] 371.2217, found: 371.2226.

Product 4-OMe 32. Pale yellow oil, 14.3 mg, 37% yield; R_f = 0.25 (silica gel, hexanes:EtOAc, 10:1); IR (film) ν_{\max} 2955, 2864, 1716, 1607, 1511, 1459, 1363, 1300, 1256, 1205, 1167, 1102, 1031, 847, 770 cm⁻¹; ¹H NMR (500 MHz, CDCl₃) δ 8.01 (d, J = 8.8 Hz, 2 H), 6.92 (d, J = 8.8 Hz, 2 H), 5.35 (s, 1 H), 4.21 (dd, J = 17.8, 10.5 Hz, 2 H), 3.86 (s, 3 H), 3.65 (s, 3 H), 2.30–2.26 (m, 1 H), 2.22–2.19 (m, 1 H), 1.70–1.57 (m, 2 H), 1.50–1.44 (m, 2 H), 1.40–1.27 (m, 2 H),

1.19 (s, 3 H), 1.00 (s, 3 H), 0.94 (s, 3 H); ^{13}C NMR (125 MHz, CDCl_3) δ 176.8, 166.4, 163.2, 144.1, 133.3, 131.5, 123.2, 113.6, 72.4, 55.5, 55.4, 51.8, 44.4, 35.8, 35.6, 34.8, 32.6, 31.9, 30.5, 29.4, 26.2; HRMS (ESI) calcd for $\text{C}_{23}\text{H}_{31}\text{O}_5^+$ $[\text{M} + \text{H}^+]$ 387.2166, found: 387.2179.

Product 3-OMe 33. Pale yellow oil, 23.2 mg, 60% yield; $R_f = 0.44$ (silica gel, hexanes:EtOAc, 10:1); IR (film) ν_{max} 2955, 2864, 2838, 1721, 1587, 1457, 1321, 1278, 1151, 1104, 1046, 977, 756 cm^{-1} ; ^1H NMR (500 MHz, CDCl_3) δ 7.65 (dt, $J = 7.7, 1.0$ Hz, 1 H), 7.58 (dd, $J = 2.6, 1.5$ Hz, 1 H), 7.35 (t, $J = 7.9$ Hz), 7.10 (ddd, $J = 8.2, 2.6, 0.8$ Hz, 1 H), 5.36 (s, 1 H), 4.24 (dd, $J = 19.1, 10.5$ Hz, 2 H), 3.86 (s, 3 H), 3.65 (s, 3 H), 2.30–2.26 (m, 1 H), 2.23–2.20 (m, 1 H), 1.70–1.58 (m, 2 H), 1.51–1.44 (m, 2 H), 1.40–1.29 (m, 2 H), 1.19 (s, 3 H), 1.00 (s, 3 H), 0.94 (s, 3 H); ^{13}C NMR (125 MHz, CDCl_3) δ 176.7, 166.5, 159.6, 144.0, 133.4, 132.0, 129.3, 121.2, 119.2, 114.2, 72.8, 55.5, 55.4, 51.9, 44.4, 35.8, 35.6, 34.8, 32.6, 31.9, 30.5, 29.4, 26.2; HRMS (ESI) calcd for $\text{C}_{23}\text{H}_{31}\text{O}_5^+$ $[\text{M} + \text{H}^+]$ 387.2166, found: 387.2173.

Product 4-CHO 34. Colorless oil, 34.1 mg, 89% yield; $R_f = 0.28$ (silica gel, hexanes:EtOAc, 10:1); IR (film) ν_{max} 2955, 2864, 1723, 1709, 1459, 1275, 1202, 1116, 1016, 759 cm^{-1} ; ^1H NMR (500 MHz, CDCl_3) δ 10.1 (s, 1 H), 8.21 (d, $J = 8.2$ Hz, 2 H), 7.96 (d, $J = 8.3$ Hz, 2 H), 5.36 (s, 1 H), 4.28 (dd, $J = 12.8, 10.6$ Hz, 2 H), 3.65 (s, 3 H), 2.31–2.27 (m, 1 H), 2.23–2.20 (m, 1 H), 1.68–1.59 (m, 2 H), 1.52–1.45 (m, 2 H), 1.40–1.25 (m, 2 H), 1.21 (s, 3 H), 0.99 (s, 3 H), 0.94 (s, 3 H); ^{13}C NMR (125 MHz, CDCl_3) δ 191.7, 176.6, 165.6, 143.9, 139.1, 135.7, 133.5, 130.1, 129.5, 73.2, 55.6, 51.9, 44.3, 35.8, 35.5, 34.7, 32.6, 31.8, 30.5, 29.3, 26.2; HRMS (ESI) calcd for $\text{C}_{23}\text{H}_{29}\text{O}_5^+$ $[\text{M} + \text{H}^+]$ 385.2010, found: 385.2009.

Product 4-CN 35. Yellow oil, 29.7 mg, 78% yield; $R_f = 0.19$ (silica gel, hexanes:EtOAc, 10:1); IR (film) ν_{max} 2955, 2864, 2231, 1724, 1458, 1275, 1206, 1152, 1118, 1018, 956, 861, 767 cm^{-1} ; ^1H NMR (500 MHz, CDCl_3) δ 8.15–8.14 (m, 2 H), 7.76–7.74 (m, 2 H), 5.34 (s, 1 H), 4.28

(dd, $J = 12.0, 10.6$ Hz, 2 H), 3.65 (s, 3 H), 2.30–2.27 (m, 1 H), 2.23–2.20 (m, 1 H), 1.70–1.57 (m, 2 H), 1.52–1.45 (m, 2 H), 1.40–1.25 (m, 2 H), 1.19 (s, 3 H), 0.99 (s, 3 H), 0.94 (s, 3 H); ^{13}C NMR (125 MHz, CDCl_3) δ 176.6, 165.0, 143.8, 134.5, 133.6, 132.2, 130.0, 118.0, 116.3, 73.4, 55.6, 51.9, 44.3, 35.8, 35.5, 34.7, 32.6, 31.8, 30.5, 29.3, 26.2; HRMS (ESI) calcd for $\text{C}_{23}\text{H}_{28}\text{O}_4\text{N}^+$ [$\text{M} + \text{H}^+$] 382.2013, found: 382.2017.

Product 4-CF₃ 36. Colorless oil, 41.1 mg, 97% yield; $R_f = 0.50$ (silica gel, hexanes:EtOAc, 10:1); IR (film) ν_{max} 2956, 2865, 1726, 1412, 1357, 1281, 1132, 1066, 1018, 862 cm^{-1} ; ^1H NMR (500 MHz, CDCl_3) δ 8.16 (d, $J = 8.1$ Hz, 2 H), 7.71 (d, $J = 8.2$ Hz, 2 H), 5.36 (s, 1 H), 4.28 (dd, $J = 15.2$ Hz, 10.6 Hz, 2 H), 3.66 (s, 3 H), 2.31–2.27 (m, 1 H), 2.23–2.20 (m, 1 H), 1.71–1.58 (m, 2 H), 1.52–1.46 (m, 2 H), 1.37–1.25 (m, 2 H), 1.20 (s, 3 H), 1.00 (s, 3 H), 0.94 (s, 3 H); ^{13}C NMR (125 MHz, CDCl_3) δ 176.6, 165.4, 143.9, 134.2, 133.9, 133.5, 139.9, 125.39, 125.36, 73.1, 55.6, 51.9, 44.3, 35.8, 35.5, 34.7, 32.6, 30.5, 29.3, 26.2; HRMS (ESI) calcd for $\text{C}_{23}\text{H}_{28}\text{O}_4\text{F}_3^+$ [$\text{M} + \text{H}^+$] 425.1934, found: 425.1942.

Product 4-COOMe 37. Yellow solid, 30.6 mg, 74% yield; $R_f = 0.37$ (silica gel, hexanes:EtOAc, 10:1); IR (film) ν_{max} 2954, 2864, 1724, 1458, 1272, 1117, 1019, 731 cm^{-1} ; ^1H NMR (500 MHz, CDCl_3) δ 8.10 (s, 4 H), 5.35 (s, 1 H), 4.26 (dd, $J = 13.7, 10.5$ Hz, 2 H), 3.94 (s, 3 H), 3.64 (s, 3 H), 2.30–2.26 (m, 1 H), 2.22–2.19 (m, 1 H), 1.70–1.58 (m, 2 H), 1.51–1.48 (m, 2 H), 1.39–1.24 (m, 2 H), 1.20 (s, 3 H), 0.98 (s, 3 H), 0.93 (s, 3 H); ^{13}C NMR (125 MHz, CDCl_3) δ 176.6, 166.3, 165.8, 143.9, 134.5, 133.8, 133.5, 129.54, 129.51, 73.1, 55.6, 52.4, 52.0, 44.3, 35.8, 35.5, 34.7, 32.6, 31.8, 30.5, 29.3, 26.2; HRMS (ESI) calcd for $\text{C}_{24}\text{H}_{31}\text{O}_6^+$ [$\text{M} + \text{H}^+$] 415.2115, found: 415.2113.

Product 4-COPh 38. Yellow oil, 42.3 mg, 92% yield; $R_f = 0.37$ (silica gel, hexanes:EtOAc, 10:1); IR (film) ν_{max} 2955, 2864, 1710, 1663, 1449, 1297, 1206, 1116, 926, 714

cm⁻¹; ¹H NMR (500 MHz, CDCl₃) δ 8.17–8.15 (m, 2 H), 7.85–7.84 (m, 2 H), 7.81–7.79 (m, 2 H), 7.63–7.60 (m, 1 H), 7.51–7.48 (m, 2 H), 5.37 (s, 1 H), 4.29 (dd, *J* = 14.7, 10.5 Hz, 2 H), 3.66 (s, 3 H), 2.31–2.27 (m, 1 H), 2.23–2.20 (m, 1 H), 1.71–1.59 (m, 2 H), 1.52–1.46 (m, 2 H), 1.40–1.29 (m, 2 H), 1.22 (s, 3 H), 1.00 (s, 3 H), 0.94 (s, 3 H); ¹³C NMR (125 MHz, CDCl₃) δ 176.7, 165.9, 143.9, 141.2, 137.0, 133.8, 133.5, 132.9, 130.1, 129.8, 129.4, 128.5, 73.1, 55.6, 51.9, 44.4, 35.8, 35.4, 34.7, 32.2, 31.8, 30.5, 29.4, 26.2; HRMS (ESI) calcd for C₂₉H₃₂O₅Na⁺ [M + Na⁺] 483.2142, found: 483.2146.

Product 4-Cl 39. Yellow oil, 27.4 mg, 70% yield; *R_f* = 0.54 (silica gel, hexanes:EtOAc, 10:1); IR (film) *v*_{max} 2956, 2864, 124, 1595, 1488, 1459, 1400, 1381, 1363, 1272, 1205, 1151, 1116, 1015, 960, 850, 760 cm⁻¹; ¹H NMR (500 MHz, CDCl₃) δ 7.98 (d, *J* = 8.5 Hz, 2 H), 7.41 (d, *J* = 8.5 Hz, 2 H), 5.34 (s, 1 H), 4.24 (dd, *J* = 14.1, 10.6 Hz, 2 H), 3.65 (s, 3 H), 2.30–2.26 (m, 1 H), 2.22–2.19 (m, 1 H), 1.69–1.58 (m, 2 H), 1.51–1.44 (m, 2 H), 1.40–1.28 (m, 2 H), 1.19 (s, 3 H), 0.99 (s, 3 H), 0.93 (s, 3 H); ¹³C NMR (125 MHz, CDCl₃) δ 176.6, 165.8, 144.0, 139.2, 133.4, 130.9, 129.6, 129.2, 128.7, 128.3, 72.8, 55.5, 51.9, 44.3, 35.8, 35.5, 34.7, 32.6, 31.8, 30.5, 29.3, 26.2; HRMS (ESI) calcd for C₂₂H₂₈O₄Cl⁺ [M + H⁺] 391.1671, found: 391.1675.

Product 4-F 40. Pale yellow oil, 33.7 mg, 90% yield; *R_f* = 0.51 (silica gel, hexanes:EtOAc, 10:1); IR (film) *v*_{max} 2955, 2864, 1724, 1604, 1508, 1458, 1274, 1152, 1116, 854, 767 cm⁻¹; ¹H NMR (500 MHz, CDCl₃) δ 8.08–8.05 (m, 2 H), 7.13–7.09 (m, 2 H), 5.34 (s, 1 H), 4.23 (dd, *J* = 13.7, 10.6 Hz, 2 H), 3.65 (s, 3 H), 2.30–2.26 (m, 1 H), 2.22–2.19 (m, 1 H), 1.70–1.57 (m, 2 H), 1.51–1.44 (m, 2 H), 1.40–1.25 (m, 2 H), 1.19 (s, 3 H), 0.99 (s, 3 H), 0.94 (s, 3 H); ¹³C NMR (125 MHz, CDCl₃) δ 176.7, 166.7, 165.7, 164.7, 144.0, 133.4, 132.1, 132.0, 127.0, 126.9, 115.5, 115.3, 72.8, 55.5, 51.9, 44.4, 35.8, 35.5, 34.7, 32.6, 31.8, 30.5, 29.4, 26.2; HRMS (ESI) calcd for C₂₂H₂₈O₄F⁺ [M + H⁺] 375.1966, found: 375.1971.

Product 3-F 41. Colorless oil, 29.9 mg, 80% yield; $R_f = 0.49$ (silica gel, hexanes:EtOAc, 10:1); IR (film) ν_{\max} 2957, 2864, 1724, 1596, 1561, 1448, 1388, 1276, 1261, 1204, 1151, 1094, 858 cm^{-1} ; ^1H NMR (500 MHz, CDCl_3) δ 7.85–7.84 (m, 1 H), 7.74–7.71 (m, 1 H), 7.44–7.40 (m, 1 H), 7.27–7.23 (m, 1 H), 5.35 (s, 1 H), 4.25 (dd, $J = 15.8, 10.5$ Hz, 2 H), 3.65 (s, 3 H), 2.30–2.26 (m, 1 H), 2.23–2.20 (m, 1 H), 1.70–1.58 (m, 2 H), 1.51–1.45 (m, 2 H), 1.40–1.25 (m, 2 H), 1.19 (s, 3 H), 1.00 (s, 3 H), 0.94 (s, 3 H); ^{13}C NMR (125 MHz, CDCl_3) δ 176.7, 165.5, 161.6, 143.9, 133.5, 130.0, 129.9, 125.3, 125.2, 119.9, 119.7, 116.5, 116.3, 73.0, 55.5, 51.9, 44.3, 35.8, 35.5, 34.7, 32.6, 31.8, 30.5, 29.3, 26.2; HRMS (ESI) calcd for $\text{C}_{22}\text{H}_{28}\text{O}_4\text{F}^+$ [$\text{M} + \text{H}^+$] 375.1966, found: 375.1971.

Product 2-F 42. Colorless oil, 32.6 mg, 87% yield; $R_f = 0.40$ (silica gel, hexanes:EtOAc, 10:1); IR (film) ν_{\max} 2955, 2864, 1724, 1613, 1486, 1458, 1363, 1297, 1153, 1128, 1033, 960, 758 cm^{-1} ; ^1H NMR (500 MHz, CDCl_3) δ 7.95 (td, $J = 7.6, 1.8$ Hz, 1 H), 7.52–7.48 (m, 1 H), 7.19 (dt, $J = 7.8, 0.5$ Hz, 1 H), 7.12 (dd, $J = 10.6, 8.7$ Hz, 1 H), 5.36 (s, 1 H), 4.25 (dd, $J = 12.8, 10.6$ Hz, 2 H), 3.64 (s, 3 H), 2.28–2.24 (m, 1 H), 2.22–2.19 (m, 1 H), 1.70–1.57 (m, 2 H), 1.50–1.43 (m, 2 H), 1.39–1.24 (m, 2 H), 1.19 (s, 3 H), 0.98 (s, 3 H), 0.93 (s, 3 H); ^{13}C NMR (125 MHz, CDCl_3) δ 176.8, 164.7, 163.0, 160.9, 143.9, 134.3, 134.2, 133.3, 132.2, 123.9, 123.8, 117.0, 116.8, 73.2, 55.6, 51.9, 44.2, 35.8, 35.6, 34.8, 32.6, 31.8, 30.5, 29.4, 26.2; HRMS (ESI) calcd for $\text{C}_{22}\text{H}_{28}\text{O}_4\text{F}^+$ [$\text{M} + \text{H}^+$] 375.1966, found: 375.1959.

Product glycine 44. Colorless oil, 29.8 mg, 68% yield; $R_f = 0.16$ (silica gel, hexanes:EtOAc, 5:1); IR (film) ν_{\max} 2925, 2857, 1778, 1749, 1724, 1457, 1417, 1388, 1206, 955, 731, 714; ^1H NMR (500 MHz, CDCl_3) δ 7.89 (dd, $J = 5.5, 3.0$ Hz, 2 H), 7.75 (dd, $J = 5.4, 3.1$ Hz, 2 H), 5.20 (s, 1 H), 4.46 (d, $J = 4.1$ Hz, 2 H), 4.09 (s, 1 H), 3.64 (s, 3 H), 2.20–2.16 (m, 2 H), 1.52–1.47 (m, 1H), 1.45–1.37 (m, 3 H), 1.34–1.29 (m, 2 H), 1.04 (s, 3 H), 0.96 (s, 3 H), 0.87 (s, 3 H);

^{13}C NMR (125 MHz, CDCl_3) δ 176.6, 167.4, 167.2, 143.8, 134.2, 133.4, 132.1, 123.6, 73.4, 55.5, 51.9, 44.0, 39.0, 35.7, 35.3, 34.8, 32.5, 31.8, 30.4, 29.7, 29.3, 25.9; HRMS (ESI) calcd for $\text{C}_{25}\text{H}_{29}\text{O}_6\text{NNa}^+$ [$\text{M} + \text{Na}^+$] 462.1887, found: 462.1889.

Product CH₂Ad 45. Colorless oil, 37.3 mg, 87% yield; $R_f = 0.34$ (silica gel, hexanes:EtOAc, 10:1); IR (film) ν_{max} 2953, 2904, 2848, 1730, 1457, 1256, 1201, 1136, 1101, 1003 cm^{-1} ; ^1H NMR (500 MHz, CDCl_3) δ 5.28 (s, 1 H), 3.96 (dd, $J = 15.3, 10.6$ Hz, 2 H), 3.65 (s, 3 H), 2.26–2.22 (m, 1 H), 2.19–2.17 (m, 1 H), 2.08 (s, 2 H), 1.96 (s, 3 H), 1.71–1.62 (m, 12 H), 1.55–1.49 (m, 2 H), 1.47–1.39 (m, 2 H), 1.37–1.27 (m, 2 H), 1.10 (s, 3 H), 0.98 (s, 3 H), 0.92 (s, 3 H); ^{13}C NMR (125 MHz, CDCl_3) δ 176.7, 172.1, 144.2, 133.1, 72.1, 55.5, 51.8, 49.3, 43.9, 42.5, 36.8, 35.8, 35.7, 34.8, 32.7, 32.6, 31.9, 30.5, 29.4; HRMS (ESI) calcd for $\text{C}_{27}\text{H}_{41}\text{O}_4^+$ [$\text{M} + \text{H}^+$] 429.2999, found: 429.2989.

Product Ad 46. Colorless oil, 38.1 mg, 92% yield; $R_f = 0.65$ (silica gel, hexanes:EtOAc, 10:1); IR (film) ν_{max} 2907, 2854, 1727, 1453, 1234, 1150, 1078, 991, 963, 738 cm^{-1} ; ^1H NMR (500 MHz, CDCl_3) δ 5.30 (s, 1 H), 3.93 (dd, $J = 12.6, 10.6$ Hz, 2 H), 3.64 (s, 3 H), 2.26–2.22 (m, 1 H), 2.19–2.16 (m, 1 H), 2.01 (s, 3 H), 1.91–1.90 (m, 6 H), 1.74–1.66 (m, 6 H), 1.54–1.47 (m, 2 H), 1.45–1.38 (m, 2 H), 1.37–1.24 (m, 2 H), 1.10 (s, 3 H), 0.99 (s, 3 H), 0.92 (s, 3 H); ^{13}C NMR (125 MHz, CDCl_3) δ 177.7, 176.8, 144.0, 133.2, 71.8, 55.4, 51.9, 44.2, 40.9, 39.0, 36.6, 35.8, 35.5, 34.7, 32.6, 31.8, 30.6, 29.4, 28.0, 26.0; HRMS calcd for $\text{C}_{26}\text{H}_{39}\text{O}_4^+$ [$\text{M} + \text{H}^+$] 415.2843, found: 415.2847.

Product pivalate 47. Pale yellow oil, 25.5 mg, 76% yield; $R_f = 0.37$ (silica gel, hexanes:EtOAc, 10:1); IR (film) ν_{max} 2957, 2866, 1728, 1597, 1563, 1481, 1448, 1396, 1286, 1217, 1150, 858, 798 cm^{-1} ; ^1H NMR (500 MHz, CDCl_3) δ 5.29 (s, 1 H), 3.94 (dd, $J = 12.2, 10.5$ Hz, 2 H), 3.64 (s, 3 H), 2.25–2.22 (m, 1 H), 2.19–2.16 (m, 1 H), 1.54–1.48 (m, 2 H), 1.46–1.39 (m, 2 H), 1.37–1.26 (m, 2 H), 1.20 (s, 9 H), 1.10 (s, 3 H), 0.98 (s, 3 H), 0.91 (s, 3 H); ^{13}C NMR (125 MHz,

CDCl₃) δ 178.6, 176.8, 144.0, 133.2, 76.8, 72.1, 55.5, 51.9, 44.2, 35.7, 35.5, 34.7, 32.6, 31.8, 30.5, 29.4, 27.3, 26.0; HRMS (ESI) calcd for C₂₀H₃₃O₄⁺ [M + H⁺] 337.2373, found: 337.2377.

Product furan 48. Yellow oil, 20.6 mg, 60% yield; R_f = 0.32 (silica gel, hexanes:EtOAc, 10:1); IR (film) ν_{\max} 2955, 2864, 1725, 1579, 1506, 1461, 1304, 1205, 1161, 1076, 980, 874 cm⁻¹; ¹H NMR (500 MHz, CDCl₃) δ 8.01 (dd, J = 1.4, 0.7 Hz, 1 H), 7.43 (t, J = 1.7 Hz, 1 H), 6.75 (dd, J = 1.8, 0.7 Hz, 1 H), 5.32 (s, 1 H), 4.18 (dd, J = 15.2, 10.6 Hz, 2 H), 3.65 (s, 3 H), 2.28–2.25 (m, 1 H), 2.22–2.19 (m, 1 H), 1.67–1.52 (m, 2 H), 1.49–1.42 (m, 2 H), 1.39–1.28 (m, 2 H), 1.15 (s, 3 H), 1.00 (s, 3 H), 0.93 (s, 3 H); ¹³C NMR (125 MHz, CDCl₃) δ 176.7, 163.2, 147.5, 144.0, 143.6, 133.3, 119.8, 109.8, 72.1, 55.5, 51.9, 44.3, 35.8, 35.4, 34.7, 32.6, 31.8, 30.5, 29.4, 26.2; HRMS (ESI) calcd for C₂₀H₂₇O₅⁺ [M + H⁺] 347.1853, found: 347.1854.

Product flourophanol 50. Pale yellow oil, 32.6 mg, 78% yield; R_f = 0.75 (silica gel, hexanes:EtOAc, 10:1); IR (film) ν_{\max} 2957, 2865, 1728, 1518, 1463, 1381, 1205, 1151, 1027, 996 cm⁻¹; ¹H NMR (500 MHz, CDCl₃) δ 5.34 (s, 1 H), 4.08 (d, J = 8.6 Hz, 1 H), 4.00 (d, J = 8.6 Hz, 1 H), 3.64 (s, 3 H), 2.27–2.23 (m, 1 H), 2.22–2.19 (m, 1 H), 1.69–1.59 (m, 2 H), 1.50–1.44 (m, 2 H), 1.39–1.27 (m, 2 H), 1.26 (s, 3 H), 0.98 (s, 3 H), 0.93 (s, 3 H); ¹³C NMR (125 MHz, CDCl₃) δ 176.6, 143.7, 142.8, 140.7, 139.1, 138.0, 137.1, 136.0, 134.5, 133.6, 83.8, 55.6, 51.8, 45.4, 35.8, 35.3, 34.8, 32.6, 31.6, 30.4, 29.3, 25.5; HRMS (ESI) calcd for C₂₁H₂₄O₃F₅⁺ [M + H⁺] 419.1640, found: 419.1649.

Product 52. Pale yellow oil, 3.3 mg, 9% yield; R_f = 0.16 (silica gel, hexanes:EtOAc, 10:1); IR (film) ν_{\max} 2956, 2864, 1725, 1462, 1376, 1358, 1263, 1239, 1206, 1153, 1088, 952, 744; ¹H NMR (500 MHz, CDCl₃) δ 8.00 (d, J = 8.4 Hz, 1 H), 7.63 (d, J = 8.3 Hz, 1 H), 7.49 (ddd, J = 8.4, 7.0, 0.7 Hz, 1 H), 7.37 (ddd, J = 8.4, 6.9, 0.9 Hz, 1 H), 5.46 (s, 1 H), 4.44 (dd, J = 43.2, 7.8 Hz, 2 H), 3.60 (s, 1 H), 2.33–2.30 (m, 1 H), 2.26–2.23 (m, 1 H), 1.79–1.69 (m, 2 H), 1.56–1.49 (m, 2 H),

1.43–1.30 (m, 2 H), 1.41 (s, 3 H), 1.03 (s, 3 H), 0.96 (s, 3 H); ^{13}C NMR (125 MHz, CDCl_3) δ 176.5, 143.7, 134.1, 127.7, 127.2, 124.5, 108.9, 88.5, 55.7, 52.0, 44.3, 35.7, 35.4, 34.7, 32.8, 31.6, 30.5, 29.3, 26.0, 20.3, 13.7; HRMS (ESI) calcd for $\text{C}_{21}\text{H}_{28}\text{O}_3\text{N}_3^+$ [$\text{M} + \text{H}^+$] 370.2125, found: 370.2113.

Product 53. Colorless oil, 24.0 mg, 96% yield; $R_f = 0.64$ (silica gel, hexanes:EtOAc, 10:1); $[\alpha]_{\text{D}}^{20} = +44.0^\circ$ ($c = 0.55$ in CHCl_3); IR (film) ν_{max} 2955, 2868, 2865, 1744, 1455, 1383, 1370, 1242, 1034 cm^{-1} ; ^1H NMR (500 MHz, CDCl_3) δ 5.30 (s, 1 H), 3.84 (d, $J = 10.4$ Hz, 1 H), 3.72 (d, $J = 10.4$ Hz, 1 H), 3.71 (s, 3 H), 2.20–2.13 (m, 1 H), 2.12–2.08 (m, 1 H), 2.07 (s, 3 H), 1.88–1.84 (m, 1 H), 1.74–1.68 (m, 1 H), 1.56 (d, $J = 13.2$ Hz, 1 H), 1.40 (d, $J = 13.2$ Hz, 1 H), 1.20–1.13 (m, 1 H), 1.16 (s, 3 H), 1.06–1.00 (m, 1 H), 1.03 (s, 3 H), 0.95 (d, $J = 7.0$ Hz, 3 H), 0.79 (s, 3 H); ^{13}C NMR (125 MHz, CDCl_3) δ 171.5, 148.2, 125.7, 73.5, 52.1, 51.0, 42.5, 39.4, 32.0, 31.2, 28.2, 24.4, 23.8, 23.0, 22.3, 21.0; HRMS (CI) calcd for $\text{C}_{16}\text{H}_{27}\text{O}_2^+$ [$\text{M} + \text{H}^+$] 251.2006, found: 251.2002.

Product 54. Yellow oil, 24.2 mg, 86% yield; $R_f = 0.26$ (silica gel, hexanes:EtOAc, 10:1); IR (film) ν_{max} 2954, 2866, 1742, 1459, 1384, 1370, 1247, 1152, 1034, 848 cm^{-1} ; ^1H NMR (500 MHz, CDCl_3) δ 5.34 (s, 1 H), 4.00 (dd, $J = 20.2, 10.5$ Hz, 2 H), 3.69 (s, 3 H), 2.27–2.23 (m, 1 H), 2.16 (d, $J = 12.9$ Hz, 1 H), 2.06 (s, 3 H), 1.90 (td, $J = 12.9, 6.9$ Hz, 1 H), 1.72 (d, $J = 13.0$ Hz, 1 H), 1.68 (ddd, $J = 13.3, 7.5, 1.0$ Hz, 1H), 1.44 (td, $J = 12.4, 7.5$ Hz, 1 H), 1.14 (s, 3 H), 1.10 (s, 6 H); ^{13}C NMR (125 MHz, CDCl_3) δ 177.5, 171.2, 10, 134.9, 70.9, 66.2, 52.2, 51.9, 50.9, 40.1, 39.6, 35.7, 30.8, 27.5, 25.9, 21.0; HRMS (CI) calcd for $\text{C}_{16}\text{H}_{25}\text{O}_4^+$ [$\text{M} + \text{H}^+$] 281.1753, found: 281.1757.

Product 55. Yellow oil, 26.4 mg, 73% yield; $R_f = 0.29$ (silica gel, hexanes:EtOAc, 10:1); IR (film) ν_{max} 2959, 2870, 1742, 1714, 1462, 1384, 1372, 1245, 1036 cm^{-1} ; ^1H NMR (500 MHz, CDCl_3) δ 5.29 (s, 1 H), 3.99 (s, 2 H), 2.73 (d, $J = 14.2$ Hz, 1 H), 2.13 (d, $J = 14.2$ Hz, 1 H), 2.08 (s, 3 H), 1.90 (td, $J = 12.9, 6.8$ Hz, 1 H), 1.72 (td, $J = 12.6, 7.1$ Hz, 1 H), 1.62–1.59 (m, 1 H), 1.53–1.51 (m, 1 H), 1.31 (s, 3 H), 1.17 (s, 3 H), 1.06 (s, 3 H), 1.00 (s, 3H); ^{13}C NMR (125 MHz, CDCl_3)

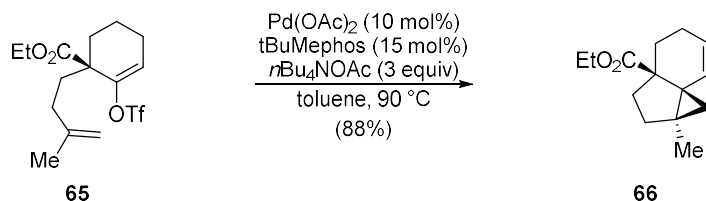
δ 213.8, 171.1, 148.6, 130.6, 71.9, 54.2, 49.2, 44.4, 38.8, 33.4, 31.0, 30.9, 30.8, 26.4, 26.0, 20.9; HRMS (ESI) calcd for $C_{16}H_{25}O_3^+$ [$M + H^+$] 265.1798, found: 265.1792.

Product O-tethered 56. Yellow oil, 24.5 mg, 83% yield; $R_f = 0.18$ (silica gel, hexanes:EtOAc = 10:1); IR (film) ν_{max} ; 1H NMR (500 MHz, $CDCl_3$) δ 5.33 (s, 1H), 3.94 (dd, $J = 35.7$, 10.9 Hz, 2 H), 3.74 (dd, $J = 54.8$, 9.4 Hz, 2 H), 3.73 (s, 3 H), 2.19–2.15 (m, 1 H), 2.04 (s, 3 H), 1.64–1.59 (m, 1 H), 1.58–1.48 (m, 2 H), 1.20 (s, 3 H), 1.06 (s, 3 H), 0.98 (s, 3 H); ^{13}C NMR (125 MHz, $CDCl_3$) δ 173.1, 171.0, 140.0, 133.8, 84.8, 76.8, 69.6, 52.3, 44.3, 33.0, 32.9, 30.3, 30.2, 29.1, 23.8, 20.9; HRMS (ESI) calcd for $C_{16}H_{24}O_5Na^+$ [$M + Na^+$] 319.1516, found: 319.1523

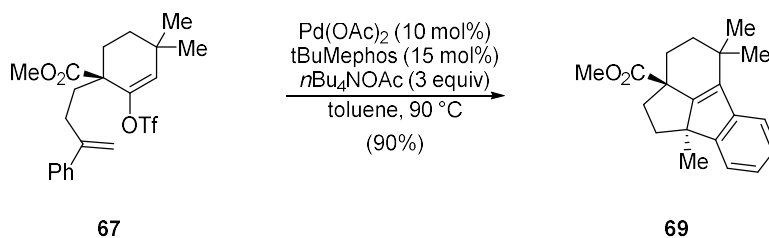
Product OBn 57. Pale yellow oil, 27.2 mg, 68% yield; $R_f = 0.31$ (silica gel, hexanes:EtOAc, 10:1); IR (film) ν_{max} 2953, 2862, 1727, 1454, 1377, 1360, 1244, 1212, 1151, 1100, 1035, 731, 698 cm^{-1} ; 1H NMR (500 MHz, $CDCl_3$) δ 7.35–7.27 (m, 5 H), 5.39 (s, 1 H), 4.50 (s, 2 H), 4.14 (dd, $J = 21.0$, 10.5 Hz, 2 H), 3.66 (s, 3 H), 3.45 (d, $J = 8.8$ Hz, 1 H), 3.23 (d, $J = 8.8$ Hz, 1 H), 2.22–2.16 (m, 2 H), 2.00 (s, 3 H), 1.90–1.86 (m, 1 H), 1.47–1.31 (m, 5 H), 0.99 (s, 3 H), 0.92 (s, 3 H); ^{13}C NMR (125 MHz, $CDCl_3$) δ 176.8, 171.0, 140.5, 138.8, 134.0, 128.3, 127.4, 127.2, 74.0, 73.2, 67.6, 55.5, 51.9, 48.1, 36.1, 34.9, 32.6, 31.5, 30.7, 30.5, 29.5, 21.0; HRMS (ESI) calcd for $C_{24}H_{32}O_5Na^+$ [$M + Na^+$] 423.2142, found: 423.2117.

Product 6-member ring 58. Pale yellow oil, 7.4 mg, 24% yield; $R_f = 0.18$ (silica gel, hexanes:EtOAc, 10:1); IR (film) ν_{max} 2951, 2863, 1726, 1453, 1386, 1371, 1241, 1154, 1058, 1031, 1000, 976 cm^{-1} ; 1H NMR (500 MHz, $CDCl_3$) δ 5.47 (s, 1 H), 3.75 (s, 2 H), 3.63 (s, 3 H), 2.44–2.41 (m, 1 H), 2.03 (s, 3 H), 1.78–1.75 (m, 1 H), 1.68–1.64 (m, 1 H), 1.57–1.47 (m, 4 H), 1.33–1.30 (m, 1 H), 1.24–1.18 (m, 1 H), 1.13–1.06 (m, 1 H), 1.09 (s, 3 H), 1.00 (s, 3 H), 0.96 (s, 3 H); ^{13}C NMR (125 MHz, $CDCl_3$) δ 176.7, 171.2, 136.9, 136.4, 69.6, 51.8, 46.6, 39.0, 37.3, 35.5, 35.4,

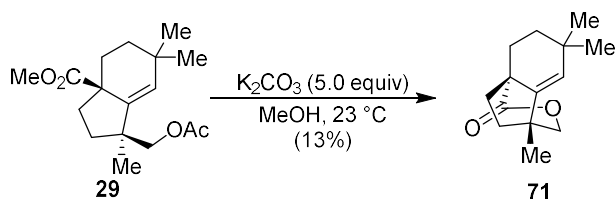
32.9, 32.5, 30.8, 29.1, 25.8, 20.9, 19.2; HRMS (ESI) calcd for $C_{18}H_{29}O_4^+$ [$M + H^+$] 309.2060, found: 309.2051.



Undesired product 66. Triflate **65** (163 mg, 43% overall yield) was prepared as described above procedure toward **S9** from **ethyl 2-oxocyclohexane carboxylate** (340 mg, 2.0 mmol, 1.0 equiv) and **4-iodo-2-methyl-but-1-ene** (431 mg, 2.2 mmol, 1.1 equiv). **65**: colorless oil. $R_f = 0.64$ (silica gel, hexanes:EtOAc, 5:1); IR (film) ν_{max} 2941, 1734, 1416, 1248, 1210, 1145, 1032, 909 cm^{-1} ; $^1\text{H NMR}$ (500 MHz, CDCl_3) δ 5.89 (dd, $J = 5.1$ Hz, 3.1 Hz, 1 H), 4.72 (s, 1H), 4.69 (s, 1H), 4.25–4.17 (m, 2H), 2.31–2.25 (m, 2H), 2.22–2.16 (m, 1H), 2.04–1.95 (m, 3 H), 1.89–1.83 (m, 1H), 1.72 (s, 3 H), 1.67–1.57 (m, 3 H), 1.29 (t, $J = 7.14$ Hz, 3 H); $^{13}\text{C NMR}$ (125 MHz, CDCl_3) δ 172.9, 148.3, 144.8, 120.1, 110.6, 61.7, 50.2, 33.5, 33.3, 32.0, 24.4, 22.4, 18.8, 14.0; HRMS (ESI) calcd for $C_{15}H_{22}OSF_3^+$ [$M + H^+$] 371.1135, found 371.1147. **Undesired product 66** (19.4 mg, 88% yield) was prepared as described above from **65** (37.0 mg, 0.1 mmol, 1.0 equiv). **66**: colorless oil. $R_f = 0.57$ (silica gel, hexanes:EtOAc, 10:1); IR (film) ν_{max} 2921, 2860, 1722, 1445, 1264, 1218, 1195, 1170, 1076, 1028, 681 cm^{-1} ; $^1\text{H NMR}$ (500 MHz, CDCl_3) δ 5.75–5.72 (m, 1 H), 5.27–5.24 (m, 1 H), 4.16–4.07 (m, 2 H), 2.21–2.13 (m, 1 H), 2.10–1.99 (m, 2 H), 1.63–1.51 (m, 3 H), 1.22 (t, $J = 7.1$ Hz), 1.19 (d, $J = 5.5$ Hz), 1.12 (s, 3 H), 0.54 (d, $J = 5.5$ Hz); $^{13}\text{C NMR}$ (125 MHz, CDCl_3) δ 176.3, 128.9, 126.6, 125.1, 60.0, 52.5, 32.3, 32.3, 30.8, 27.0, 23.7, 22.2, 19.6, 14.3; HRMS (ESI) $C_{14}H_{21}O_2^+$ [$M + H^+$] 221.1536, found 221.1534.



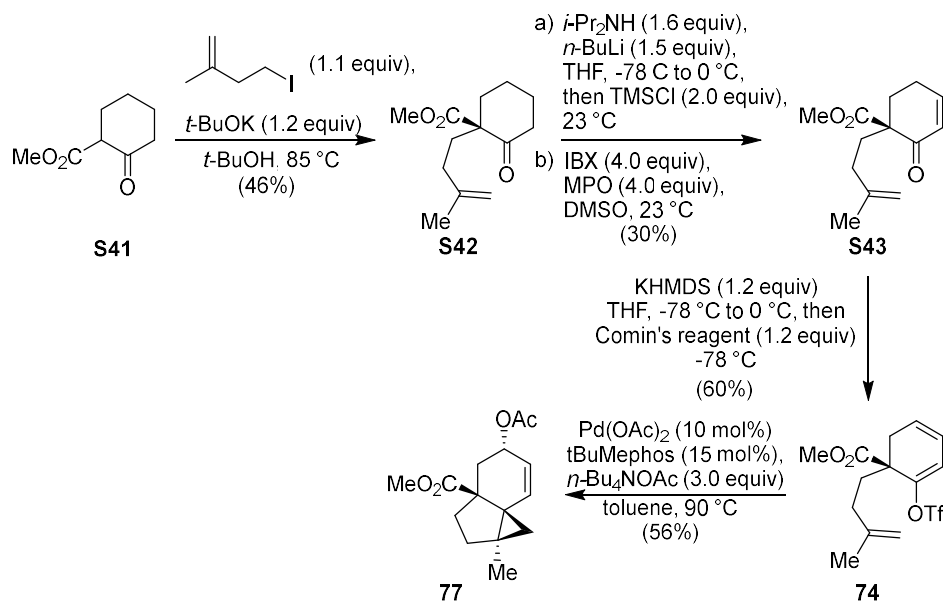
Undesired product 69. Triflate **67** (94.2 mg, 11% overall yield) was prepared as described above procedure toward **28** from **S1** (1.84g, 10.0 mmol, 1.0 equiv) and **phenyl vinyl ketone** (1.98 g, 15.0 mmol, 1.5 equiv). **67**: colorless oil. $R_f = 0.58$ (silica gel, hexanes:EtOAc, 10:1); $^1\text{H NMR}$ (500 MHz, CDCl_3) δ 7.39–7.37 (m, 2 H), 7.34–7.31 (m, 2 H), 7.29–7.26 (m, 1 H), 5.64 (s, 1 H), 5.30 (s, 1 H), 5.09 (s, 1 H), 3.72 (s, 3 H), 2.54–2.45 (m, 2 H), 2.23 (dt, $J = 14.0$ Hz, 4.2 Hz, 1 H), 2.02–1.96 (m, 1 H), 1.90–1.83 (m, 2 H), 1.51–1.49 (m, 2 H), 1.11 (s, 3 H), 1.07 (s, 3H). **Undesired product 69** (26.6 mg, 90% yield) was prepared as described above from **67** (44.6 mg, 0.1 mmol, 1.0 equiv). **69**: colorless oil. $R_f = 0.66$ (silica gel, hexanes:EtOAc, 10:1); $^1\text{H NMR}$ (500 MHz, DMSO-d_6) δ 7.36 (dd, $J = 13.2$ Hz, 7.5 Hz, 2H), 7.21 (td, $J = 7.6$ Hz, 0.9 Hz, 1 H), 7.11 (td, $J = 7.6$ Hz, 0.9 Hz, 1 H), 3.66 (s, 3 H), 2.71–2.68 (m, 1 H), 2.22 (dd, $J = 13.6$ Hz, 8.4 Hz, 1 H), 2.14 (dt, $J = 13.2$ Hz, 2.7 Hz, 1 H), 1.35–1.30 (m, 1 H), 1.33 (s, 3 H), 1.29 (s, 3 H), 1.15 (s, 3H), 0.87 (dt, $J = 8.7$ Hz, 11.6 Hz, 1H); $^{13}\text{C NMR}$ (125 MHz, DMSO-d_6) δ 176.7, 156.2, 153.4, 143.0, 141.7, 126.9, 124.8, 123.1, 122.1, 56.3, 52.4, 47.6, 42.4, 38.7, 33.8, 33.0, 32.1, 29.2, 25.4, 19.7.



Bicycle 71. To a solution of cyclized product **29** (0.294 g, 1.00 mmol, 1.0 equiv) in MeOH (10 mL) was added K_2CO_3 (0.690 g, 5.00 mmol, 5.0 equiv). The resultant mixture was then stirred

for 12 h at 23 °C. The reaction contents were diluted with EtOAc (10 mL) and quenched by the addition of saturated aqueous NH₄Cl (10 mL). The mixture was poured into a separatory funnel and the layers were separated. The aqueous layer was then extracted with EtOAc (2 × 10 mL). The combined organic layers were washed with H₂O (10 mL) and brine (10 mL) before being dried (Na₂SO₄), filtered, and concentrated. The resultant residue was purified by flash column chromatography (silica gel, hexanes:Et₂O, 20:1→10:1) to provide bicycle **71** (28.4 mg, 13% yield) as a yellow solid. **71**: R_f = 0.66 (silica gel, hexanes:EtOAc, 5:1); IR (film) ν_{max} 2955, 2870, 1743, 1597, 1563, 1467, 1395, 1351, 1219, 1126, 1104, 1054, 1034, 992, 880, 864 cm⁻¹; ¹H NMR (500 MHz, CDCl₃) δ 5.28 (s, 1 H), 4.02 (d, *J* = 9.6 Hz, 1 H), 3.83 (dd, *J* = 9.6 Hz, 1.7 Hz, 1 H), 2.60–2.53 (m, 1 H), 2.21 (ddd, *J* = 13.1, 9.6, 4.4 Hz, 1 H), 1.95 (ddd, *J* = 14.7, 9.5, 5.5 Hz, 1 H), 1.74 (tdd, *J* = 12.6, 4.3, 2.0 Hz, 1 H), 1.61 (td, *J* = 18.1 Hz, 5.4 Hz, 1 H), 1.48–1.42 (m, 1 H), 1.40–1.32 (m, 2 H), 1.13 (s, 3 H), 0.99 (s, 3 H), 0.97 (s, 3 H); ¹³C NMR (125 MHz, CDCl₃) δ 174.9, 139.5, 126.6, 82.2, 50.7, 42.2, 36.8, 35.8, 35.0, 31.6, 30.6, 28.1, 26.2, 16.6; HRMS (ESI) calcd for C₁₄H₂₁O₂⁺ [M + H⁺] 221.1536, found: 221.1539. See 2.8 for 2D NMR.

D-labeled 71 was prepared by the described procedure toward **S9**. The D-labeled side chain (**E**)-4-iodo-2-methylbut-1-ene-1-*d* is prepared according to Prof. Brown's method^[22]. **D-labeled 71**: ¹H NMR (500 MHz, CDCl₃) δ 5.28 (s, 1 H), 3.81 (s, 1 H), 2.62–2.54 (m, 1 H), 2.21 (ddd, *J* = 13.1, 9.6, 4.4 Hz, 1 H), 1.95 (ddd, *J* = 14.7, 9.5, 5.5 Hz, 1 H), 1.75 (tdd, *J* = 12.6, 4.3, 2.0 Hz, 1 H), 1.61 (td, *J* = 18.1 Hz, 5.4 Hz, 1 H), 1.49–1.42 (m, 1 H), 1.40–1.35 (m, 2 H), 1.13 (s, 3 H), 0.99 (s, 3 H), 0.98 (s, 3 H).

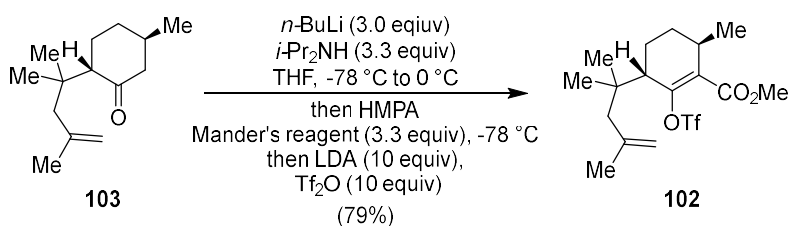


Cyclopropane 77. Ketone **S42** (0.103 g, 46% yield) was prepared as a colorless oil from commercial **methyl 2-oxocyclohexane-1-carboxylate (S41)** (0.156 g, 1.00 mmol, 1.0 equiv) and **4-iodo-2-methylbut-1-ene** (0.216 g, 1.10 mmol, 1.1 equiv) by the procedure described above. **S42**: $R_f = 0.29$ (silica gel, hexanes:EtOAc, 10:1); IR (film) ν_{\max} 2949, 2865, 1714, 1650, 1450, 1215, 1135, 889 cm^{-1} ; ^1H NMR (500 MHz, CDCl_3) δ 4.70 (s, 1 H), 4.68 (s, 1 H), 3.73 (s, 1 H), 2.54–2.51 (m, 1 H), 2.46–2.43 (m, 1 H), 2.05–2.00 (m, 2 H), 1.96–1.84 (m, 1 H), 1.78–1.65 (m, 3 H), 1.72 (s, 3 H), 1.50–1.44 (m, 1 H); ^{13}C NMR (125 MHz, CDCl_3) δ 207.7, 127.5, 145.3, 110.0, 60.1, 52.2, 41.1, 36.0, 32.9, 32.3, 27.6, 22.54, 22.52; HRMS (ESI) calcd for $\text{C}_{13}\text{H}_{21}\text{O}_3^+$ [$\text{M} + \text{H}^+$] 225.1485, found: 225.1472. Next, to a solution of *i*-Pr₂NH (0.440 mL, 3.20 mmol, 1.6 equiv) in THF (5 mL) at 0 °C was added *n*-BuLi (1.20 mL, 2.5 M in hexanes, 3.00 mmol, 1.5 equiv), The resultant solution was then stirred for 15 min at 0 °C. The reaction contents were then cooled to –78 °C and a solution of ketone **S42** (0.448 g, 2.00 mmol, 1.0 equiv) in THF (5 mL) was added dropwise. After warming to 0 °C and stirring at that temperature for 90 min, the solution was recooled to –78 °C and TMSCl (0.510 mL, 4.00 mmol, 2.0 equiv) was added. The resultant solution was slowly warmed to 23 °C and stirred for 12 h. Upon completion, the reaction contents

were quenched by the addition of saturated aqueous NH_4Cl (10 mL). The mixture was then poured into a separatory funnel and the layers were separated. The aqueous layer was extracted with EtOAc (2×10 mL). The combined organic layers were then washed with brine (10 mL) before being dried (Na_2SO_4), filtered, and concentrated to provide the desired crude TMS enol ether intermediate (2.00 mmol assumed) directly used for next step without any further purification. Passing forward, IBX (2.24 g, 8.00 mmol, 4.0 equiv) and MPO (1.00 g, 5.00 mmol, 2.5 equiv) was dissolved in DMSO (20 mL). This solution was then added to a solution of the crude enol ether (2.00 mmol assumed, 1.0 equiv) in DMSO (2.5 mL) at 23 °C. The resultant mixture was stirred at 23 °C for 6 h. Upon completion, the reaction contents were diluted with Et_2O (10 mL) and quenched by the addition of saturated aqueous NaHCO_3 (10 mL). The mixture was poured into a separatory funnel and the layers were separated. The aqueous layer was then extracted with Et_2O (2×10 mL). The combined organic layers were washed with saturated aqueous NaHCO_3 (10 mL), H_2O (10 mL) and brine (10 mL) before being dried (Na_2SO_4), filtered, and concentrated. The resultant residue was purified by flash column chromatography (silica gel, hexanes:EtOAc, 20:1 \rightarrow 10:1) to provide enone **S43** (0.242 g, 30% yield from ketone **S42**)^[9] as a pale-yellow oil. **S43**: R_f = 0.46 (silica gel, hexanes:EtOAc, 5:1); IR (film) ν_{max} 2953, 1723, 1686, 1449, 1386, 1245, 1205, 1081, 889 cm^{-1} ; ^1H NMR (500 MHz, CDCl_3) δ 6.91–6.88 (m, 1 H), 6.01 (dt, J = 10.1, 1.7 Hz, 1 H), 4.69 (s, 1 H), 4.68 (s, 1 H), 3.68 (s, 3 H), 2.54–2.46 (m, 2 H), 2.37–2.30 (m, 1 H), 2.09–2.03 (m, 1 H), 1.99–1.94 (m, 1 H), 1.87–1.81 (m, 1 H), 1.72 (s, 3 H); ^{13}C NMR (125 MHz, CDCl_3) δ 196.2, 172.0, 149.5, 145.2, 129.1, 110.2, 56.8, 52.4, 32.6, 31.9, 29.9, 23.6, 22.5; HRMS (ESI) calcd for $\text{C}_{13}\text{H}_{19}\text{O}_3^+$ [$\text{M} + \text{H}^+$] 223.1329, found: 223.1333. Next, triflate **74** was prepared as described above starting from **S43** (0.111 g, 5.00 mmol, 1.0 equiv) to afford triflate **74** (0.104 g, 60% yield) as a colorless oil. **74**: R_f = 0.62 (silica gel, hexanes:EtOAc, 10:1); IR (film) ν_{max} 2956,

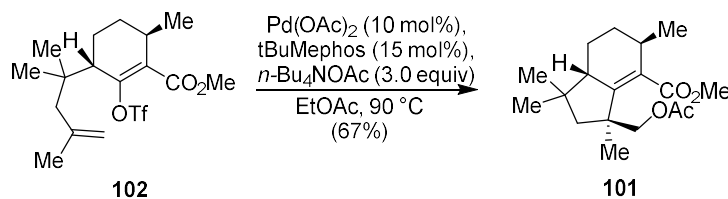
1714, 1654, 1416, 1213, 1143, 1060, 891, 702 cm^{-1} ; ^1H NMR (500 MHz, CDCl_3) δ 6.05–6.04 (m, 1 H), 5.87–5.82 (m, 1 H), 4.73 (s, 1 H), 4.71 (s, 1 H), 3.74 (s, 3 H), 3.07 (dd, $J = 17.7, 4.2$ Hz, 1 H), 2.53–2.49 (m, 1 H), 2.04–1.98 (m, 3 H), 1.94–1.88 (m, 1 H), 1.72 (s, 3 H); ^{13}C NMR (125 MHz, CDCl_3) δ 172.7, 148.1, 144.5, 126.1, 120.8, 115.35, 115.34, 115.33, 110.6, 52.6, 49.5, 33.7, 32.7, 32.5, 22.4; HRMS (ESI) calcd for $\text{C}_{14}\text{H}_{18}\text{O}_5\text{F}_3\text{S}^+$ [$\text{M} + \text{H}^+$] 355.0822, found: 355.0822. Finally, **77** was prepared following the general Pd-cyclization procedure defined above from **74** (35.4 mg, 0.1 mmol) to provide **77** (14.9 mg, 56% yield) as a pale-yellow oil. **77**: $R_f = 0.29$ (silica gel, hexanes:EtOAc, 10:1); IR (film) ν_{max} 2952, 2926, 2863, 1738, 1442, 1370, 1240, 1152, 1113, 1055, 1023 cm^{-1} ; ^1H NMR (500 MHz, CDCl_3) δ 5.67–5.65 (m, 1 H), 5.59–5.55 (m, 1 H), 5.36 (dd, $J = 10.0, 1.5$ Hz, 1 H), 3.69 (s, 3H), 2.35–2.32 (m, 1 H), 2.06 (s, 3 H), 1.92–1.82 (m, 2 H), 1.71–1.62 (m, 2 H), 1.53–1.49 (m, 1 H), 1.29 (d, $J = 5.8$ Hz, 1 H), 1.16 (s, 3 H), 0.62 (d, $J = 5.7$ Hz, 1 H); ^{13}C NMR (125 MHz, CDCl_3) δ 176.0, 170.6, 132.2, 126.7, 69.6, 54.2, 51.9, 35.82, 35.76, 32.6, 31.7, 31.3, 22.3, 21.3, 19.6; HRMS (ESI) calcd for $\text{C}_{15}\text{H}_{20}\text{O}_4\text{Na}^+$ [$\text{M} + \text{Na}^+$] 287.1254, found: 287.1246.

Synthetic application

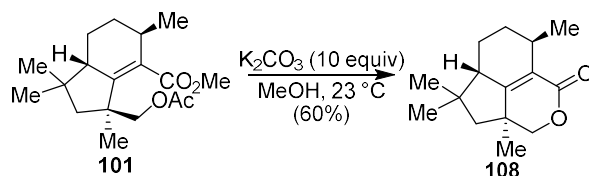


Triflate 102. To a solution of $i\text{-Pr}_2\text{NH}$ (0.440 mL, 3.20 mmol, 3.2 equiv) in THF (3 mL) at $0\text{ }^\circ\text{C}$ was added $n\text{-BuLi}$ (1.20 mL, 2.5 M in hexanes, 3.00 mmol, 3.0 equiv) and the resultant solution was stirred at $0\text{ }^\circ\text{C}$ for 15 min. The reaction contents were then cooled to $-78\text{ }^\circ\text{C}$ and a solution of **103** (0.208 g, 1.00 mmol, 1.0 equiv) in THF (5 mL) was added dropwise over the course of 5 min. The reaction mixture was then warmed to $0\text{ }^\circ\text{C}$ and kept at that temperature for 90 min

before being cooled to $-78\text{ }^{\circ}\text{C}$. HMPA (0.50 mL) and a solution of Mander's reagent (0.280 g, 3.30 mmol, 3.3 equiv) in THF (2 mL) were then added sequentially. After stirring the resultant mixture at $-78\text{ }^{\circ}\text{C}$ for 1 h, LDA (prepared from *n*-BuLi and *i*-Pr₂NH as described, 10.0 mL, 1.0 M in THF, 10.0 mmol, 10 equiv) was added at $-78\text{ }^{\circ}\text{C}$ and was followed by the addition of Tf₂O (1.68 mL, 10.0 mmol, 10 equiv). The reaction contents were stirred at $-78\text{ }^{\circ}\text{C}$ for 4 h before being quenched by the addition of saturated aqueous NaHCO₃ (10 mL). The mixture was then warmed to $23\text{ }^{\circ}\text{C}$, H₂O (10 mL) was added to dissolve any remaining solids, and the reaction contents was poured into a separatory funnel and the layers were separated. The aqueous layer was then extracted with Et₂O (2 × 10 mL). The combined organic layers were washed with saturated aqueous NaHCO₃ (100 mL), H₂O (10 mL), and brine (10 mL), dried (Na₂SO₄), filtered, and concentrated. The resultant residue was purified by flash column chromatography (Et₃N-buffered silica gel, hexanes) to provide triflate **102** (0.314 g, 79% yield) as a colorless oil. **102**: $R_f = 0.65$ (silica gel, hexanes:EtOAc, 15:1); $[\alpha]_D^{20} = +94.0^{\circ}$ ($c = 0.53$, CHCl₃); IR (film) ν_{max} 2968, 1731, 1421, 1245, 1203, 1142, 1065, 1048, 961, 891, 810 cm⁻¹; ¹H NMR (500 MHz, CDCl₃) δ 4.91 (s, 1 H), 4.68 (s, 1 H), 3.80 (s, 3 H), 2.75–2.72 (m, 1 H), 2.41–2.39 (m, 1 H), 2.16 (d, $J = 13.1$ Hz, 1 H), 1.98 (d, $J = 13.1$ Hz, 1 H), 1.91–1.87 (m, 3 H), 1.78 (s, 3 H), 1.38–1.33 (m, 1 H), 1.18 (d, $J = 7.0$ Hz, 3 H), 1.05 (s, 3 H), 1.01 (s, 3 H); ¹³C NMR (125 MHz, CDCl₃) δ 165.6, 151.4, 142.5, 131.9, 122.1, 119.6, 117.0, 115.3, 114.5, 52.0, 48.2, 45.9, 39.0, 31.5, 27.5, 27.2, 26.6, 25.5, 22.3, 20.9; HRMS (ESI) calcd for C₁₇H₂₆O₅F₃S⁺ [M + H⁺] 399.1448, found: 399.1436.

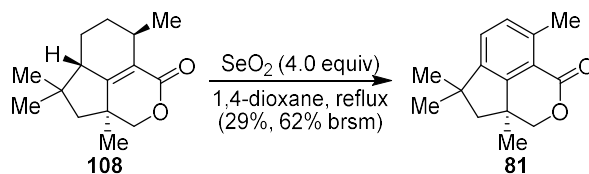


Acetate 101. To a flame-dried sealed tube was added Pd(OAc)₂ (0.112 g, 0.500 mmol, 10 mol %), *t*-BuMephos (0.234 g, 0.75 mmol, 15 mol %), and *n*-Bu₄NOAc (4.52 g, 15.0 mmol, 3.0 equiv). A solution of triflate **102** (1.99 g, 5.00 mmol, 1.0 equiv) in EtOAc (50 mL) was then added at 23 °C. The mixture was heated to 90 °C and stirred at this temperature for 12 h. Upon completion, the reaction contents were cooled to 23 °C and filtered through a pad of silica gel (EtOAc). The filtrate was concentrated, and the resultant residue was purified by flash column chromatography (silica gel, hexanes:EtOAc, 30:1) to provide acetate **101** (1.03 g, 67% yield) as a pale-yellow oil. **101**: R_f = 0.48 (silica gel, hexanes:EtOAc, 10:1); [α]_D²⁰ = +23.0° (*c* = 1.0 in CHCl₃); IR (film) ν_{max} 2932, 2865, 1742, 1726, 1373, 1226, 1061, 1030 cm⁻¹; ¹H NMR (500 MHz, CDCl₃) δ 4.24 (d, *J* = 10.3 Hz, 1 H), 4.05 (d, *J* = 10.3 Hz, 1 H), 3.71 (s, 3 H), 2.56–2.48 (m, 1 H), 2.15–2.11 (m, 1 H), 2.06 (s, 3 H), 1.93–1.88 (m, 1 H), 1.73 (d, *J* = 13.3 Hz, 1 H), 1.71–1.66 (m, 1 H), 1.40 (d, *J* = 13.2 Hz, 1 H), 1.25–1.06 (m, 2 H), 1.11 (s, 3 H), 1.00 (s, 3 H), 0.91 (d, *J* = 6.8 Hz, 3 H), 0.78 (s, 3 H); ¹³C NMR (125 MHz, CDCl₃) δ 171.2, 170.8, 149.8, 131.5, 73.1, 53.7, 53.1, 51.1, 44.1, 39.0, 33.1, 31.4, 27.9, 23.7, 22.9, 22.1, 21.0, 20.1; HRMS (ESI) calcd for C₁₈H₂₉O₄⁺ [M + H⁺] 309.2060, found: 309.2063.



Lactone 108. To a solution of acetate **101** (0.616 g, 2.00 mmol, 1.0 equiv) in MeOH (20 mL) at 23 °C was added K₂CO₃ (2.76 g, 20 mmol, 10 equiv), and the resultant mixture was stirred at 23 °C for 10 h. Upon completion, the reaction contents were diluted with EtOAc (10 mL) and quenched by the addition of saturated aqueous NH₄Cl (10 mL). The mixture was poured into a separatory funnel and the layers were separated. The aqueous layer was then extracted with EtOAc (2 × 10 mL). The combined organic layers were washed with H₂O (2 × 10 mL) and brine (10 mL)

before being dried (Na₂SO₄), filtered, and concentrated. The resultant residue was purified by flash column chromatography (silica gel, hexanes:EtOAc, 20:1→10:1) to provide lactone **108** (0.281 g, 60% yield) as a colorless solid. **108**: *R_f* = 0.58 (silica gel, hexanes:EtOAc, 5:1); [α]_D²⁰ = -53.0° (*c* = 0.30 in CHCl₃); IR (film) ν_{max} 2957, 2869, 1714, 1462, 1388, 1368, 1256, 1209, 1165, 1070, 1049, 1000, 974, 788 cm⁻¹; ¹H NMR (500 MHz, CDCl₃) δ 4.08 (d, *J* = 10.4 Hz, 1 H), 4.01 (d, *J* = 10.4 Hz, 1 H), 2.78–2.72 (m, 1 H), 2.45 (ddd, *J* = 10.3, 6.8, 2.3 Hz, 1 H), 1.68–1.58 (m, 1 H), 1.56 (d, *J* = 14.1 Hz, 1 H), 1.50, (d, *J* = 13.9 Hz, 1 H), 1.39–1.31 (m, 1 H), 1.24 (s, 3 H), 1.16 (s, 3 H), 1.02, (d, *J* = 7.0 Hz, 1 H), 0.83 (s, 3 H); ¹³C NMR (125 MHz, CDCl₃) δ 166.5, 124.6, 50.4, 49.1, 41.6, 38.4, 29.3, 29.2, 25.9, 25.1, 25.0, 19.3, 16.1; HRMS (ESI) calcd for C₁₅H₂₃O₂⁺ [*M* + H⁺] 235.1693, found: 235.1700.

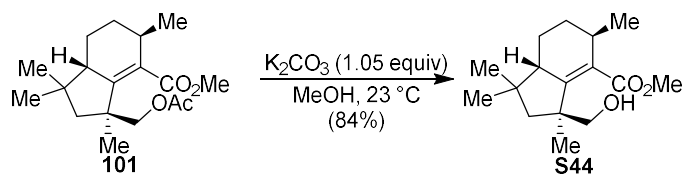


10-oxodehydrodihydrobotrydial (81). Lactone **108** (46.8 mg, 0.200 mmol, 1.0 equiv) in 1,4-dioxane (2 mL) was treated with SeO₂ (88.8 mg, 0.800 mmol, 4.0 equiv). The resultant mixture was then heated at 110 °C for 12 h. Upon completion, the resultant mixture was cooled to 23 °C and filtered through a pad of silica gel (EtOAc). The filtrate was then concentrated, and the resultant residue was purified by column chromatography (hexanes: EtOAc = 100:1→50:1) to provide recovered starting material (24.9 mg) along with 10-oxodehydrodihydrobotrydial (**81**, 13.3 mg, 29% yield, 62% yield brsm) as a white solid. **81**: *R_f* = 0.68 (silica gel, hexanes: EtOAc = 5:1); [α]_D²⁰ = -95.0° (*c* = 1.0 in CHCl₃); lit: [α]_D²⁸ = -166.5° (*c* = 1.0 in CHCl₃)^[28a]; as such the rotation data showed this should be the enantiomer of natural **81** ([α]_D²⁸ = +87° (*c* = 3.9 mg/ml in CHCl₃)^[28b]); IR (film) ν_{max} 2961, 2927, 2867, 1759, 1584, 1484, 1245, 1067, 829, 801, 532 cm⁻¹; ¹H NMR (500 MHz, CDCl₃) δ 7.21 (d, *J* = 7.7 Hz, 1 H), 7.15 (d, *J* = 7.7 Hz, 1 H), 4.36 (d, *J* = 10.0

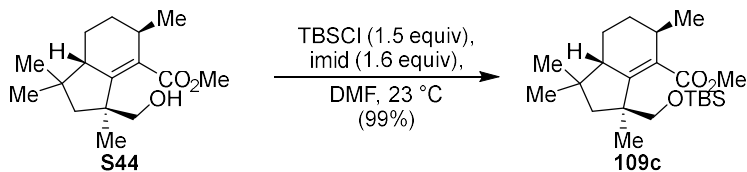
Hz, 1 H), 4.13 (d, $J = 10.0$ Hz, 1 H), 2.62 (s, 3 H), 1.98 (d, $J = 12.8$ Hz, 1 H), 1.86 (d, $J = 12.9$ Hz, 1 H), 1.51 (s, 3 H), 1.45 (s, 3 H), 1.32 (s, 3 H); ^{13}C NMR (125 MHz, CDCl_3) δ 164.0, 151.4, 147.0, 139.6, 131.7, 127.2, 119.6, 79.2, 52.1, 45.1, 40.8, 30.8, 30.7, 24.7, 20.2; HRMS (ESI) calcd for $\text{C}_{15}\text{H}_{19}\text{O}_2^+$ [$\text{M} + \text{H}^+$] 231.1380, found: 231.1388.

Table 2-S1: Comparison of literature and obtained spectral values for *10-oxodehydrodihydrobotrydial (58)*^[28b]

^1H NMR in CDCl_3	
Natural (Collado, 400 MHz)	Synthetic (This work, 500 MHz)
7.20 (d, $J = 7.8$ Hz, 1 H)	7.21 (d, $J = 7.7$ Hz, 1 H)
7.14 (d, $J = 7.8$ Hz, 1 H)	7.15 (d, $J = 7.7$ Hz, 1 H)
4.35 (d, $J = 10.1$ Hz, 1 H)	4.36 (d, $J = 10.0$ Hz, 1 H)
4.12 (d, $J = 10.1$ Hz, 1 H)	4.13 (d, $J = 10.0$ Hz, 1 H)
2.61 (s, 3 H)	2.62 (s, 3 H)
1.96 (d, $J = 13.0$ Hz, 1 H)	1.98 (d, $J = 13.0$ Hz, 1 H)
1.84 (d, $J = 13.0$ Hz, 1 H)	1.86 (d, $J = 13.0$ Hz, 1 H)
1.50 (s, 3 H)	1.50 (s, 3 H)
1.44 (s, 3 H)	1.45 (s, 3 H)
1.31 (s, 3 H)	1.32 (s, 3 H)
^{13}C NMR in CDCl_3	
Natural (Collado, 100 MHz)	Synthetic (This work, 125 MHz)
164.0	164.0
151.4	151.4
147.0	147.0
139.6	139.6
131.7	131.7
127.3	127.2
119.6	119.6
79.2	79.2
52.1	52.1
45.1	45.1
40.4	40.5
30.7	30.8
30.7	30.7
24.7	24.7
20.3	20.2

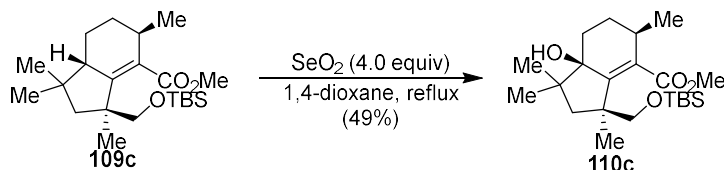


Alcohol S44. To a solution of acetate **101** (1.23 g, 4.00 mmol, 1.0 equiv) in MeOH (40 mL) at 23 °C was added K_2CO_3 (0.580 g, 4.20 mmol, 1.05 equiv), and the resultant mixture was stirred at 23 °C for 10 h. Upon completion, the reaction mixture was quenched by the addition of saturated aqueous NH_4Cl (30 mL), poured into a separatory funnel, and the resultant layers were separated. The aqueous layer was then extracted with EtOAc (2×30 mL). The combined organic layers were washed with H_2O (30 mL) and brine (30 mL) before being dried (Na_2SO_4), filtered, and concentrated. The resultant residue was purified by flash column chromatography (silica gel, hexanes:EtOAc, 10:1 \rightarrow 5:1) to provide alcohol **S44** (0.894 g, 84% yield) as a white solid. **S44**: $R_f = 0.33$ (silica gel, hexanes: EtOAc = 5:1); $[\alpha]_D^{20} = -19.0^\circ$ ($c = 0.53$ in $CHCl_3$); IR (film) ν_{max} 3438, 2921, 2864, 1718, 1656, 1446, 1366, 1261, 1225, 1060, 981, 750 cm^{-1} ; 1H NMR (500 MHz, $CDCl_3$) 3.75 (s, 3 H), 3.45–3.40 (m, 2 H), 3.21 (dd, $J = 6.6, 5.8$ Hz, 1 H), 2.56–2.48 (m, 1 H), 2.14–2.10 (m, 1 H), 1.93–1.88 (m, 1 H), 1.72–1.68 (m, 1 H), 1.59 (d, $J = 13.0$ Hz, 1 H), 1.40 (d, $J = 12.9$ Hz, 1 H), 1.25–1.08 (m, 2 H), 1.10 (s, 3 H), 1.01 (s, 3 H), 0.93 (d, $J = 6.8$ Hz, 3 H), 0.79 (s, 3 H); ^{13}C NMR (125 MHz, $CDCl_3$) δ 172.2, 151.2, 131.2, 71.7, 54.1, 53.0, 51.6, 46.6, 38.5, 33.2, 31.4, 27.8, 23.9, 22.4, 22.1, 20.2; HRMS (ESI) calcd for $C_{16}H_{27}O_3^+ [M + H^+]$ 267.1955, found: 267.1965.



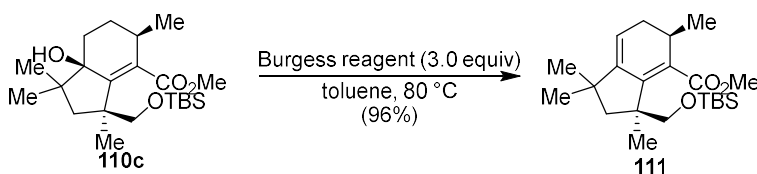
TBS-Protected Ether 109c. To a solution of alcohol **S46** (0.266 g, 1.00 mmol, 1.0 equiv) in DMF (10 mL) at 23 °C was sequentially added imidazole (0.109 g, 1.60 mmol, 1.6 equiv) and TBSCl (0.225 g, 1.50 mmol, 1.5 equiv). The resultant solution was stirred at 23 °C for 12 h. Upon

completion, the reaction contents were quenched by the addition of saturated aqueous NH_4Cl (10 mL), poured into a separatory funnel, and the resultant layers were separated. The aqueous layer was then extracted with Et_2O (2×10 mL). The combined organic layers were washed with H_2O (4×10 mL) and brine (10 mL) before being dried (Na_2SO_4), filtered, and concentrated. The resultant residue was purified by flash column chromatography (silica gel, hexanes:EtOAc, 20:1) to provide **109c** (0.380 g, 99% yield). **109c**: $R_f = 0.39$ (silica gel, hexanes:EtOAc, 20:1); $[\alpha]_D^{20} = +30.0^\circ$ ($c = 1.0$ in CHCl_3); IR (film) ν_{max} 2930, 2857, 1727, 1432, 1385, 1254, 1226, 1212, 1060, 837 cm^{-1} ; ^1H NMR (500 MHz, CDCl_3) δ 3.80 (d, $J = 8.7$ Hz, 1 H), 3.70 (s, 3 H), 3.39 (d, $J = 8.7$ Hz, 1 H), 2.54–2.48 (m, 1 H), 2.18–2.14 (m, 1 H), 2.04 (d, $J = 12.7$ Hz, 1 H), 1.90–1.86 (m, 1 H), 1.68–1.63 (m, 1 H), 1.23 (d, $J = 12.8$ Hz, 1 H), 1.17–1.08 (m, 2 H), 1.001 (s, 3 H), 0.995 (s, 3 H), 0.90 (d, $J = 6.8$ Hz, 3 H), 0.88 (s, 9 H), 0.77 (s, 3 H), 0.04 (s, 3 H), 0.03 (s, 3 H); ^{13}C NMR (125 MHz, CDCl_3) δ 171.2, 152.2, 129.4, 71.6, 53.9, 52.7, 50.9, 46.3, 38.9, 33.1, 27.9, 24.0, 22.8, 22.3, 20.1, 18.2, -5.3 , -5.4 ; HRMS (ESI) calcd for $\text{C}_{22}\text{H}_{41}\text{O}_3\text{Si}^+ [\text{M} + \text{H}^+]$ 381.2819, found: 381.2826.



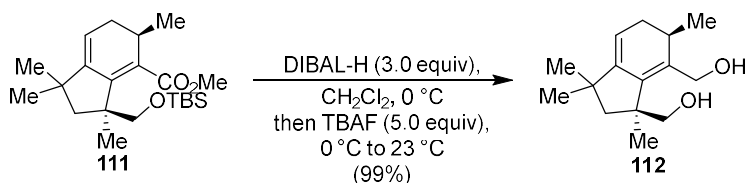
Allylic alcohol 110c. To a solution of **109c** (0.076 g, 0.200 mmol, 1.0 equiv) in 1,4-dioxane (2 mL) at 23 °C was added SeO_2 (0.089 g, 0.800 mmol, 4.0 equiv). The resultant mixture was then heated at 110 °C for 12 h. Upon completion, the resultant mixture was cooled to 23 °C and filtered through a pad of silica gel (EtOAc). The filtrate was then concentrated, and the resultant residue was purified by column chromatography (hexanes: EtOAc = 100:1→50:1) to provide allylic alcohol **110c** (0.039 g, 49% yield) as a colorless oil. **110c**: $R_f = 0.53$ (silica gel, hexanes:EtOAc, 10:1); $[\alpha]_D^{20} = +10.0^\circ$ ($c = 0.21$ in CHCl_3); IR (film) ν_{max} 3426, 2951, 2931, 2860, 1720, 1471, 1251, 1215, 1065, 782 cm^{-1} ; ^1H NMR (500 MHz, CDCl_3) δ 4.59 (d, $J = 2.0$ Hz, 1 H), 4.02 (d, $J =$

8.5 Hz, 1 H), 3.72 (s, 3 H), 3.21 (d, $J = 8.5$ Hz, 1 H), 2.49–2.44 (m, 1 H), 2.20 (d, $J = 12.8$ Hz, 1 H), 1.73–1.54 (m, 2 H), 1.39–1.34 (m, 1 H), 1.15 (d, $J = 12.8$ Hz, 1 H), 1.023 (s, 3 H), 1.017 (s, 3 H), 0.93 (d, $J = 6.9$ Hz, 1 H), 0.92 (s, 3 H), 0.85 (s, 3 H), 0.113 (s, 3 H), 0.111 (s, 3 H); ^{13}C NMR (125 MHz, CDCl_3) δ 170.8, 148.8, 134.2, 79.6, 71.8, 51.1, 49.7, 45.2, 43.4, 33.7, 27.0, 26.9, 26.7, 25.9, 23.3, 21.6, 19.8, 18.4, -5.5 , -5.6 ; HRMS (ESI) calcd for $\text{C}_{22}\text{H}_{39}\text{O}_3\text{Si}^+$ [$\text{M} + \text{H}^+ - \text{H}_2\text{O}$] 379.2663, found: 379.2665.

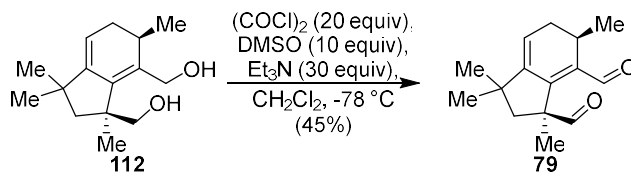


Diene 111. To a solution of allylic alcohol **110c** (0.070 g, 0.180 mmol, 1.0 equiv) in toluene (8 mL) at 23 °C was added the Burgess reagent (0.120 g, 0.500 mmol, 3.0 equiv). The resultant flask was then immersed in a pre-heated oil bath at 80 °C and stirring of the mixture was continued at 80 °C for 12 h. Upon completion, the reaction contents were cooled to 23 °C, diluted with Et_2O (10 mL), and poured into a separatory funnel. This organic layer was washed with saturated aqueous NaHCO_3 (10 mL) and brine (10 mL). The organic phase was then dried (Na_2SO_4) and concentrated. The resultant residue was purified by flash column chromatography (silica gel, hexanes:EtOAc, 10:1) to provide diene **111** (0.064 g, 96% yield) as a pale-yellow oil. **111**: $R_f = 0.62$ (silica gel, hexanes: EtOAc = 10:1); $[\alpha]_{\text{D}}^{20} = +28^\circ$ ($c = 1.0$ in CHCl_3); IR (film) ν_{max} 2955, 2928, 2859, 1711, 1463, 1432, 1230, 1096, 852, 836, 775 cm^{-1} ; ^1H NMR (500 MHz, CDCl_3) δ 5.59 (dd, $J = 5.6, 3.5$ Hz, 1 H), 3.78 (d, $J = 9.4$ Hz, 1 H), 3.74 (s, 3 H), 3.58 (d, $J = 9.4$ Hz, 1 H), 2.67–2.62 (m, 1 H), 2.42–2.36 (m, 1 H), 2.20 (d, $J = 13.2$ Hz, 1 H), 2.06–2.00 (m, 1 H), 1.40 (d, $J = 13.2$ Hz, 1 H), 1.24 (s, 3 H), 1.16 (s, 3 H), 1.11 (s, 3 H), 1.04 (d, $J = 6.9$ Hz, 1 H), 0.04 (s, 3 H), 0.02 (s, 3 H); ^{13}C NMR (125 MHz, CDCl_3) δ 169.4, 151.4, 151.0, 126.0, 118.5, 67.7, 52.3, 50.69,

47.9, 38.5, 31.4, 31.3, 31.0, 30.2, 25.8, 24.1, 18.4, -5.5, -5.6; HRMS (ESI) calcd for $C_{22}H_{39}O_3Si^+$ [M + H⁺] 379.2663, found: 379.2669.



Botrydienol (112). To a solution of diene **111** (37.8 mg, 0.100 mmol, 1.0 equiv) in CH_2Cl_2 (0.7 mL) at 0 °C was added DIBAL-H (0.3 mL, 1.0 M in CH_2Cl_2 , 0.300 mmol, 3.0 equiv) dropwise over the course of 10 min. The resultant solution was then stirred at 0 °C for 1 h. Next, TBAF (0.500 mL, 1.0 M in THF, 0.500 mmol, 5.0 equiv) was added dropwise over the course of 10 min. The resultant mixture was then warmed to 23 °C and stirred at that temperature for an additional 3 h. Upon completion, the reaction contents were quenched by the slow addition of solid $Na_2SO_4 \cdot 10H_2O$ at 0 °C until no further gas was released. The mixture was then filtered through Celite, and the residual solids were washed with EtOAc (5 × 1 mL). The combined organic layers were washed with saturated H_2O (3 × 5 mL) and brine (5 mL) before being dried (Na_2SO_4), filtered, and concentrated. The resultant residue was purified by flash column chromatography (silica gel, hexanes:EtOAc, 5:1 → 1:1) to provide botrydienol (**112**, 23.6 mg, 99% yield) as a white solid. **112**: $R_f = 0.21$ (silica gel, hexanes: EtOAc, 3:1); $[\alpha]_D^{20} = +53.0^\circ$ ($c = 0.80$ in $CHCl_3$); IR (film) ν_{max} 3337, 2955, 2924, 2862, 1597, 1563, 1449, 1395, 1219, 1119, 1034 cm^{-1} ; 1H NMR (500 MHz, $CDCl_3$) δ 5.38 (dd, $J = 5.3, 3.1$ Hz, 1 H), 4.35 (d, $J = 11.3$ Hz, 1 H), 4.20 (d, $J = 11.3$ Hz, 1 H), 3.65 (d, $J = 11.1$ Hz, 1 H), 3.53 (d, $J = 11.1$ Hz, 1 H), 2.42–2.30 (m, 2 H), 2.02 (dt, $J = 17.1, 4.5$ Hz, 1 H), 1.95 (d, $J = 13.2$ Hz, 1 H), 1.46 (d, $J = 13.2$ Hz, 1 H), 1.25 (s, 3 H), 1.15 (s, 3 H), 1.10 (s, 3 H), 1.08 (d, $J = 6.8$ Hz, 1 H); ^{13}C NMR (125 MHz, $CDCl_3$) δ 150.3, 141.1, 134.2, 114.1, 70.4, 61.9, 53.1, 47.0, 39.7, 32.6, 31.7, 31.0, 30.7, 25.8, 18.8; HRMS (CI) calcd for $C_{15}H_{23}O^+$ [M + H⁺ - H_2O] 219.1743, found: 219.1746.



Botrydienal (79). To a solution of $(\text{COCl})_2$ (0.110 mL, 1.28 mmol, 20 equiv) in CH_2Cl_2 (0.5 mL) at $-78\text{ }^\circ\text{C}$ was added DMSO (0.045 mL, 0.64 mmol, 10 equiv) dropwise over the course of 5 min. After stirring the resultant solution at $-78\text{ }^\circ\text{C}$ for 15 min, a solution of botrydienol (**112**, 15.0 mg, 0.064 mmol, 1.0 equiv) in CH_2Cl_2 (0.5 mL) was added dropwise over the course of 10 min. The resultant solution was then stirred at $-78\text{ }^\circ\text{C}$ for 3 h, at which time Et_3N (0.26 mL, 1.9 mmol, 30 equiv) was added the reaction contents were stirred at $-78\text{ }^\circ\text{C}$ for another 30 min. Upon completion, the reaction contents were quenched by the addition of H_2O (2 mL), warmed to $23\text{ }^\circ\text{C}$, poured into a separatory funnel, and the resultant layers were separated. The aqueous layer was then extracted with CH_2Cl_2 (2×2 mL). The combined organic layers were washed with H_2O (3 mL) before being dried (Na_2SO_4), filtered, and concentrated. The resultant residue was purified by flash column chromatography (silica gel, hexanes:EtOAc, 10:1 \rightarrow 5:1) to provide botrydienal (**79**, 8.2 mg, 55% yield) as a white solid. **79**: $R_f = 0.55$ (silica gel, hexanes:EtOAc, 5:1); $[\alpha]_D^{20} = +167.0^\circ$ ($c = 0.18$ in *n*-hexane); lit: $[\alpha]_D^{30} = +190^\circ$ ($c = 0.18$ in *n*-hexane)^[26]; IR (film) ν_{max} 2959, 2926, 2853, 2819, 2807, 2712, 1723, 1658, 1640, 1570, 1453, 1143, 924, 797, 404 cm^{-1} ; ^1H NMR (500 MHz, CDCl_3) δ 9.74 (s, 1 H), 9.67 (s, 1 H), 5.88 (dd, $J = 6.3, 2.8$ Hz, 1 H), 3.00–2.97 (m, 1 H), 2.45 (ddd, $J = 18.4, 8.7, 6.7$ Hz, 1 H); 2.23 (d, $J = 13.6$ Hz, 1H), 2.20 (dd, $J = 19.2, 6.3$ Hz, 1 H); 1.63 (d, $J = 13.7$ Hz, 1 H); 1.53 (s, 3 H), 1.26 (s, 3 H), 1.06 (s, 3 H), 0.92 (d, $J = 7.0$ Hz, 1 H); ^{13}C NMR (125 MHz, CDCl_3) δ 199.9, 189.8, 155.2, 149.5, 136.0, 123.7, 56.5, 50.6, 40.5, 30.62, 30.61, 28.8, 25.3, 24.2, 18.3; HRMS (ESI) calcd for $\text{C}_{15}\text{H}_{21}\text{O}_2^+ [\text{M} + \text{H}^+]$ 233.1536, found: 233.1542.

Table 2-S2: Comparison of literature and obtained spectral values for *botrydial* (**79**)^[26]

¹ H NMR in CDCl ₃	
Natural (Marumo, 100 MHz)	Synthetic (This work, 500 MHz)
9.74 (s, 1 H)	9.74 (s, 1 H)
9.67 (s, 1 H)	9.67 (s, 1 H)
5.87 (ddd, <i>J</i> = 6.1, 3.2, 0.5 Hz, 1 H)	5.88 (dd, <i>J</i> = 6.3, 2.8 Hz, 1 H)
2.98 (m, 1 H)	3.00–2.97 (m, 1 H)
2.48 (ddd, <i>J</i> = 18, 8.1, 6.1 Hz, 2 H)	2.48 (ddd, <i>J</i> = 18.4, 8.7, 6.7 Hz, 2 H)
2.24 (d, <i>J</i> = 13.9 Hz, 2 H)	2.24 (d, <i>J</i> = 13.6 Hz, 2 H)
1.63 (d, <i>J</i> = 13.9 Hz, 3 H)	1.63 (d, <i>J</i> = 13.7 Hz, 3 H)
1.54 (s, 3 H)	1.53 (s, 3 H)
1.25 (s, 3 H)	1.26 (s, 3 H)
1.06 (s, 3 H)	1.06 (s, 3 H)
¹³ C NMR in CDCl ₃	
Natural (Marumo, 25 MHz)	Synthetic (This work, 125 MHz)
199.5	199.9
189.5	189.8
154.9	155.2
149.2	149.5
135.7	136.0
123.5	123.7
56.3	56.5
50.4	50.6
40.4	40.5
30.5	30.6
30.5	30.5
28.7	28.8
25.3	25.3
24.2	24.2
18.3	18.3

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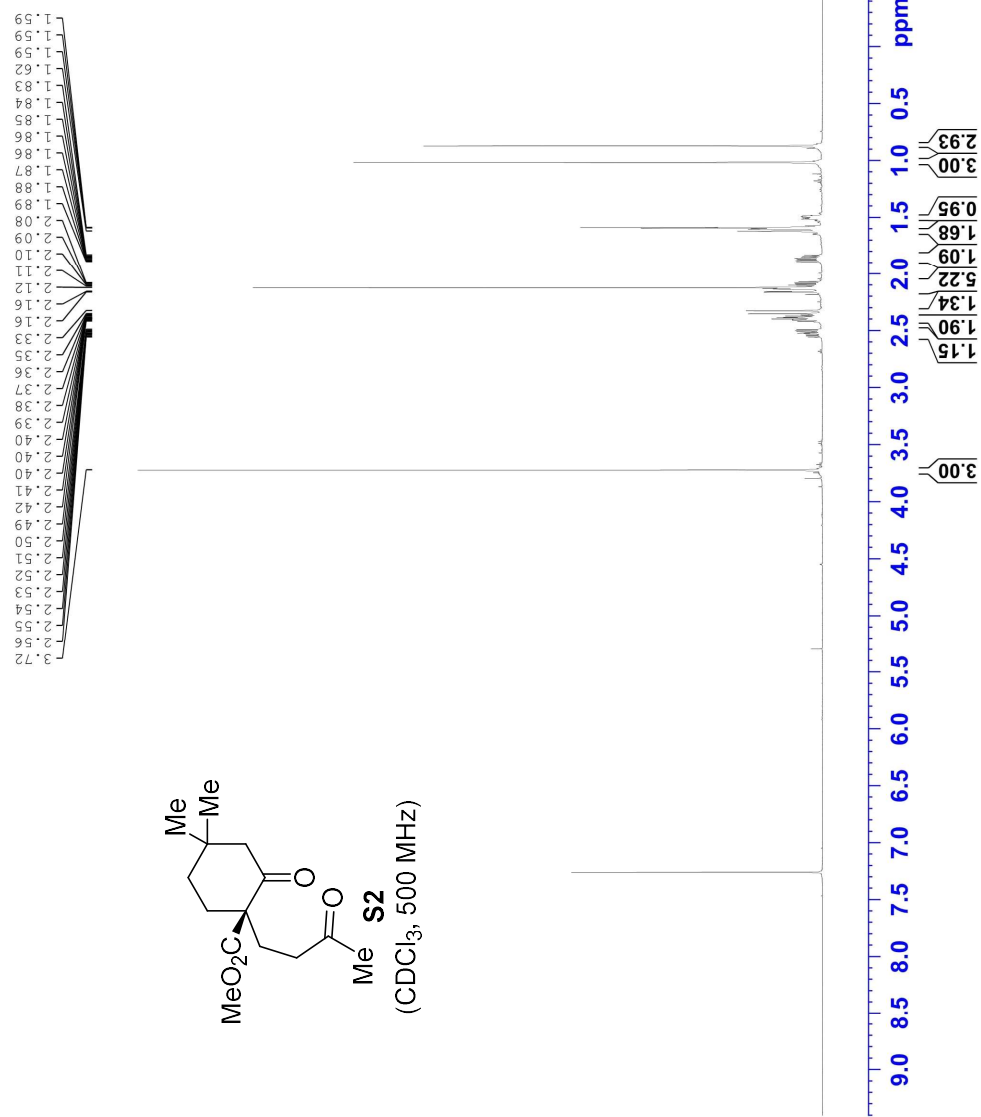
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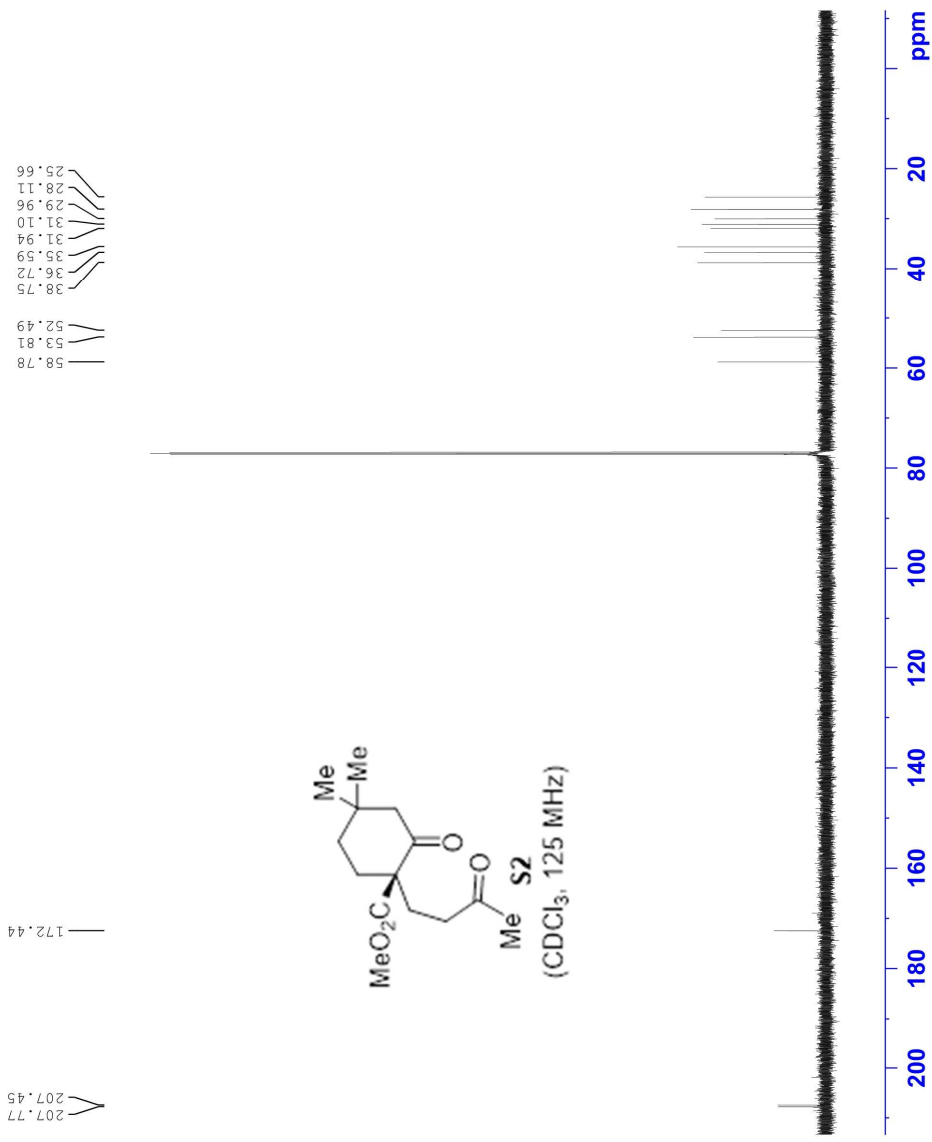
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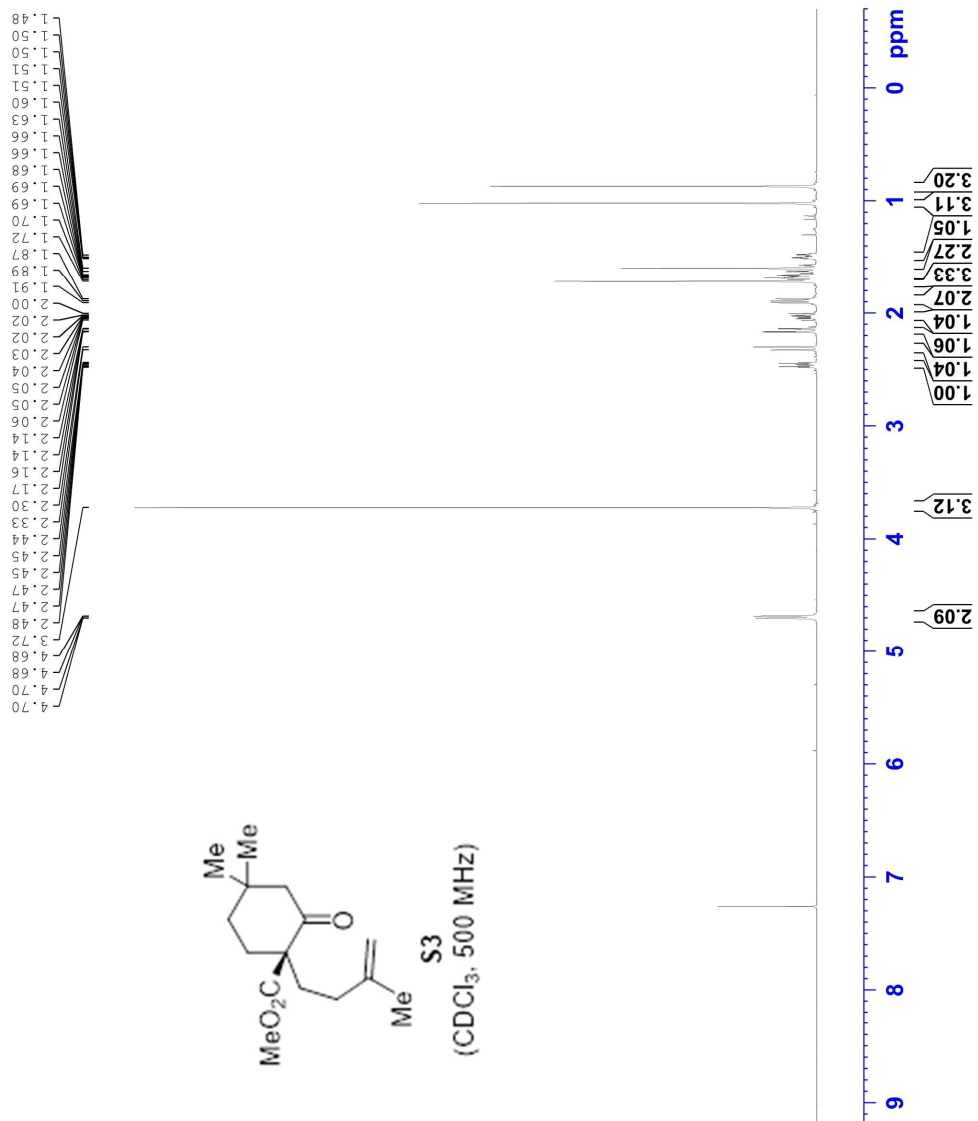


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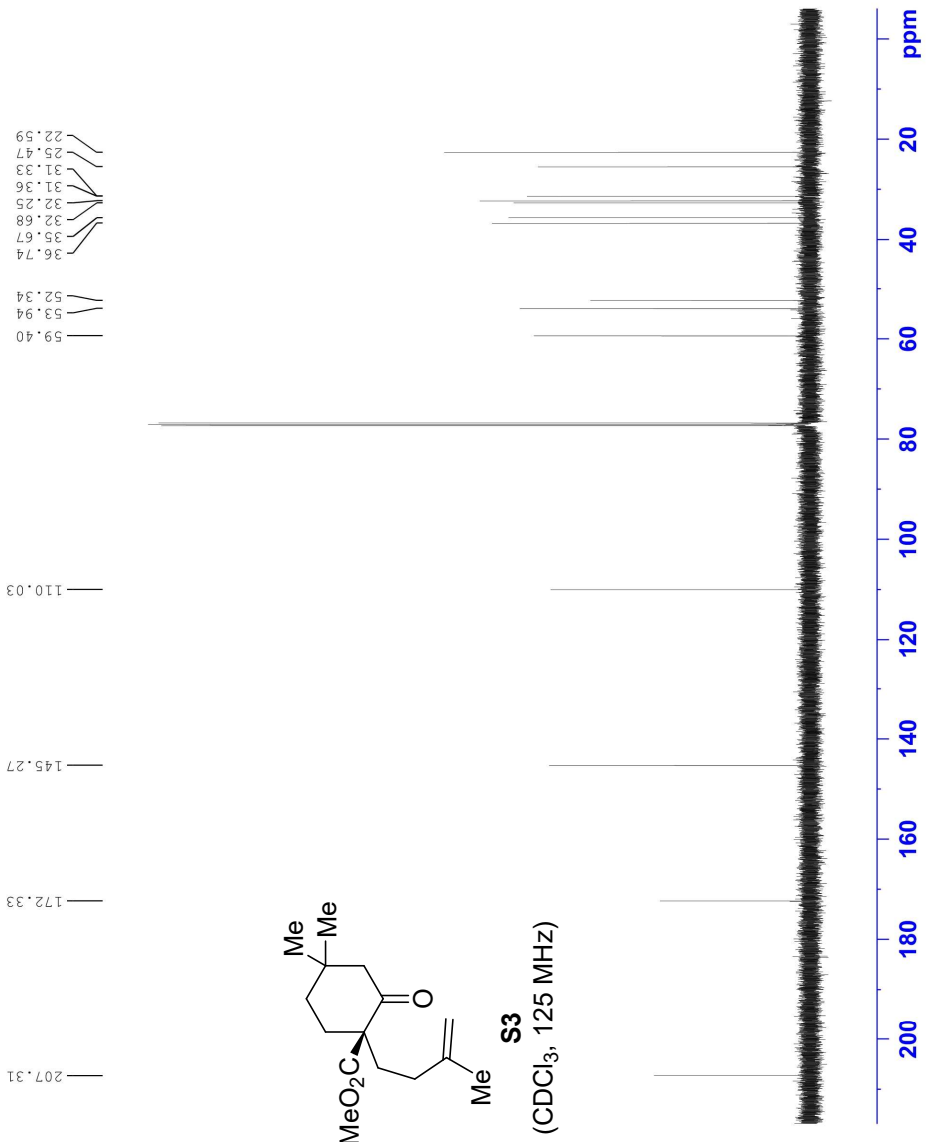
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 FC 1.40



```

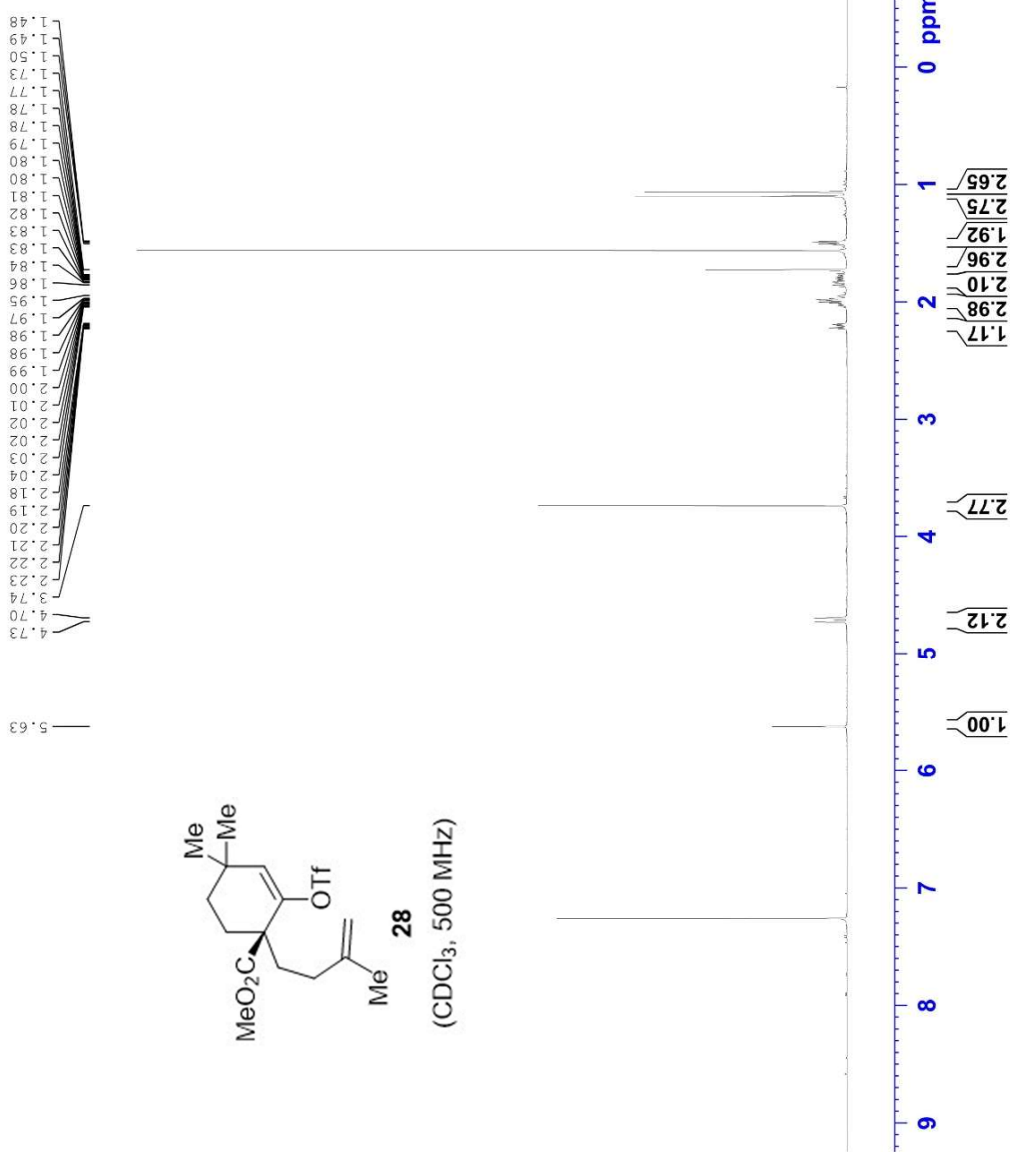
Current Data Parameters
NAME      OTf (SM)
EXPNO    1
PROCNO   1

F2 - Acquisition Parameters
Date_    20180730
Time     14.32
INSTRUM spect
PROBHD   5 mm PABBO BB/
PULPROG zg
TD       59998
SOLVENT  CDCl3
NS       16
DS       0
SWH      10000.000 Hz
FIDRES   0.166672 Hz
AQ       2.9999001 sec
RG       118.99
DW       50.000 usec
DE       6.50 usec
TE       295.5 K
D1       4.00000000 sec
TDO      1

===== CHANNEL f1 =====
SFO1    499.8730869 MHz
NUC1    1H
PL      3.58 usec
PLW1    18.25000000 W

F2 - Processing parameters
SI       65536
SF       499.8700123 MHz
WDW      EM
SSB      0
LB       0.30 Hz
GB       0
PC       1.00

```



```

Current Data Parameters
NAME          OTf(SM)
EXPNO         2
PROCNO        2

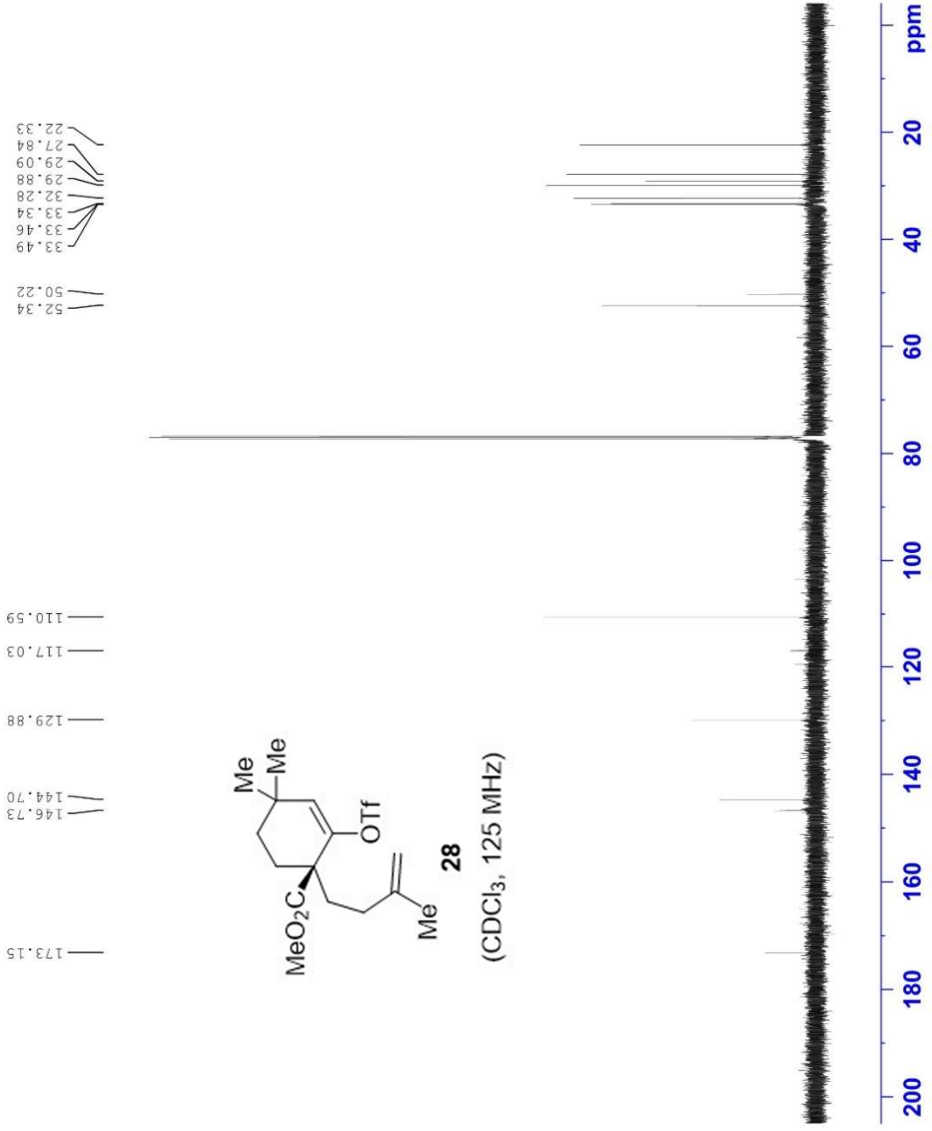
F2 - Acquisition Parameters
Date_         20180730
Time_         22.15
INSTRUM       spect
PROBHD        5 mm FAPBBO BB/
PULPROG       zgpgc
TD            187496
SOLVENT       CDCl3
NS            490
DS            0
SWH           31250.000 Hz
FIDRES        0.166670 Hz
AQ            2.9999361 sec
RG            2050
DW            16.000 usec
DE            6.50 usec
TE            297.7 K
D1            3.00000000 sec
D11           0.03000000 sec
TD0           1

===== CHANNEL f1 =====
SF01         125.7049802 MHz
NUC1         13C
P1           10.00 usec
PLW1         72.83999634 W

===== CHANNEL f2 =====
SF02         499.8724993 MHz
NUC2         1H
CPDPRG[2]    waltz16
PCPD2        80.00 usec
PLW2         19.00000000 W
PLW12        0.29688001 W

F2 - Processing parameters
SI           1048576
SF           125.6924115 MHz
WDW          EM
SSB          0
LB           0.30 Hz
GB           0
PC           1.40

```



```

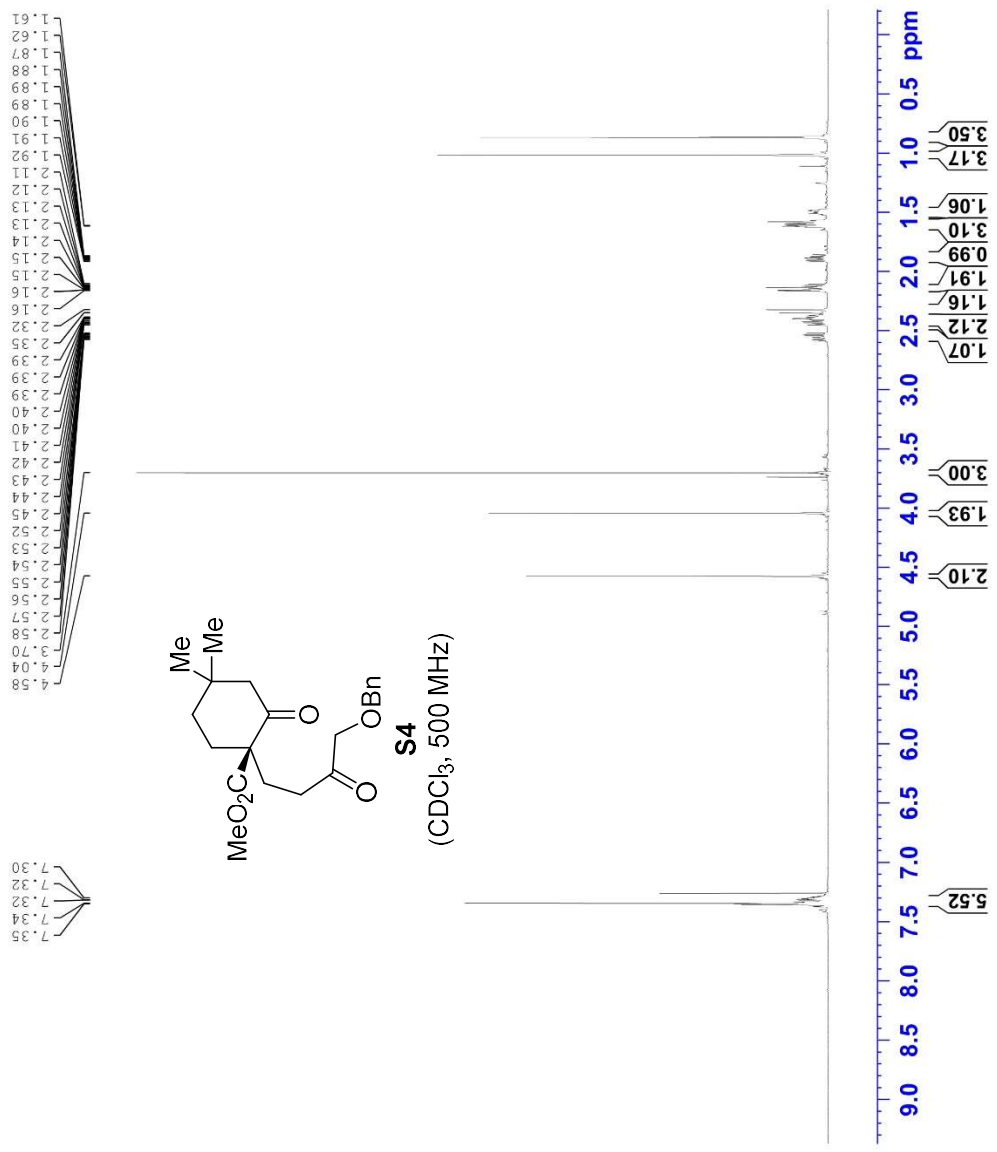
Current Data Parameters
NAME      yn-2-160-b
EXPNO     1
PROCNO    1

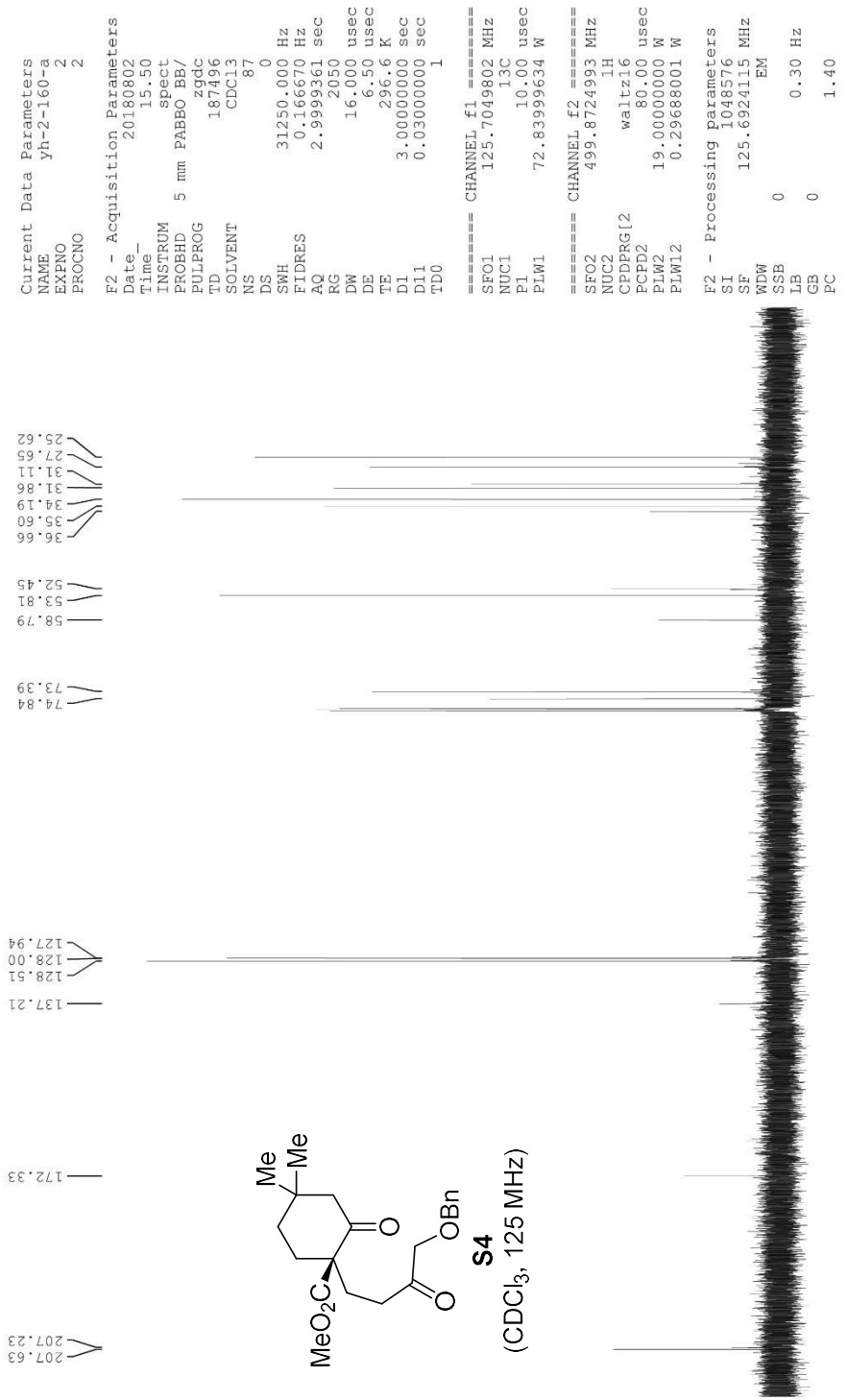
F2 - Acquisition Parameters
Date_     20191020
Time_     18.31
INSTRUM   Spect
PROBHD    5 mm PAXI LH/
PULPROG   zg
TD         59998
SOLVENT   CDC13
NS         1
DS         0
SWH        10000.000 Hz
FIDRES     0.166672 Hz
AQ         2.9999001 sec
RG         110.37
DW         50.000 usec
DE         10.00 usec
TE         297.0 K
D1         2.00000000 sec
TD0        1

===== CHANNEL f1 =====
SFO1      500.1330885 MHz
NUC1      1H
PI        9.90 usec
PLW1      12.19999981 W

F2 - Processing parameters
SI         65536
SF         500.1300133 MHz
WDW        EM
SSB        0
LB         0
GB         0
PC         1.00

```



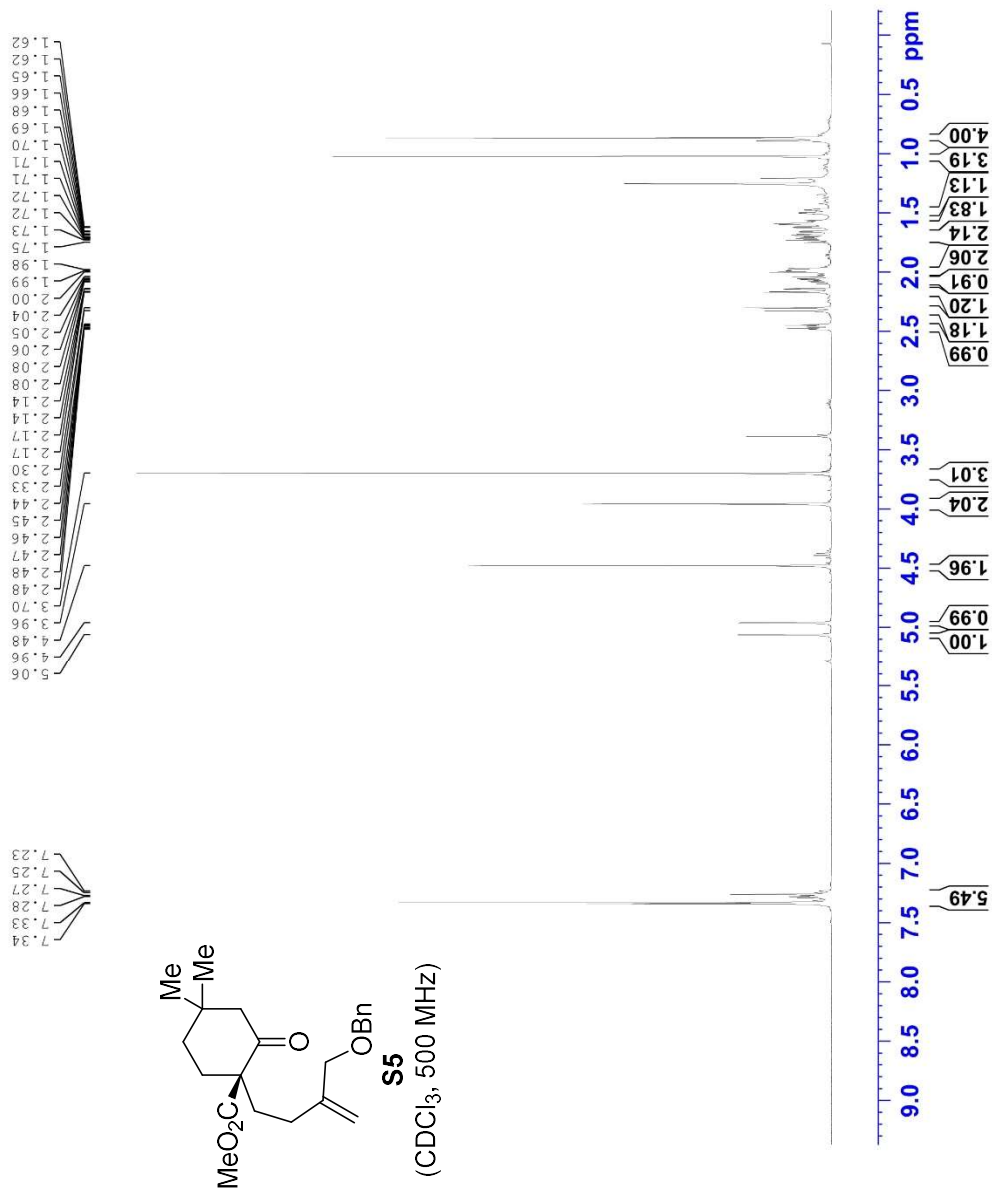


Current Data Parameters
 NAME YH-3-1-b
 EXPNO 1
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20191020
 Time_ 18.37
 INSTRUM Spect
 PROBHD 5 mm PAXI LH/
 PULPROG zg
 TD 59998
 SOLVENT CDCl3
 NS 2
 DS 0
 SWH 10000.000 Hz
 FIDRES 0.166672 Hz
 AQ 2.9999001 sec
 RG 71.76
 DW 50.000 usec
 DE 10.00 usec
 TE 296.9 K
 DI 2.00000000 sec
 TD0 1

==== CHANNEL f1 =====
 SFO1 500.1330885 MHz
 NUC1 1H
 P1 9.90 usec
 PLW1 12.1999981 W

F2 - Processing parameters
 SI 65536
 SF 500.1300134 MHz
 WDM EM
 SSB 0
 LB 0
 GB 0
 PC 1.00



```

Current Data Parameters
NAME      Yh-3-1-a
EXPNO    2
PROCNO   2

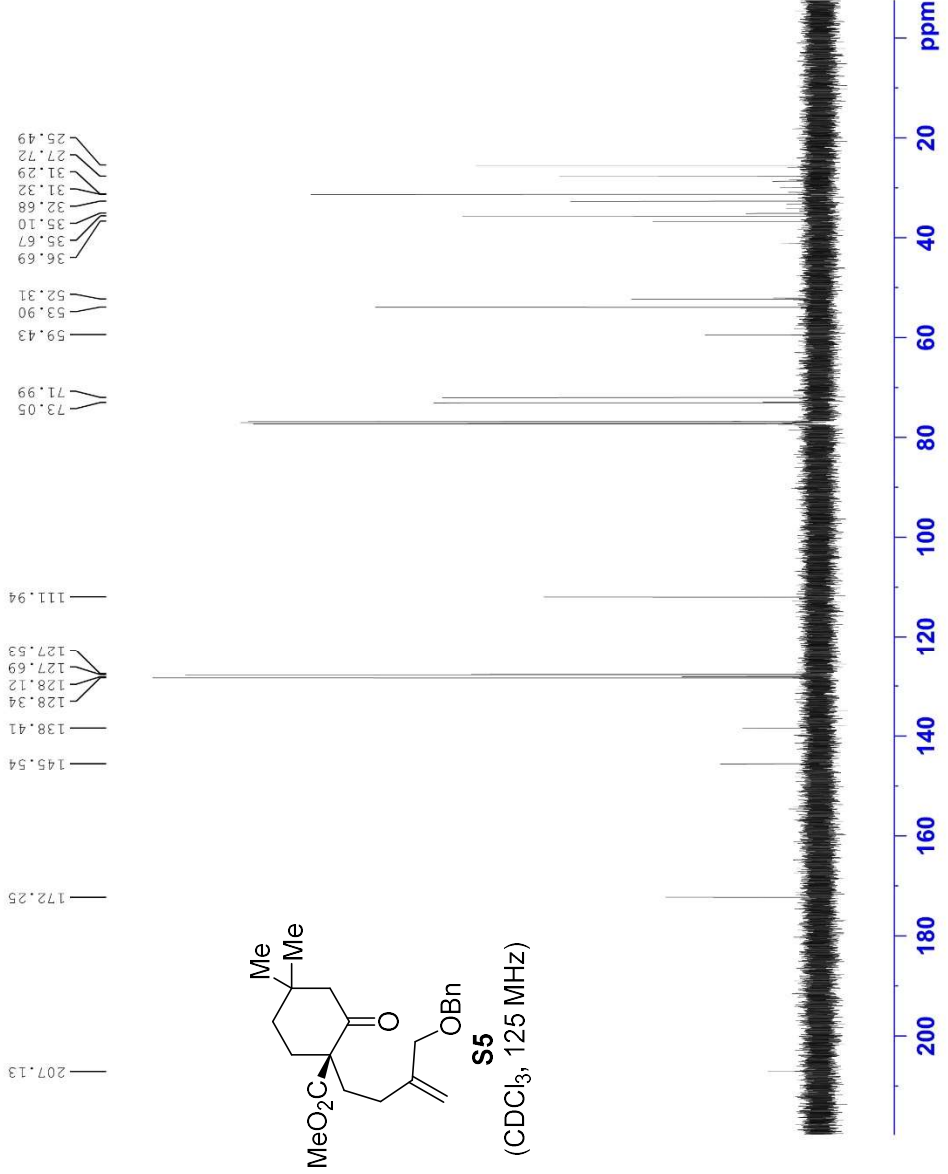
F2 - Acquisition Parameters
Date_     20180802
Time_    16.08
INSTRUM  Spect
PROBHD   5 mm PABBO BB/
PULPROG  zgdc
TD       187496
SOLVENT  CDCl3
NS       121
DS       0
SWH      31250.000 Hz
FIDRES   0.166670 Hz
AQ       2.9999361 sec
RG       2050
DW       16.000 usec
DE       6.50 usec
TE       297.0 K
D1       3.00000000 sec
D11      0.03000000 sec
TD0      1

===== CHANNEL f1 =====
SFO1    125.7049802 MHz
NUC1     13C
P1       10.00 usec
PLW1     72.83999634 W

===== CHANNEL f2 =====
SFO2    499.8724993 MHz
NUC2     1H
CPDPRG2 waltz16
PCPD2    80.00 usec
PLW2    19.00000000 W
PLW12   0.29688001 W

F2 - Processing parameters
SI       1048576
SF       125.6924115 MHz
WDW      EM
SSB      0
LB       0.30 Hz
GB       0
PC       1.40

```



S5
(CDCl₃, 125 MHz)

```

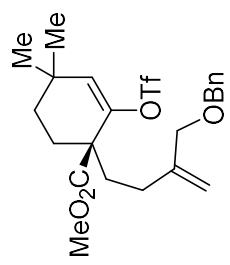
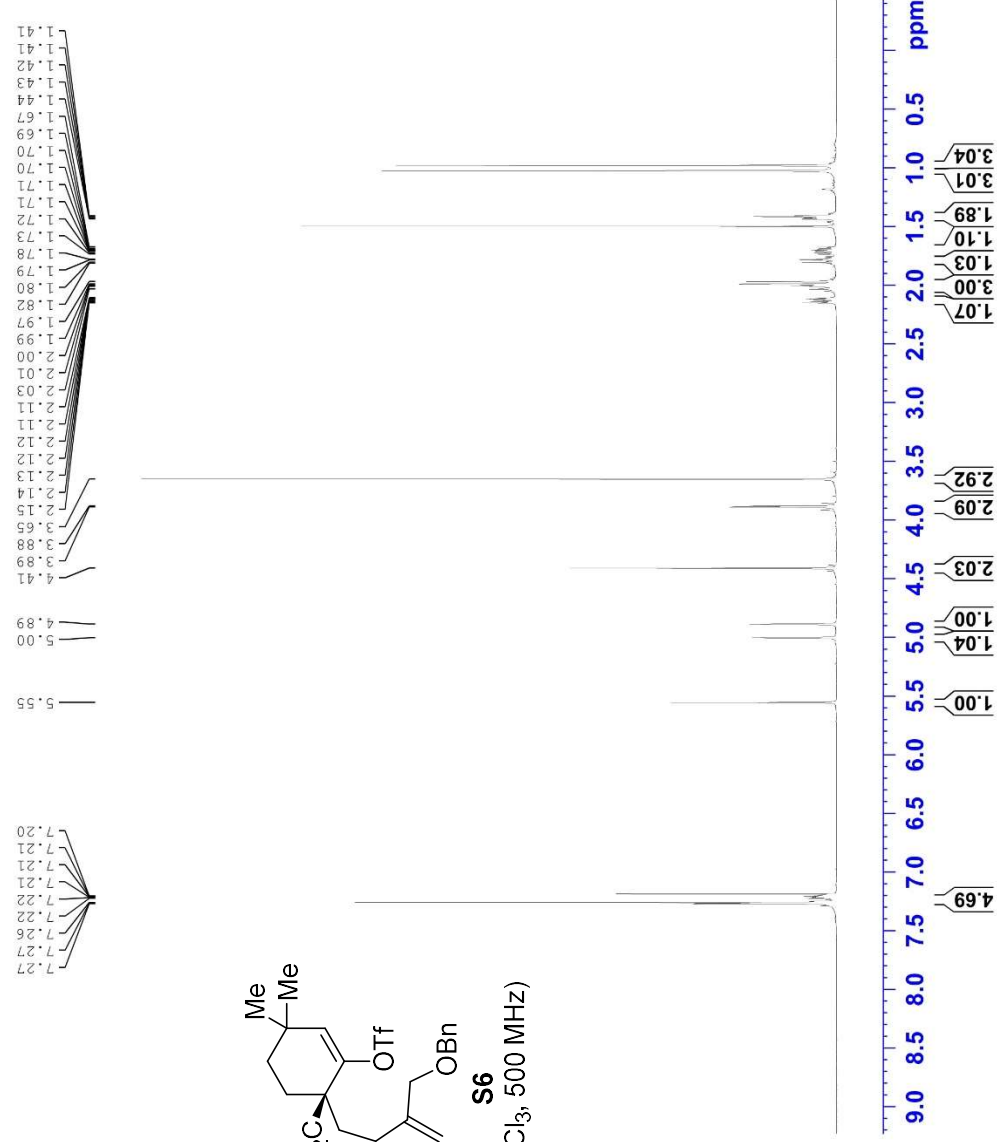
Current Data Parameters
NAME      YH-3-2-r
EXPNO    1
PROCNO   1

F2 - Acquisition Parameters
Date_    20180730
Time_    19.39
INSTRUM  Spect
PROBHD   5 mm PABBO BB/
PULPROG  zg
TD       59998
SOLVENT  CDCl3
NS       16
DS       0
SWH      10000.000 Hz
FIDRES   0.166672 Hz
AQ       2.9999001 sec
RG       86.16
DW       50.000 usec
DE       6.50 usec
TE       295.8 K
D1       4.00000000 sec
TD0      1

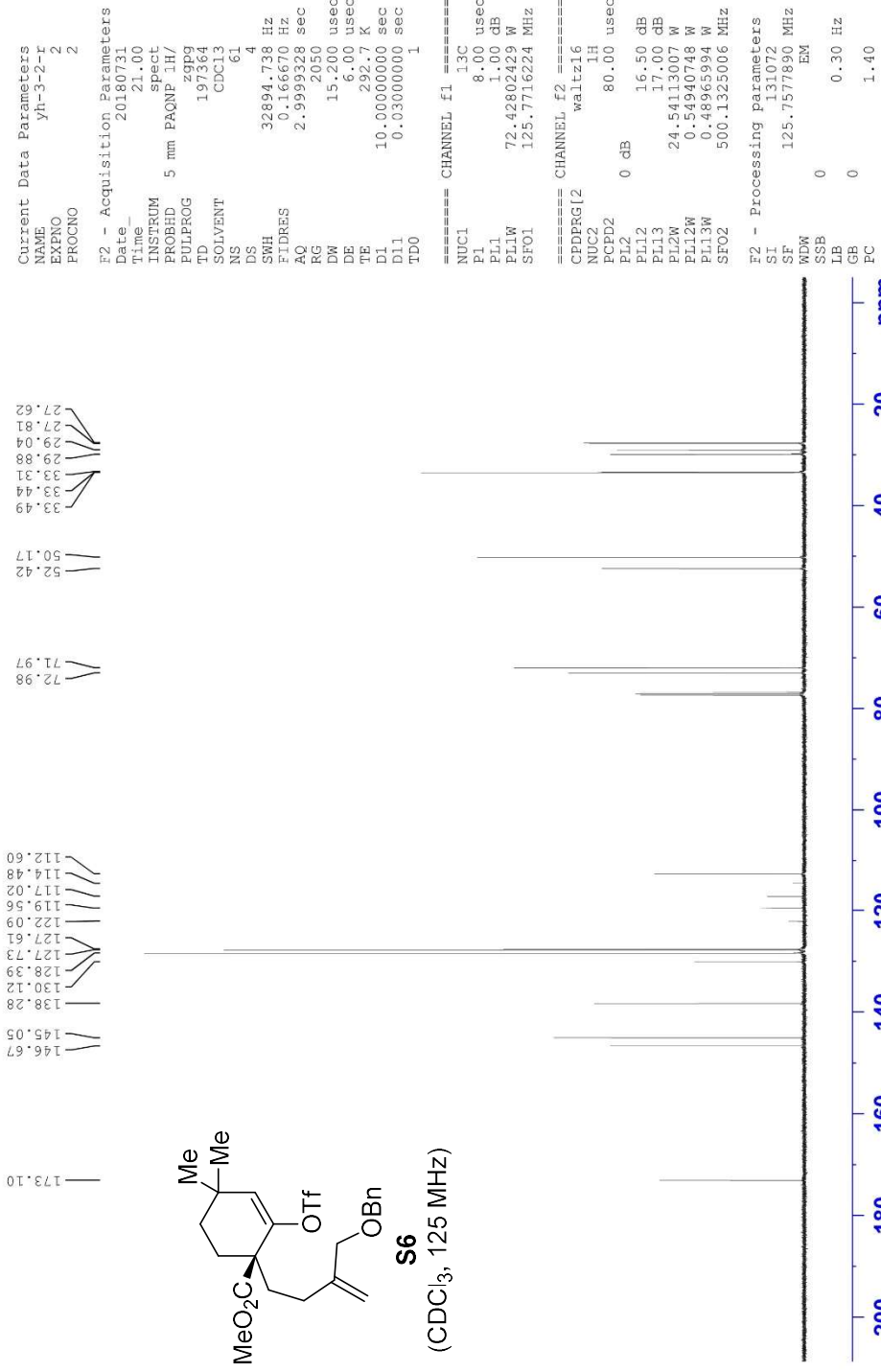
===== CHANNEL f1 =====
SFO1     499.8730869 MHz
NUC1     1H
P1       3.58 usec
PLW1     18.25000000 W

F2 - Processing parameters
SI       65536
SF       499.8700490 MHz
WDW      EM
SSB      0
LB       0.30 Hz
GB       0
PC       1.00

```



S6
(CDCl₃, 500 MHz)



```

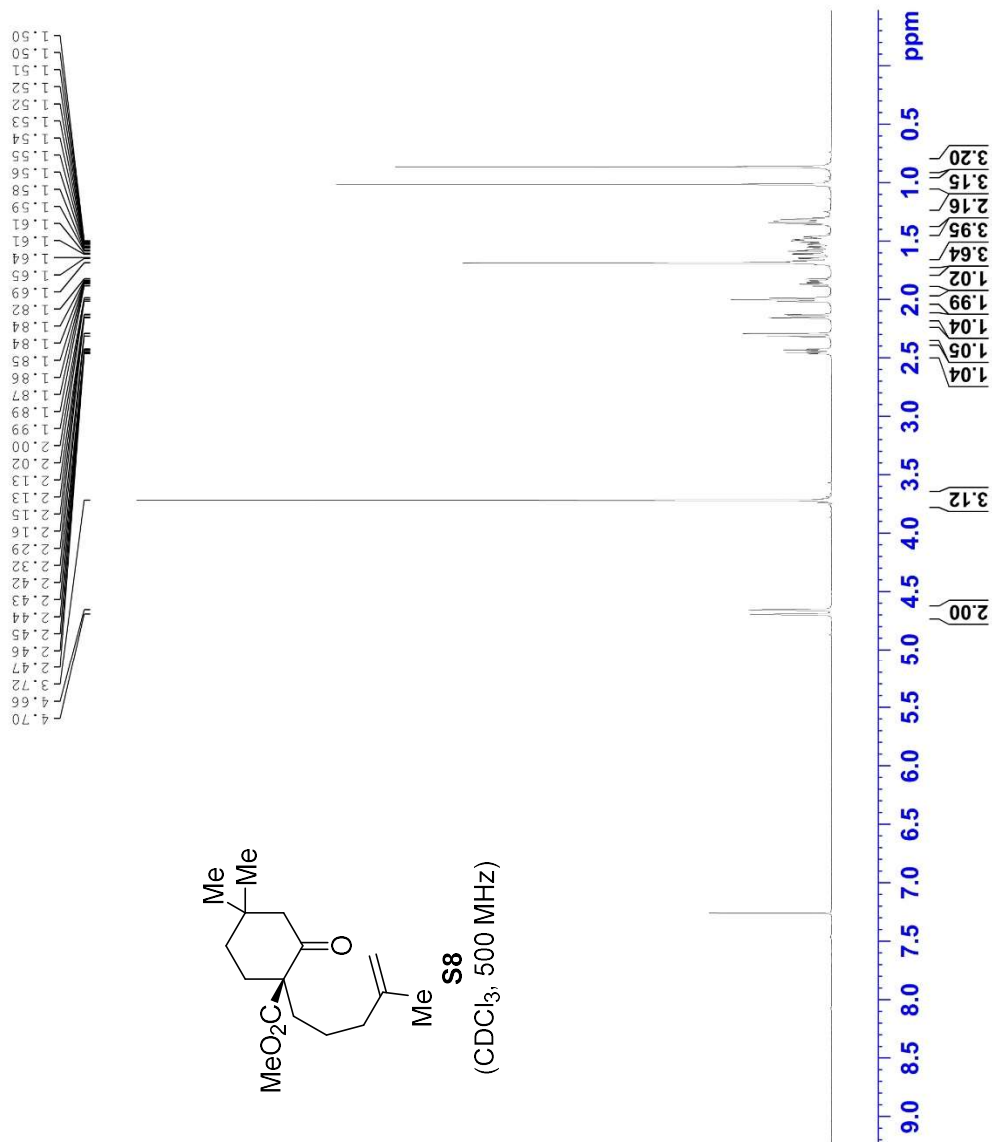
Current Data Parameters
NAME       Yh-5-17-a
EXPNO     1
PROCNO    1

F2 - Acquisition Parameters
Date_     20180812
Time_    20.00
INSTRUM  Spect
PROBHD   5 mm PAXI LH/
PULPROG  zg
TD        59998
SOLVENT  CDCl3
NS        3
DS        0
SWH       10000.000 Hz
FIDRES    0.166672 Hz
AQ         2.9999001 sec
RG         141.13
DW         50.000 usec
DE         10.00 usec
TE         294.3 K
D1         3.00000000 sec
TD0        1

===== CHANNEL f1 =====
SFO1     500.1330885 MHz
NUC1      1H
P1        2.67 usec
PLW1     12.1999981 W

F2 - Processing parameters
SI        65536
SF        500.1300138 MHz
WDW       EM
SSB       0
LB        0.30 Hz
GB        0
PC        1.00

```



```

Current Data Parameters
NAME      yh-5-17-a
EXENO    2
PROCNO    2

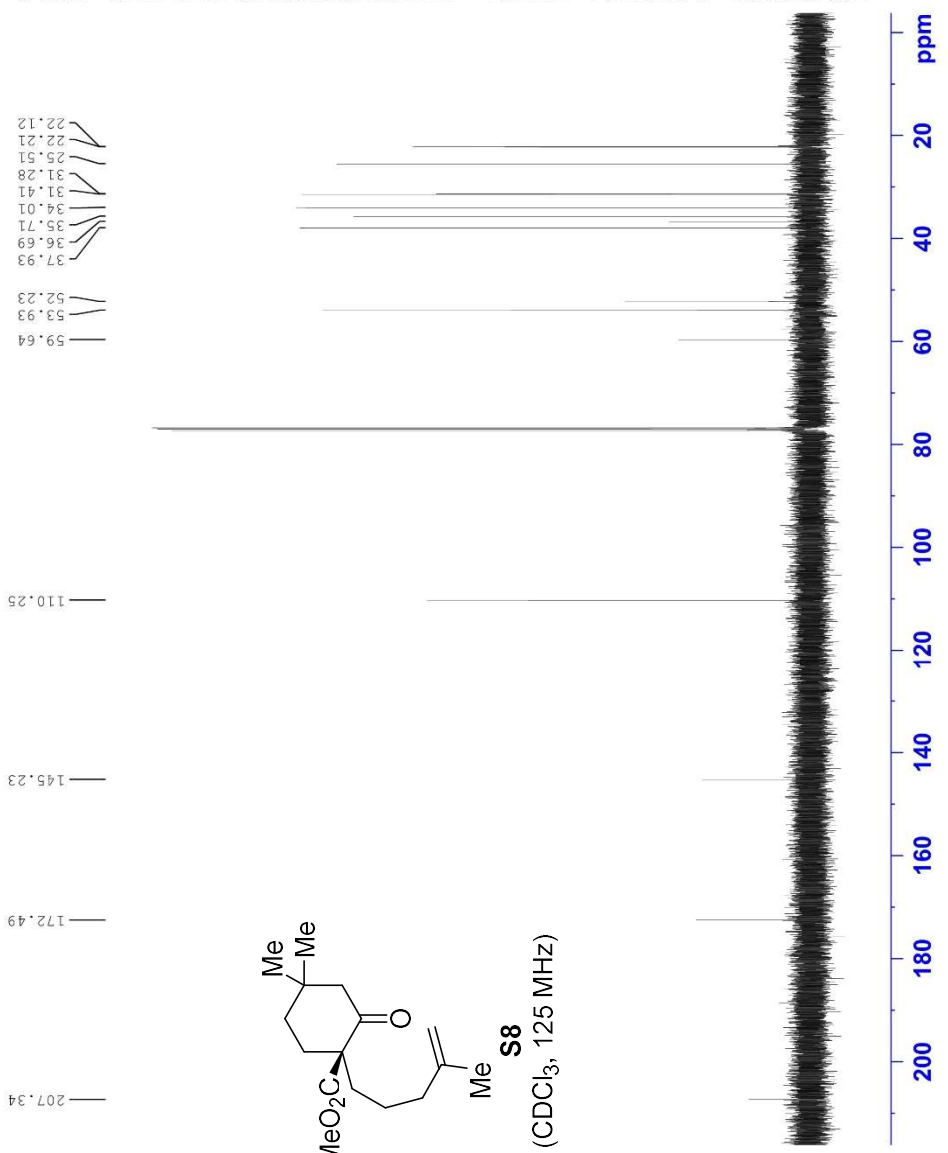
F2 - Acquisition Parameters
Date      20180812
Time      21.14
INSTRUM   spect
PROBHD    5 mm PABBO BB/
PULPROG   zgpgc
TD        187496
SOLVENT   CDCl3
NS        153
DS        0
SWH       31250.000 Hz
FIDRES    0.166670 Hz
AQ        2.9999361 sec
RG        2050
DW        16.000 usec
DE        6.50 usec
TE        297.2 K
D1        3.0000000 sec
D11       0.0300000 sec
TDO       1

===== CHANNEL f1 =====
SF01     125.7049802 MHz
NUC1     13C
P1       10.00 usec
PLW1     72.83999634 W

===== CHANNEL f2 =====
SF02     499.8724993 MHz
NUC2     1H
waltz16
CPDPRG[2]
PCPD2    80.00 usec
PLW2     19.0000000 W
PLW12    0.29688001 W

F2 - Processing parameters
SI       1048576
SF       125.6924115 MHz
WDW      EM
SSB      0
LB       0.30 Hz
GB       0
PC       1.40

```

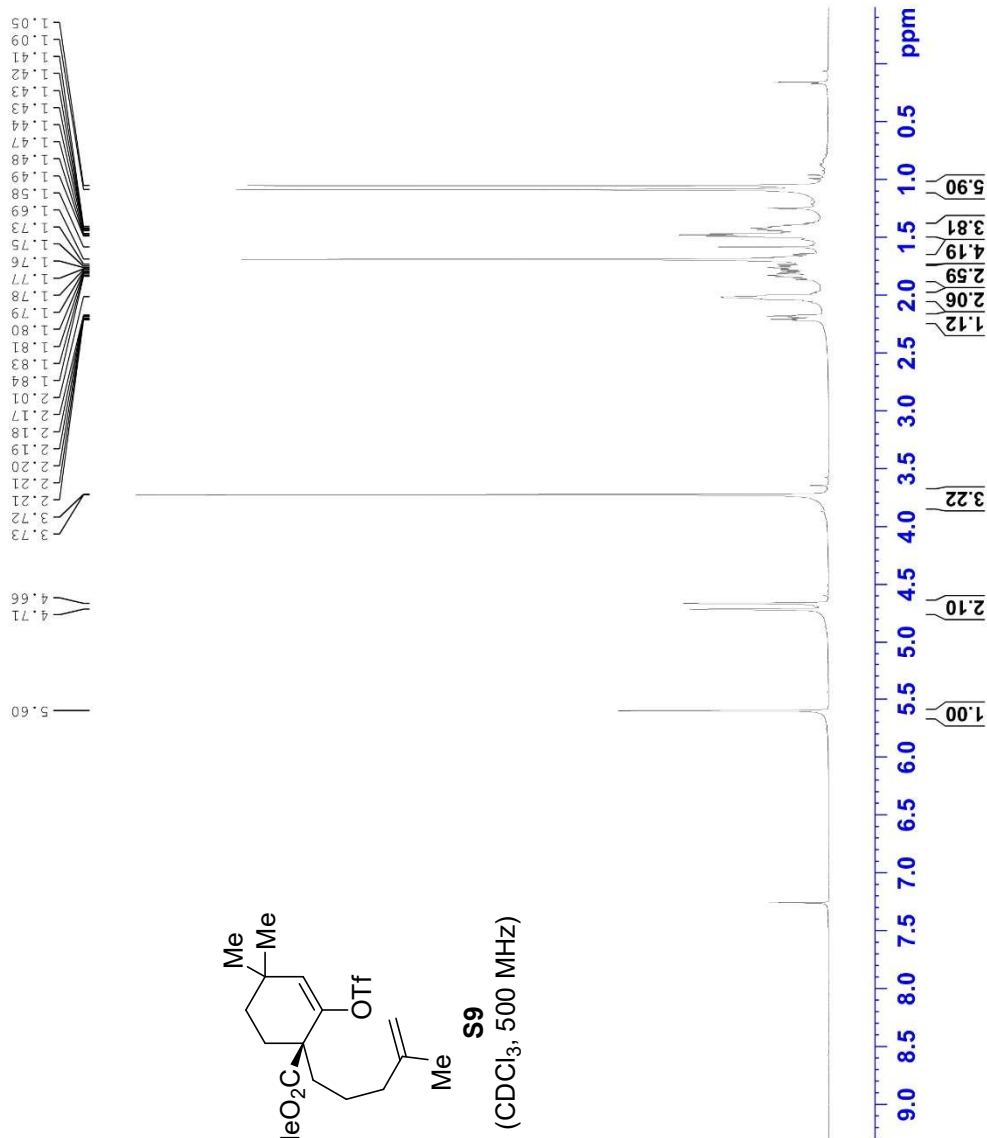


Current Data Parameters
 NAME yh-5-18-f
 EXPNO 1
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20180730
 Time_ 15.21
 INSTRUM spect
 PROBHD 5 mm PABBO BE/
 PULPROG zg
 TD 59998
 SOLVENT CDCl3
 NS 16
 DS 0
 SWH 10000.000 Hz
 FIDRES 0.166672 Hz
 AQ 2.9999001 sec
 RG 51.11
 DW 50.000 usec
 DE 6.50 usec
 TE 295.6 K
 D1 4.0000000 sec
 TDO 1

=====
 CHANNEL f1
 SF01 499.8730869 MHz
 NUC1 1H
 P1 3.58 usec
 PLW1 18.25000000 W

F2 - Processing parameters
 SI 65536
 SF 499.8700129 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00



```

Current Data Parameters
NAME      yh-5-18-r
EXENO    2
PROCNO   2

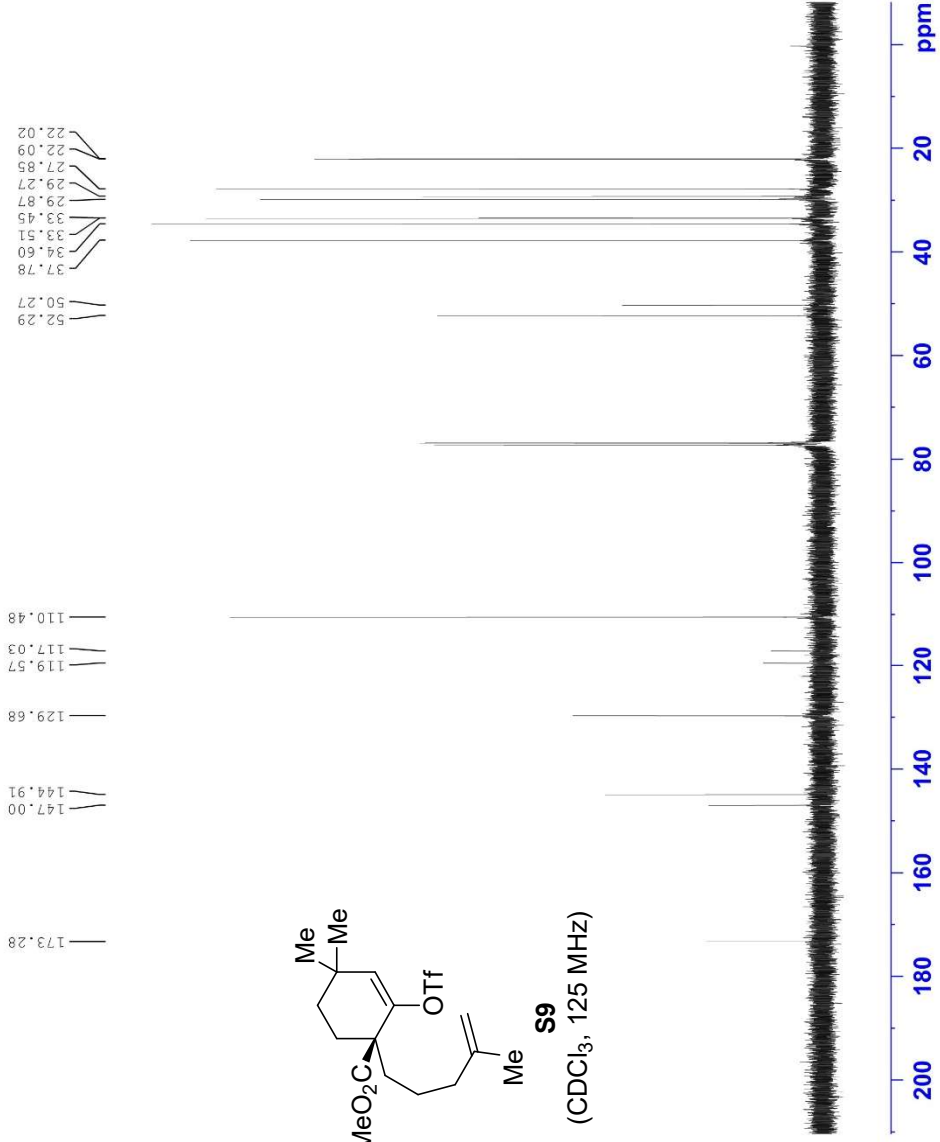
F2 - Acquisition Parameters
Date_    20180730
Time_    21.08
INSTRUM  spect
PROBHD   5 mm PABBO BE/
PULPROG  zgpgc
TD       187496
SOLVENT  CDCl3
NS       191
DS       0
SWH      31250.000 Hz
FIDRES   0.166670 Hz
AQ       2.9999361 sec
RG       2050
DW       16.000 usec
DE       6.50 usec
TE       297.3 K
D1       3.0000000 sec
D11      0.0300000 sec
TDO      1

===== CHANNEL f1 =====
SF01    125.7049802 MHz
NUC1     13C
P1       10.00 usec
PLW1    72.83999634 W

===== CHANNEL f2 =====
SF02    499.8724993 MHz
NUC2     1H
waltz16
CPDPRG[2]
PCPD2    80.00 usec
PLW2    19.0000000 W
PLW12   0.29688001 W

F2 - Processing parameters
SI       1048576
SF       125.6924115 MHz
WDW      EM
SSB      0
LB       0.30 Hz
GB       0
PC       1.40

```



```

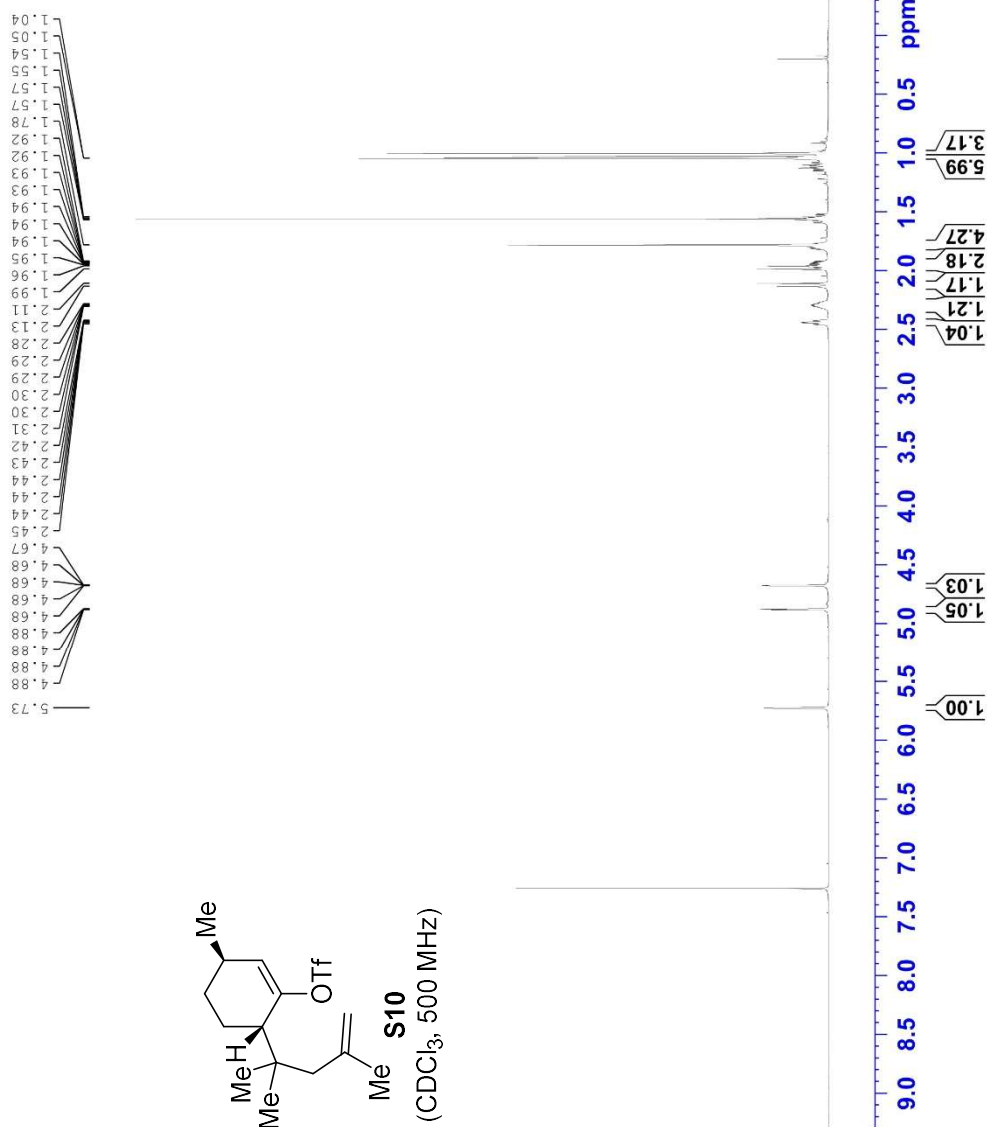
Current Data Parameters
NAME      yh-4-1-f
EXPNO    1
PROCNO   1

F2 - Acquisition Parameters
Date_    20180730
Time_    19.46
INSTRUM  spect
PROBHD   5 mm PABBO BE/
PULPROG  zg
TD        59998
SOLVENT  CDCl3
NS        16
DS        0
SWH       10000.000 Hz
FIDRES    0.166672 Hz
AQ         2.9999001 sec
RG         100.96
DW         50.000 usec
DE         6.50 usec
TE         295.8 K
D1         4.0000000 sec
TD0        1

===== CHANNEL f1 =====
SFO1     499.8730869 MHz
NUC1      1H
P1        3.58 usec
PLW1     18.25000000 W

F2 - Processing parameters
SI        65536
SF        499.8700122 MHz
WDW       EM
SSB       0
LB        0.30 Hz
GB        0
PC        1.00

```



```

Current Data Parameters
NAME      yn-4-1-r
EXENO    3
PROCNO   3

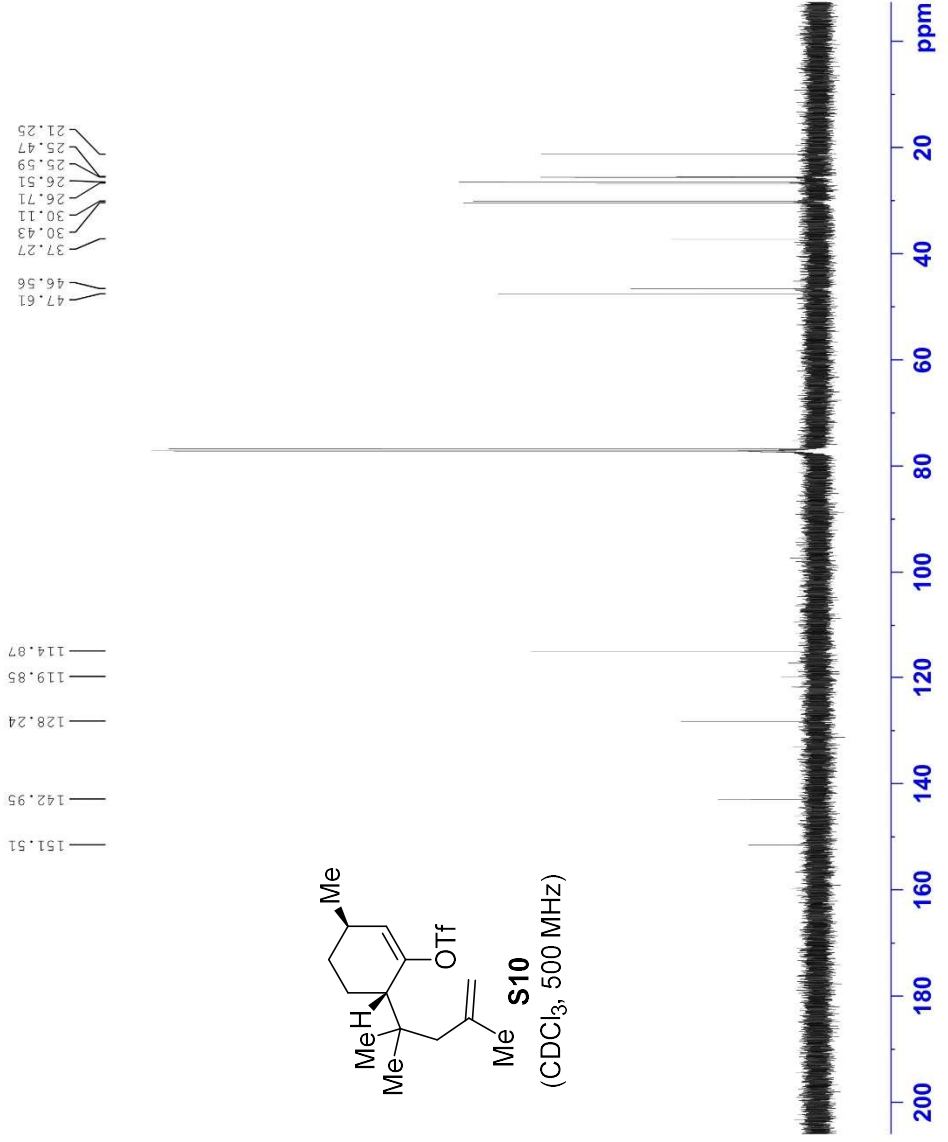
F2 - Acquisition Parameters
Date_    20180730
Time_    21.45
INSTRUM  spect
PROBHD   5 mm PABBO BE/
PULPROG  zgpgc
TD       187496
SOLVENT  CDCl3
NS       306
DS       0
SWH      31250.000 Hz
FIDRES   0.166670 Hz
AQ       2.9999361 sec
RG       2050
DW       16.000 usec
DE       6.50 usec
TE       297.7 K
D1       3.0000000 sec
D11      0.0300000 sec
TDO      1

===== CHANNEL f1 =====
SF01    125.7049802 MHz
NUC1    13C
P1      10.00 usec
PLW1    72.83999634 W

===== CHANNEL f2 =====
SF02    499.8724993 MHz
NUC2    1H
waltz16
CPDPRG[2]
PCPD2   80.00 usec
PLW2    19.0000000 W
PLW12   0.29688001 W

F2 - Processing parameters
SI      1048576
SF      125.6924115 MHz
WDW     EM
SSB     0
LB      0.30 Hz
GB      0
PC      1.40

```

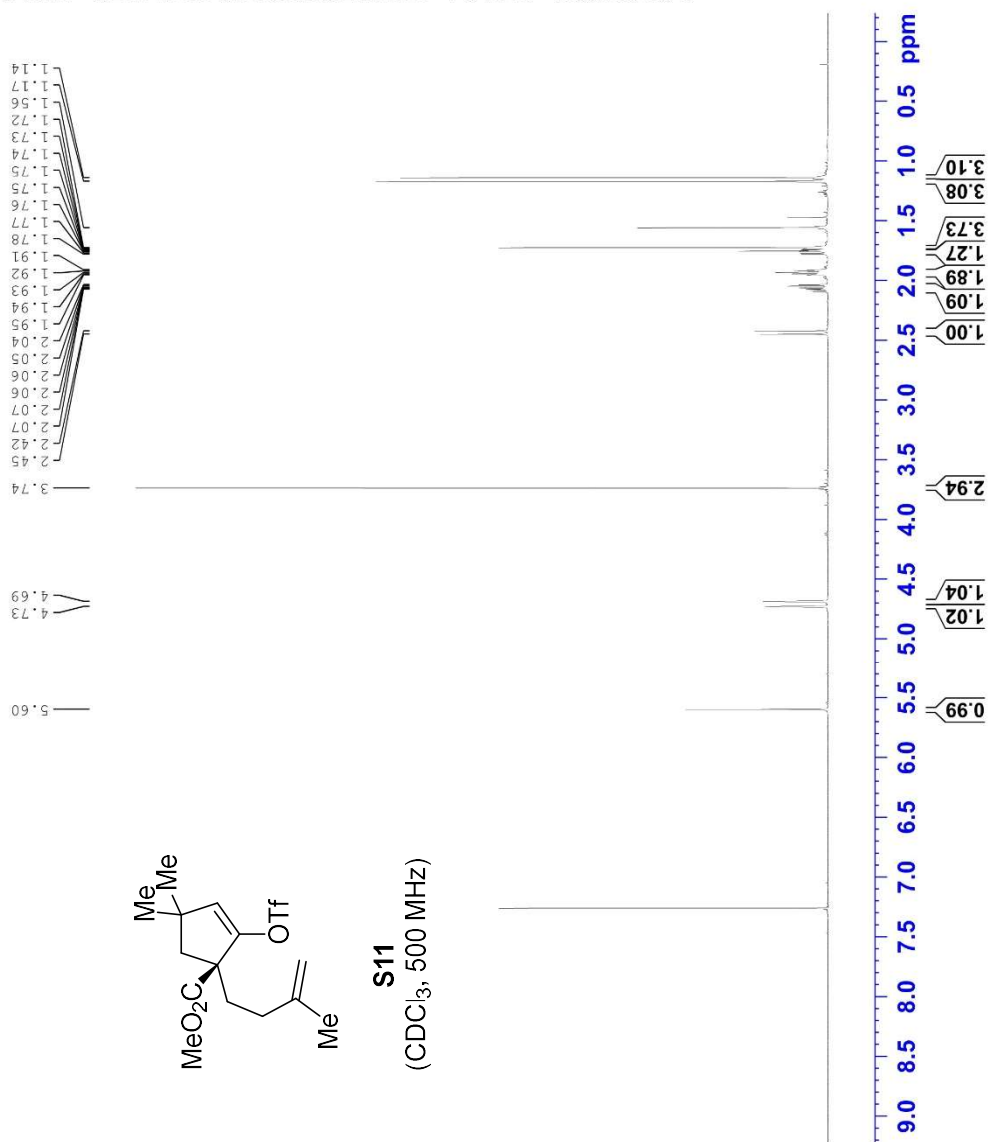


Current Data Parameters
 NAME 5-member-Ring-OTf
 EXPNO 1
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20190812
 Time_ 14.30
 INSTRUM spect
 PROBHD 5 mm PAXYI 1H/
 PULPROG zg
 TD 59998
 SOLVENT CDCl3
 NS 4
 DS 0
 SWH 10000.000 Hz
 FIDRES 0.166672 Hz
 AQ 2.9999001 sec
 RG 196.79
 DW 50.000 usec
 DE 10.00 usec
 TE 293.4 K
 D1 4.0000000 sec
 TDO 1

=====
 CHANNEL f1
 SF01 500.1330885 MHz
 NUC1 1H
 P1 3.30 usec
 PLW1 12.19999981 W

F2 - Processing parameters
 SI 65536
 SF 500.1300138 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00



```

Current Data Parameters
NAME      5-member-Ring-OTf
EXPNO    2
PROCNO   2

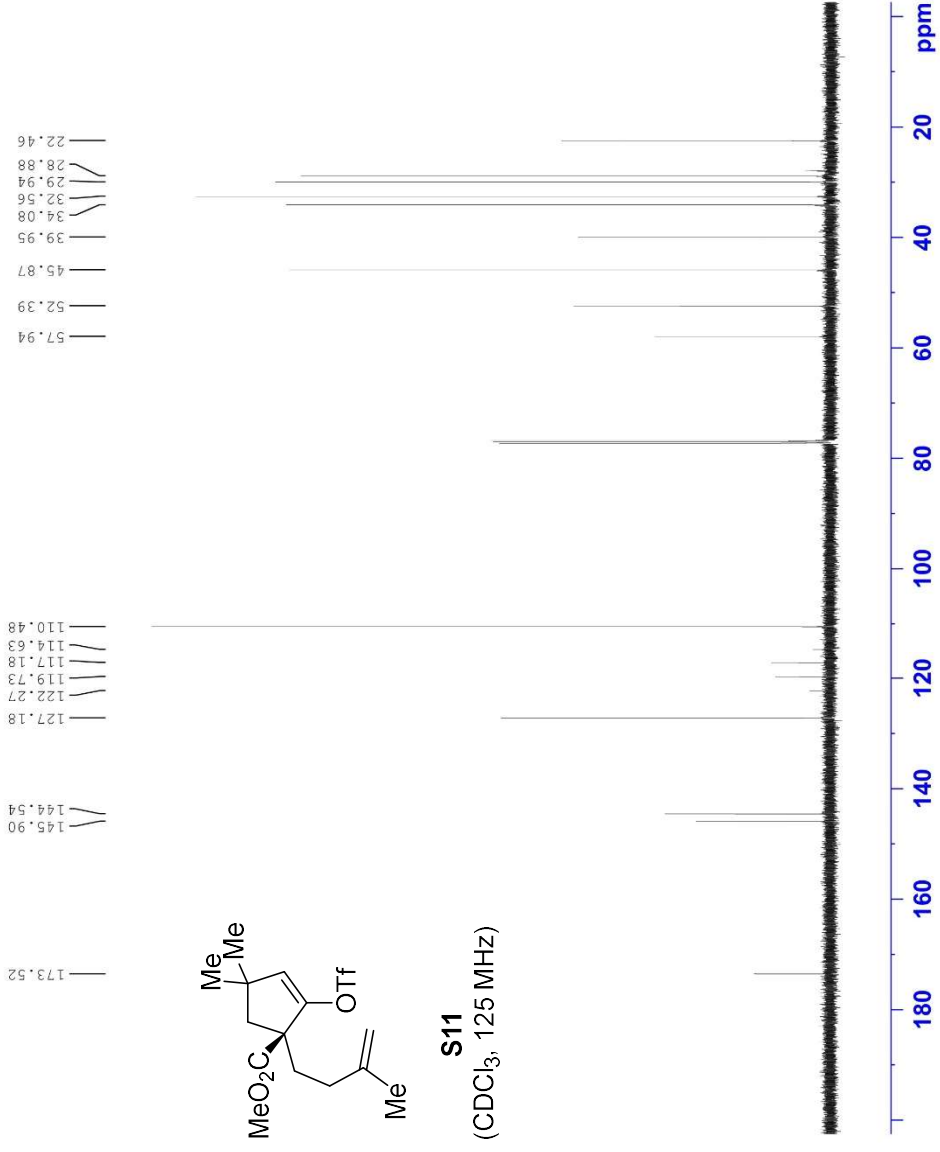
F2 - Acquisition Parameters
Date_    20190812
Time_    21.48
INSTRUM  spect
PROBHD   5 mm PABBO BB/
PULPROG  zgpgc
TD        187496
SOLVENT  CDCl3
NS        329
DS        0
SWH       31250.000 Hz
FIDRES    0.166670 Hz
AQ         2.9999361 sec
RG         2050
DW         16.000 usec
DE         6.50 usec
TE         296.3 K
D1         3.00000000 sec
D11        0.03000000 sec
TDO       1

===== CHANNEL f1 =====
SF01     125.7049802 MHz
NUC1      13C
P1        10.00 usec
PLW1     72.83999634 W

===== CHANNEL f2 =====
SF02     499.8724993 MHz
NUC2      1H
waltz16
CPDPRG2  waltz16
PCPD2    80.00 usec
PLW2     19.00000000 W
PLW12    0.29688001 W

F2 - Processing parameters
SI        1048576
SF        125.6924115 MHz
WDW       EM
SSB       0
LB        0.30 Hz
GB        0
PC        1.40

```



S11
(CDCl₃, 125 MHz)

Current Data Parameters
 NAME yh-5-36-f
 EXENO 1
 PROCNO 1

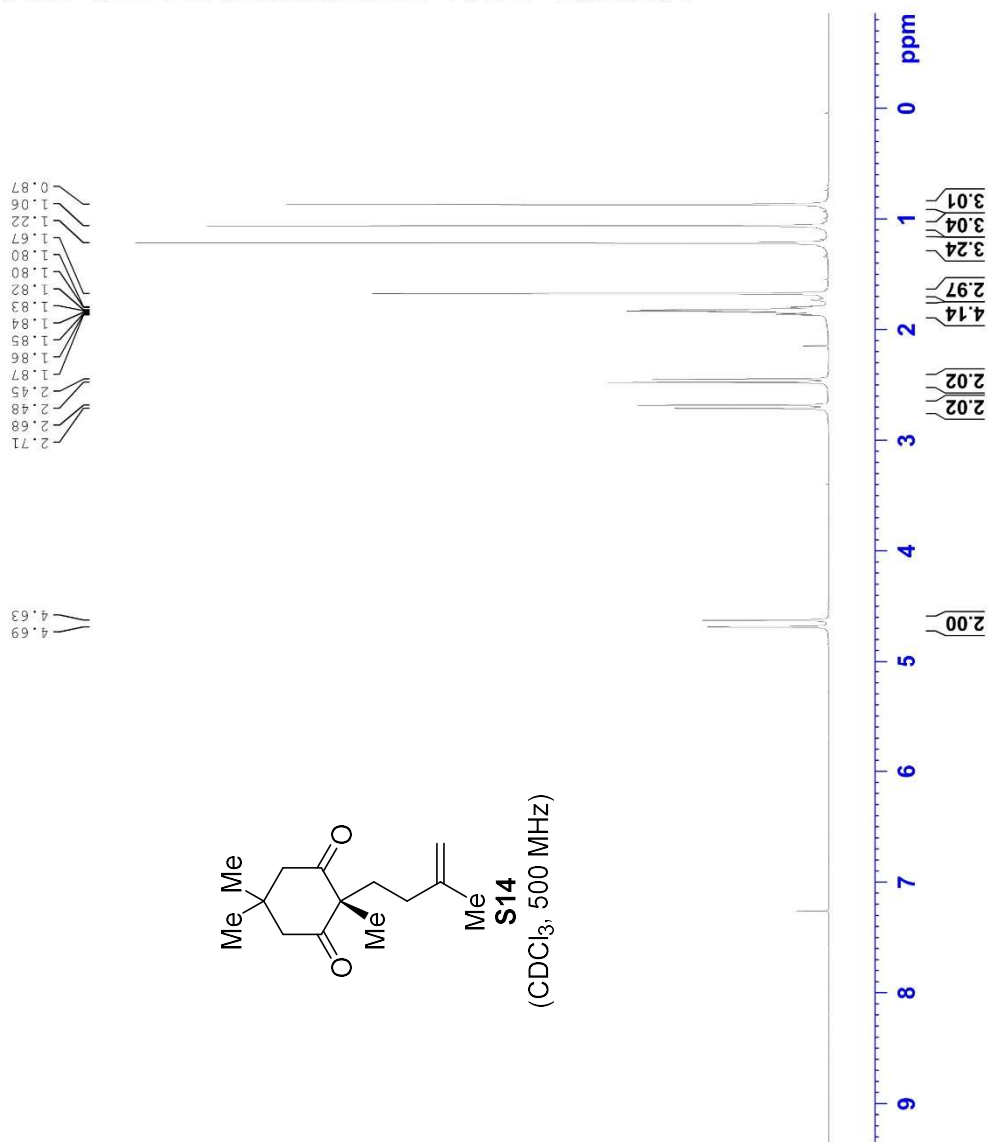
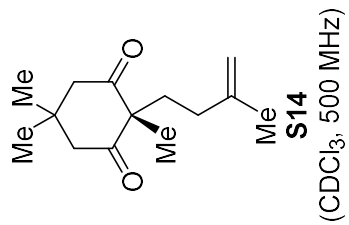
F2 - Acquisition Parameters
 Date_ 20180801
 Time_ 13.14
 INSTRUM spect
 PROBHD 5 mm PAXI 1H/
 PULPROG zg
 TD 59998
 SOLVENT CDCl3
 NS 6
 DS 0
 SWH 10000.000 Hz
 FIDRES 0.166672 Hz
 AQ 2.9999001 sec
 RG 30.11
 DW 50.000 usec
 DE 10.00 usec
 TE 294.1 K
 D1 3.00000000 sec
 TD0 1

==== CHANNEL f1 =====
 SF01 500.1330885 MHz
 NUC1 1H
 P1 2.67 usec
 PLW1 12.19999981 W

F2 - Processing parameters
 SI 65536
 SF 500.1300136 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00

4.69
4.63

2.71
2.68
2.64
2.48
2.45
1.97
1.86
1.85
1.84
1.83
1.82
1.80
1.80
1.67
1.22
1.06
0.87



```

Current Data Parameters
NAME      yh-5-36-r
EXPNO    2
PROCNO   2

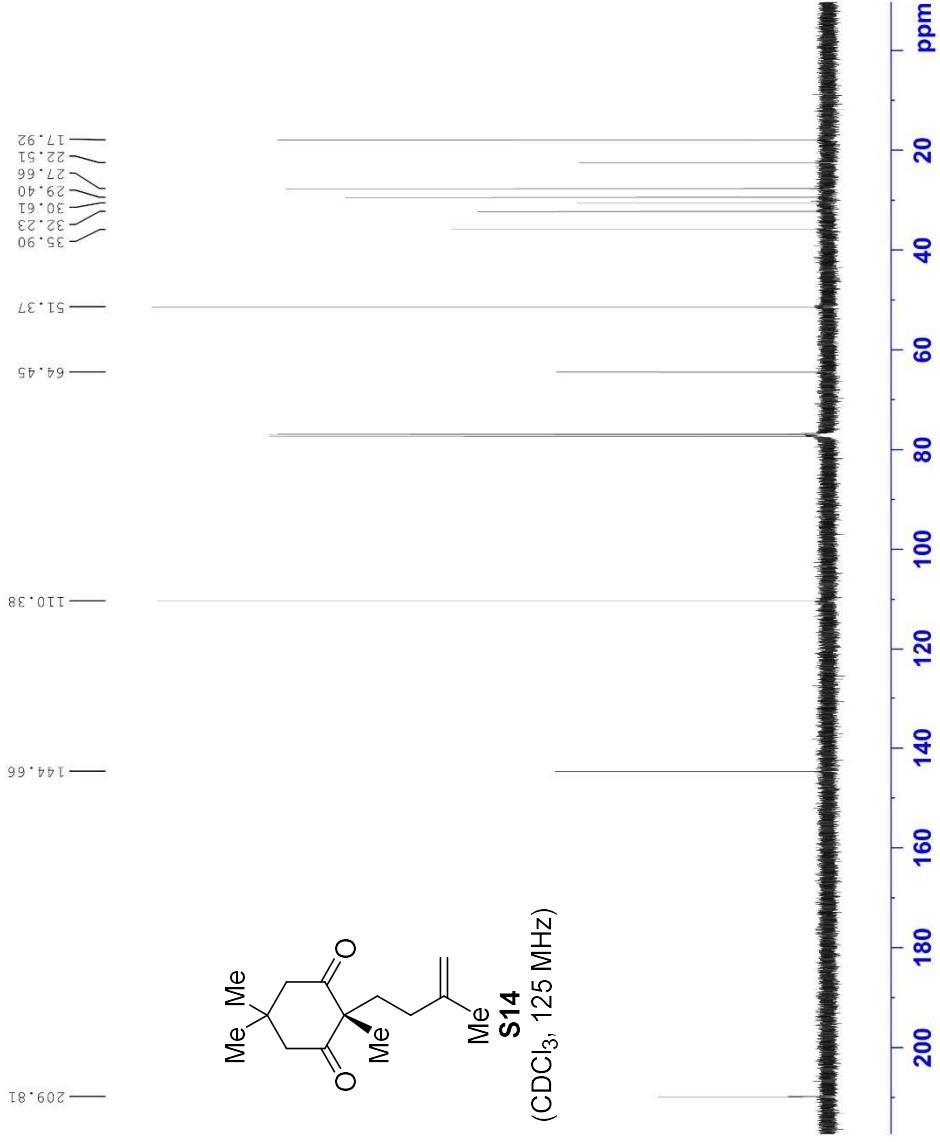
F2 - Acquisition Parameters
Date_     20180801
Time_    13.41
INSTRUM  spect
PROBHD   5 mm PABBO BB/
PULPROG  zgpgc
TD       187496
SOLVENT  CDCl3
NS       97
DS       0
SWH      31250.000 Hz
FIDRES   0.166670 Hz
AQ       2.9999361 sec
RG       2050
RW       16.000 usec
DE       6.50 usec
TE       296.6 K
D1       4.0000000 sec
D11      0.0300000 sec
TDO      1

===== CHANNEL f1 =====
SF01    125.7049802 MHz
NUC1    13C
P1      3.58 usec
PLW1    72.83999634 W

===== CHANNEL f2 =====
SF02    499.8724993 MHz
NUC2    1H
waltz16
PCPD2   80.00 usec
PLW2    19.0000000 W
PLW12   0.29688001 W

F2 - Processing parameters
SI      1048576
SF      125.6924115 MHz
WDW     EM
SSB     0
LB      0.30 Hz
GB      0
PC      1.40

```

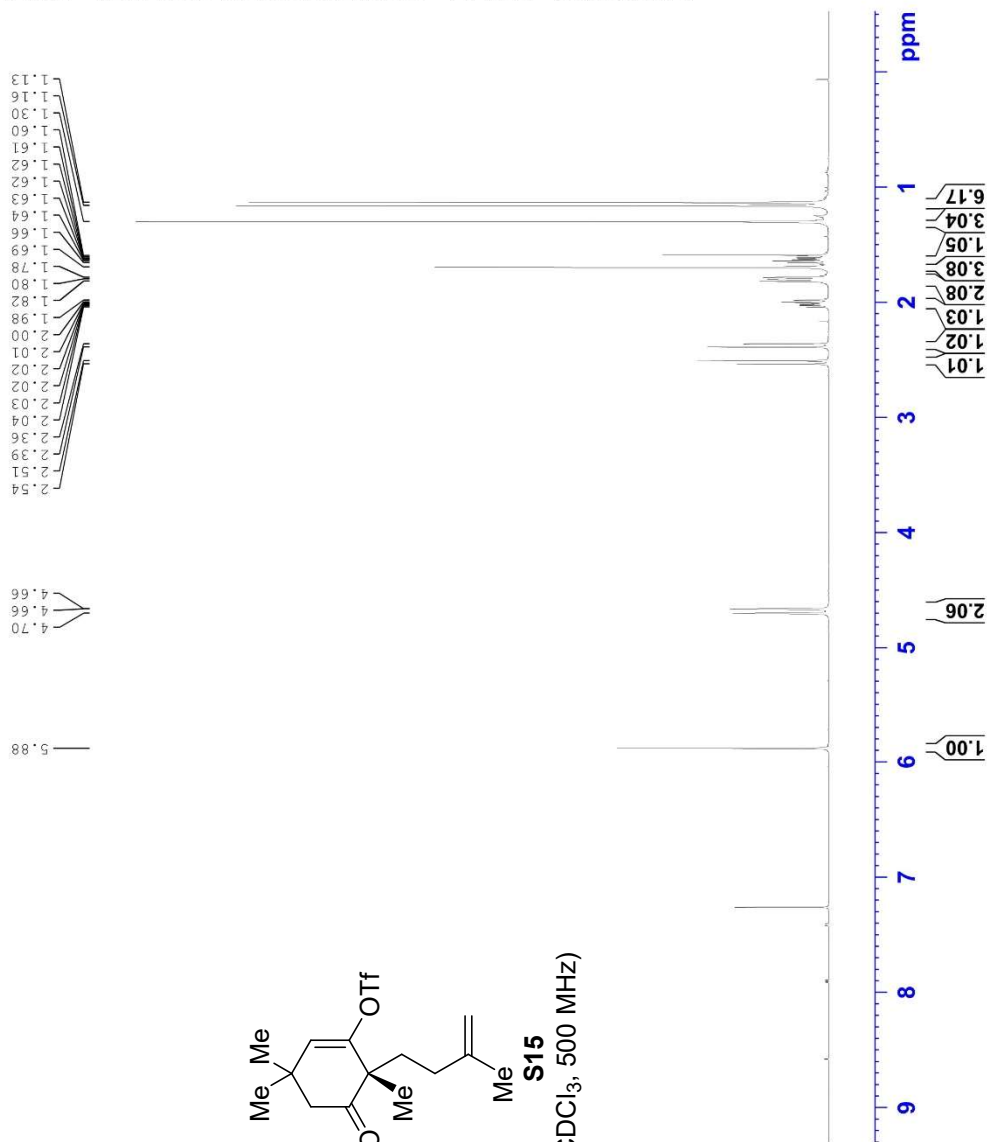


Current Data Parameters
 NAME yh-5-38-1
 EXPNO 1
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20180801
 Time_ 13.54
 INSTRUM spect
 PROBHD 5 mm PAXI 1H/
 PULPROG zg
 TD 59998
 SOLVENT CDCl3
 NS 8
 DS 0
 SWH 10000.000 Hz
 FIDRES 0.166672 Hz
 AQ 2.9999001 sec
 RG 97.37
 DW 50.000 usec
 DE 10.00 usec
 TE 294.2 K
 D1 3.00000000 sec
 TDO 1

=====
 CHANNEL f1
 SF01 500.1330885 MHz
 NUC1 1H
 P1 2.67 usec
 PLW1 12.19999981 W

F2 - Processing parameters
 SI 65536
 SF 500.1300138 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00



```

Current Data Parameters
NAME      yh-5-38-1
EXENO    2
PROCNO   2

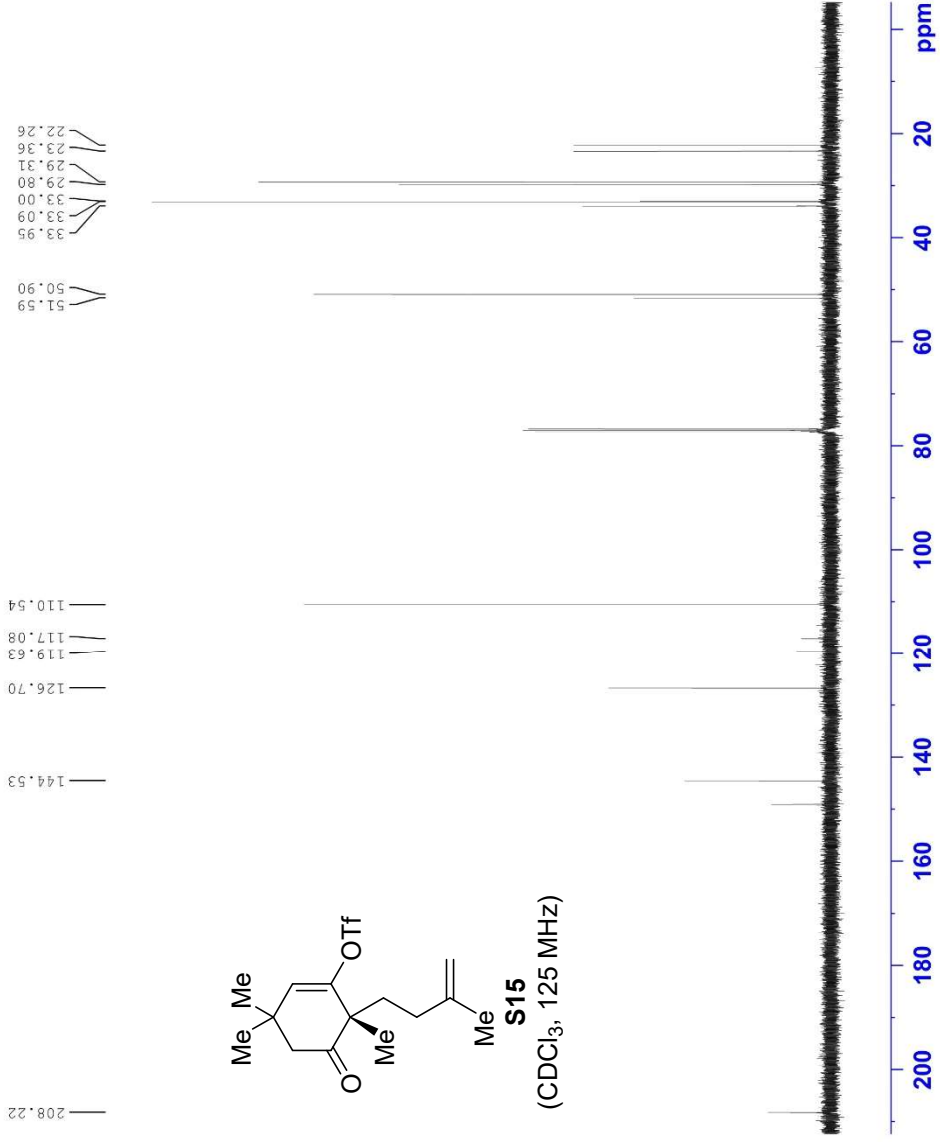
F2 - Acquisition Parameters
Date_    20180801
Time_    14.37
INSTRUM  spect
PROBHD   5 mm PABBO BB/
PULPROG  zgpgc
TD       187496
SOLVENT  CDCl3
NS       158
DS       0
SWH      31250.000 Hz
FIDRES   0.166670 Hz
AQ       2.9999361 sec
RG       2050
DW       16.000 usec
DE       6.50 usec
TE       297.1 K
D1       3.0000000 sec
D11      0.0300000 sec
TDO      1

===== CHANNEL f1 =====
SF01     125.7049802 MHz
NUC1     13C
P1       10.00 usec
PLW1     72.83999634 W

===== CHANNEL f2 =====
SF02     499.8724993 MHz
NUC2     1H
waltz16
CPDPRG[2]
PCPD2    80.00 usec
PLW2     19.0000000 W
PLW12    0.29688001 W

F2 - Processing parameters
SI       1048576
SF       125.6924115 MHz
WDW      EM
SSB      0
LB       0.30 Hz
GB       0
PC       1.40

```

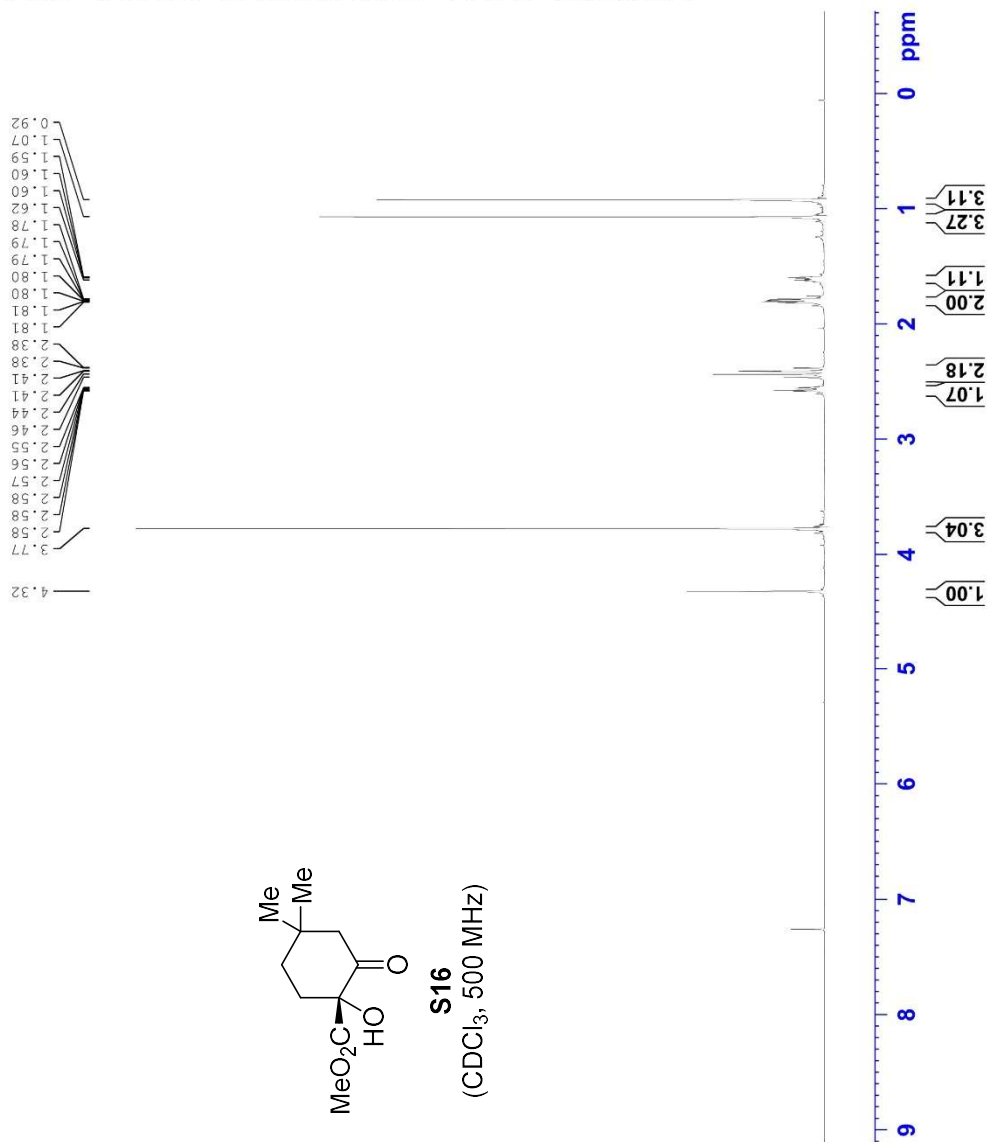


Current Data Parameters
 NAME yh-5-52-a
 EXPNO 1
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20190711
 Time_ 23:52
 INSTRUM spect
 PROBHD 5 mm PABBO BE/
 PULPROG zg
 TD 59998
 SOLVENT CDCl3
 NS 1
 DS 0
 SWH 10000.000 Hz
 FIDRES 0.166672 Hz
 AQ 2.9999001 sec
 RG 56.75
 DW 50.000 usec
 DE 6.50 usec
 TE 296.1 K
 D1 4.0000000 sec
 TDO 1

=====
 CHANNEL f1
 SF01 499.8730869 MHz
 NUC1 1H
 P1 3.58 usec
 PLW1 18.25000000 W

F2 - Processing parameters
 SI 65536
 SF 499.8700118 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00



```

Current Data Parameters
NAME      yh-5-52-a
EXENO    2
PROCNO   2

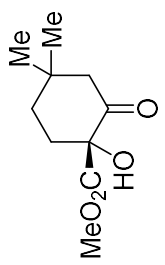
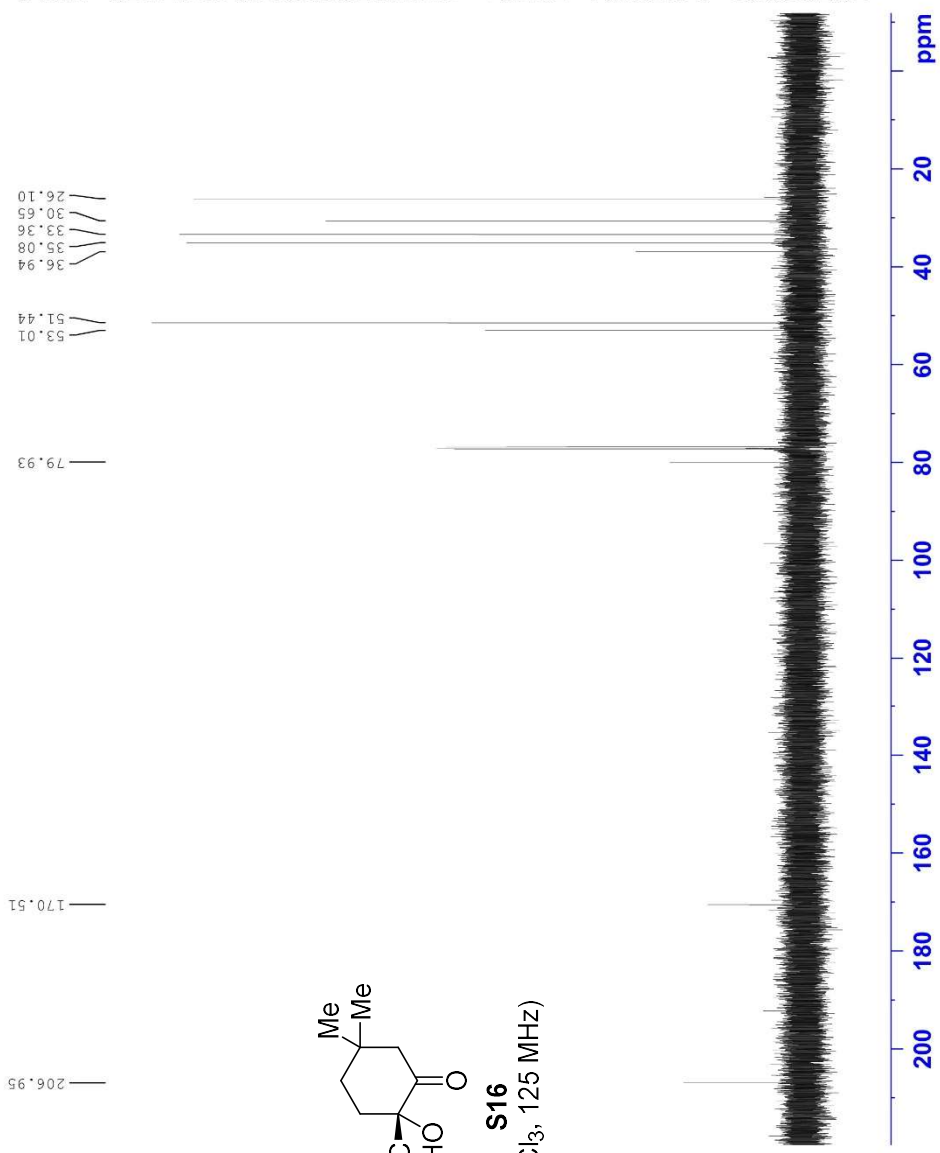
F2 - Acquisition Parameters
Date_    20190711
Time_    23:58
INSTRUM  spect
PROBHD   5 mm PABBO BB/
PULPROG  zgpgc
TD       187496
SOLVENT  CDCl3
NS       7
DS       0
SWH      31250.000 Hz
FIDRES   0.166670 Hz
AQ       2.9999361 sec
RG       2050
DW       16.000 usec
DE       6.50 usec
TE       296.3 K
D1       3.0000000 sec
D11      0.0300000 sec
TDO      1

===== CHANNEL f1 =====
SF01    125.7049802 MHz
NUC1     13C
P1       10.00 usec
PLW1    72.83999634 W

===== CHANNEL f2 =====
SF02    499.8724993 MHz
NUC2     1H
waltz16
CPDPRG2  waltz16
PCPD2    80.00 usec
PLW2     19.0000000 W
PLW12    0.29688001 W

F2 - Processing parameters
SI       1048576
SF       125.6924115 MHz
WDW      EM
SSB      0
LB       0.30 Hz
GB       0
PC       1.40

```



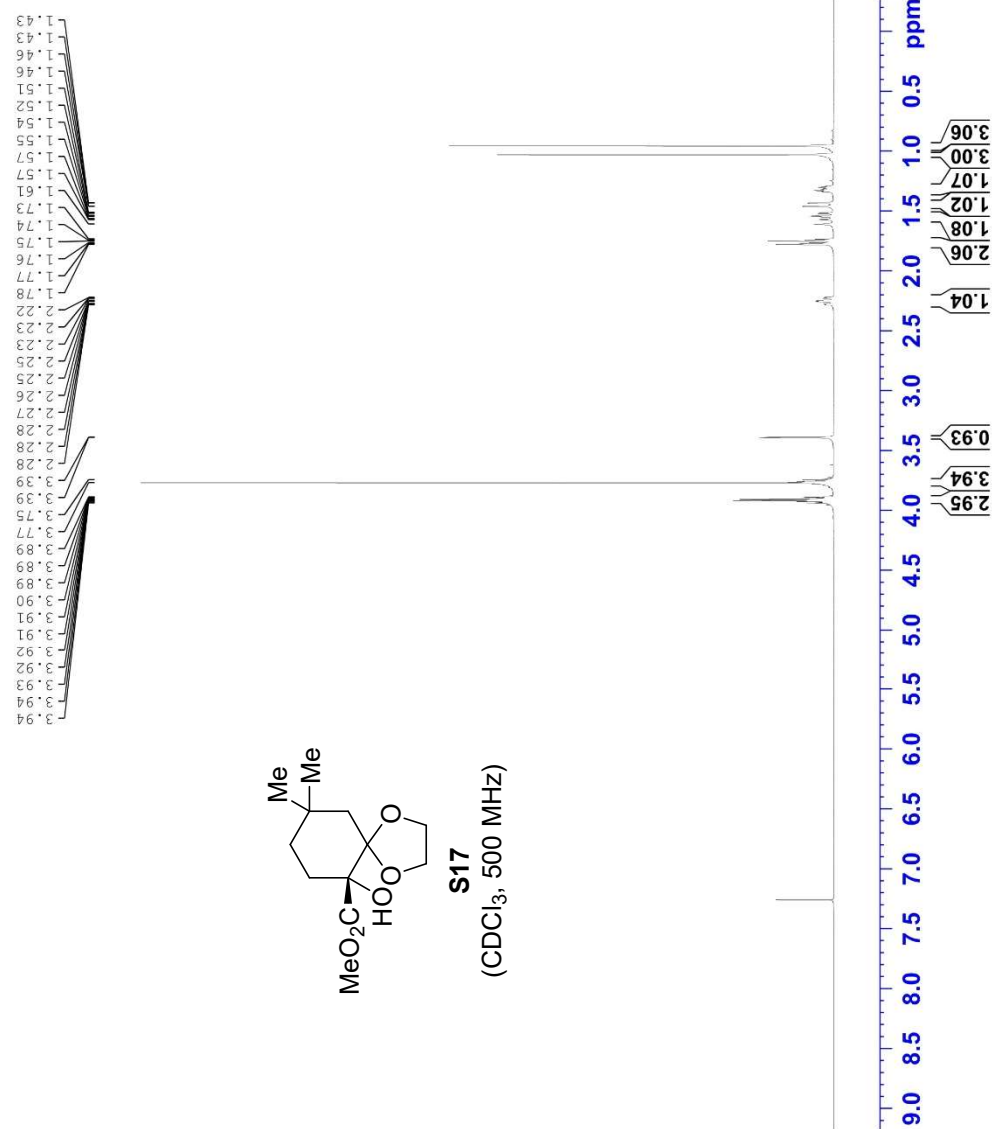
S16
(CDCl₃, 125 MHz)

Current Data Parameters
 NAME yh-5-77-a
 EXENO 1
 PROCNO 1

F2 - Acquisition Parameters
 Date 20190711
 Time 14.56
 INSTRUM spect
 PROBHD 5 mm PABBO BE/
 PULPROG zg
 TD 59998
 SOLVENT CDCl3
 NS 2
 DS 0
 SWH 10000.000 Hz
 FIDRES 0.166672 Hz
 AQ 2.9999001 sec
 RG 70.49
 DW 50.000 usec
 DE 6.50 usec
 TE 295.4 K
 D1 4.0000000 sec
 TD0 1

==== CHANNEL f1 =====
 SF01 499.8730869 MHz
 NUC1 1H
 P1 3.58 usec
 PLW1 18.25000000 W

F2 - Processing parameters
 SI 65536
 SF 499.8700119 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00



```

Current Data Parameters
NAME      yh-5-77-a
EXPNO    2
PROCNO   2

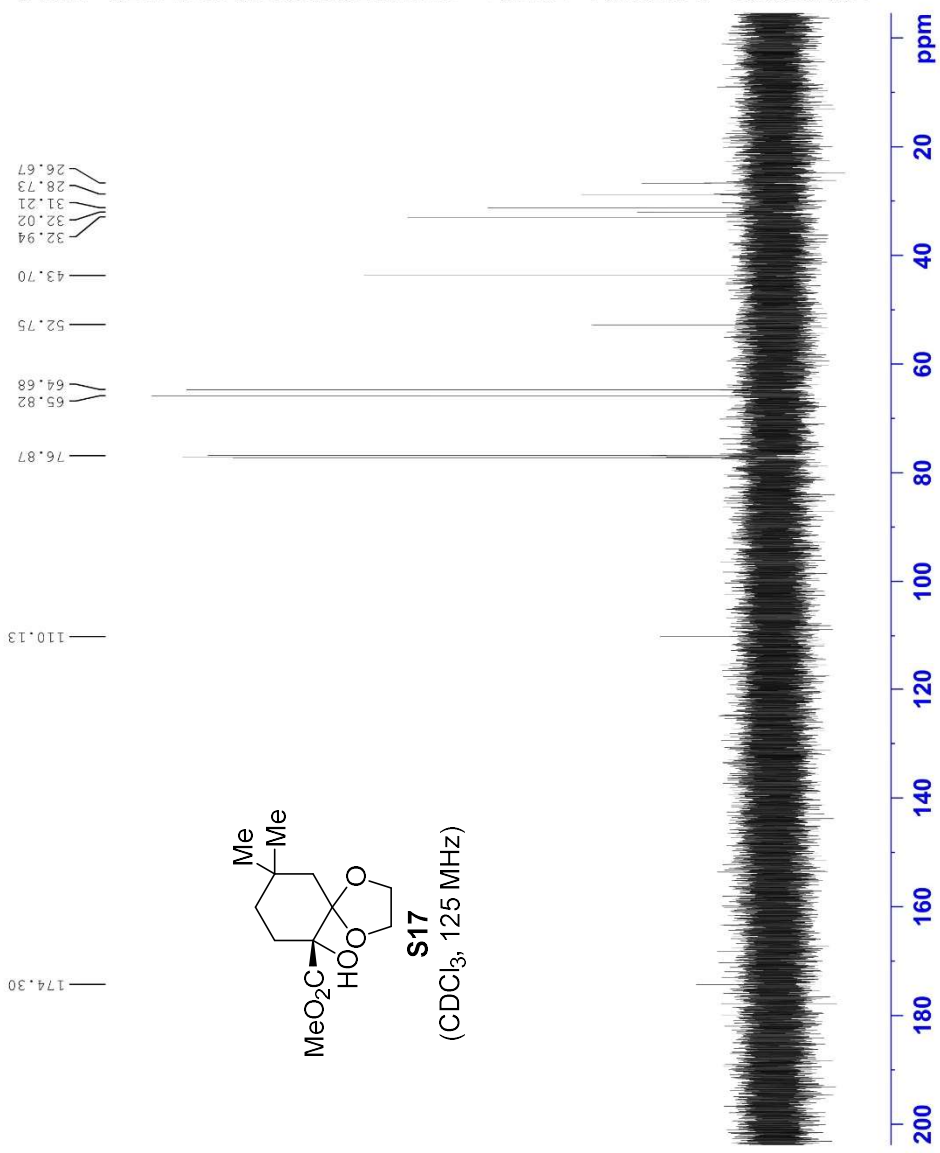
F2 - Acquisition Parameters
Date_    20190711
Time     15.02
INSTRUM  spect
PROBHD   5 mm PABBO BB/
PULPROG  zgpgc
TD       187496
SOLVENT  CDCl3
NS       21
DS       0
SWH      31250.000 Hz
FIDRES   0.166670 Hz
AQ       2.9999361 sec
RG       2050
DW       16.000 usec
DE       6.50 usec
TE       295.8 K
D1       3.0000000 sec
D11      0.0300000 sec
TDO      1

===== CHANNEL f1 =====
SF01    125.7049802 MHz
NUC1    13C
P1      10.00 usec
PLW1    72.83999634 W

===== CHANNEL f2 =====
SF02    499.8724993 MHz
NUC2    1H
waltz16
CPDPRG2
PCPD2   80.00 usec
PLW2    19.0000000 W
PLW12   0.29688001 W

F2 - Processing parameters
SI      1048576
SF      125.6924115 MHz
WDW     EM
SSB     0
LB      0.30 Hz
GB      0
PC      1.40

```

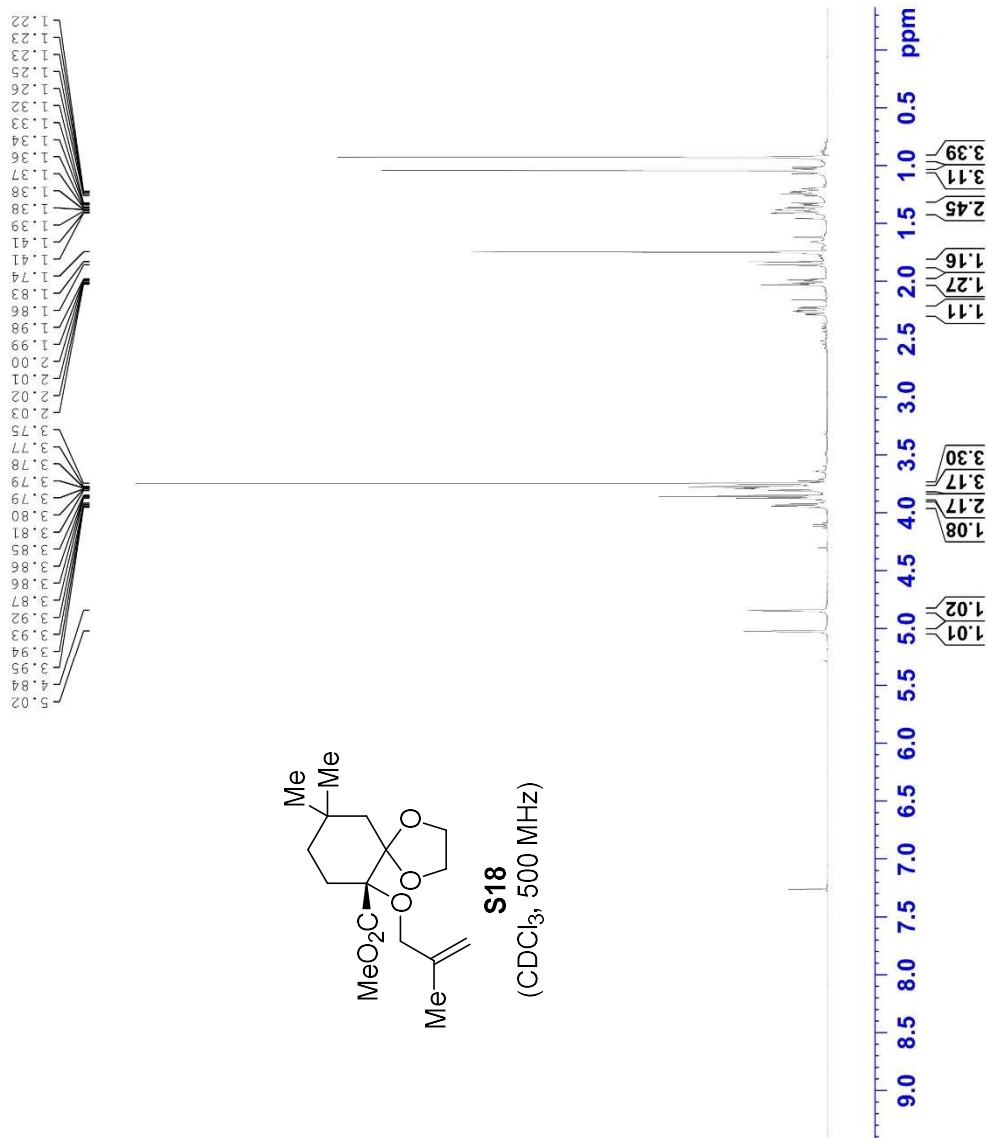


Current Data Parameters
 NAME yh-5-78-c
 EXPNO 1
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20190712
 Time_ 17.20
 INSTRUM spect
 PROBHD 5 mm PABBO BE/
 PULPROG zg
 TD 59998
 SOLVENT CDCl3
 NS 1
 DS 0
 SWH 10000.000 Hz
 FIDRES 0.166672 Hz
 AQ 2.9999001 sec
 RG 44.57
 DW 50.000 usec
 DE 6.50 usec
 TE 297.0 K
 D1 4.0000000 sec
 TDO 1

=====
 CHANNEL f1
 SF01 499.8730869 MHz
 NUC1 1H
 P1 3.58 usec
 PLW1 18.25000000 W

F2 - Processing parameters
 SI 65536
 SF 499.8700120 MHz
 EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00



```

Current Data Parameters
NAME      yh-5-78-c
EXPNO    2
PROCNO   2

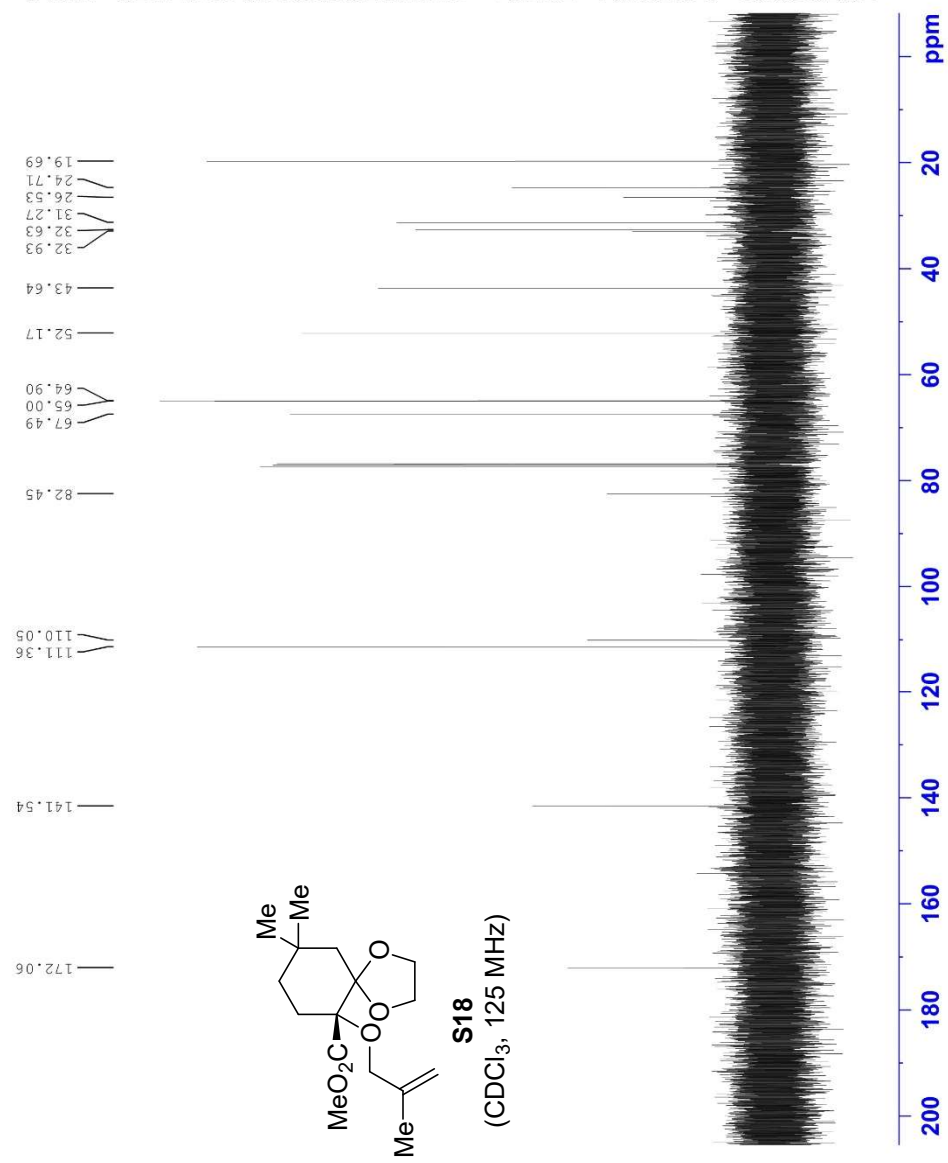
F2 - Acquisition Parameters
Date_    20190712
Time     17.23
INSTRUM  spect
PROBHD   5 mm PABBO BB/
PULPROG  zgpgc
TD        187496
SOLVENT  CDCl3
NS        3
DS        0
SWH       31250.000 Hz
FIDRES    0.166670 Hz
AQ         2.9999361 sec
RG         2050
DW         16.000 usec
DE         6.50 usec
TE        297.1 K
D1         3.00000000 sec
D11        0.03000000 sec
TDO       1

===== CHANNEL f1 =====
SFO1     125.7049802 MHz
NUC1      13C
P1        10.00 usec
PLW1     72.83999634 W

===== CHANNEL f2 =====
SFO2     499.8724993 MHz
NUC2      1H
waltz16
CPDPRG2  waltz16
PCPD2    80.00 usec
PLW2     19.00000000 W
PLW12    0.29688001 W

F2 - Processing parameters
SI        1048576
SF        125.6924115 MHz
WDW       EM
SSB       0
LB        0.30 Hz
GB        0
PC        1.40

```

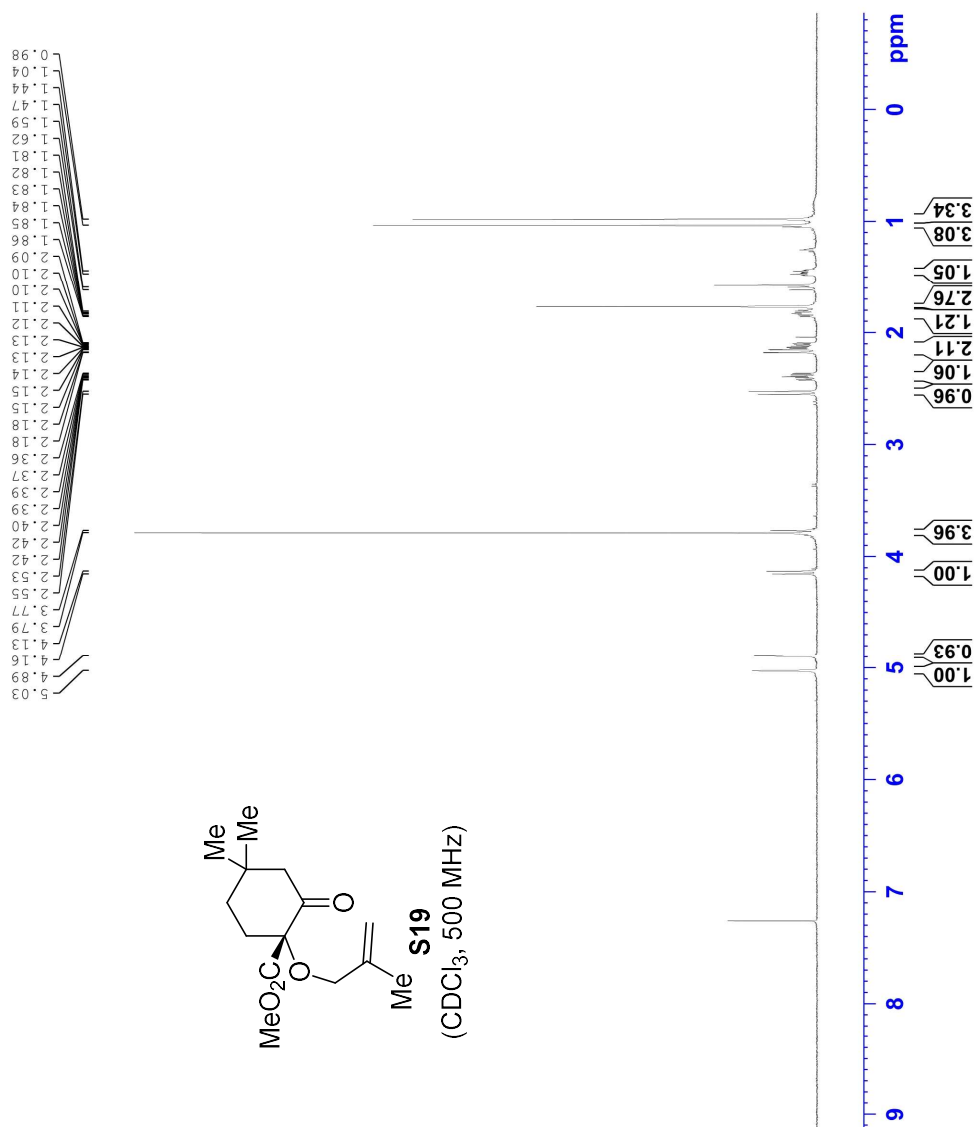


Current Data Parameters
 NAME Yh-5-86-b
 EXPNO 1
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20190712
 Time_ 17.30
 INSTRUM spect
 PROBHD 5 mm PABBO BB/
 PULPROG zg
 TD 59998
 SOLVENT CDCl3
 NS 2
 DS 0
 SWH 10000.000 Hz
 FIDRES 0.166672 Hz
 AQ 2.9999001 sec
 RG 0.98
 DW 50.000 usec
 DE 6.50 usec
 TE 297.1 K
 DL 4.0000000 sec
 TDO 1

===== CHANNEL f1 =====
 SFO1 499.8730869 MHz
 NUC1 1H
 P1 3.58 usec
 PLW1 18.25000000 W

F2 - Processing parameters
 SI 65536
 SF 499.8700121 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00



```

Current Data Parameters
NAME      Yh-5-86-b
EXPNO     2
PROCNO    2

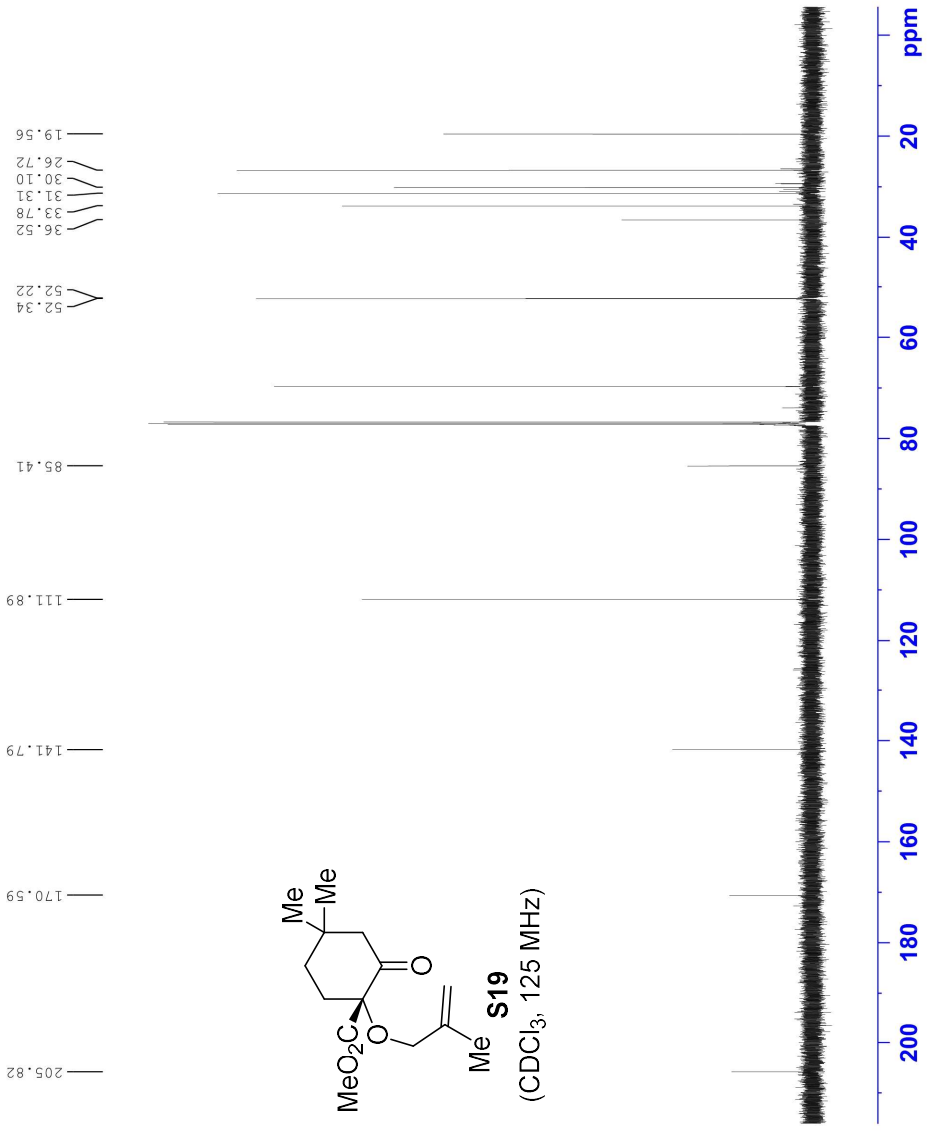
F2 - Acquisition Parameters
Date_     20190712
Time      22.36
INSTRUM   spect
PROBHD    5 mm PABBO BB/
PULPROG   zgpg
TD         187496
SOLVENT   CDCl3
NS         367
DS         0
SWH        31250.000 Hz
FIDRES     0.166670 Hz
AQ         2.9999361 sec
RG         2050
DW         16.000 usec
DE         6.50 usec
TE         299.3 K
D1         3.0000000 sec
d11        0.03000000 sec
TD0        1

===== CHANNEL f1 =====
SFO1      125.7049802 MHz
NUC1       13C
P1         10.00 usec
PLW1      72.83999634 W

===== CHANNEL f2 =====
SFO2      499.8724993 MHz
NUC2       1H
PCPRG[2]  waltz16
PCPD2     80.00 usec
PLW2      19.0000000 W
PLWL2     0.29688001 W

F2 - Processing parameters
SI         1048576
SF         125.6924115 MHz
WDW        EM
SSB        0
LB         0.30 Hz
GB         0
PC         1.40

```



```

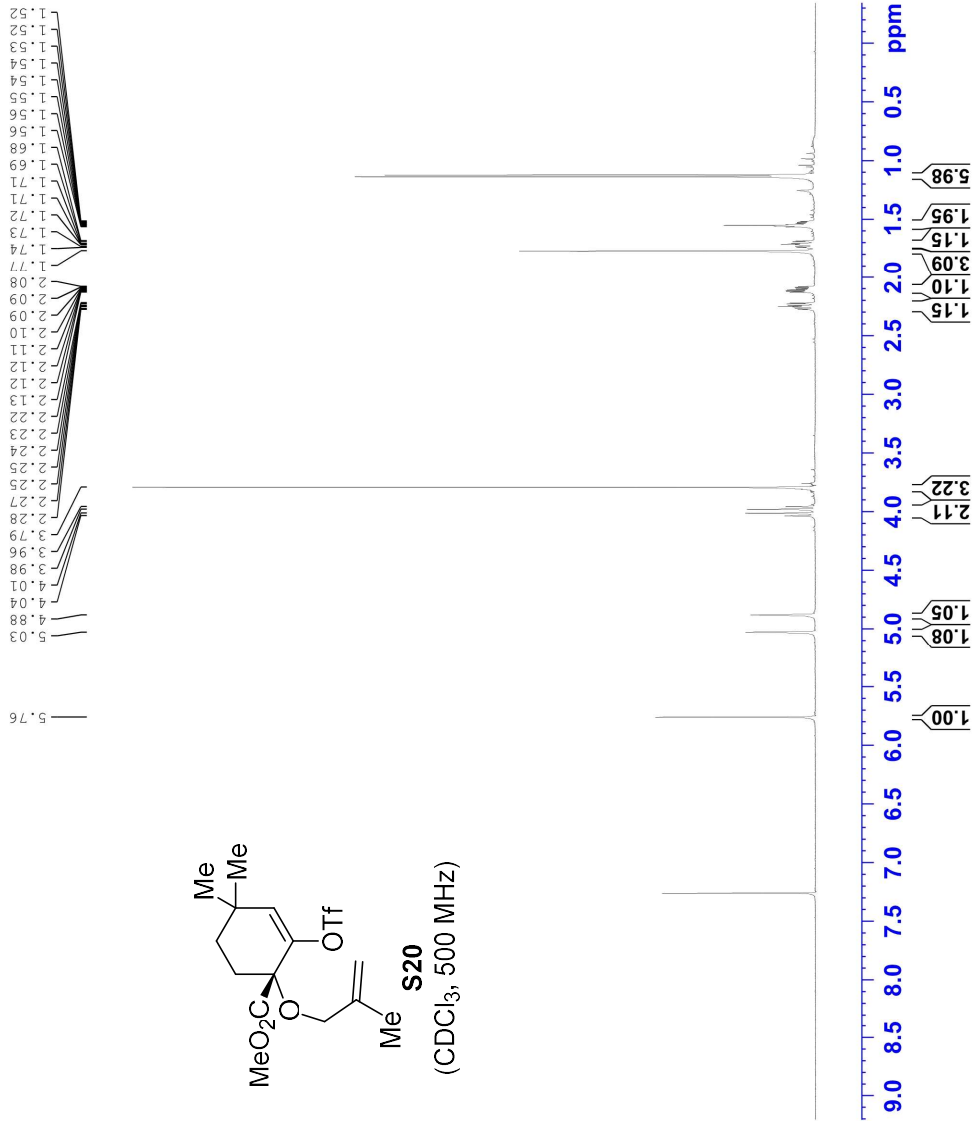
Current Data Parameters
NAME      Yh-5-87-c
EXPNO     1
PROCNO    1

F2 - Acquisition Parameters
Date_     20190712
Time      21.21
INSTRUM   spect
PROBHD    5 mm PABBO BB/
PULPROG   zg
TD         59998
SOLVENT   CDCl3
NS         2
DS         0
SWH        10000.000 Hz
FIDRES     0.166672 Hz
AQ         2.9999001 sec
RG         100.96
DW         50.000 usec
DE         6.50 usec
TE         297.5 K
DL         4.0000000 sec
TD0        1

===== CHANNEL f1 =====
SFO1      499.8730869 MHz
NUC1       1H
P1         3.58 usec
PLW1      18.25000000 W

F2 - Processing parameters
SI         65536
SF         499.8700122 MHz
WDW        EM
SSB        0
LB         0.30 Hz
GB         0
PC         1.00

```



```

Current Data Parameters
NAME      Yh-5-87-C
EXPNO     3
PROCNO    3

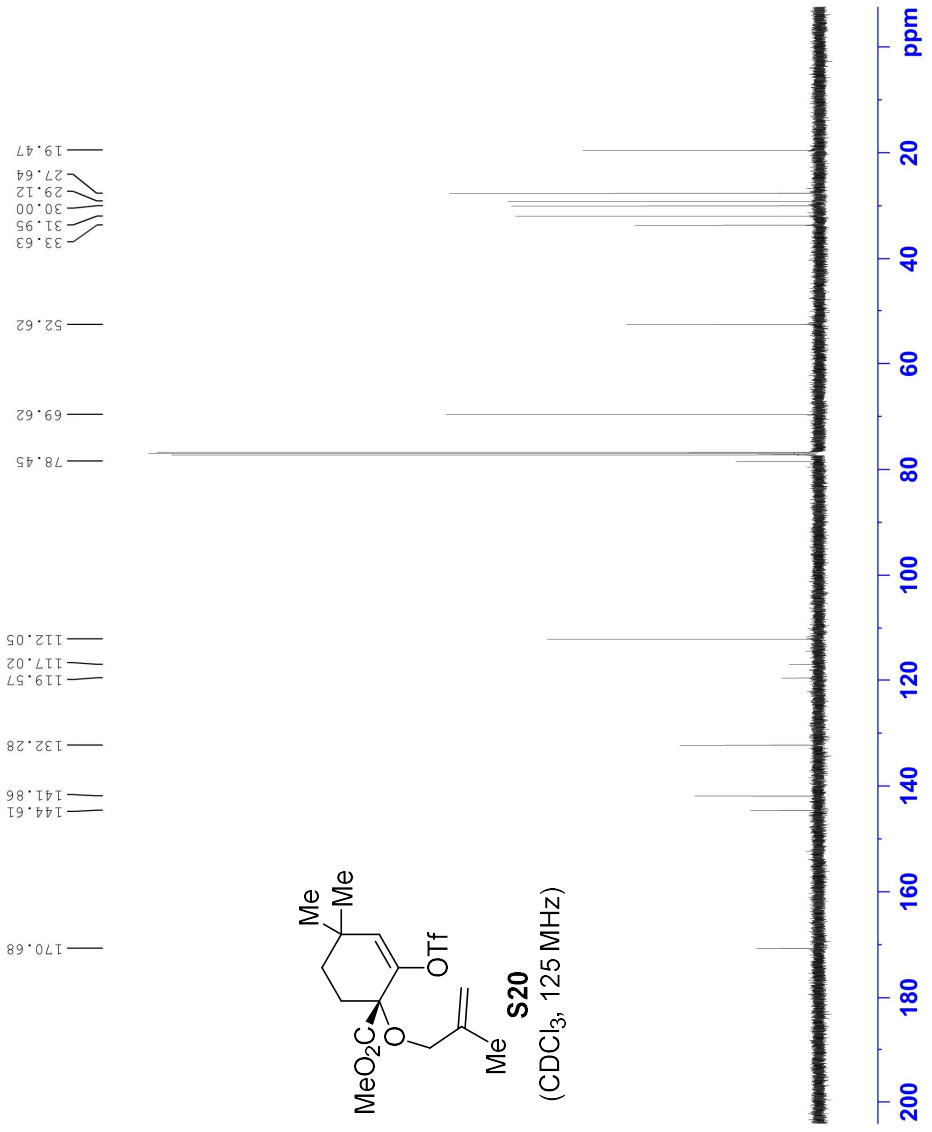
F2 - Acquisition Parameters
Date_     20190713
Time      0.02
INSTRUM   spect
PROBHD    5 mm PABBO BB/
PULPROG   zgpg
TD         187496
SOLVENT   CDCl3
NS         803
DS         0
SWH        31250.000 Hz
FIDRES     0.166670 Hz
AQ         2.9999361 sec
RG         2050
DE         16.000 usec
TE         299.5 K
DL         3.0000000 sec
D11        0.03000000 sec
TD0        1

===== CHANNEL f1 =====
SFO1      125.7049802 MHz
NUC1       13C
P1         10.00 usec
PLW1      72.83999634 W

===== CHANNEL f2 =====
SFO2      499.8724993 MHz
NUC2       1H
PCPD2     waltz16
PLW2      19.0000000 W
PLW12     0.29688001 W

F2 - Processing parameters
SI         1048576
SF         125.6924115 MHz
WDW        EM
SSB        0
LB         0.30 Hz
GB         0
PC         1.40

```

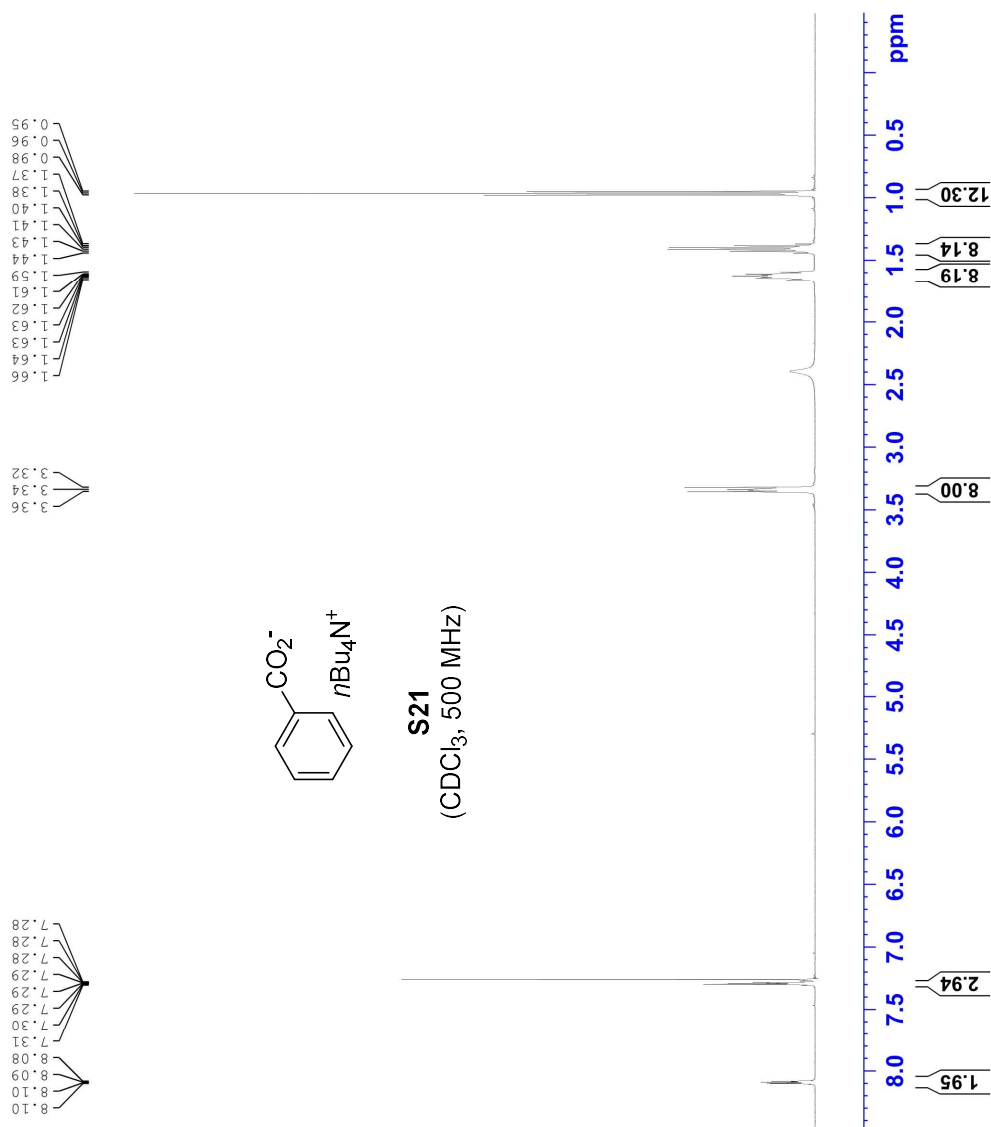


Current Data Parameters
 NAME Yh-7-32-a
 EXPNO 1
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20191010
 Time 13.42
 INSTRUM spect
 PROBHD 5 mm PATXI 1H/
 PULPROG zg
 TD 5998
 SOLVENT CDC13
 NS 4
 DS 0
 SWH 10000.000 Hz
 FIDRES 0.166672 Hz
 AQ 2.9999001 sec
 RG 196.79
 DW 50.000 usec
 DE 10.00 usec
 TE 297.0 K
 D1 4.0000000 sec
 TD0 1

===== CHANNEL f1 =====
 SFO1 500.1330885 MHz
 NUC1 1H
 P1 3.30 usec
 PLW1 12.19999981 W

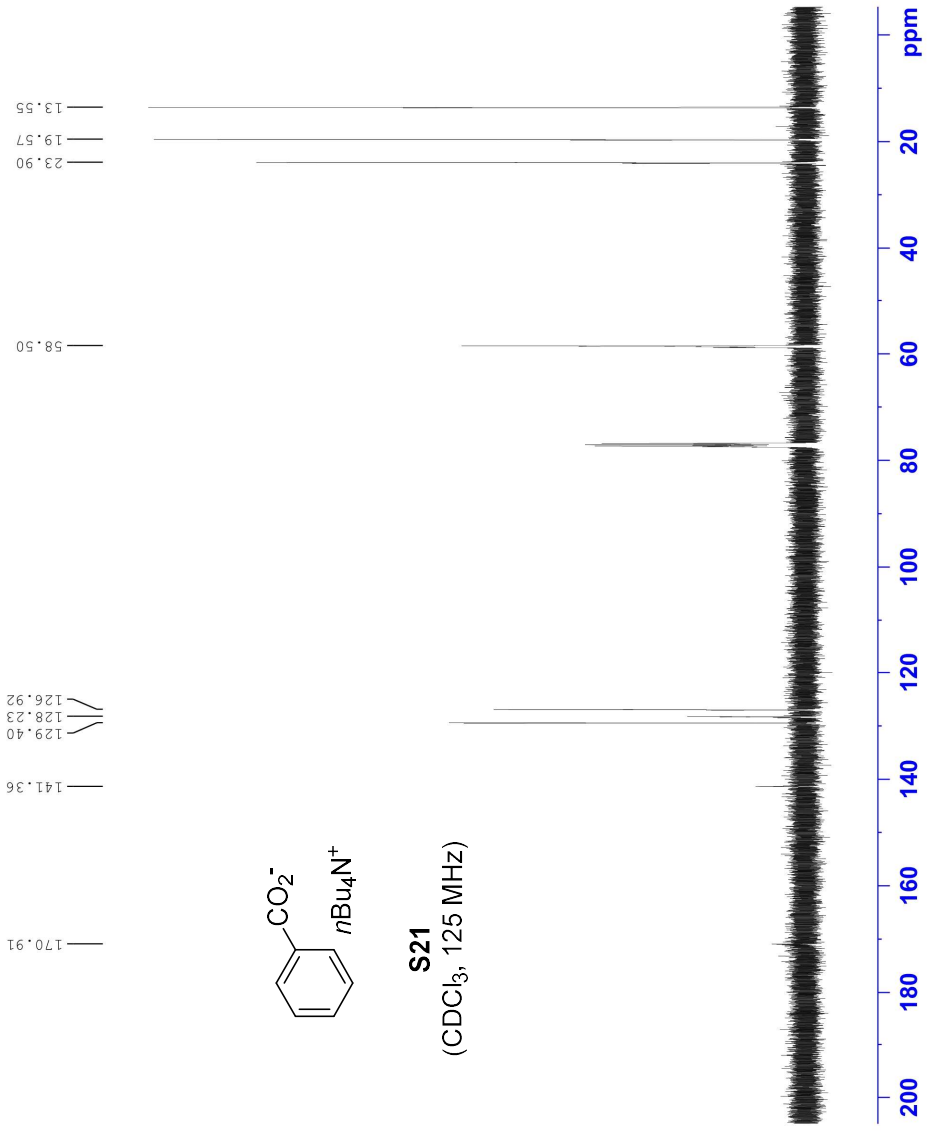
F2 - Processing parameters
 SI 65536
 SF 500.1300135 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00



Current Data Parameters
 NAME yh-7-32
 EXPNO 2
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20191005
 Time_ 15.02 h
 INSTRUM spect
 PROBHD z113652_0187 (
 PULPROG zgdc
 TD 187496
 SOLVENT CDCl3
 NS 153
 DS 0
 SWH 31250.000 Hz
 FIDRES 0.333340 Hz
 AQ 2.9999361 sec
 RG 2050
 DW 16.000 usec
 DE 6.50 usec
 TE 296.7 K
 D1 3.0000000 sec
 D11 0.03000000 sec
 TDO 1
 SFO1 125.7049802 MHz
 NUC1 13C
 P1 10.00 usec
 PLW1 72.83999634 W
 SFO2 499.8724993 MHz
 NUC2 1H
 CPDPRG2 waltz16
 PCPD2 80.00 usec
 PLW2 19.00000000 W
 PLW12 0.29688001 W

F2 - Processing parameters
 SI 1048576
 SF 125.6924115 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.40

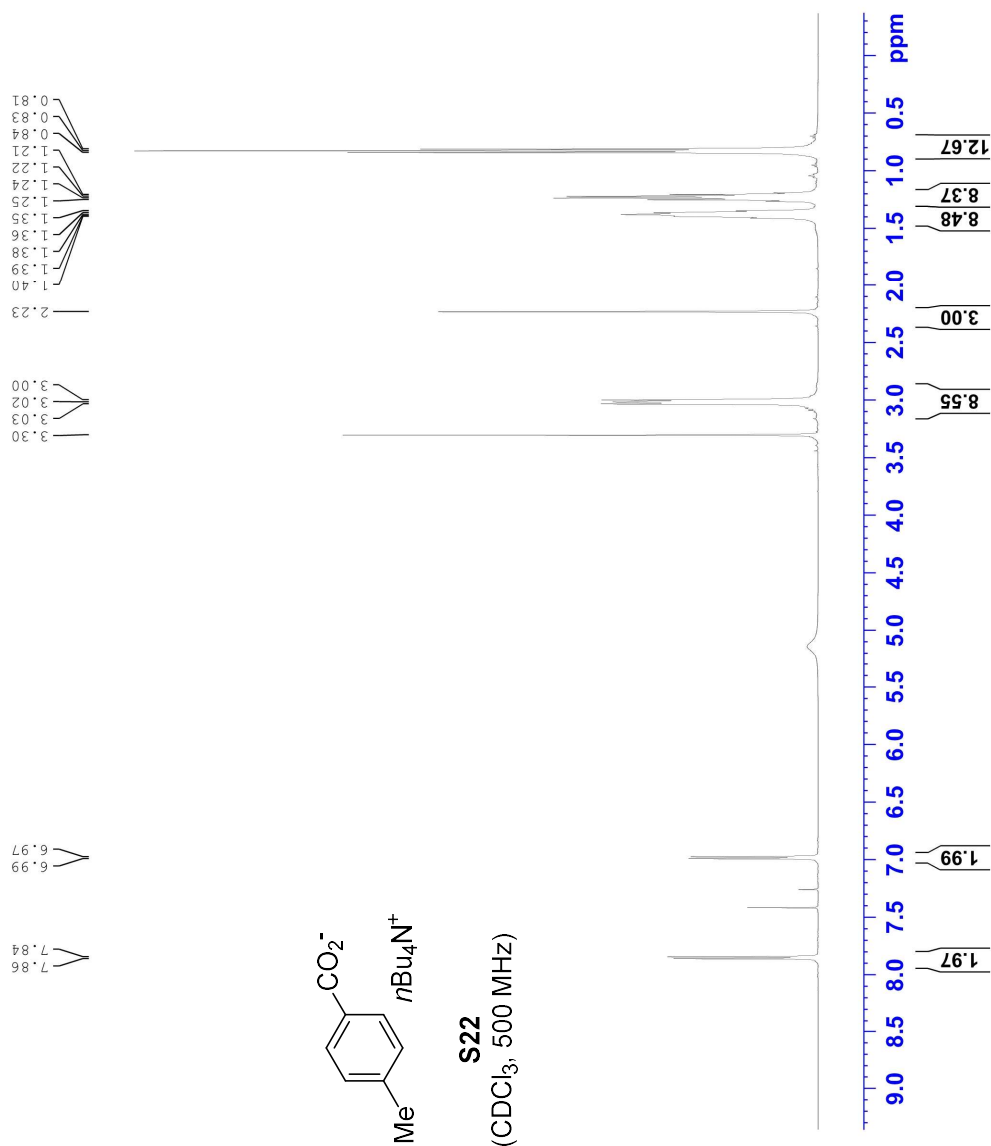


Current Data Parameters
 NAME yh-2-41
 EXPNO 1
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20180814
 Time 14.39
 INSTRUM spect
 PROBHD 5 mm PATXI 1H/
 PULPROG zg
 TD 59998
 SOLVENT CDCl3
 NS 4
 DS 0
 SWH 10000.000 Hz
 FIDRES 0.166672 Hz
 AQ 2.9999001 sec
 RG 9.88
 DW 50.000 usec
 DE 10.00 usec
 TE 294.3 K
 D1 4.0000000 sec
 TDO 1

===== CHANNEL f1 =====
 SFO1 500.1330885 MHz
 NUC1 1H
 P1 3.30 usec
 PLW1 12.19999981 W

F2 - Processing parameters
 SI 65536
 SF 500.1299379 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00



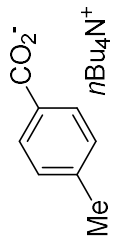
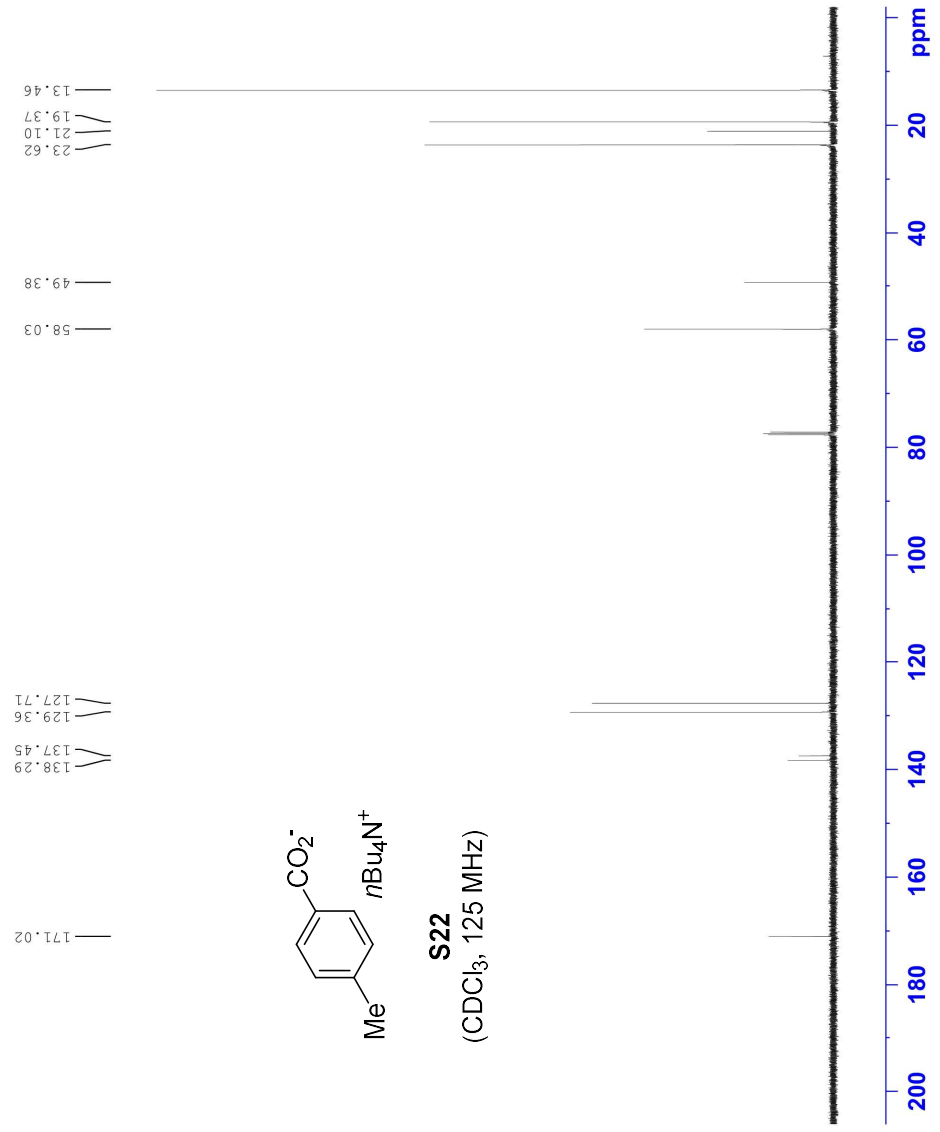
Current Data Parameters
 NAME yh-2-41
 EXPNO 2
 PROCNO 2

F2 - Acquisition Parameters
 Date_ 20180814
 Time 15.10
 INSTRUM spect
 PROBHD 5 mm PABBO BB/
 PULPROG zgpg
 TD 187496
 SOLVENT CDC13
 NS 5
 DS 0
 SWH 31250.000 Hz
 FIDRES 0.166670 Hz
 AQ 2.9999361 sec
 RG 2050
 DW 16.000 usec
 DE 6.50 usec
 TE 295.9 K
 TL 3.0000000 sec
 DL1 0.03000000 sec
 TD0 1

===== CHANNEL f1 =====
 SFO1 125.7049802 MHz
 NUC1 13C
 P1 10.00 usec
 PLW1 72.83999634 W

===== CHANNEL f2 =====
 SFO2 499.8724993 MHz
 NUC2 1H
 CPDPRG[2] waltz16
 PCPD2 80.00 usec
 PLW2 19.0000000 W
 PLW12 0.29688001 W

F2 - Processing parameters
 SI 1048576
 SF 125.6924115 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.40



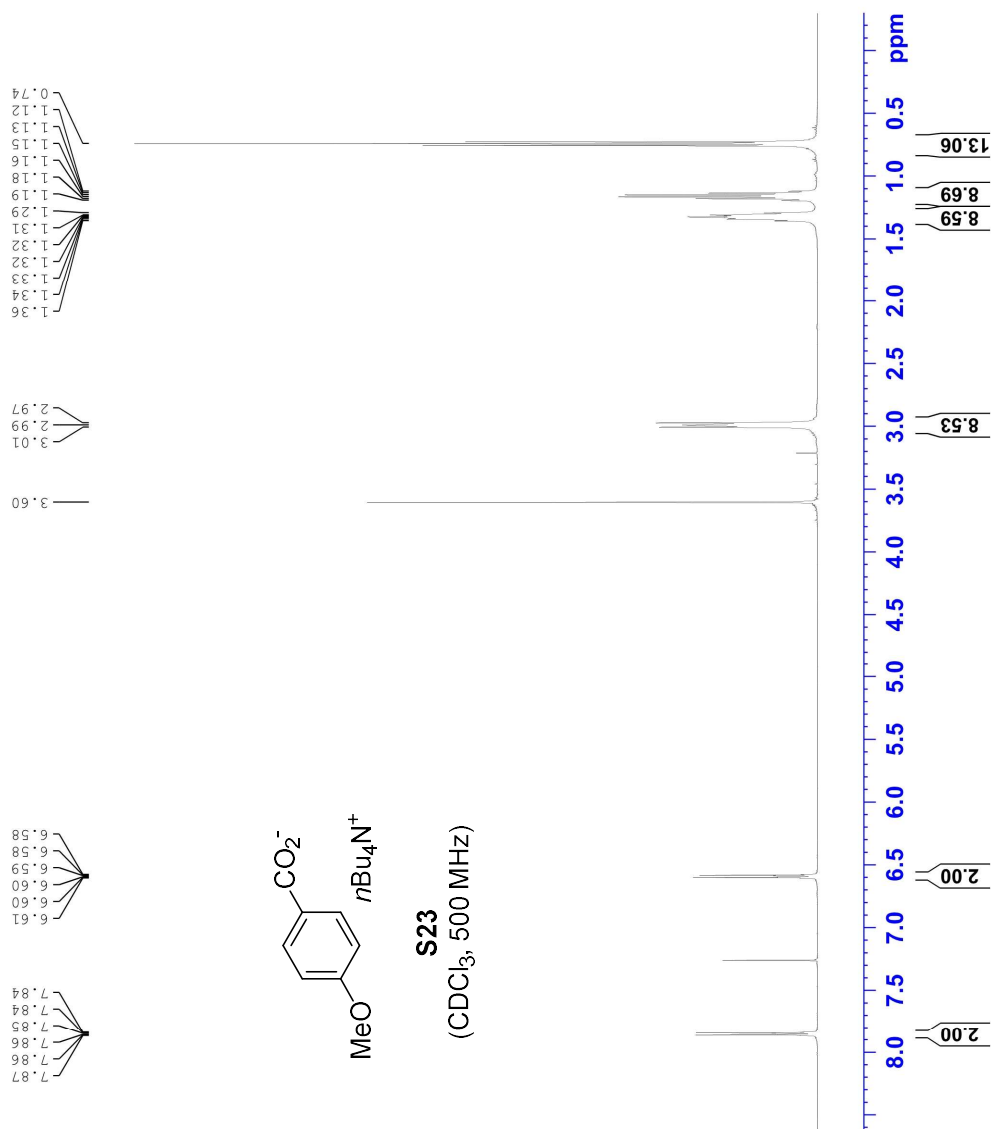
S22
 (CDCl₃, 125 MHz)

Current Data Parameters
 NAME yh-2-29
 EXPNO 1
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20180827
 Time_ 20.40
 INSTRUM spect
 PROBHD 5 mm PABBO BB/
 PULPROG zg
 TD 59998
 SOLVENT CDCl3
 NS 1
 DS 0
 SWH 10000.000 Hz
 FIDRES 0.166672 Hz
 AQ 2.9999001 sec
 RG 13.94
 DW 50.000 usec
 DE 6.50 usec
 TE 296.3 K
 D1 4.0000000 sec
 TD0 1

===== CHANNEL f1 =====
 SFO1 499.8730869 MHz
 NUC1 1H
 P1 3.58 usec
 PLW1 18.25000000 W

F2 - Processing parameters
 SI 65536
 SF 499.8700148 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00



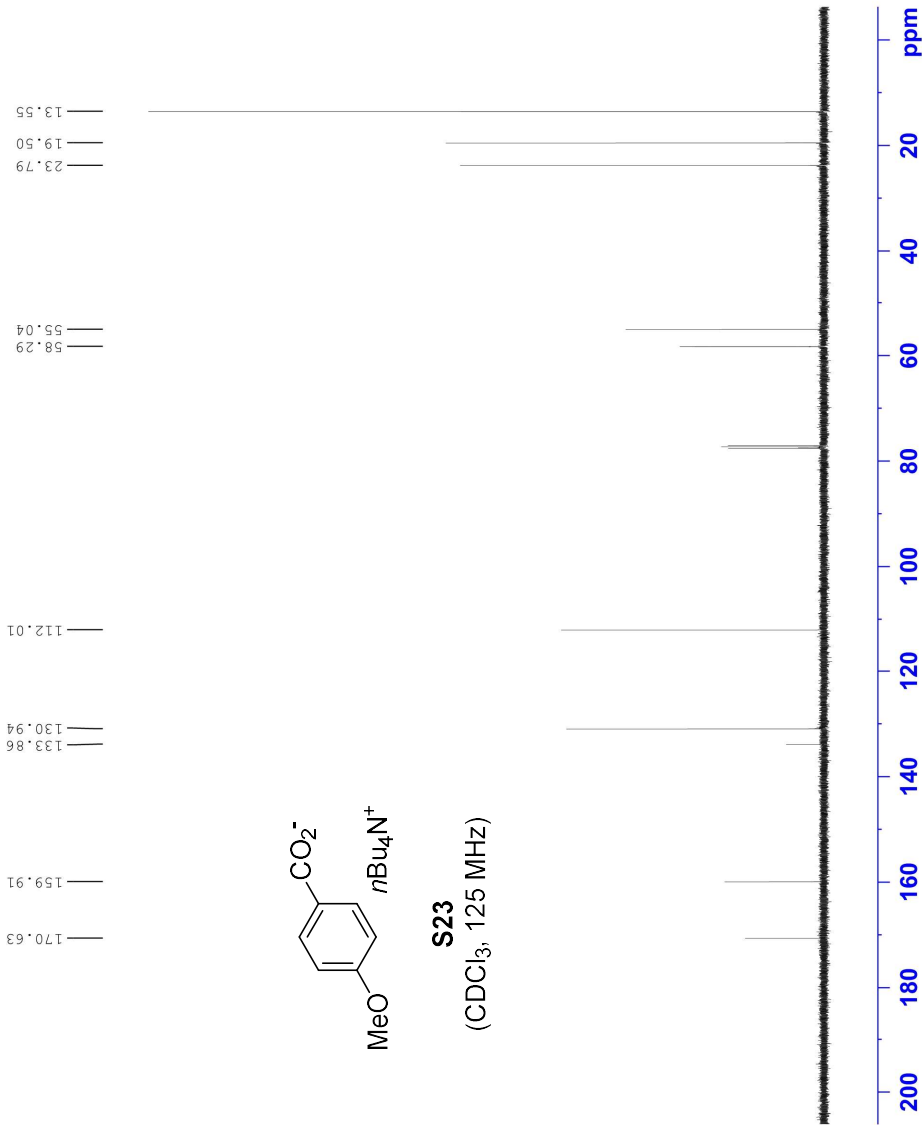
Current Data Parameters
 NAME yh-2-29
 EXPNO 2
 PROCNO 2

F2 - Acquisition Parameters
 Date_ 20180827
 Time_ 20.44
 INSTRUM spect
 PROBHD 5 mm PABBO BB/
 PULPROG zgpg
 TD 187496
 SOLVENT CDCl3
 NS 4
 DS 0
 SWH 31250.000 Hz
 FIDRES 0.166670 Hz
 AQ 2.9999361 sec
 RG 2050
 DW 16.000 usec
 DE 6.50 usec
 TE 296.3 K
 TL 3.0000000 sec
 DLI 0.03000000 sec
 TDO 1

===== CHANNEL f1 =====
 SFO1 125.7049802 MHz
 NUC1 13C
 P1 10.00 usec
 PLW1 72.83999634 W

===== CHANNEL f2 =====
 SFO2 499.8724993 MHz
 NUC2 1H
 CPDPRG[2] waltz16
 PCPD2 80.00 usec
 PLW2 19.0000000 W
 PLW12 0.29688001 W

F2 - Processing parameters
 SI 1048576
 SF 125.6924115 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.40




```

Current Data Parameters
NAME      yh-2-58
EXPNO     2
PROCNO    2

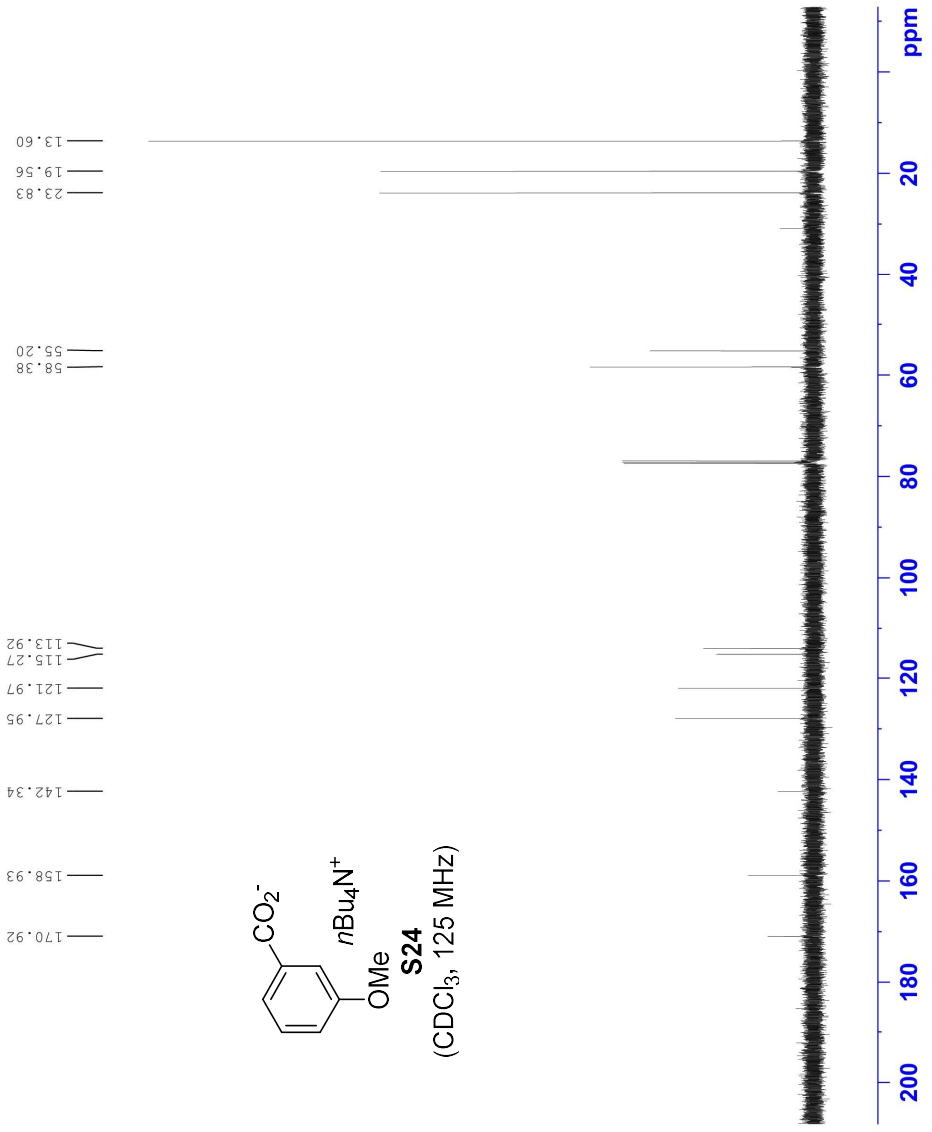
F2 - Acquisition Parameters
Date_     20180814
Time      21.25
INSTRUM   spect
PROBHD    5 mm PABBO BB/
PULPROG   zgpg
TD         187496
SOLVENT   CDCl3
NS         6
DS         0
SWH        31250.000 Hz
FIDRES     0.166670 Hz
AQ         2.9999361 sec
RG         2050
DW         16.000 usec
DE         6.50 usec
TE         296.0 K
D1         3.0000000 sec
d11        0.03000000 sec
TD0        1

===== CHANNEL f1 =====
SFO1      125.7049802 MHz
NUC1       13C
P1         10.00 usec
PLW1      72.83999634 W

===== CHANNEL f2 =====
SFO2      499.8724993 MHz
NUC2       1H
CPDPRG[2] waltz16
PCPD2     80.00 usec
PLW2      19.0000000 W
PLW12     0.29688001 W

F2 - Processing parameters
SI         1048576
SF         125.6924115 MHz
WDW        EM
SSB        0
LB         0.30 Hz
GB         0
PC         1.40

```

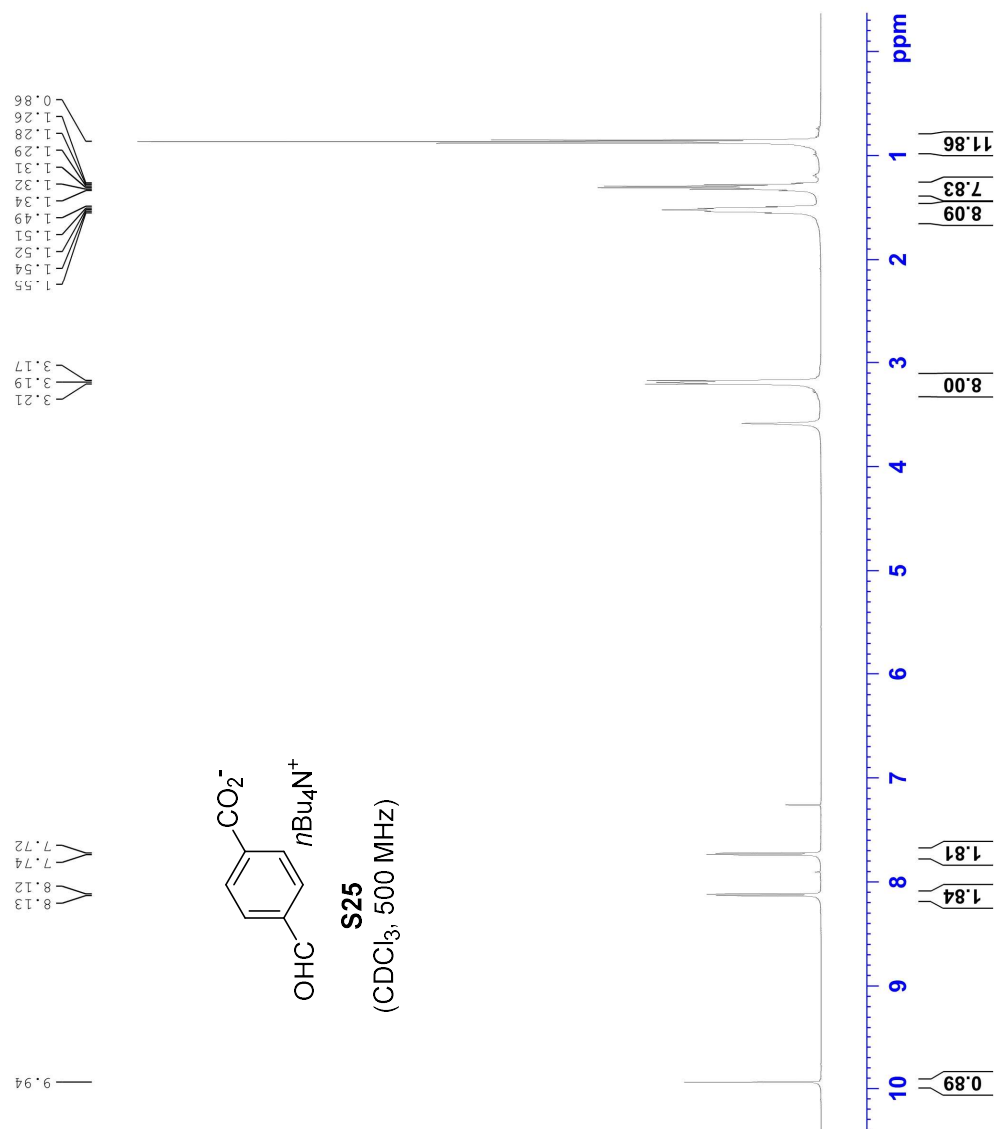


Current Data Parameters
 NAME yh-2-75
 EXPNO 1
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20180814
 Time_ 22.45
 INSTRUM spect
 PROBHD 5 mm PABBO BB/
 PULPROG zg
 TD 59998
 SOLVENT CDCl3
 NS 1
 DS 0
 SWH 10000.000 Hz
 FIDRES 0.166672 Hz
 AQ 2.9999001 sec
 RG 25.24
 DW 50.000 usec
 DE 6.50 usec
 TE 296.1 K
 D1 4.0000000 sec
 TDO 1

===== CHANNEL f1 =====
 SFO1 499.8730869 MHz
 NUC1 1H
 P1 3.58 usec
 PLW1 18.25000000 W

F2 - Processing parameters
 SI 65536
 SF 499.8700126 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00



```

Current Data Parameters
NAME      yh-2-75
EXPNO     2
PROCNO    2

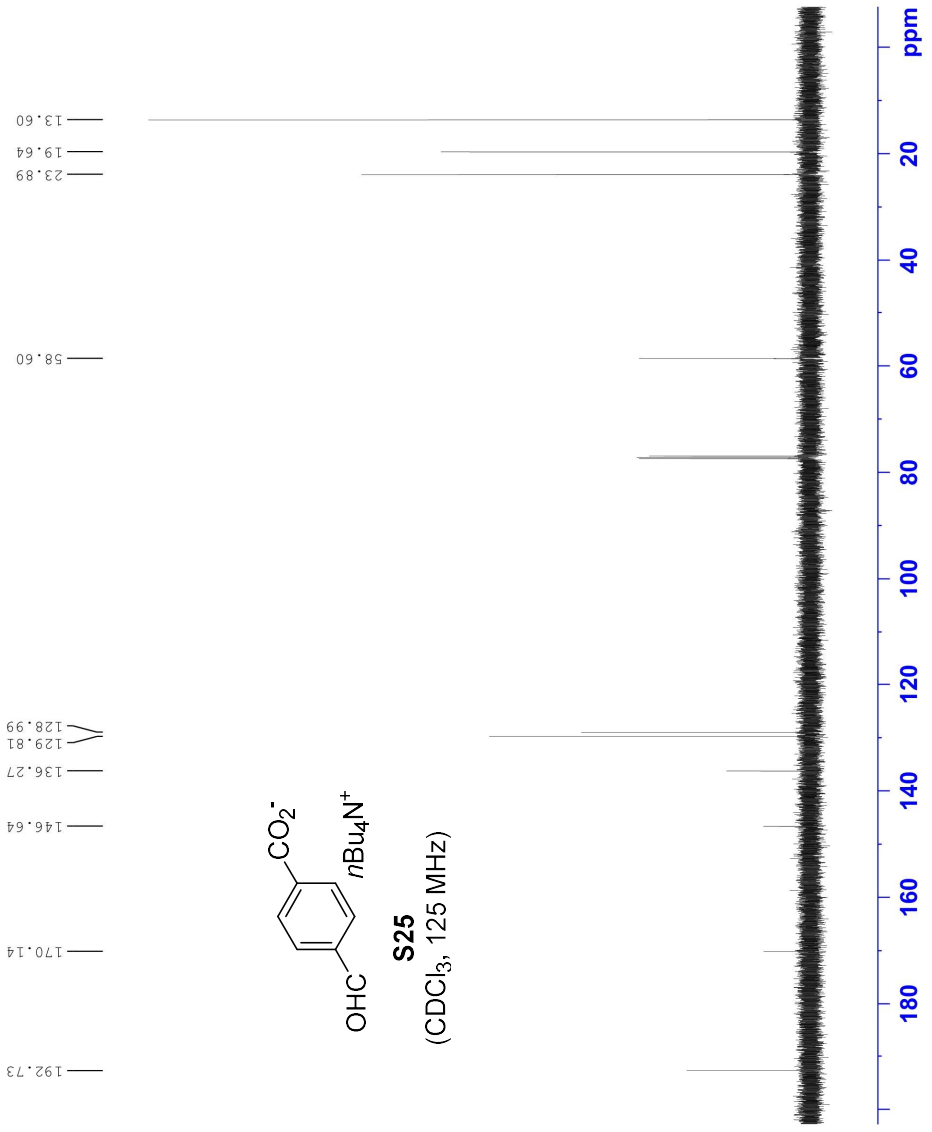
F2 - Acquisition Parameters
Date_     20180814
Time      22.48
INSTRUM   spect
PROBHD    5 mm PABBO BB/
PULPROG   zgpg
TD         187496
SOLVENT   CDC13
NS         3
DS         0
SWH        31250.000 Hz
FIDRES     0.166670 Hz
AQ         2.9999361 sec
RG         2050
DW         16.000 usec
DE         6.50 usec
TE         296.1 K
D1         3.0000000 sec
DL1        0.03000000 sec
TD0        1

===== CHANNEL f1 =====
SFO1      125.7049802 MHz
NUC1      13C
P1         10.00 usec
PLW1      72.83999634 W

===== CHANNEL f2 =====
SFO2      499.8724993 MHz
NUC2      1H
CFPRG[2]  waltz16
PCPD2     80.00 usec
PLW2      19.0000000 W
PLWL2     0.29688001 W

F2 - Processing parameters
SI         1048576
SF         125.6924115 MHz
WDW        EM
SSB        0
LB         0.30 Hz
GB         0
PC         1.40

```



Current Data Parameters
 NAME yh-2-53
 EXPNO 1
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20180814
 Time 16.03
 INSTRUM spect
 PROBHD 5 mm PABBO BB/
 PULPROG zg
 TD 59998
 SOLVENT CDCl3
 NS 1
 DS 0
 SWH 10000.000 Hz
 FIDRES 0.166672 Hz
 AQ 2.9999001 sec
 RG 13.94
 DW 50.000 usec
 DE 6.50 usec
 TE 295.8 K
 DL 4.0000000 sec
 TD0 1

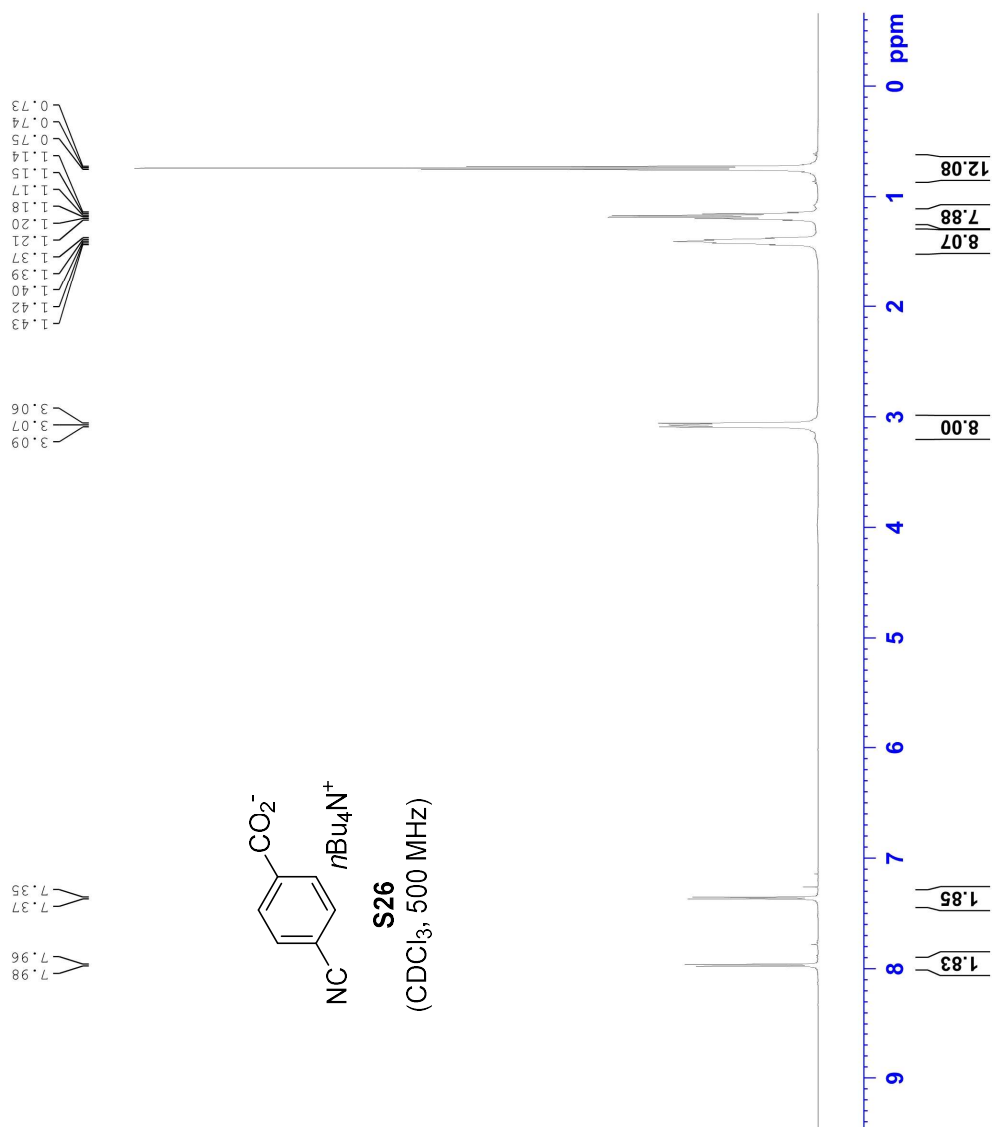
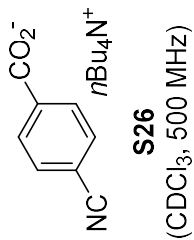
===== CHANNEL f1 =====
 SFO1 499.8730869 MHz
 NUC1 1H
 P1 3.58 usec
 PLW1 18.25000000 W

F2 - Processing parameters
 SI 65536
 SF 499.8700143 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00

1.43
1.42
1.40
1.39
1.37
1.21
1.20
1.18
1.17
1.15
1.14
0.75
0.74
0.73

3.09
3.07
3.06

7.96
7.97
7.98



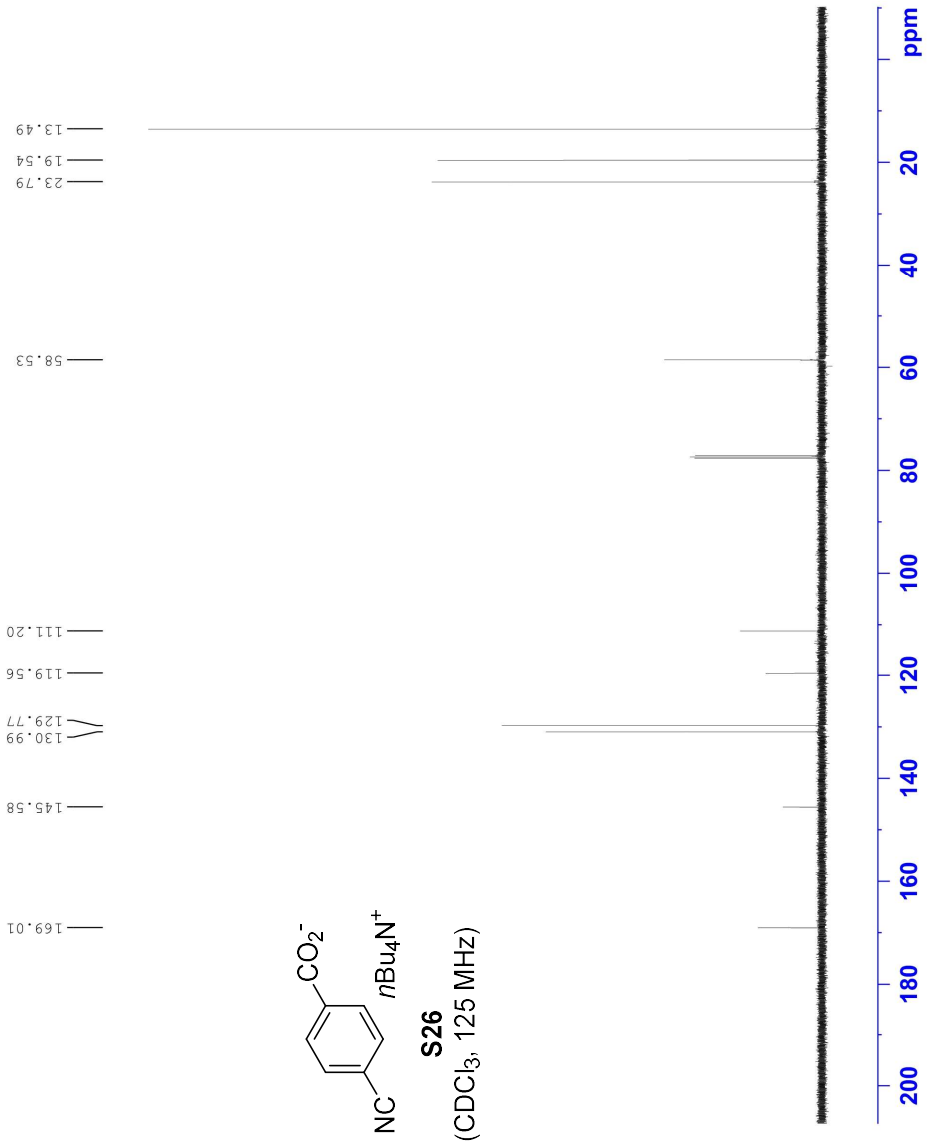
Current Data Parameters
 NAME yh-2-53
 EXPNO 2
 PROCNO 2

F2 - Acquisition Parameters
 Date_ 20180814
 Time_ 16.07
 INSTRUM spect
 PROBHD 5 mm PABBO BB/
 PULPROG zgpg
 TD 187496
 SOLVENT CDC13
 NS 3
 DS 0
 SWH 31250.000 Hz
 FIDRES 0.166670 Hz
 AQ 2.9999361 sec
 RG 2050
 DW 16.000 usec
 DE 6.50 usec
 TE 295.8 K
 D1 3.0000000 sec
 D11 0.03000000 sec
 TDO 1

===== CHANNEL f1 =====
 SFO1 125.7049802 MHz
 NUC1 13C
 P1 10.00 usec
 PLW1 72.83999634 W

===== CHANNEL f2 =====
 SFO2 499.8724993 MHz
 NUC2 1H
 CPDPRG[2] waltz16
 PCPD2 80.00 usec
 PLW2 19.0000000 W
 PLW12 0.29688001 W

F2 - Processing parameters
 SI 1048576
 SF 125.6924115 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.40

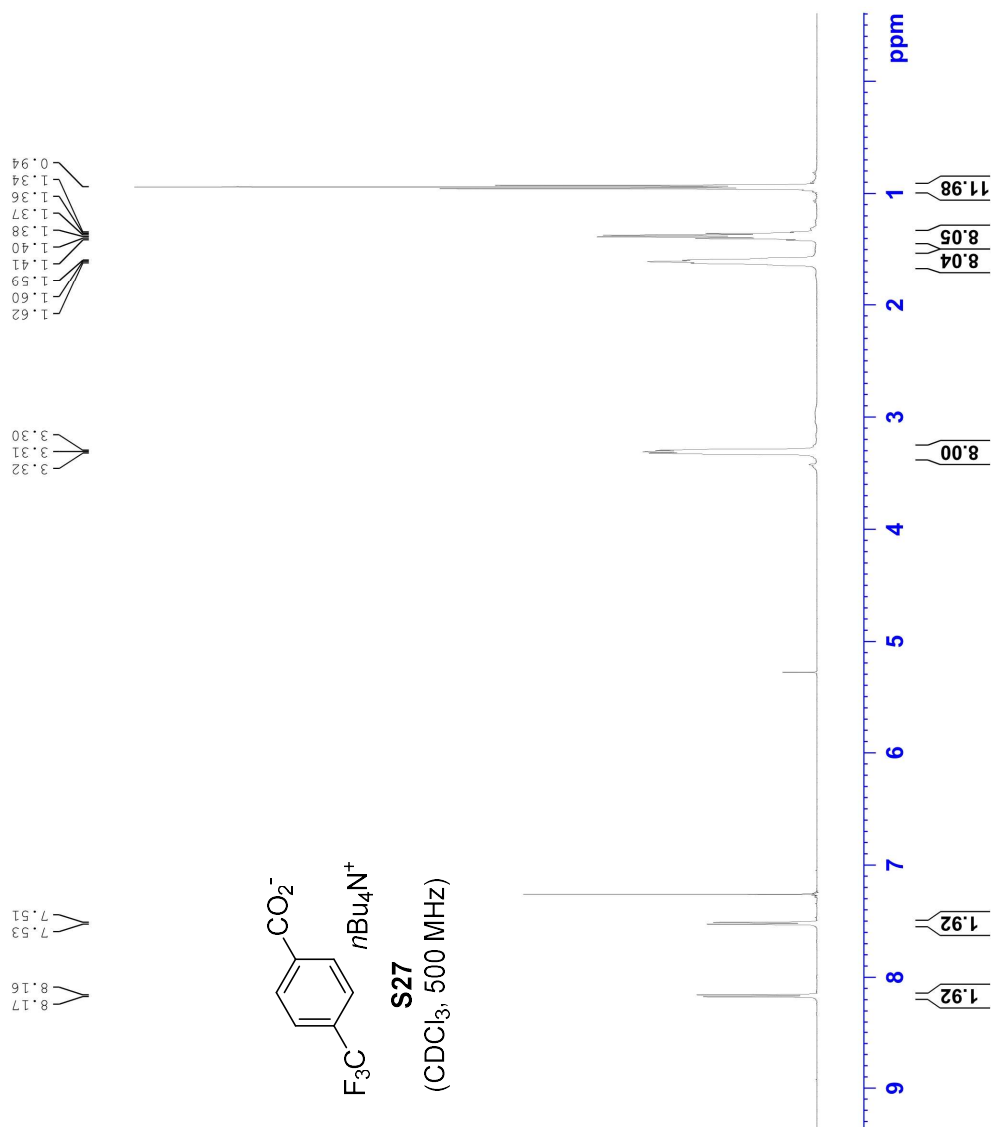


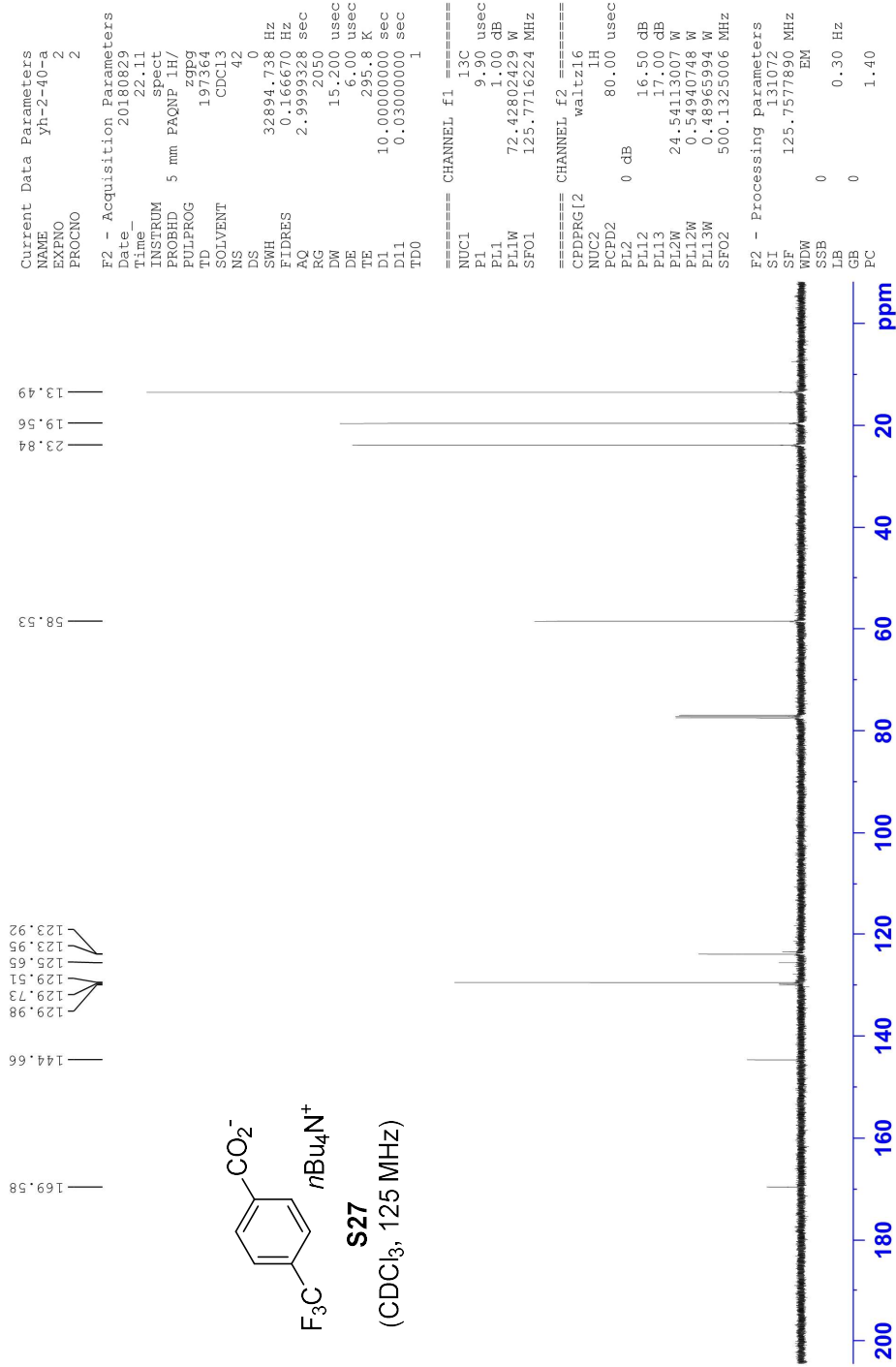
Current Data Parameters
 NAME Yh-2-40-a
 EXPNO 1
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20180829
 Time 22.24
 INSTRUM spect
 PROBHD 5 mm PATXI 1H/
 PULPROG zg
 TD 59998
 SOLVENT CDCl3
 NS 1
 DS 0
 SWH 10000.000 Hz
 FIDRES 0.166672 Hz
 AQ 2.9999001 sec
 RG 97.37
 DW 50.000 usec
 DE 10.00 usec
 TE 294.1 K
 D1 4.0000000 sec
 TD0 1

===== CHANNEL f1 =====
 SFO1 500.1330885 MHz
 NUC1 1H
 P1 3.30 usec
 PLW1 12.19999981 W

F2 - Processing parameters
 SI 65536
 SF 500.1300140 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00



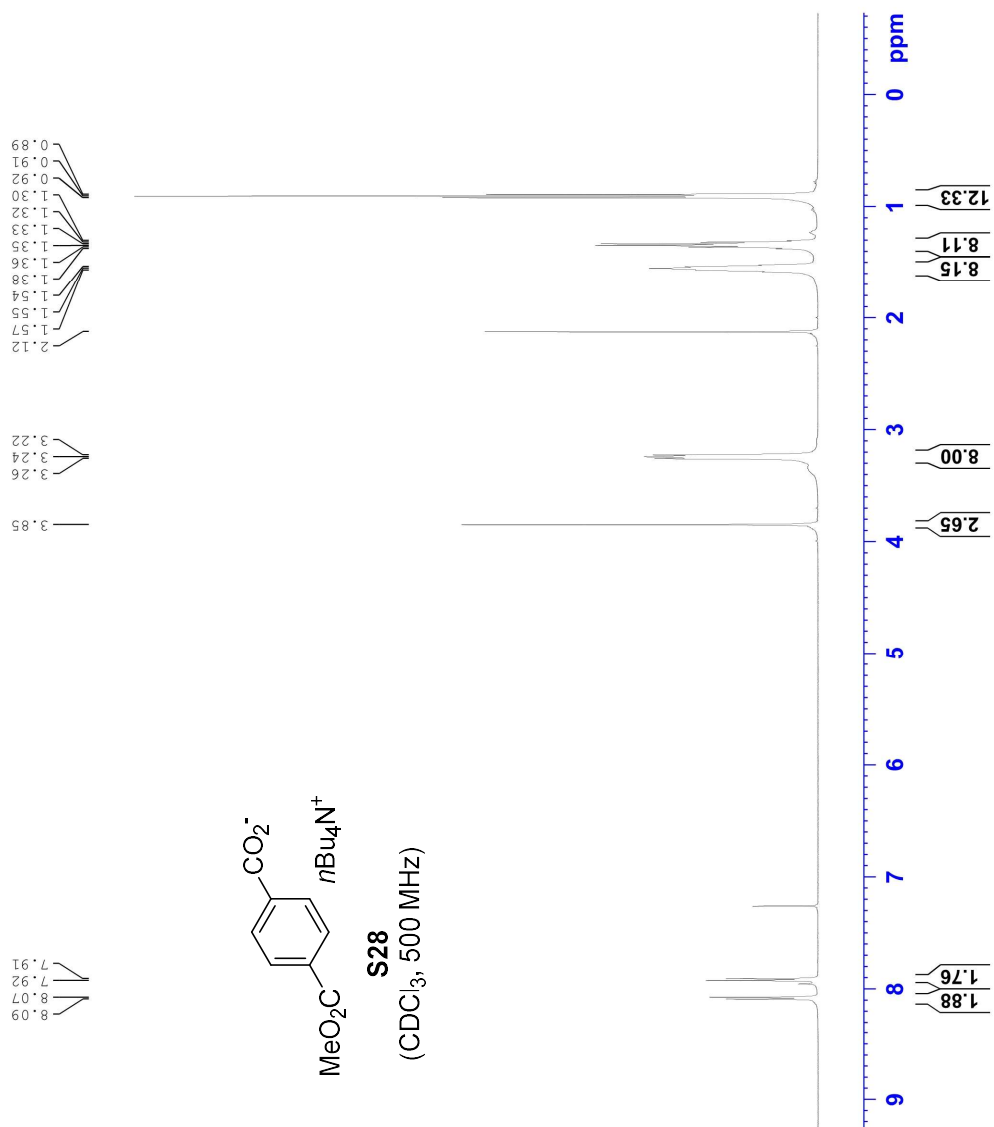


Current Data Parameters
 NAME yh-2-65
 EXPNO 1
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20180814
 Time 21:31
 INSTRUM spect
 PROBHD 5 mm PABBO BB/
 PULPROG zg
 TD 59998
 SOLVENT CDCl3
 NS 2
 DS 0
 SWH 10000.000 Hz
 FIDRES 0.166672 Hz
 AQ 2.9999001 sec
 RG 44.57
 DW 50.000 usec
 DE 6.50 usec
 TE 296.0 K
 DL 4.0000000 sec
 TDO 1

===== CHANNEL f1 =====
 SFO1 499.8730869 MHz
 NUC1 1H
 P1 3.58 usec
 PLW1 18.25000000 W

F2 - Processing parameters
 SI 65536
 SF 499.8700123 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00



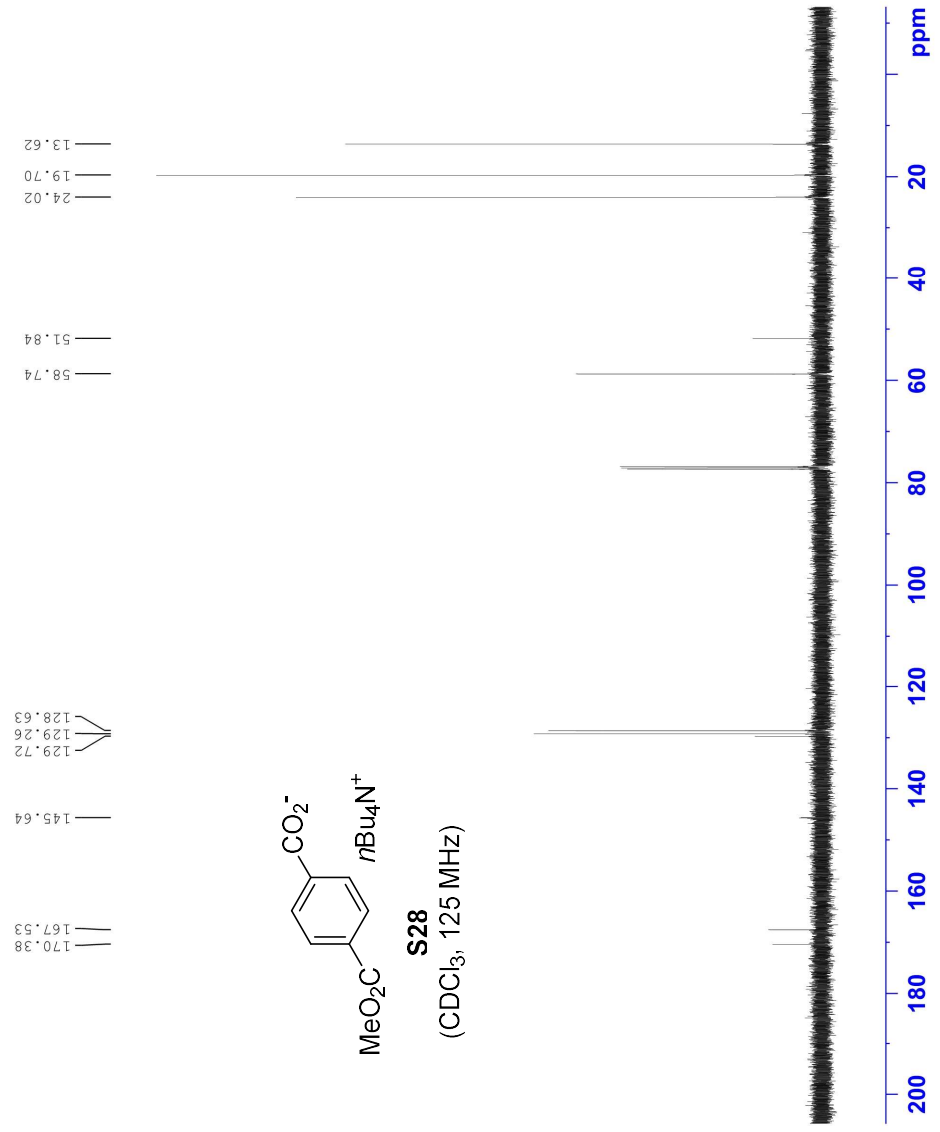
Current Data Parameters
 NAME yh-2-65
 EXPNO 2
 PROCNO 2

F2 - Acquisition Parameters
 Date_ 20180814
 Time_ 21.45
 INSTRUM spect
 PROBHD 5 mm PABBO BB/
 PULPROG zgpg
 TD 187496
 SOLVENT CDCl3
 NS 47
 DS 0
 SWH 31250.000 Hz
 FIDRES 0.166670 Hz
 AQ 2.9999361 sec
 RG 2050
 DW 16.000 usec
 DE 6.50 usec
 TE 296.8 K
 D1 3.0000000 sec
 DL1 0.03000000 sec
 TDO 1

===== CHANNEL f1 =====
 SFO1 125.7049802 MHz
 NUC1 13C
 P1 10.00 usec
 PLW1 72.83999634 W

===== CHANNEL f2 =====
 SFO2 499.8724993 MHz
 NUC2 1H
 CPDPRG[2] waltz16
 PCPD2 80.00 usec
 PLW2 19.0000000 W
 PLW12 0.29688001 W

F2 - Processing parameters
 SI 1048576
 SF 125.6924115 MHz
 WDW EM
 SSB 0
 LB 0 0.30 Hz
 GB 0
 PC 1.40




```

Current Data Parameters
NAME      yh-2-70
EXPNO    2
PROCNO   2

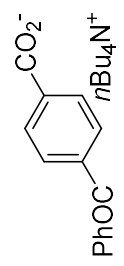
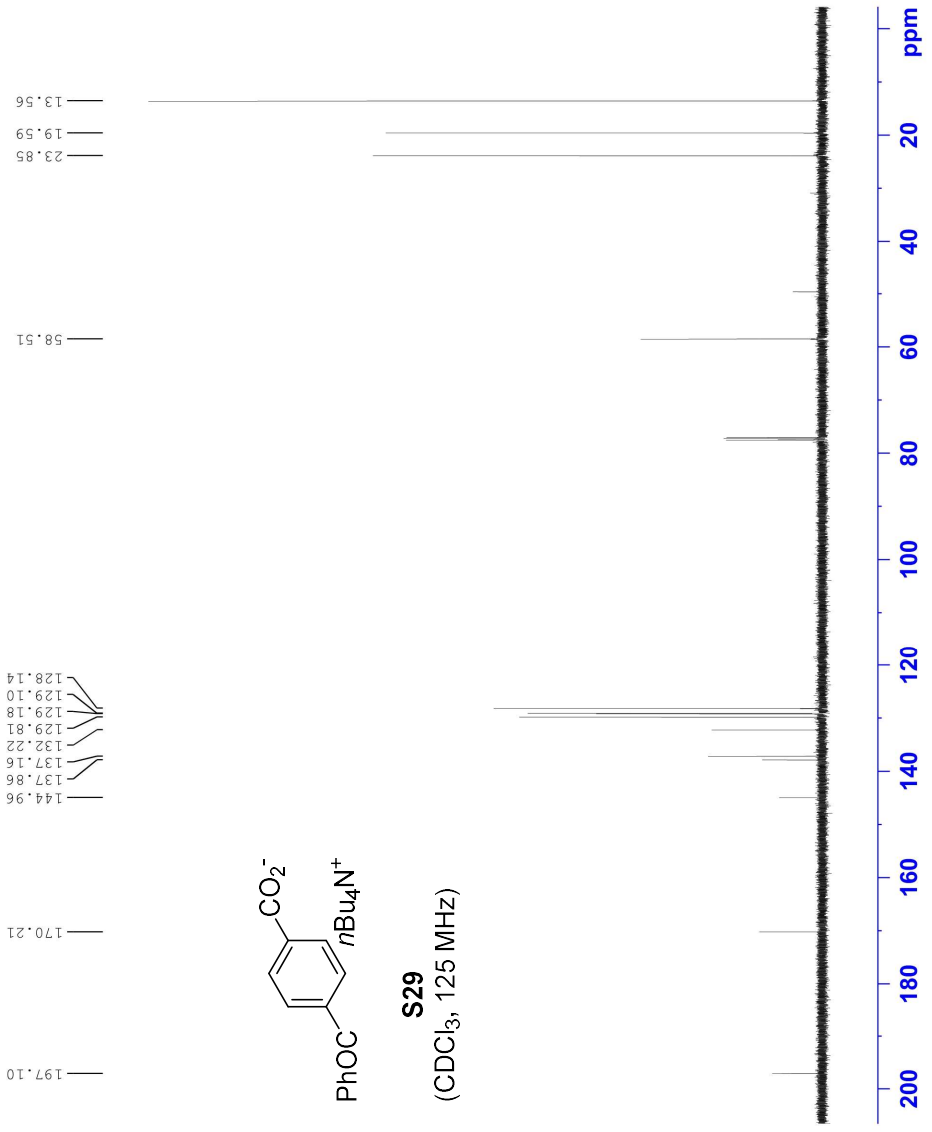
F2 - Acquisition Parameters
Date_    20180814
Time     21.59
INSTRUM spect
PROBHD   5 mm PABBO BB/
PULPROG zgpg
TD       187496
SOLVENT  CDCl3
NS       6
DS       0
SWH      31250.000 Hz
FIDRES   0.166670 Hz
AQ       2.9999361 sec
RG       2050
DW       16.000 usec
DE       6.50 usec
TE       296.4 K
D1       3.0000000 sec
D11      0.03000000 sec
TD0      1

===== CHANNEL f1 =====
SFO1    125.7049802 MHz
NUC1    13C
P1      10.00 usec
PLW1    72.83999634 W

===== CHANNEL f2 =====
SFO2    499.8724993 MHz
NUC2    1H
PCPD2   waitz16
PLW2    19.0000000 W
PLWL2   0.29688001 W

F2 - Processing parameters
SI      1048576
SF      125.6924115 MHz
WDW     EM
SSB     0
LB      0.30 Hz
GB      0
PC      1.40

```



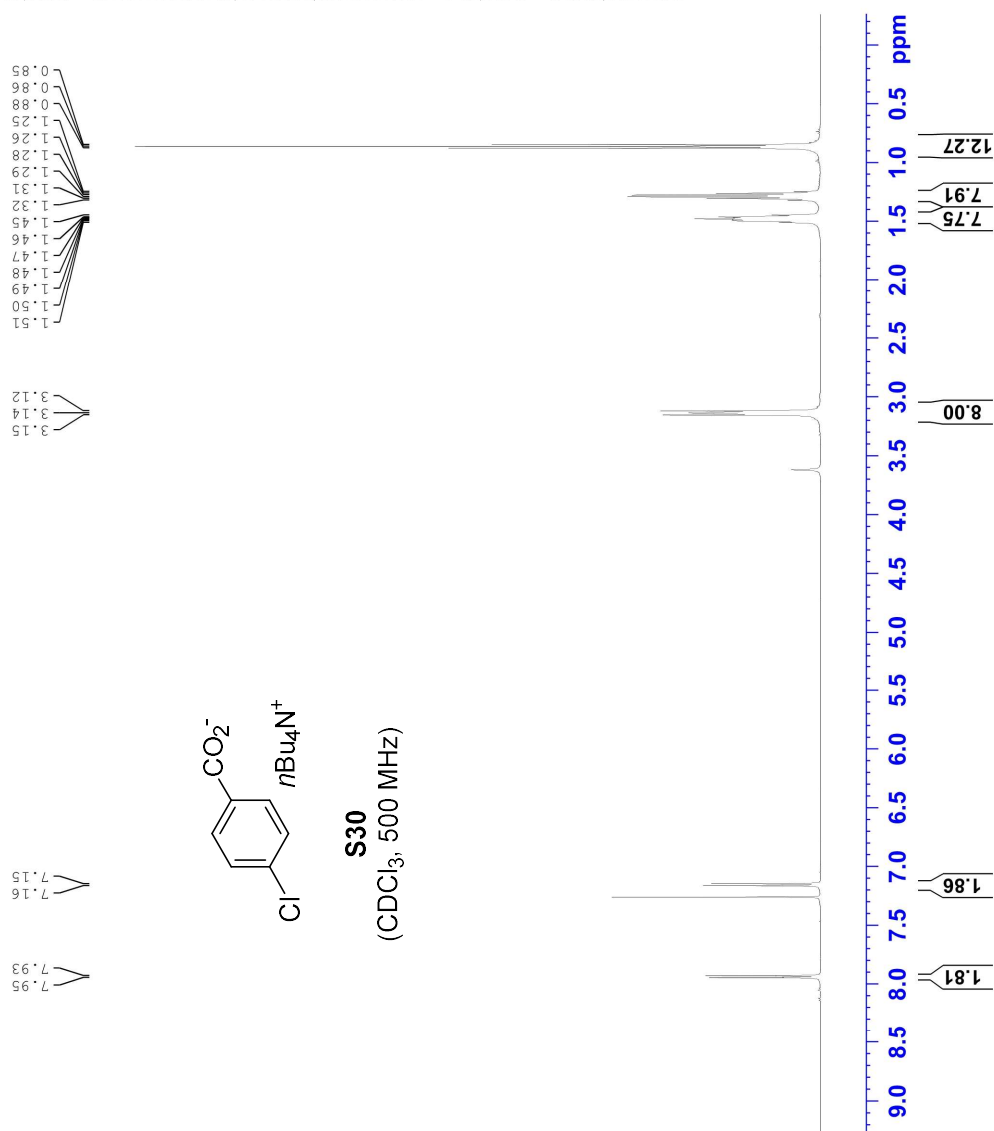
S29
(CDCl₃, 125 MHz)

Current Data Parameters
 NAME yh-2-44
 EXPNO 1
 PROCNO 1

F2 - Acquisition Parameters
 Date 20180814
 Time 15.54
 INSTRUM spect
 PROBHD 5 mm PABBO BB/
 PULPROG zg
 TD 5998
 SOLVENT CDCl3
 NS 6
 DS 0
 SWH 10000.000 Hz
 FIDRES 0.166672 Hz
 AQ 2.9999001 sec
 RG 25.24
 DW 50.000 usec
 DE 6.50 usec
 TE 295.7 K
 DL 4.0000000 sec
 TDO 1

===== CHANNEL f1 =====
 SFO1 499.8730869 MHz
 NUC1 1H
 P1 3.58 usec
 PLW1 18.25000000 W

F2 - Processing parameters
 SI 65536
 SF 499.8700130 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00



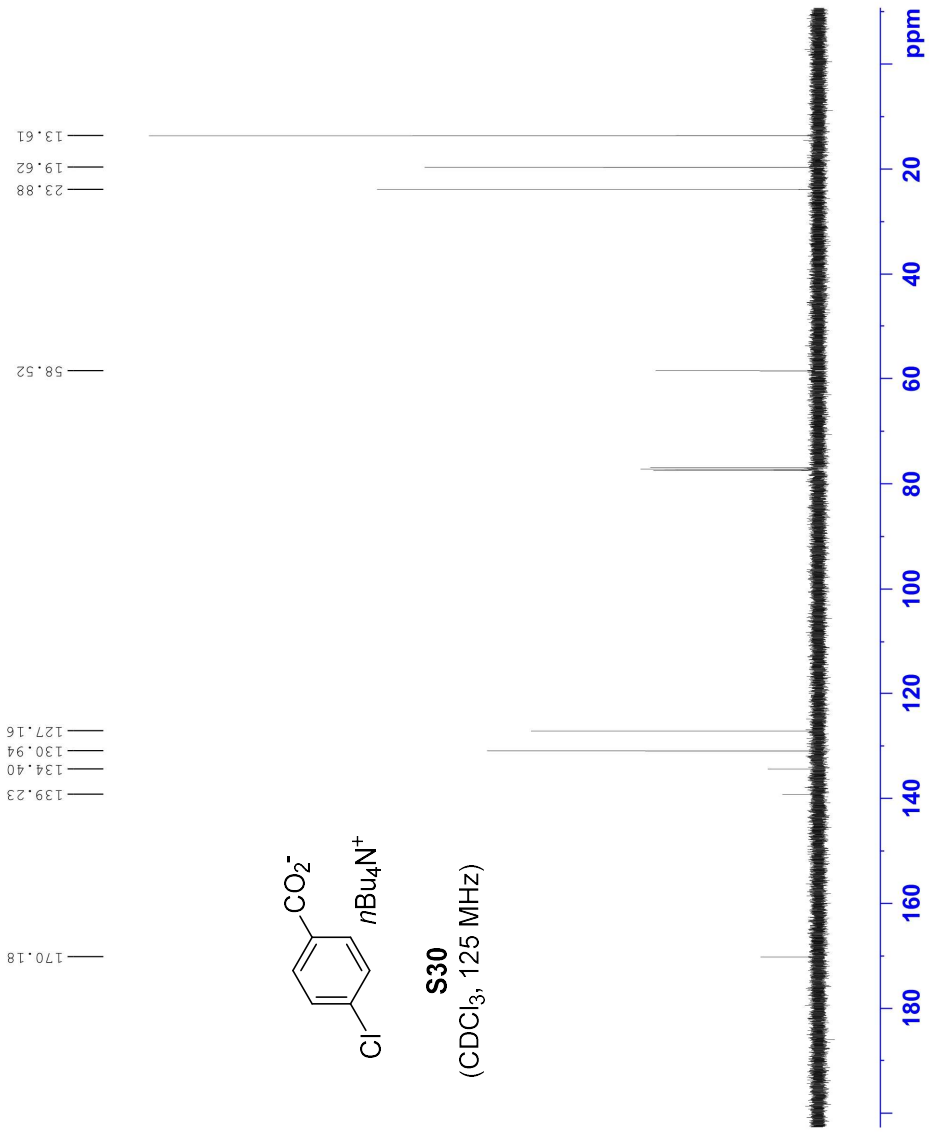
Current Data Parameters
 NAME yh-2-44
 EXPNO 2
 PROCNO 2

F2 - Acquisition Parameters
 Date_ 20180814
 Time_ 15.58
 INSTRUM spect
 PROBHD 5 mm PABBO BB/
 PULPROG zgpg
 TD 187496
 SOLVENT CDC13
 NS 4
 DS 0
 SWH 31250.000 Hz
 FIDRES 0.166670 Hz
 AQ 2.9999361 sec
 RG 2050
 DW 16.000 usec
 DE 6.50 usec
 TE 295.8 K
 D1 3.0000000 sec
 D11 0.0300000 sec
 TDO 1

==== CHANNEL f1 =====
 SFO1 125.7049802 MHz
 NUC1 13C
 P1 10.00 usec
 PLW1 72.83999634 W

==== CHANNEL f2 =====
 SFO2 499.8724993 MHz
 NUC2 1H
 CPDPRG12 waitz16
 PCPD2 80.00 usec
 PLW2 19.0000000 W
 PLW12 0.29688001 W

F2 - Processing parameters
 SI 1048576
 SF 125.6924115 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.40



```

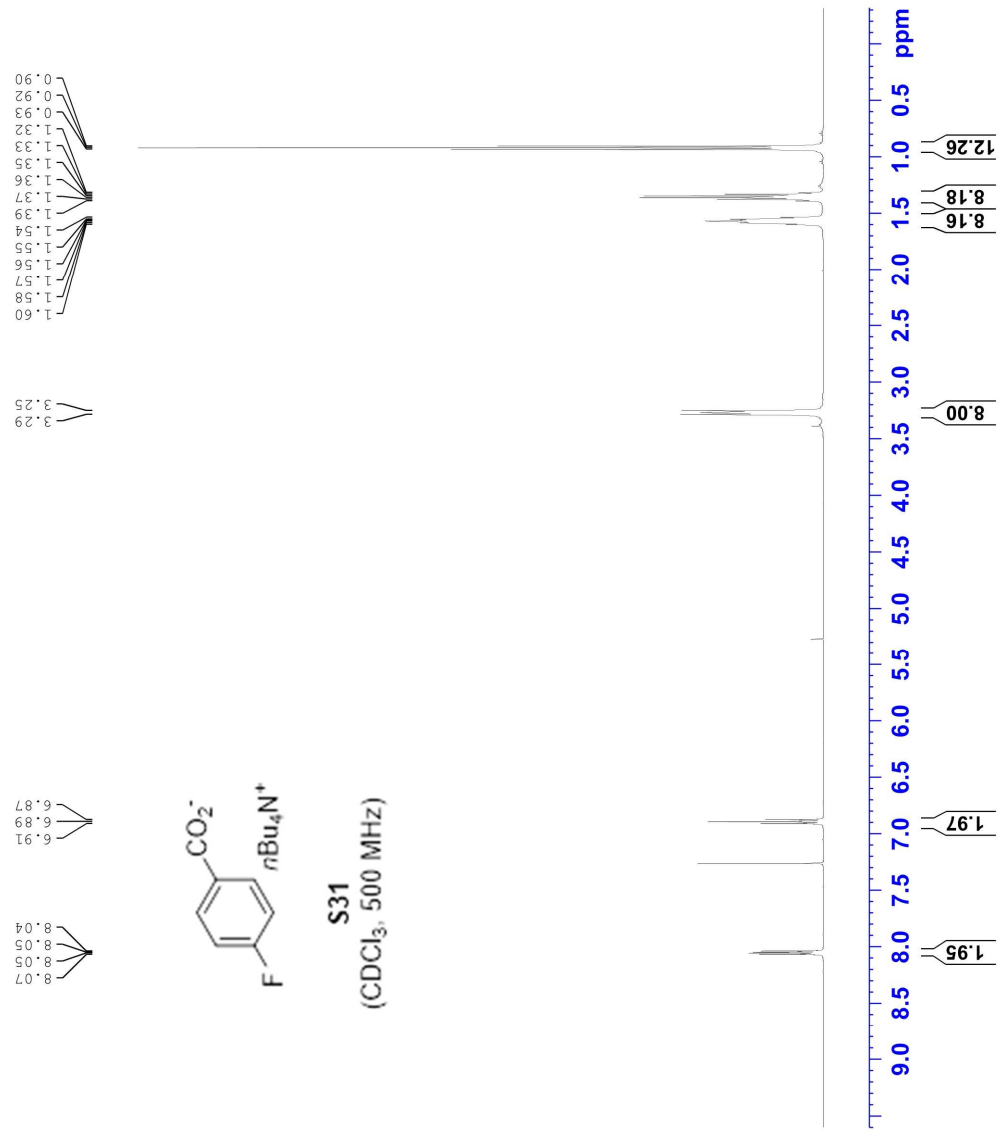
Current Data Parameters
NAME      Yh-2-51-a
EXPNO    1
PROCNO   1

F2 - Acquisition Parameters
Date_    20180829
Time     19.37
INSTRUM  spect
PROBHD   5 mm PATXI 1H/
PULPROG  zg
TD        59998
SOLVENT  CDCl3
NS        2
DS        0
SWH       10000.000 Hz
FIDRES    0.166672 Hz
AQ         2.9999001 sec
RG         71.78
DW         50.000 usec
DE         10.00 usec
TE         294.2 K
D1         4.0000000 sec
TD0        1

===== CHANNEL f1 =====
SFO1     500.1330885 MHz
NUC1      1H
P1         3.30 usec
PLW1     12.19999981 W

F2 - Processing parameters
SI        65536
SF        500.1300143 MHz
WDW       EM
SSB       0
LB        0.30 Hz
GB        0
PC        1.00

```



```

Current Data Parameters
NAME      Yh-2-51-a
EXPNO    2
PROCNO   2

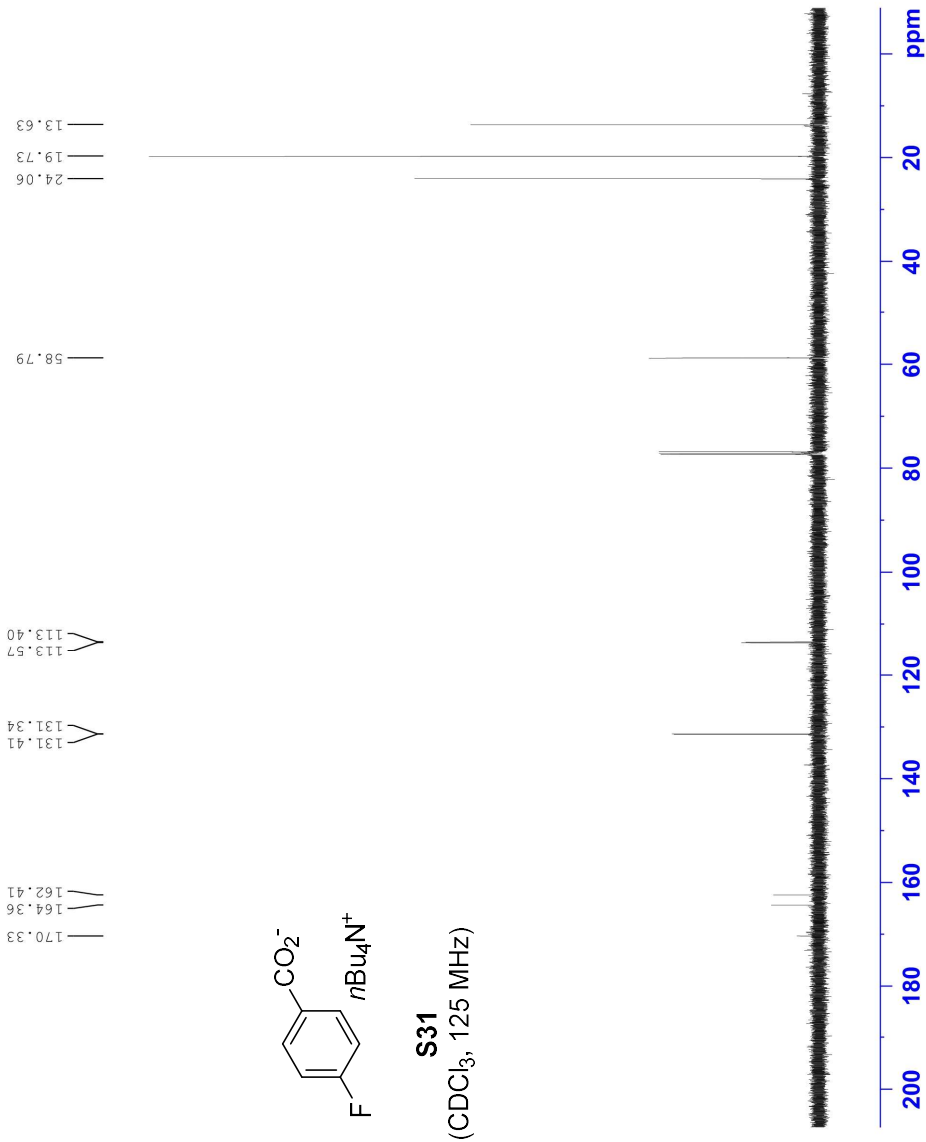
F2 - Acquisition Parameters
Date_     20180829
Time      19.56
INSTRUM   spect
PROBHD    5 mm PABBO BB/
PULPROG   zgpg
TD         187496
SOLVENT   CDC13
NS         52
DS         0
SWH        31250.000 Hz
FIDRES     0.166670 Hz
AQ         2.9999361 sec
RG         2050
DW         16.000 usec
DE         6.50 usec
TE         296.7 K
D1         3.0000000 sec
D11        0.03000000 sec
TDO        1

===== CHANNEL f1 =====
SFO1      125.7049802 MHz
NUC1       13C
P1         10.00 usec
PLW1      72.83999634 W

===== CHANNEL f2 =====
SFO2      499.8724993 MHz
NUC2       1H
CFDPERG12 waitz16
PCPD2     80.00 usec
PLW2      19.0000000 W
PLW12     0.29688001 W

F2 - Processing parameters
SI         1048576
SF         125.6924115 MHz
WDW        EM
SSB        0
LB         0.30 Hz
GB         0
PC         1.40

```

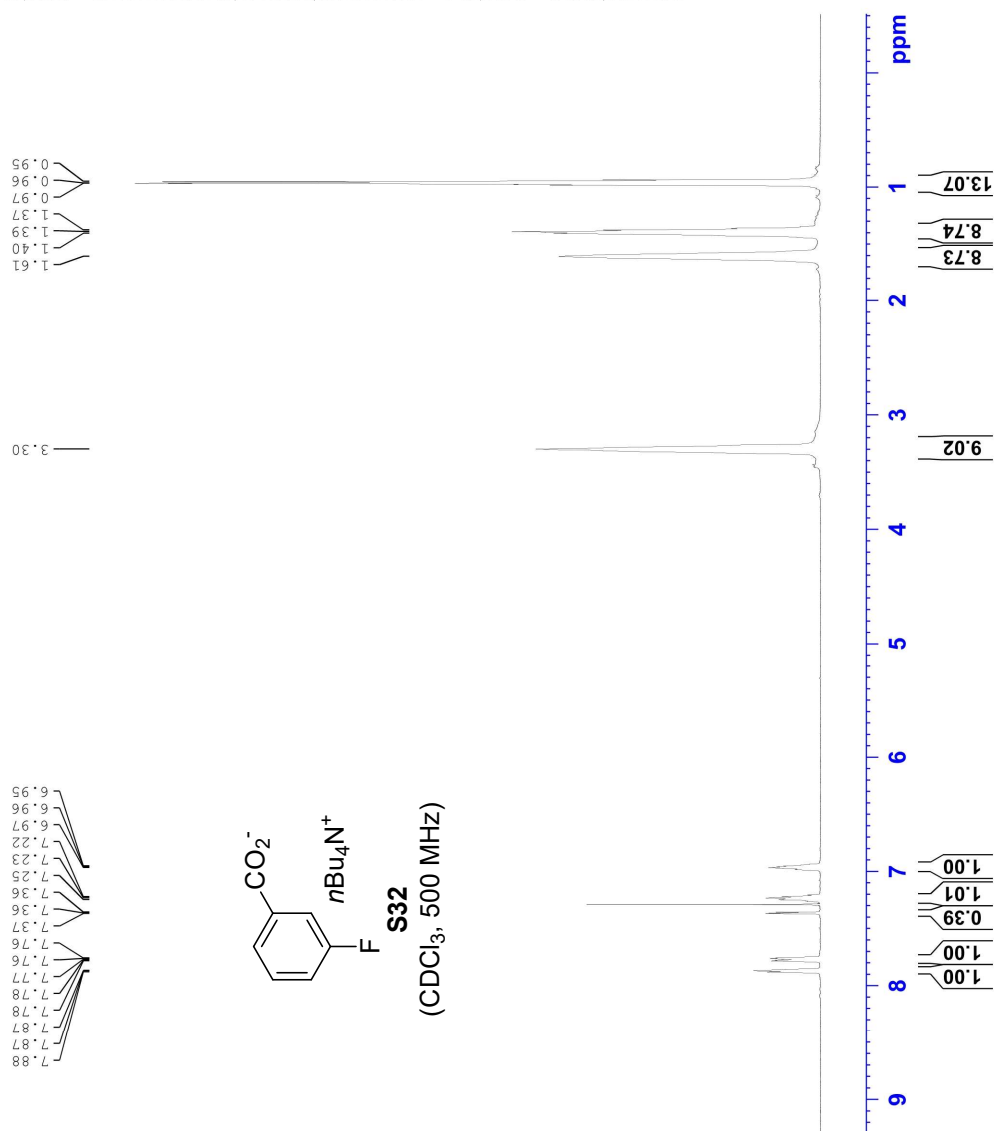


Current Data Parameters
 NAME Yh-2-66-a
 EXPNO 1
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20180829
 Time_ 22.43
 INSTRUM spect
 PROBHD 5 mm PATXI 1H/
 PULPROG zg
 TD 5998
 SOLVENT CDCl3
 NS 2
 DS 0
 SWH 10000.000 Hz
 FIDRES 0.166672 Hz
 AQ 2.9999001 sec
 RG 79.04
 DW 50.000 usec
 DE 10.00 usec
 TE 294.2 K
 DL 4.0000000 sec
 TDO 1

===== CHANNEL f1 =====
 SFO1 500.1330885 MHz
 NUC1 1H
 PL 3.30 usec
 PLW1 12.19999981 W

F2 - Processing parameters
 SI 65536
 SF 500.1300000 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00



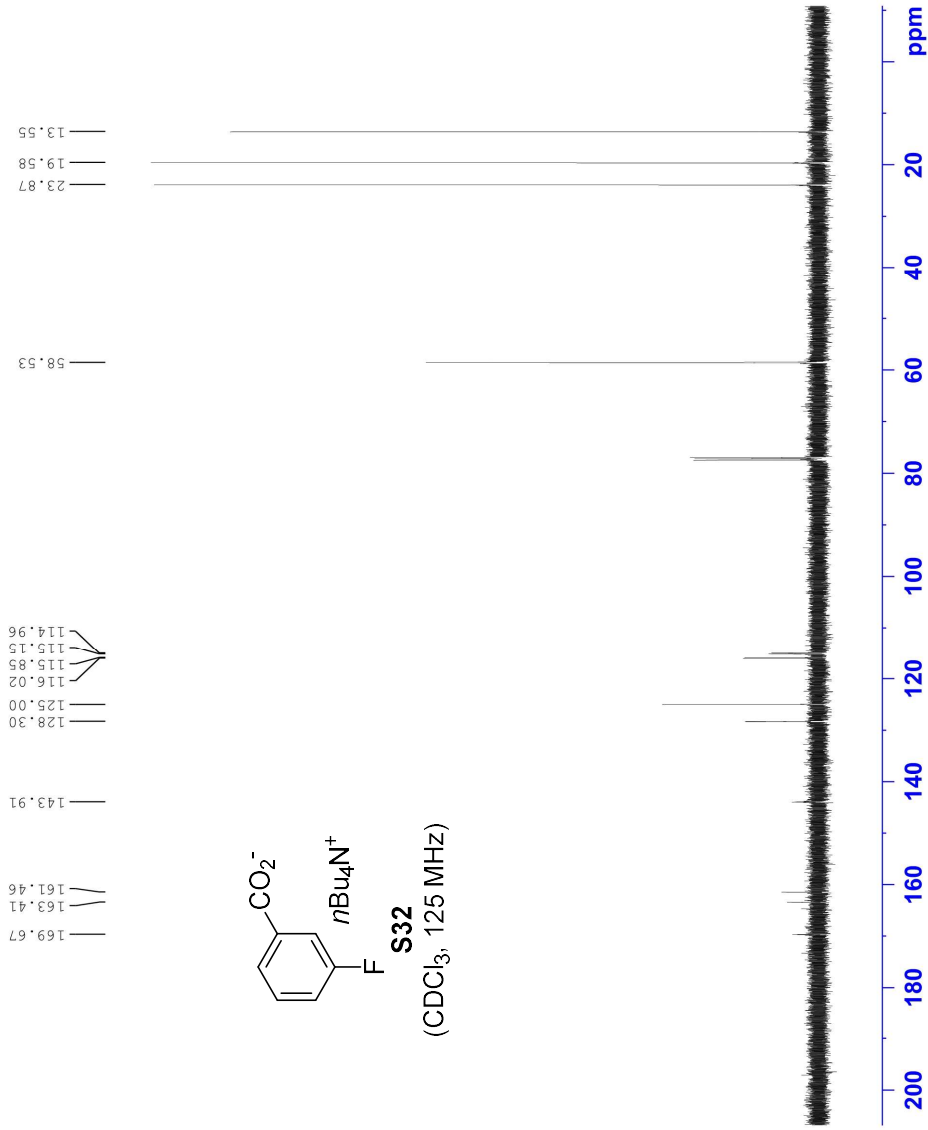
Current Data Parameters
 NAME yh-2-66
 EXPNO 2
 PROCNO 2

F2 - Acquisition Parameters
 Date_ 20180827
 Time_ 13.47
 INSTRUM spect
 PROBHD 5 mm PABBO BB/
 PULPROG zgpg
 TD 187496
 SOLVENT CDC13
 NS 43
 DS 0
 SWH 31250.000 Hz
 FIDRES 0.166670 Hz
 AQC 2.9999361 sec
 RG 2050
 DW 16.000 usec
 DE 6.50 usec
 TE 296.4 K
 D1 3.0000000 sec
 D11 0.0300000 sec
 TDO 1

==== CHANNEL f1 =====
 SFO1 125.7049802 MHz
 NUC1 13C
 P1 10.00 usec
 PLW1 72.83999634 W

==== CHANNEL f2 =====
 SFO2 499.8724993 MHz
 NUC2 1H
 CPDPRG12 waitz16
 PCPD2 80.00 usec
 PLW2 19.0000000 W
 PLWI2 0.29688001 W

F2 - Processing parameters
 SI 1048576
 SF 125.6924115 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.40



```

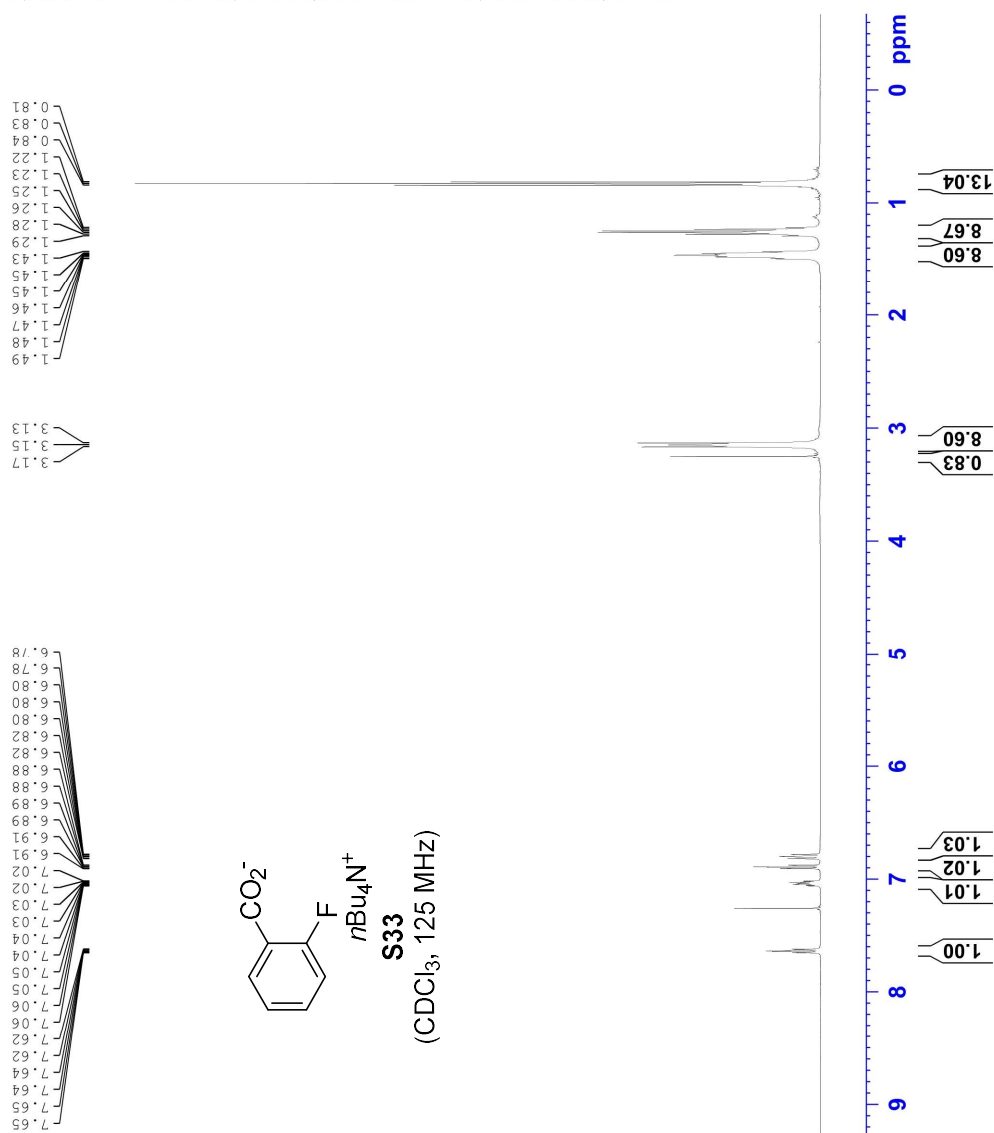
Current Data Parameters
NAME      yh-2-33
EXPNO    1
PROCNO   1

F2 - Acquisition Parameters
Date_    20180814
Time     14.28
INSTRUM  spect
PROBHD   5 mm PATXI 1H/
PULPROG  zg
TD       59998
SOLVENT  CDCl3
NS       1
DS       0
SWH      10000.000 Hz
FIDRES   0.166672 Hz
AQ       2.9999001 sec
RG       20.66
DW       50.000 usec
DE       10.00 usec
TE       294.3 K
DL       4.0000000 sec
TDO      1

===== CHANNEL f1 =====
SFO1    500.1330885 MHz
NUC1     1H
P1       3.30 usec
PLW1    12.19999981 W

F2 - Processing parameters
SI       65536
SF       500.1300150 MHz
WDW      EM
SSB      0
LB       0.30 Hz
GB       0
PC       1.00

```



```

Current Data Parameters
NAME      Yh-2-33
EXPNO    2
PROCNO   2

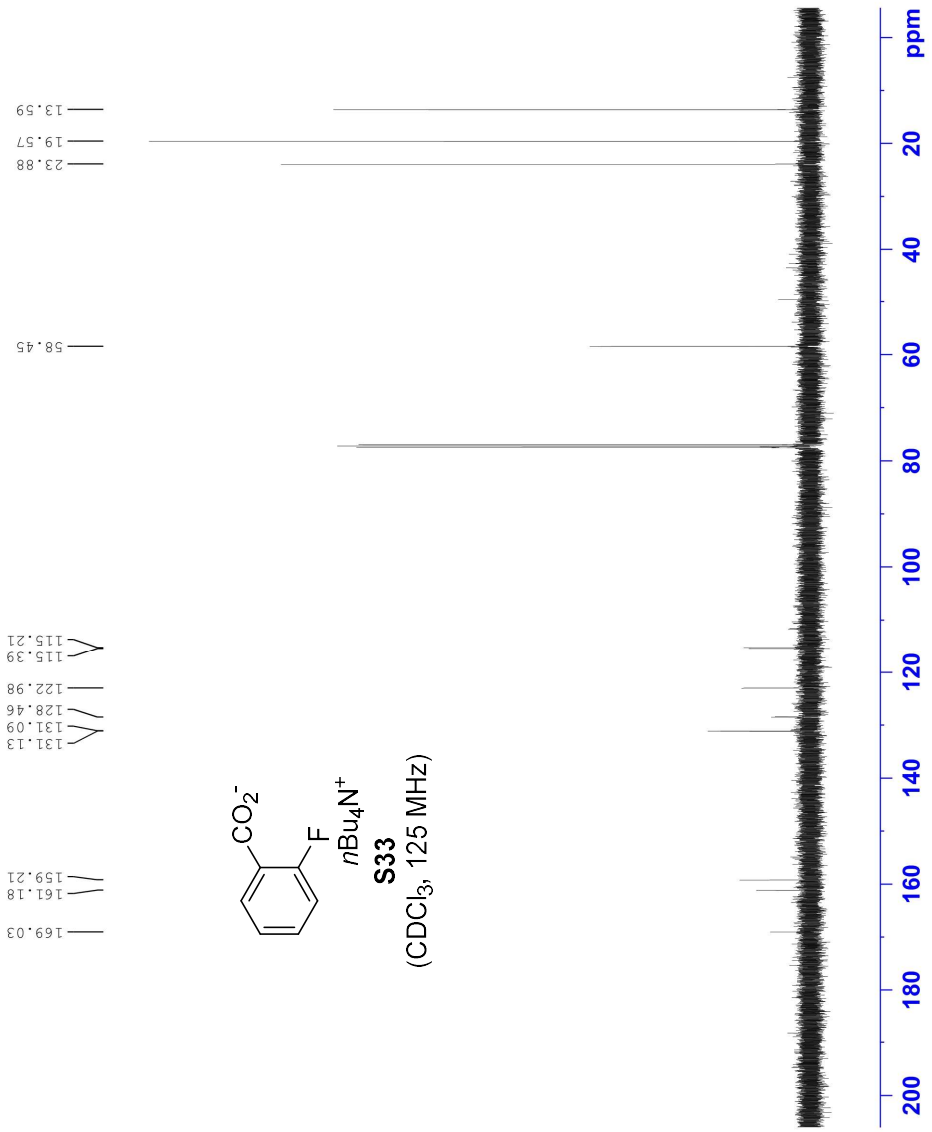
F2 - Acquisition Parameters
Date_     20180814
Time      14.55
INSTRUM   spect
PROBHD    5 mm PABBO BB/
PULPROG   zgpg
TD         187496
SOLVENT   CDC13
NS         17
DS         0
SWH        31250.000 Hz
FIDRES     0.166670 Hz
AQ         2.9999361 sec
RG         2050
DW         16.000 usec
DE         6.50 usec
TE         296.0 K
D1         4.0000000 sec
D11        0.03000000 sec
TDO        1

===== CHANNEL f1 =====
SFO1      125.7049802 MHz
NUC1      13C
P1         3.33 usec
PLW1      72.83999634 W

===== CHANNEL f2 =====
SFO2      499.8724993 MHz
NUC2      1H
CPDPRG12  waitz16
PCPD2     80.00 usec
PLW2      19.0000000 W
PLW12     0.29688001 W

F2 - Processing parameters
SI         1048576
SF         125.6924115 MHz
WDW        EM
SSB        0
LB         0.30 Hz
GB         0
PC         1.40

```

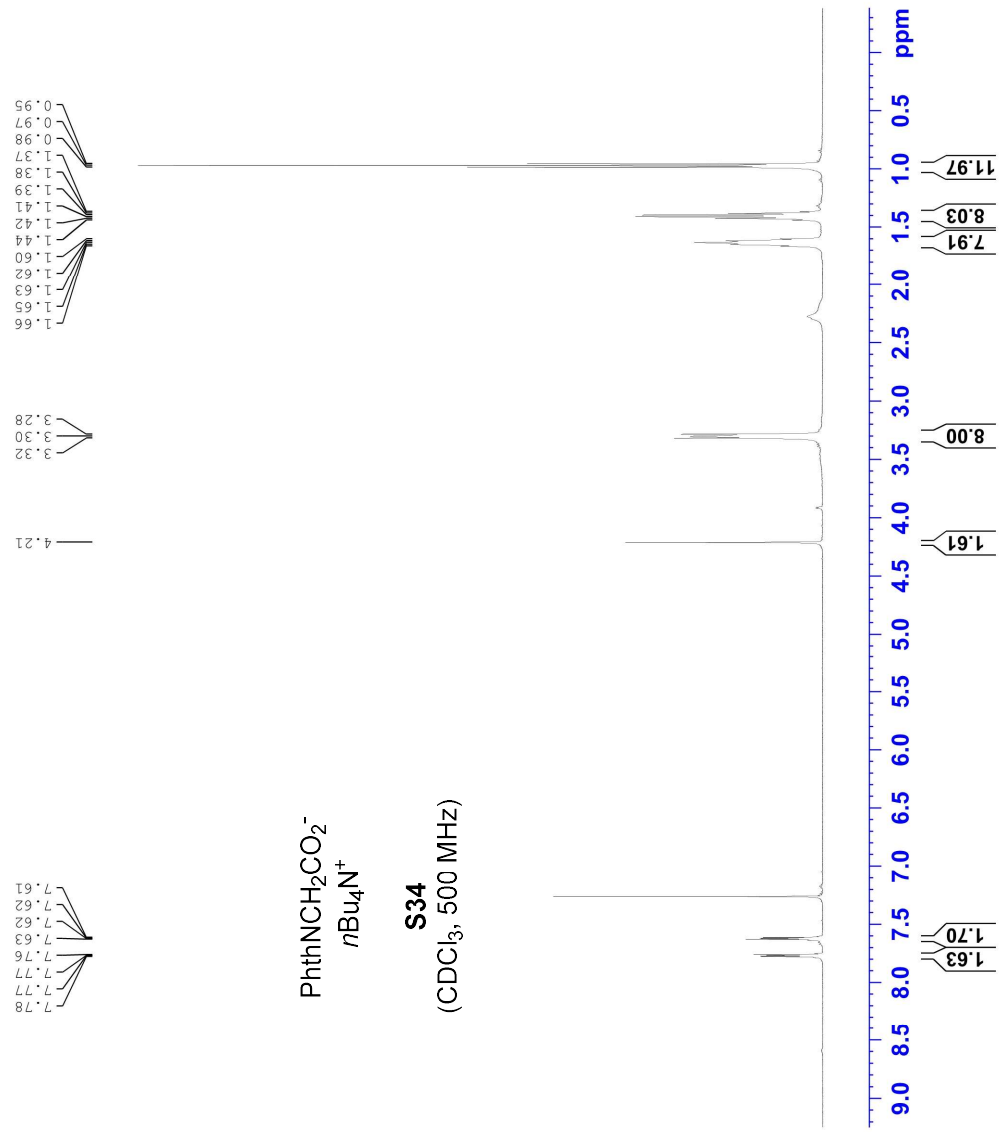


Current Data Parameters
 NAME glycinate
 EXPNO 1
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20190715
 Time_ 11.52
 INSTRUM spect
 PROBHD 5 mm PABBO BB/
 PULPROG zg
 TD 5998
 SOLVENT CDCl3
 NS 4
 DS 0
 SWH 1000.000 Hz
 FIDRES 0.166672 Hz
 AQC 2.9999001 sec
 RG 159.35
 DW 50.000 usec
 DE 6.50 usec
 TE 295.7 K
 DL 4.0000000 sec
 TDO 1

===== CHANNEL f1 =====
 SFO1 499.8730869 MHz
 NUC1 1H
 PL 3.58 usec
 PLW1 18.25000000 W

F2 - Processing parameters
 SI 65536
 SF 499.8700120 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00



```

Current Data Parameters
NAME      glycinate
EXPNO    2
PROCNO   2

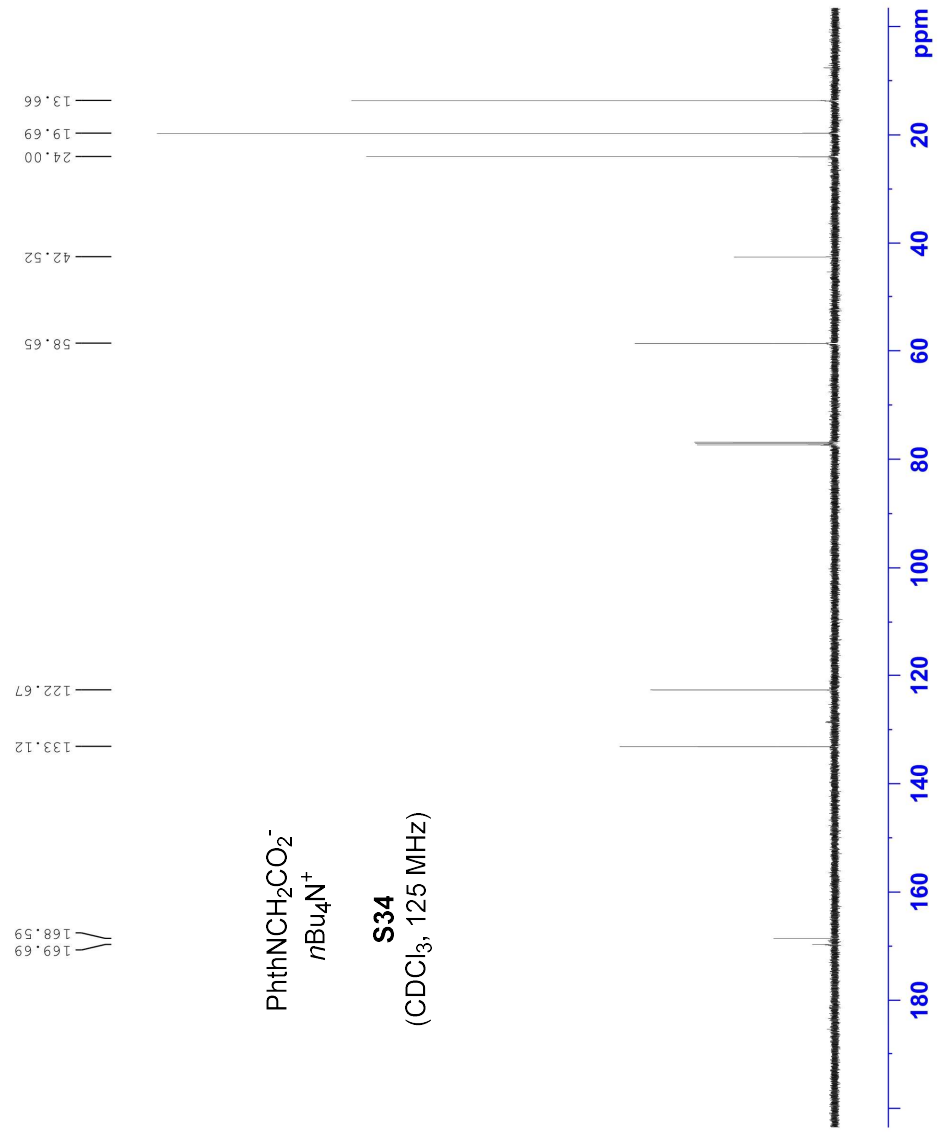
F2 - Acquisition Parameters
Date_     20190715
Time      12.27
INSTRUM   spect
PROBHD    5 mm PABBO BB/
PULPROG   zgpg
TD         187496
SOLVENT   CDC13
NS         102
DS         0
SWH        31250.000 Hz
FIDRES     0.166670 Hz
AQ         2.9999361 sec
RG         2050
DW         16.000 usec
DE         6.50 usec
TE         297.5 K
D1         3.00000000 sec
D11        0.03000000 sec
TD0        1

===== CHANNEL f1 =====
SFO1      125.7049802 MHz
NUC1      13C
P1         10.00 usec
PLW1      72.83999634 W

===== CHANNEL f2 =====
SFO2      499.8724993 MHz
NUC2      1H
CPDPRG12  waitz16
PCPD2     80.00 usec
PLW2      19.00000000 W
PLW12     0.29688001 W

F2 - Processing parameters
SI         1048576
SF         125.6924115 MHz
WDW        EM
SSB        0
LB         0.30 Hz
GB         0
PC         1.40

```

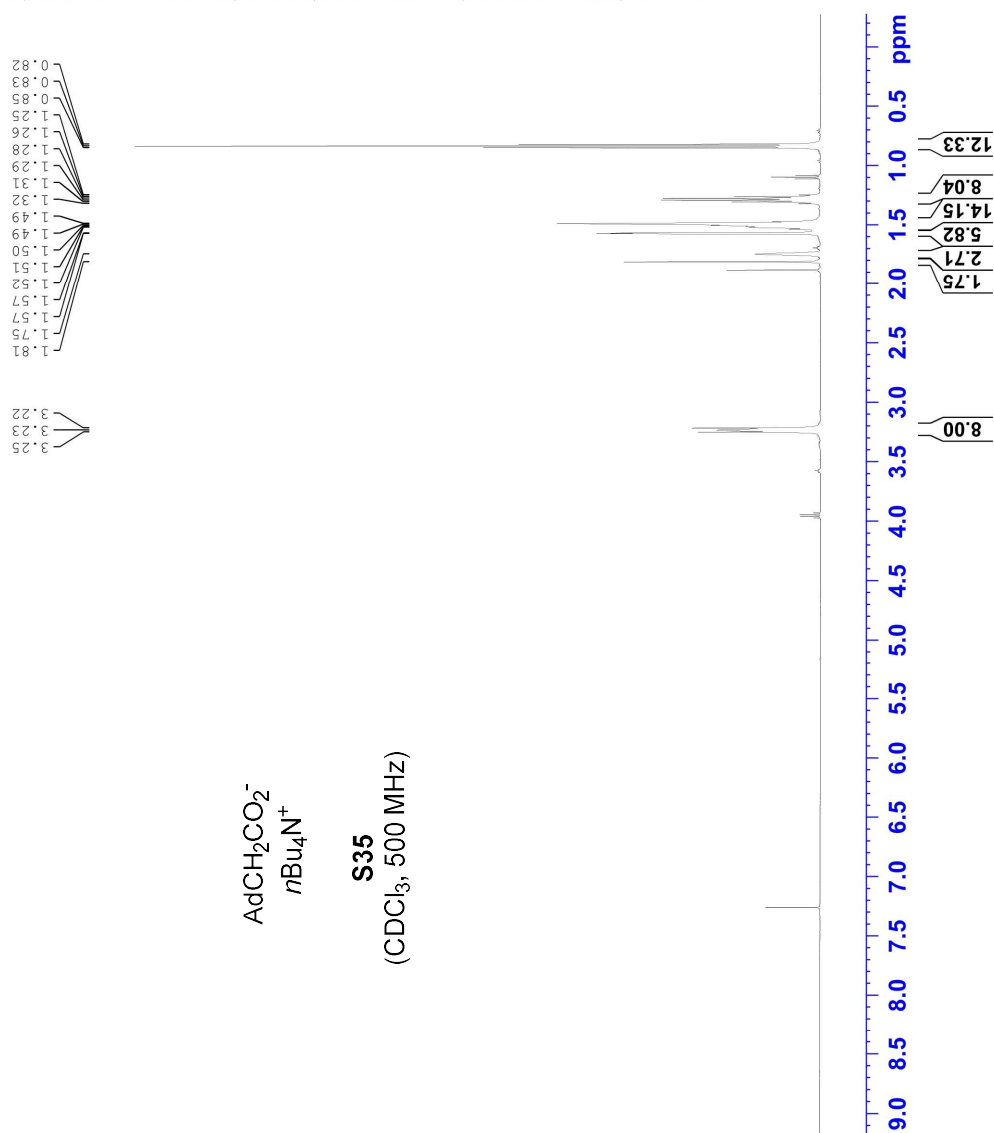


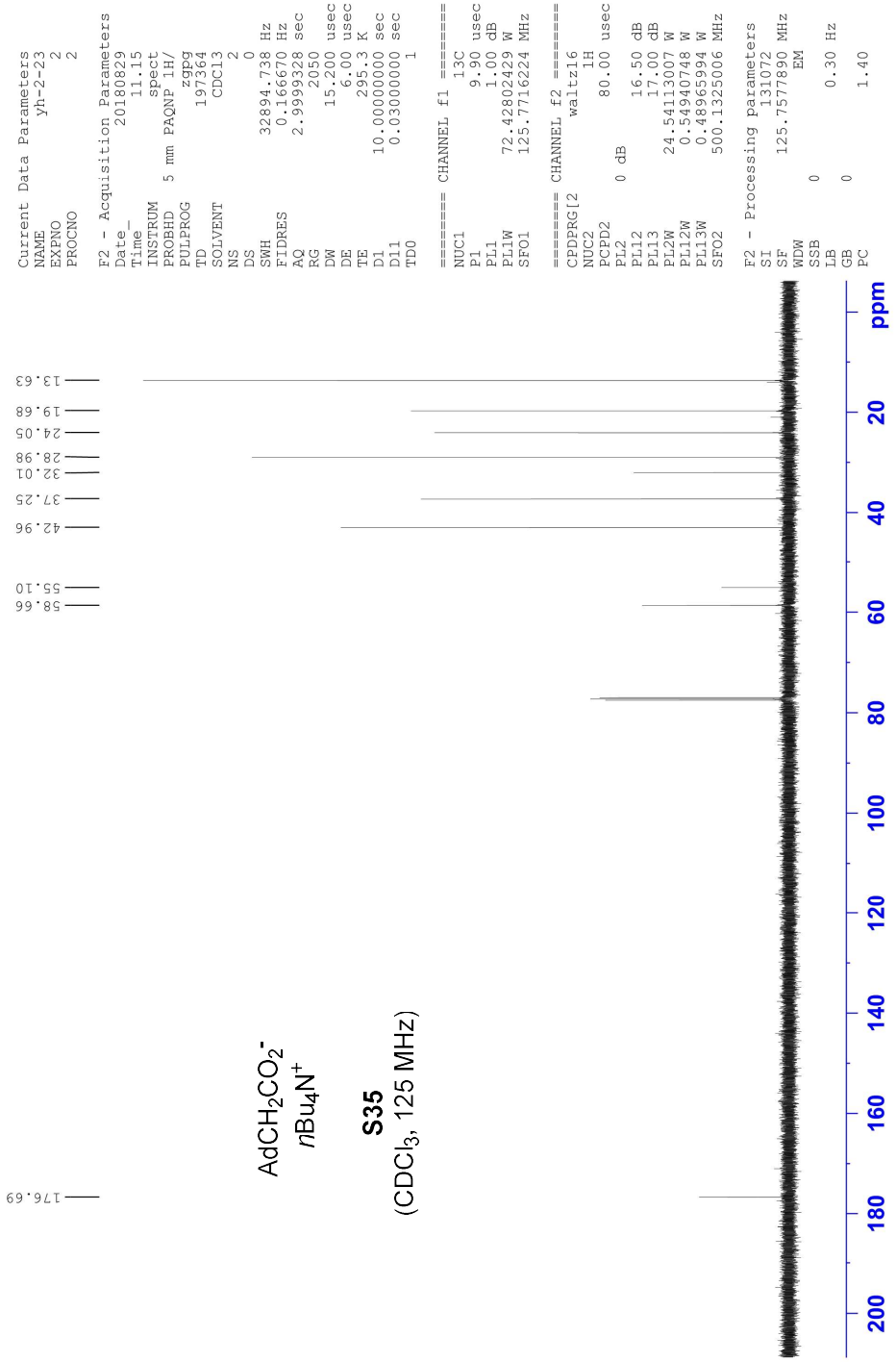
Current Data Parameters
 NAME yh-2-23
 EXPNO 1
 PROCNO 1

F2 - Acquisition Parameters
 Date 20180829
 Time 11.09
 INSTRUM spect
 PROBHD 5 mm PAQNP 1H/
 PULPROG zg
 TD 4498
 SOLVENT CDCl3
 NS 1
 DS 0
 SWH 7500.000 Hz
 FIDRES 0.166674 Hz
 AQ 2.9998667 sec
 RG 16
 DW 66.667 usec
 DE 71.43 usec
 TE 295.0 K
 DL 3.0000000 sec
 TDO 1

===== CHANNEL f1 =====
 NUC1 1H
 P1 14.50 usec
 PL1 0 dB
 PL1W 24.54113007 W
 SF01 500.1330008 MHz

F2 - Processing parameters
 SI 32768
 SF 500.1300168 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00



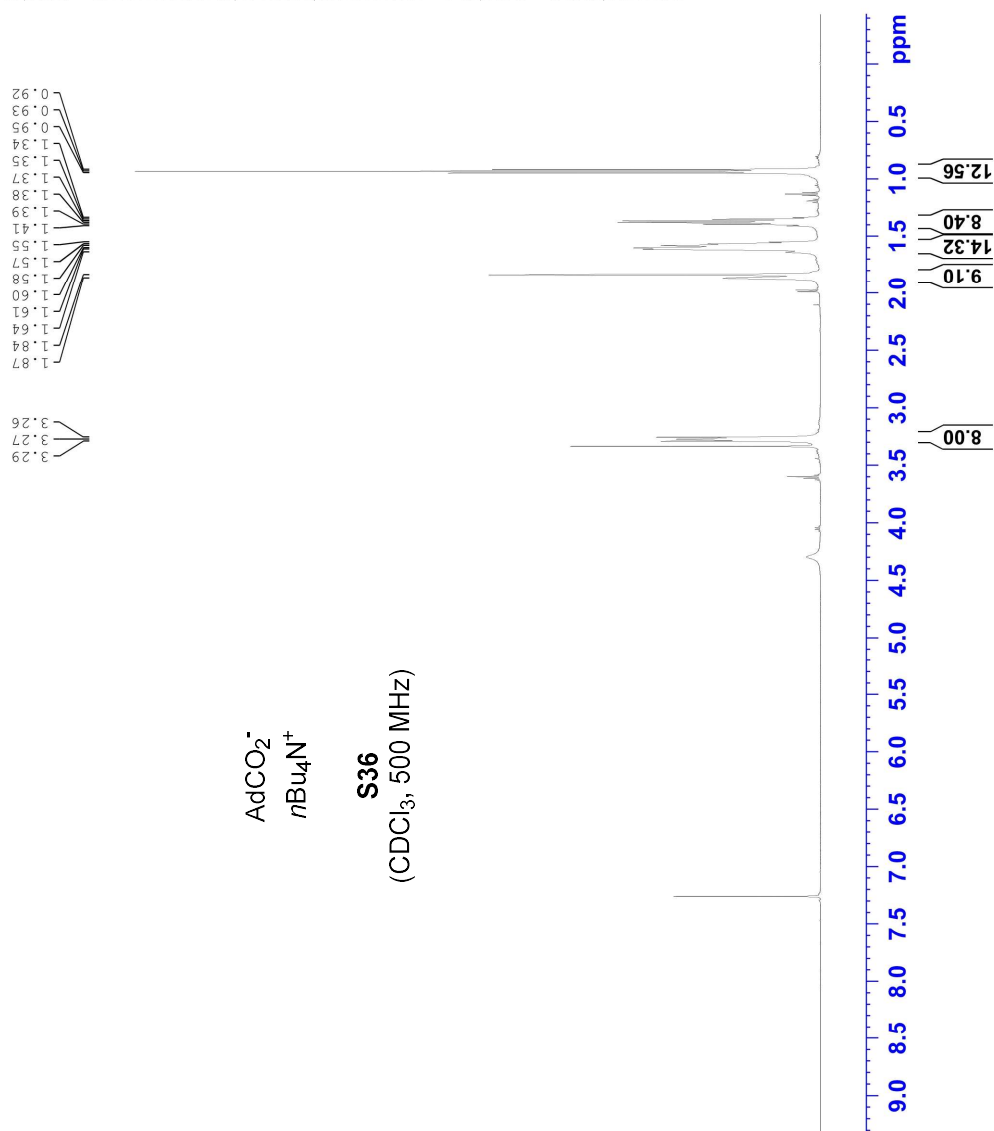


Current Data Parameters
 NAME yh-2-71
 EXPNO 1
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20180814
 Time_ 22.35
 INSTRUM spect
 PROBHD 5 mm PABBO BB/
 PULPROG zg
 TD 5998
 SOLVENT CDCl3
 NS 2
 DS 0
 SWH 10000.000 Hz
 FIDRES 0.166672 Hz
 AQ 2.9999001 sec
 RG 22.37
 DW 50.000 usec
 DE 6.50 usec
 TE 296.0 K
 DL 4.0000000 sec
 TDO 1

===== CHANNEL f1 =====
 SFO1 499.8730869 MHz
 NUC1 1H
 P1 3.58 usec
 PLW1 18.25000000 W

F2 - Processing parameters
 SI 65536
 SF 499.8700132 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00



```

Current Data Parameters
NAME      yh-2-71
EXPNO     2
PROCNO    2

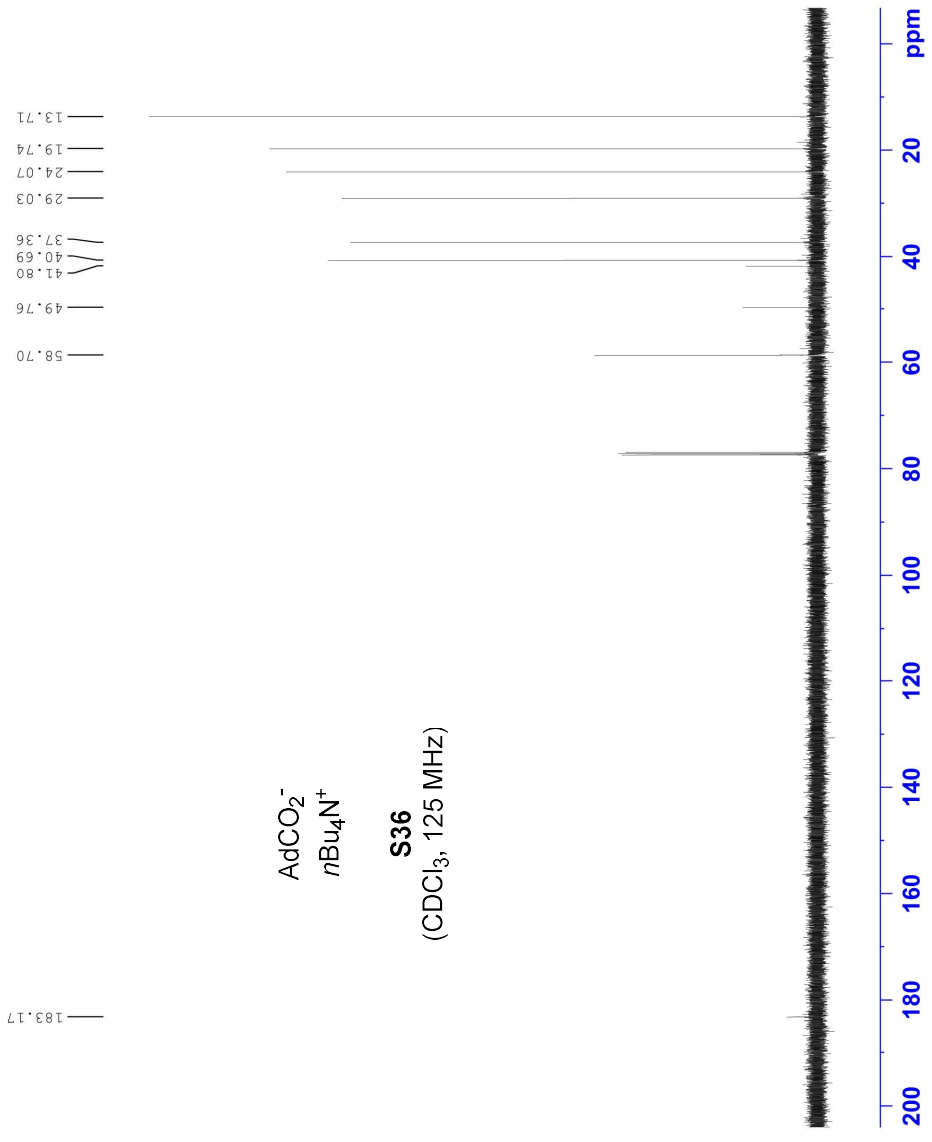
F2 - Acquisition Parameters
Date_     20180814
Time      22.40
INSTRUM   spect
PROBHD    5 mm PABBO BB/
PULPROG   zgpg
TD         187496
SOLVENT   CDC13
NS         7
DS         0
SWH        31250.000 Hz
FIDRES     0.166670 Hz
AQ         2.9999361 sec
RG         2050
DW         16.000 usec
DE         6.50 usec
TE         296.1 K
D1         3.00000000 sec
D11        0.03000000 sec
TD0        1

===== CHANNEL f1 =====
SFO1      125.7049802 MHz
NUC1       13C
P1         10.00 usec
PLW1       72.83999634 W

===== CHANNEL f2 =====
SFO2      499.8724993 MHz
NUC2       1H
CFPRG12   waitz16
PCPD2     80.00 usec
PLW2      19.00000000 W
PLW12     0.29688001 W

F2 - Processing parameters
SI         1048576
SF         125.6924115 MHz
WDW        EM
SSB        0
LB         0.30 Hz
GB         0
PC         1.40

```

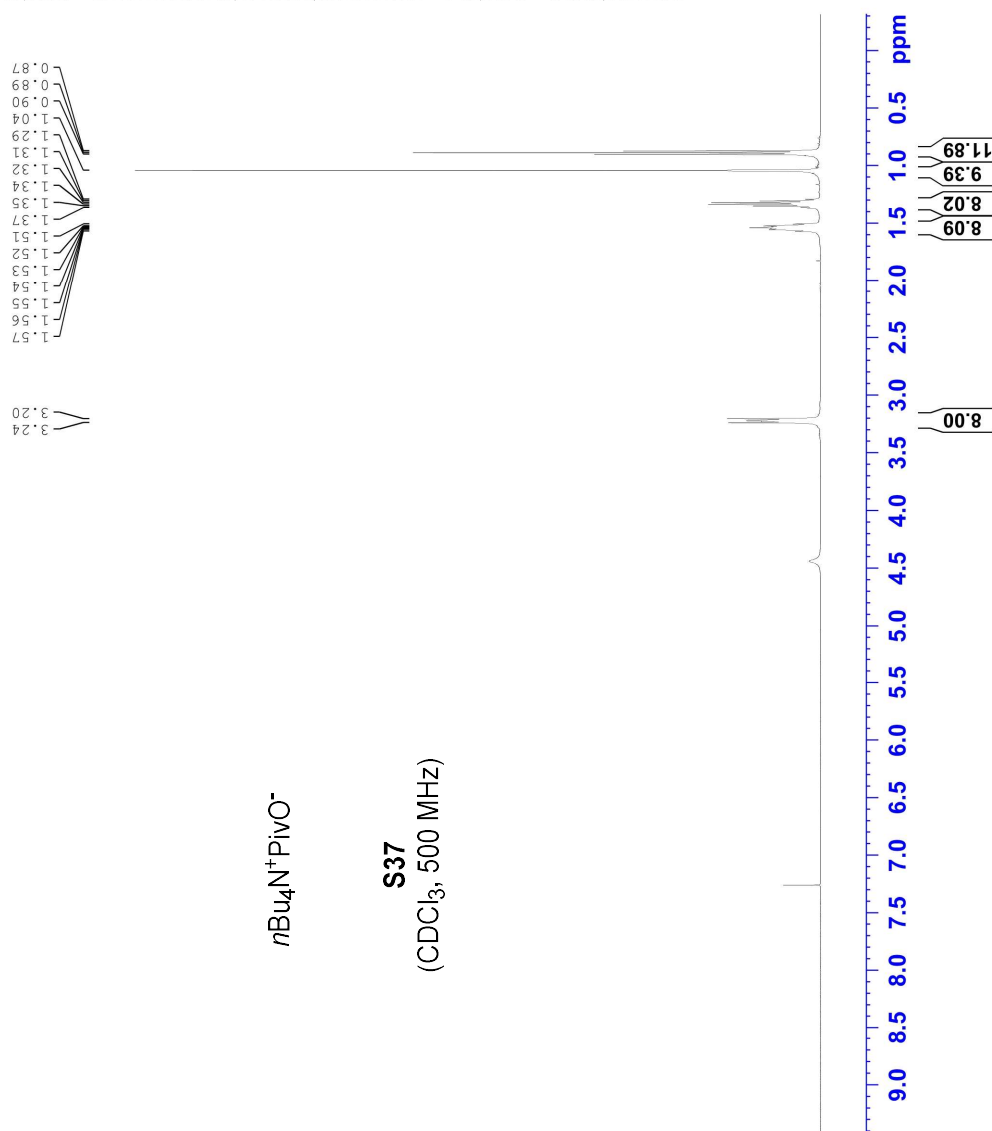


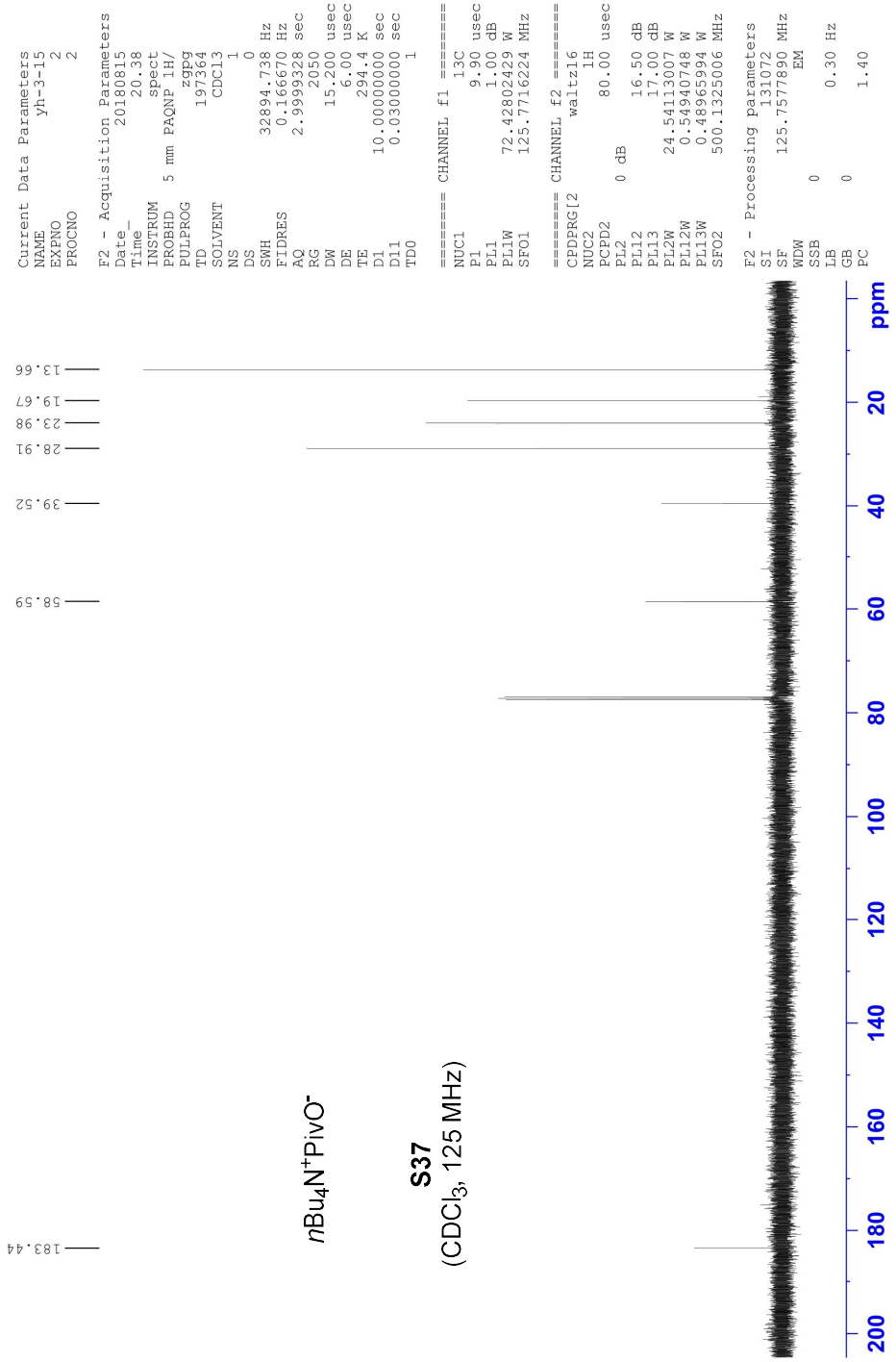
Current Data Parameters
 NAME yh-3-15
 EXPNO 1
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20180815
 Time_ 20.23
 INSTRUM spect
 PROBHD 5 mm PABBO BB/
 PULPROG zg
 TD 5998
 SOLVENT CDC13
 NS 3
 DS 0
 SWH 10000.000 Hz
 FIDRES 0.166672 Hz
 AQ 2.9999001 sec
 RG 19.64
 DW 50.000 usec
 DE 6.50 usec
 TE 296.0 K
 DL 4.00000000 sec
 TDO 1

===== CHANNEL f1 =====
 SFO1 499.8730869 MHz
 NUC1 1H
 PL 3.58 usec
 PLW1 18.25000000 W

F2 - Processing parameters
 SI 65536
 SF 499.8700134 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00





```

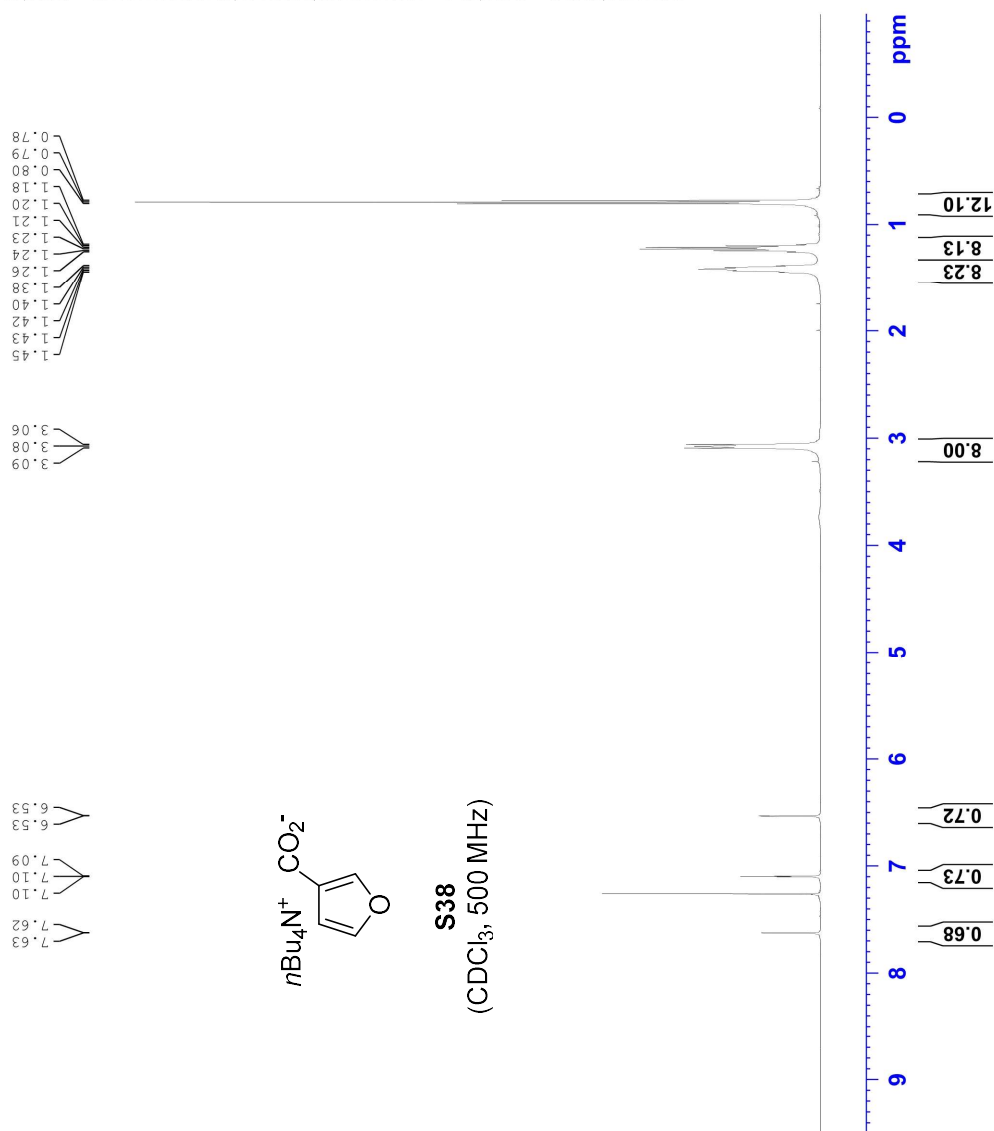
Current Data Parameters
NAME      yn-2-139
EXPNO    1
PROCNO   1

F2 - Acquisition Parameters
Date_    20180814
Time     22.52
INSTRUM  spect
PROBHD   5 mm PABBO BB/
PULPROG  zg
TD        59998
SOLVENT  CDCl3
NS        2
DS        0
SWH       10000.000 Hz
FIDRES    0.166672 Hz
AQ         2.9999001 sec
RG         5.87
DW         50.000 usec
DE         6.50 usec
TE         296.1 K
DL         3.0000000 sec
TDO        1

===== CHANNEL f1 =====
SFO1     499.8730869 MHz
NUC1      1H
P1        10.75 usec
PLW1     18.25000000 W

F2 - Processing parameters
SI        65536
SF        499.8700140 MHz
WDW       EM
SSB       0
LB        0.30 Hz
GB        0
PC        1.00

```



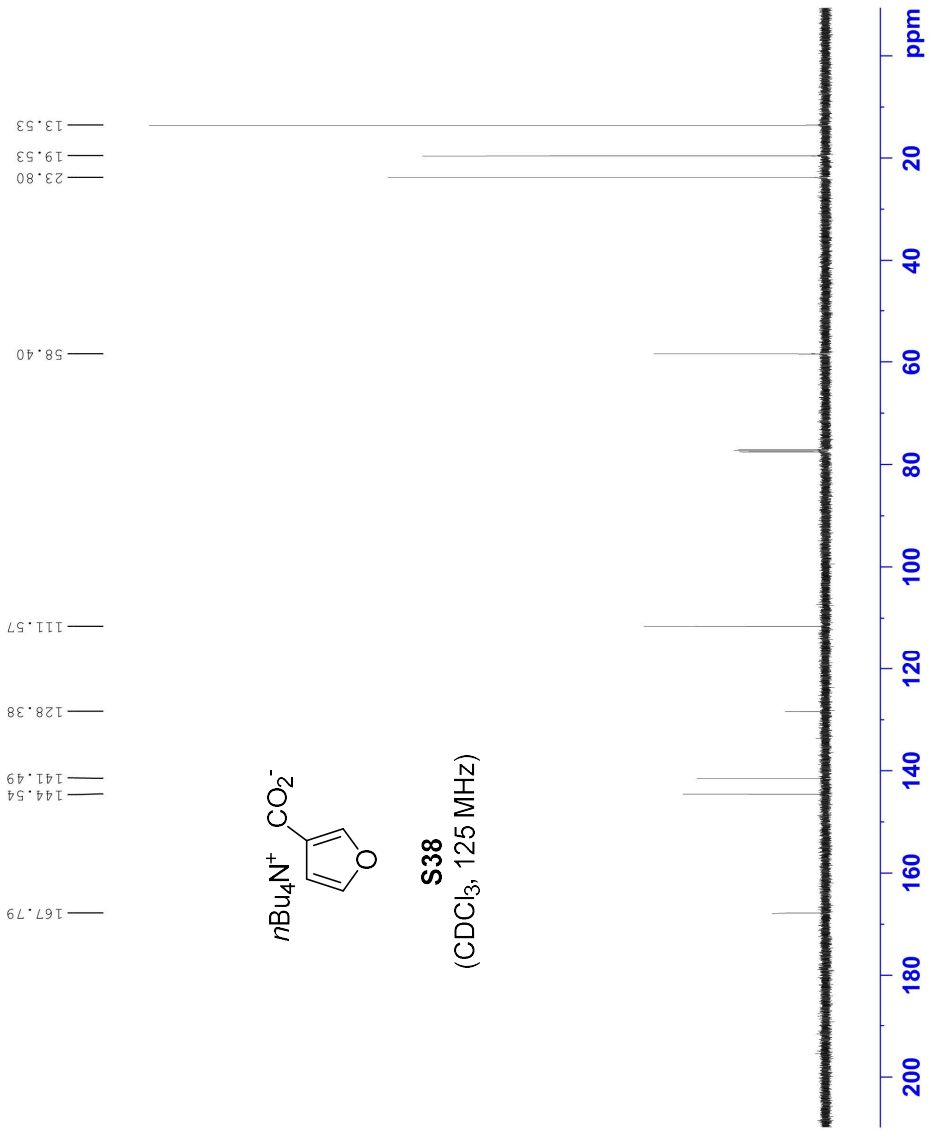
Current Data Parameters
 NAME yn-2-139
 EXPNO 2
 PROCNO 2

F2 - Acquisition Parameters
 Date_ 20180814
 Time_ 22.55
 INSTRUM spect
 PROBHD 5 mm PABBO BB/
 PULPROG zgpg
 TD 187496
 SOLVENT CDCl3
 NS 4
 DS 0
 SWH 31250.000 Hz
 FIDRES 0.166670 Hz
 AQ 2.9999361 sec
 RG 2050
 DW 16.000 usec
 DE 6.50 usec
 TE 296.1 K
 D1 3.0000000 sec
 D11 0.0300000 sec
 TDO 1

==== CHANNEL f1 =====
 SFO1 125.7049802 MHz
 NUC1 13C
 P1 10.00 usec
 PLW1 72.83999634 W

==== CHANNEL f2 =====
 SFO2 499.8724993 MHz
 NUC2 1H
 CPDPRG12 waitz16
 PCPD2 80.00 usec
 PLW2 19.0000000 W
 PLW12 0.29688001 W

F2 - Processing parameters
 SI 1048576
 SF 125.6924115 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.40

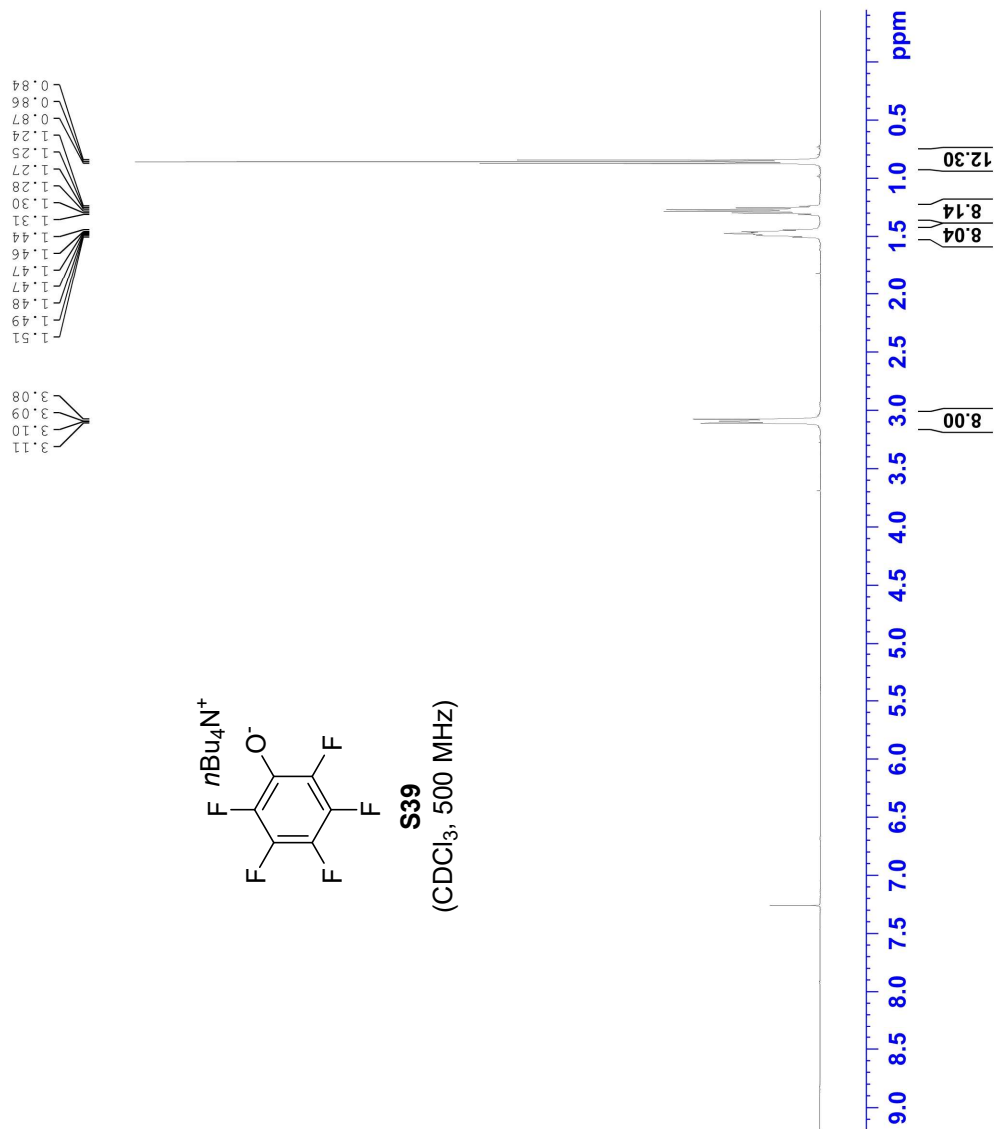


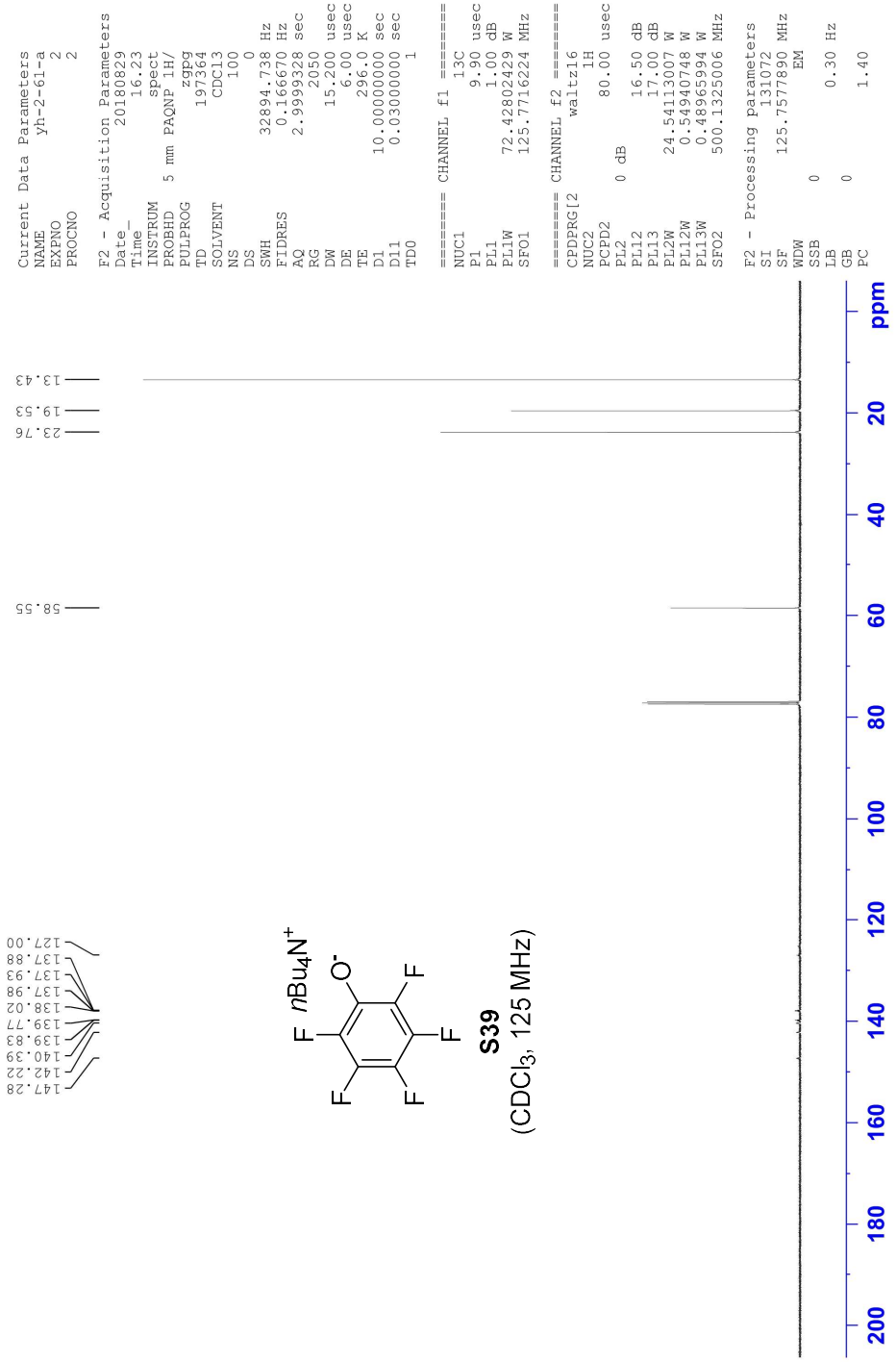
Current Data Parameters
 NAME Yh-2-61-a
 EXPNO 1
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20180829
 Time_ 15.56
 INSTRUM spect
 PROBHD 5 mm P4QNP 1H/
 PULPROG zg
 TD 4498
 SOLVENT CDCl3
 NS 2
 DS 0
 SWH 7500.000 Hz
 FIDRES 0.166674 Hz
 AQ 2.9998667 sec
 RG 22.6
 DW 66.667 usec
 DE 71.43 usec
 TE 295.2 K
 DL 3.0000000 sec
 TDO 1

===== CHANNEL f1 =====
 NUC1 1H
 P1 14.50 usec
 PL1 0 dB
 PL1W 24.54113007 W
 SF01 500.1330008 MHz

F2 - Processing parameters
 SI 32768
 SF 500.1300156 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00



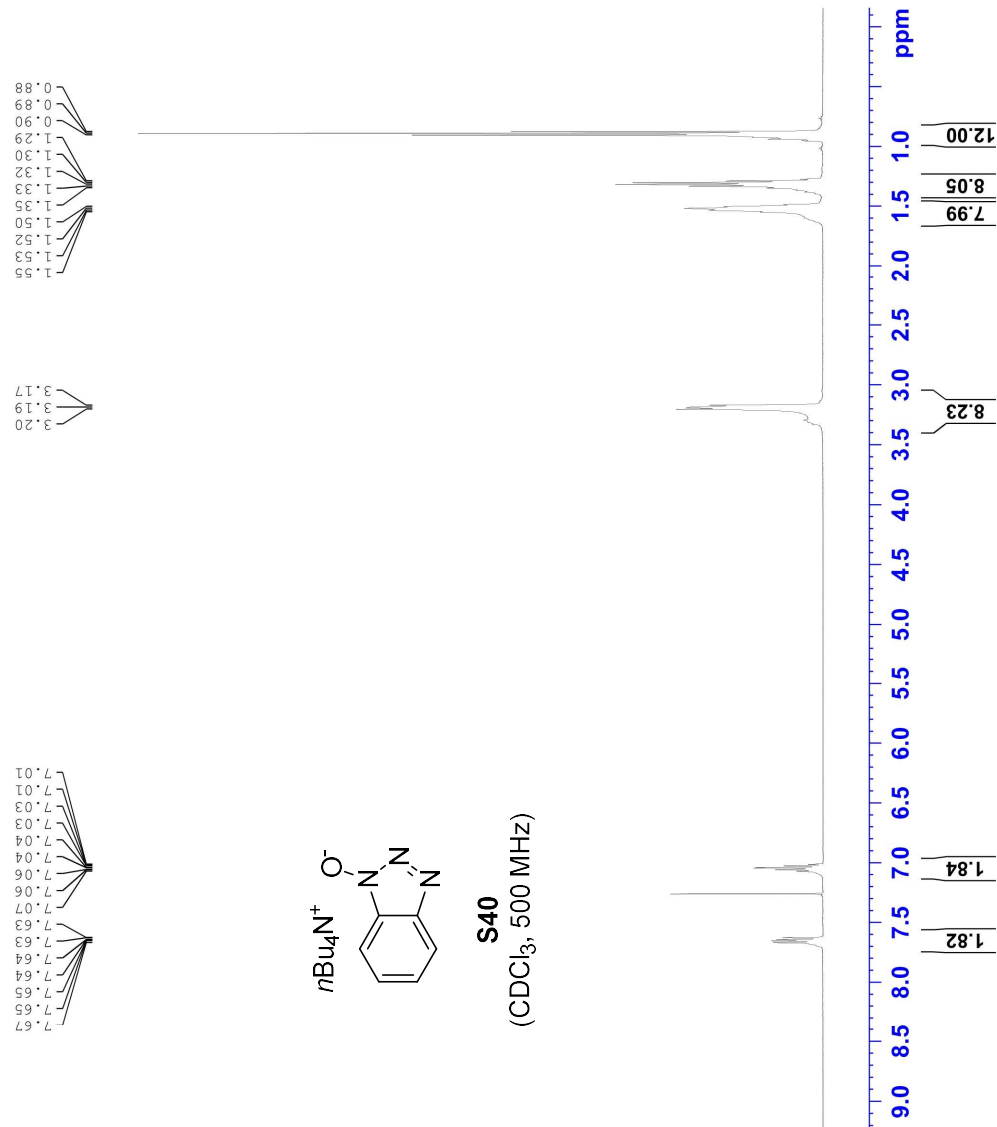


Current Data Parameters
 NAME HOBt salt
 EXPNO 1
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20190927
 Time_ 16.44
 INSTRUM spect
 PROBHD 5 mm PATXI 1H/
 PULPROG zg
 TD 5998
 SOLVENT CDCl3
 NS 4
 DS 0
 SWH 10000.000 Hz
 FIDRES 0.166672 Hz
 AQ 2.9999001 sec
 RG 110.37
 DW 50.000 usec
 DE 10.00 usec
 TE 297.0 K
 DL 4.0000000 sec
 TDO 1

===== CHANNEL f1 =====
 SFO1 500.1330885 MHz
 NUC1 1H
 P1 3.30 usec
 PLW1 12.19999981 W

F2 - Processing parameters
 SI 65536
 SF 500.1300141 MHz
 EM
 WDW 0
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00



```

Current Data Parameters
NAME      HOBt salt
EXPNO    2
PROCNO   2

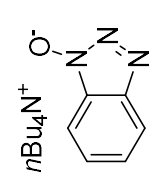
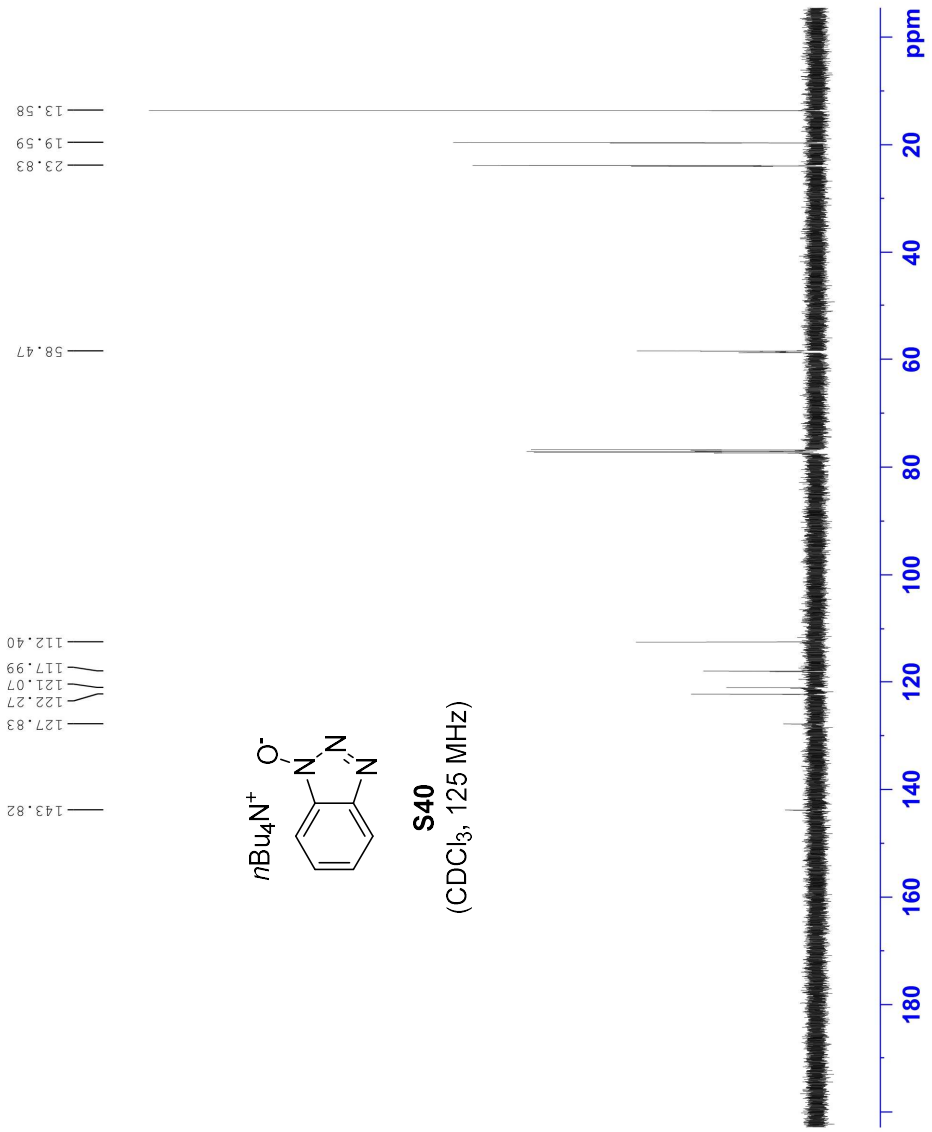
F2 - Acquisition Parameters
Date_    20190930
Time     16.46
INSTRUM  spect
PROBHD   5 mm PABBO BB/
PULPROG  zgpg30
TD        187496
SOLVENT  CDCl3
NS        151
DS        0
SWH       31250.000 Hz
FIDRES    0.166670 Hz
AQ        2.9999361 sec
RG        2050
DW        16.000 usec
DE        6.50 usec
TE        298.0 K
D1        3.0000000 sec
D11       0.03000000 sec
TDO       1

===== CHANNEL f1 =====
SFO1     125.7049802 MHz
NUC1     13C
P1       10.00 usec
PLW1     72.83999634 W

===== CHANNEL f2 =====
SFO2     499.8724993 MHz
NUC2     1H
CFPRG12  waitz16
PCPD2    80.00 usec
PLW2     19.0000000 W
PLW12    0.29688001 W

F2 - Processing parameters
SI       1048576
SF       125.6924115 MHz
WDW      EM
SSB      0
LB       0.30 Hz
GB       0
PC       1.40

```



S40
 (CDCl₃, 125 MHz)

```

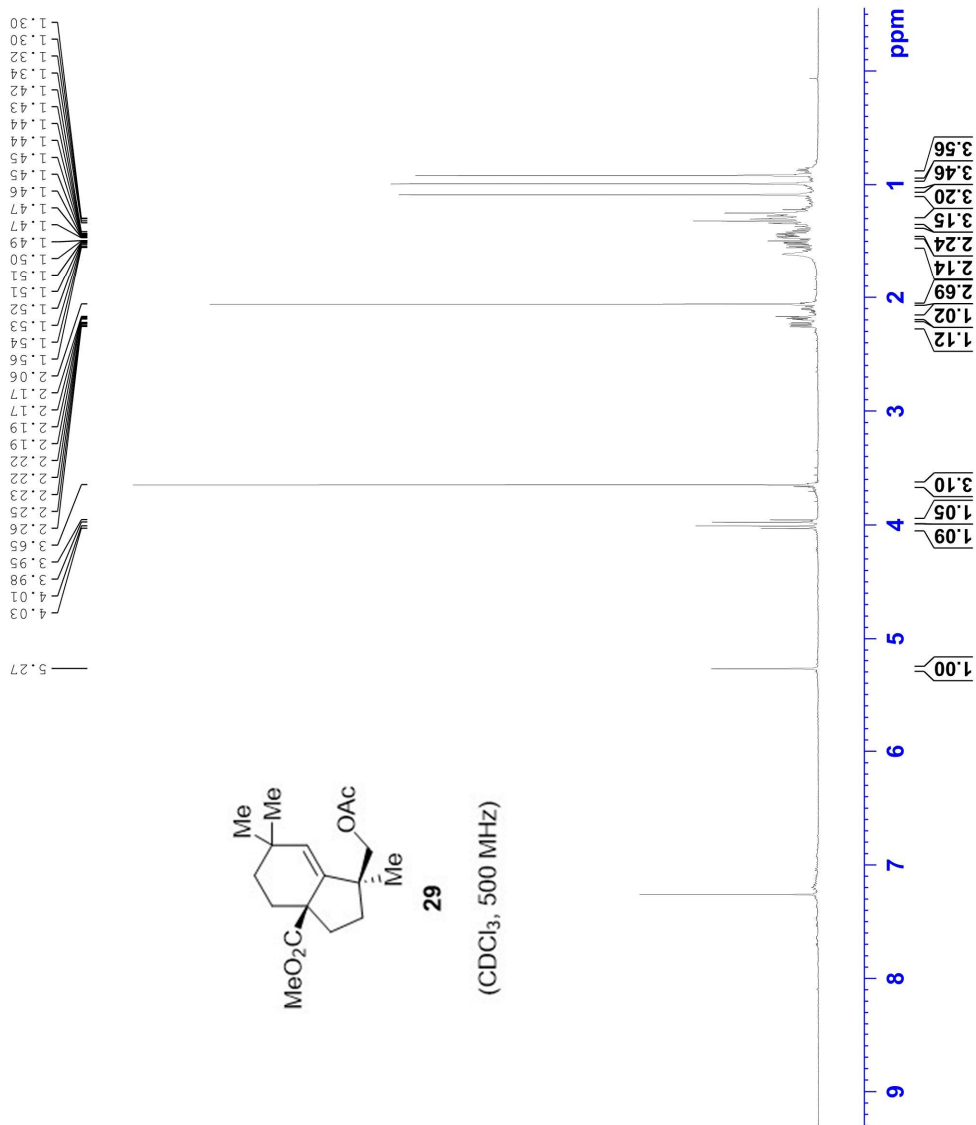
Current Data Parameters
NAME acetate product
EXPNO 1
PROCNO 1

F2 - Acquisition Parameters
Date_ 20180820
Time_ 15.39
INSTRUM spect
PROBHD 5 mm PABBO BB/
PULPROG zg
TD 5998
SOLVENT CDCl3
NS 2
DS 0
SWH 10000.000 Hz
FIDRES 0.166672 Hz
AQ 2.9999001 sec
RG 77.07
DM 50.000 usec
DE 6.50 usec
TE 296.0 K
DL 4.00000000 sec
TD0 1

===== CHANNEL f1 =====
SFO1 499.8730869 MHz
NUC1 1H
P1 3.58 usec
PLW1 18.25000000 W

F2 - Processing parameters
SI 65536
SF 499.8700122 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00

```



```

Current Data Parameters
NAME acetate Product
EXPNO 2
PROCNO 2

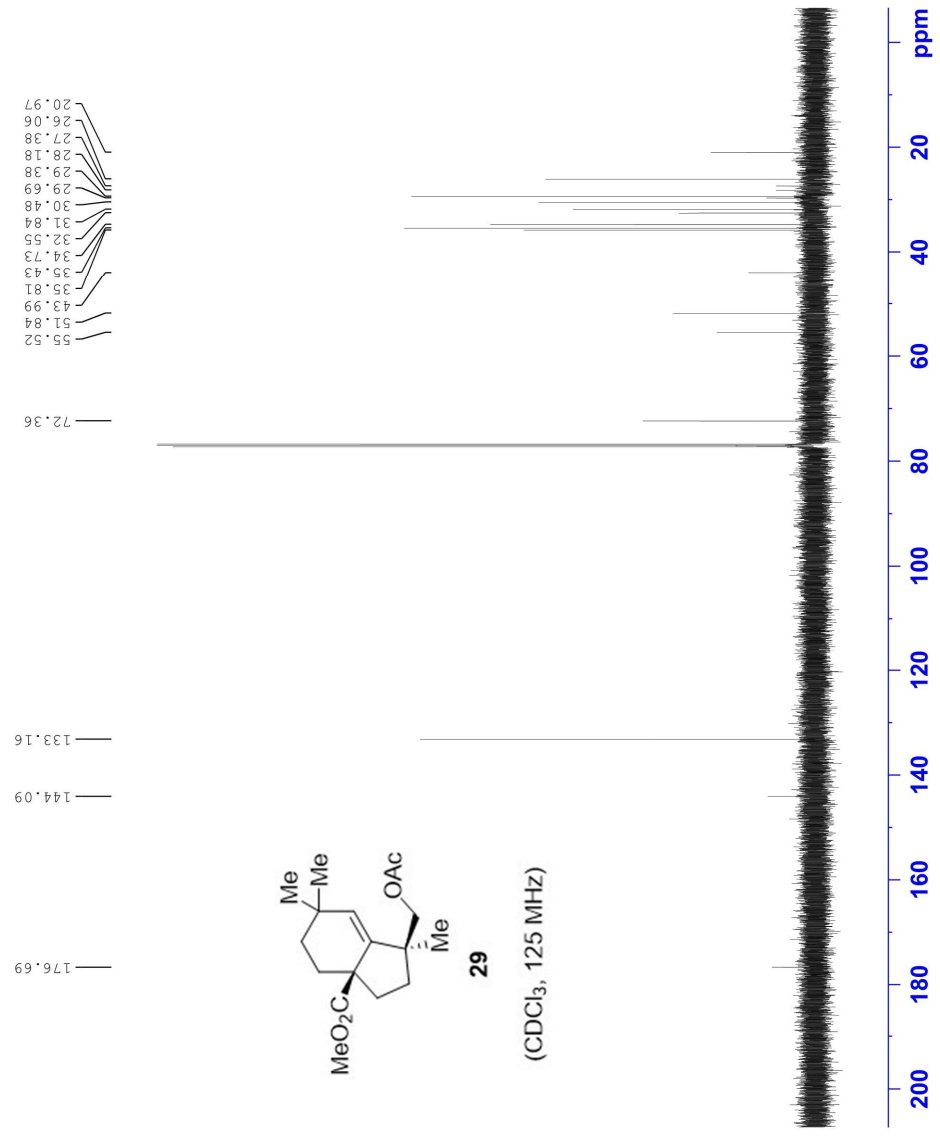
F2 - Acquisition Parameters
Date_ 20180820
Time_ 15.59
INSTRUM spect
PROBHD 5 mm PABBO BB/
PULPROG zgpg
TD 187496
SOLVENT CDCl3
NS 159
DS 0
SWH 31250.000 Hz
FIDRES 0.166670 Hz
AQ 2.9999361 sec
RG 2050
DM 16.000 usec
DE 6.50 usec
TE 297.3 K
D1 3.0000000 sec
D11 0.0300000 sec
TDO 1

===== CHANNEL f1 =====
SFO1 125.7049802 MHz
NUC1 13C
P1 10.00 usec
PLW1 72.83999634 W

===== CHANNEL f2 =====
SFO2 499.8724993 MHz
NUC2 1H
CFPRG12 waitz16
PCPD2 80.00 usec
PLW2 19.0000000 W
PLW12 0.29688001 W

F2 - Processing parameters
SI 1048576
SF 125.6924115 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.40

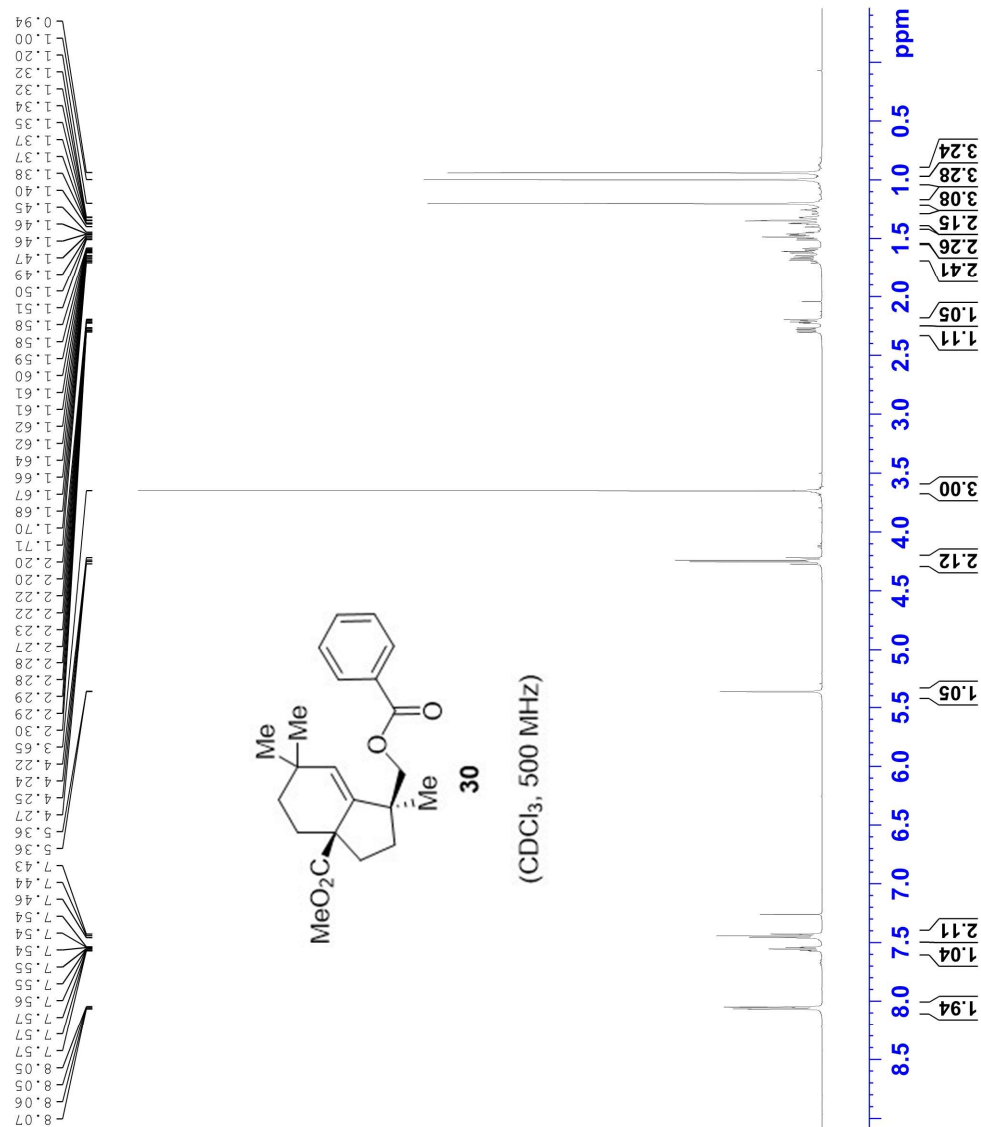
```



Current Data Parameters
 NAME Yh-7-35-b
 EXPNO 1
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20191005
 Time_ 15.10 h
 INSTRUM spect
 PROBHD Z113652_0187 (zg)
 PULPROG 5998
 SOLVENT CDCl3
 NS 4
 DS 0
 SWH 10000.000 Hz
 FIDRES 0.333344 Hz
 AQ 2.9999001 sec
 RG 37.92
 DW 50.000 usec
 DE 6.50 usec
 TE 296.0 K
 TL 3.0000000 sec
 TD0 1
 SF01 499.8730869 MHz
 NUC1 1H
 P1 10.75 usec
 PLW1 18.2500000 W

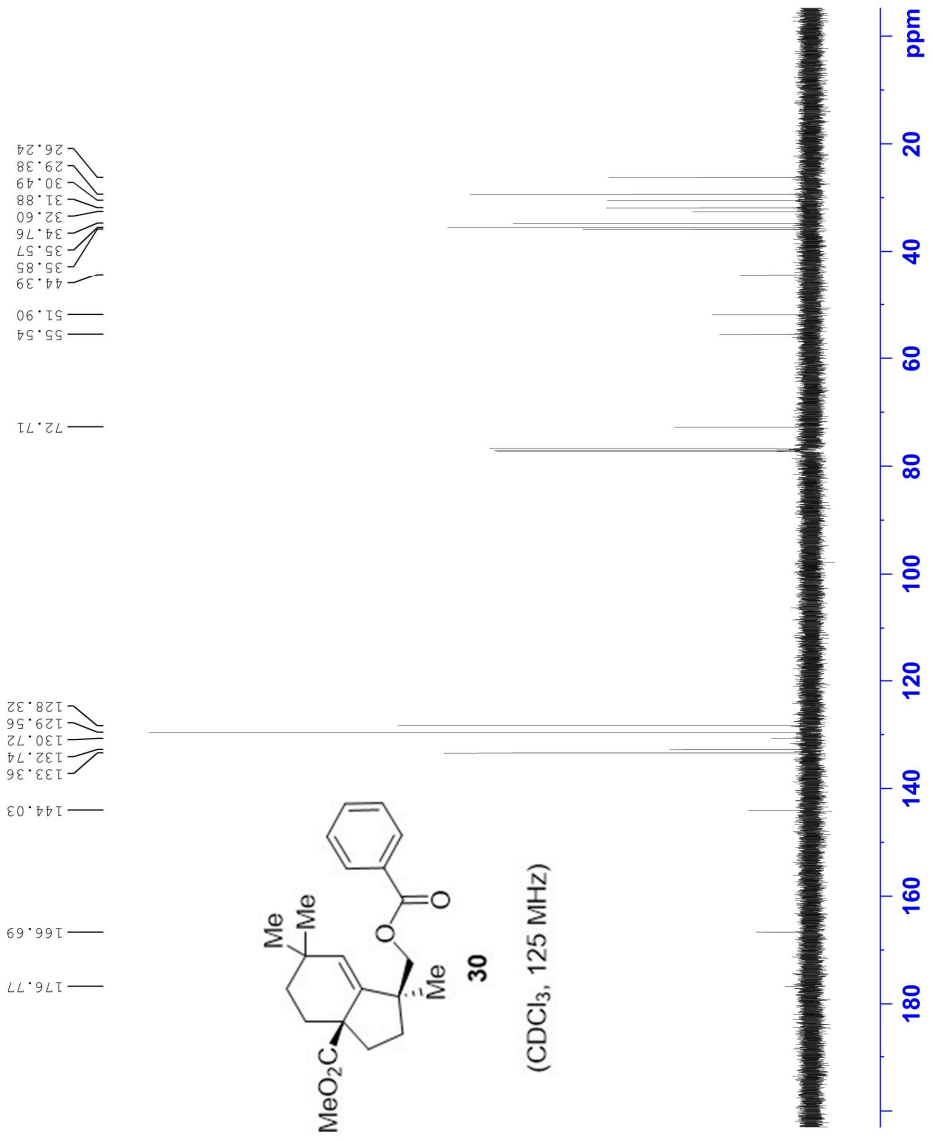
F2 - Processing parameters
 SI 65536
 SF 499.8700125 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00



Current Data Parameters
 NAME Yh-7-35-B
 EXPNO 2
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20191005
 Time_ 15.28 h
 INSTRUM spect
 PROBHD Z113652_0187 (zdc
 PULPROG zgpg30
 TD 187496
 SOLVENT CDCl3
 NS 56
 DS 0
 SWH 31250.000 Hz
 FIDRES 0.333340 Hz
 AQ 2.9999361 sec
 RG 2050
 DW 16.000 usec
 DE 6.50 usec
 TE 296.7 K
 D1 3.0000000 sec
 D11 0.03000000 sec
 TD0 1
 SFO1 125.7049802 MHz
 NUC1 13C
 P1 10.00 usec
 PLW1 72.83999634 W
 SFO2 499.8724993 MHz
 NUC2 1H
 CPDPRG2 waltz16
 PCPD2 80.00 usec
 PLW2 19.00000000 W
 PLWI2 0.29688001 W

F2 - Processing parameters
 SI 1048576
 SF 125.6924115 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.40

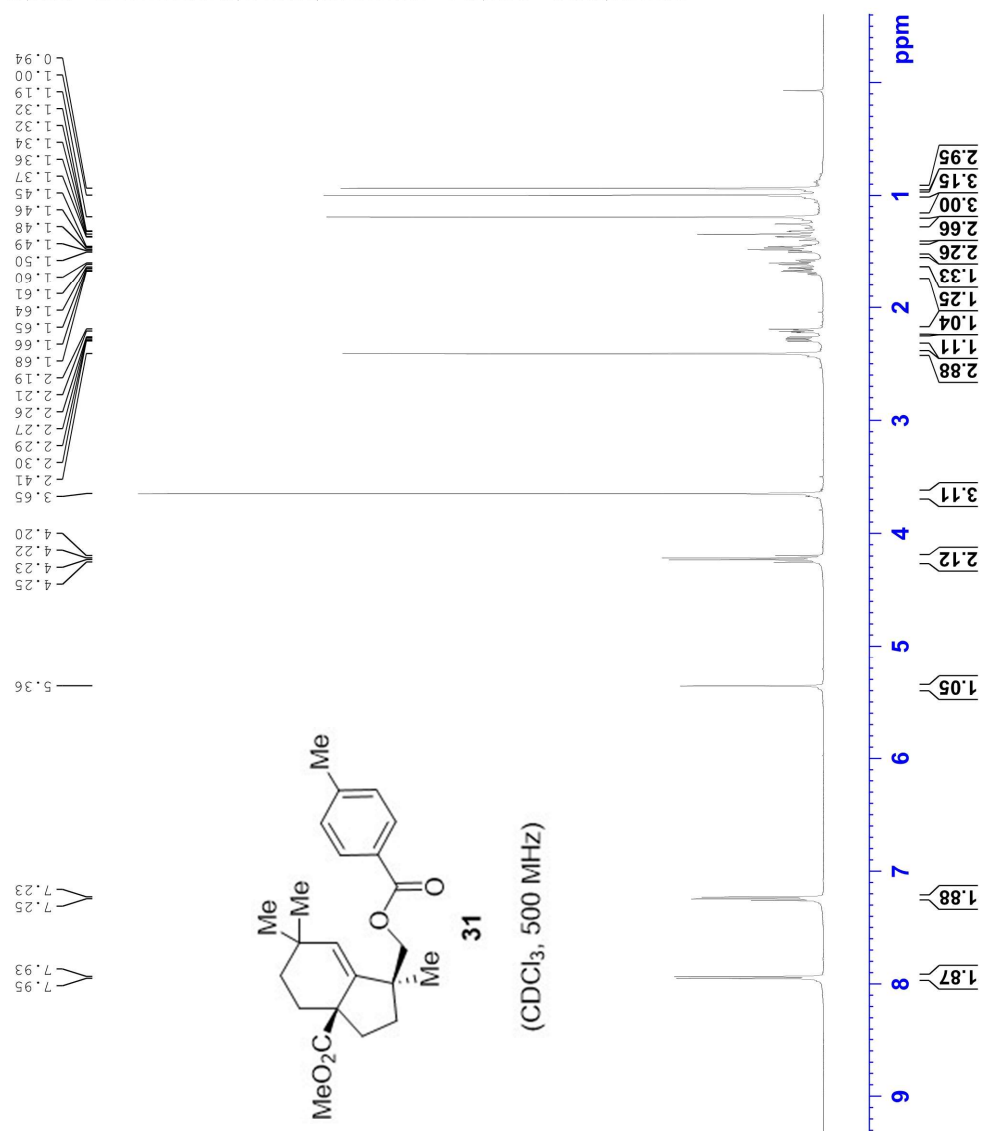


Current Data Parameters
 NAME Yh-2-42-a
 EXPNO 1
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20171226
 Time_ 21.24
 INSTRUM spect
 PROBHD 5 mm PABBO BB/
 PULPROG zg
 TD 5998
 SOLVENT CDCl3
 NS 4
 DS 0
 SWH 10000.000 Hz
 FIDRES 0.166672 Hz
 AQ 2.9999001 sec
 RG 28.76
 DW 50.000 usec
 DE 6.50 usec
 TE 296.0 K
 DL 3.00000000 sec
 TDO 1

===== CHANNEL f1 =====
 SFO1 499.8730869 MHz
 NUC1 1H
 P1 10.75 usec
 PLW1 18.25000000 W

F2 - Processing parameters
 SI 65536
 SF 499.8700123 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00



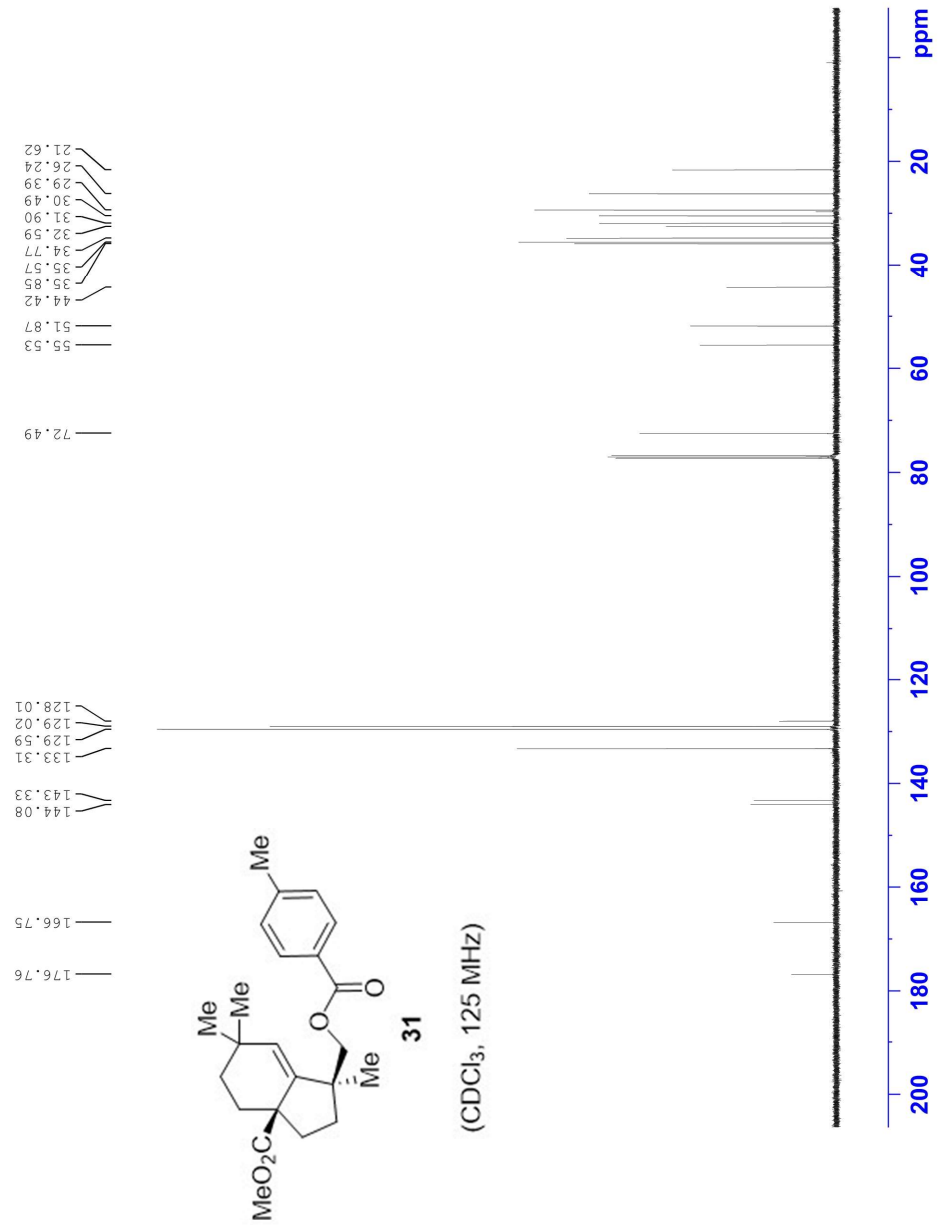
Current Data Parameters
 NAME Yh-2-42-a
 EXPNO 2
 PROCNO 2

F2 - Acquisition Parameters
 Date_ 20171226
 Time_ 22.26
 INSTRUM spect
 PROBHD 5 mm PABBO BB/
 PULPROG zgpg
 TD 187496
 SOLVENT CDC13
 NS 482
 DS 0
 SWH 31250.000 Hz
 FIDRES 0.166670 Hz
 AQ 2.9999361 sec
 RG 2050
 DW 16.000 usec
 DE 6.50 usec
 TE 297.7 K
 D1 3.0000000 sec
 D11 0.0300000 sec
 TDO 1

==== CHANNEL f1 =====
 SFO1 125.7049802 MHz
 NUC1 13C
 P1 10.00 usec
 PLW1 72.83999634 W

==== CHANNEL f2 =====
 SFO2 499.8724993 MHz
 NUC2 1H
 CPDPRG12 waitz16
 PCPD2 80.00 usec
 PLW2 19.0000000 W
 PLW12 0.29688001 W

F2 - Processing parameters
 SI 1048576
 SF 125.6924115 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.40

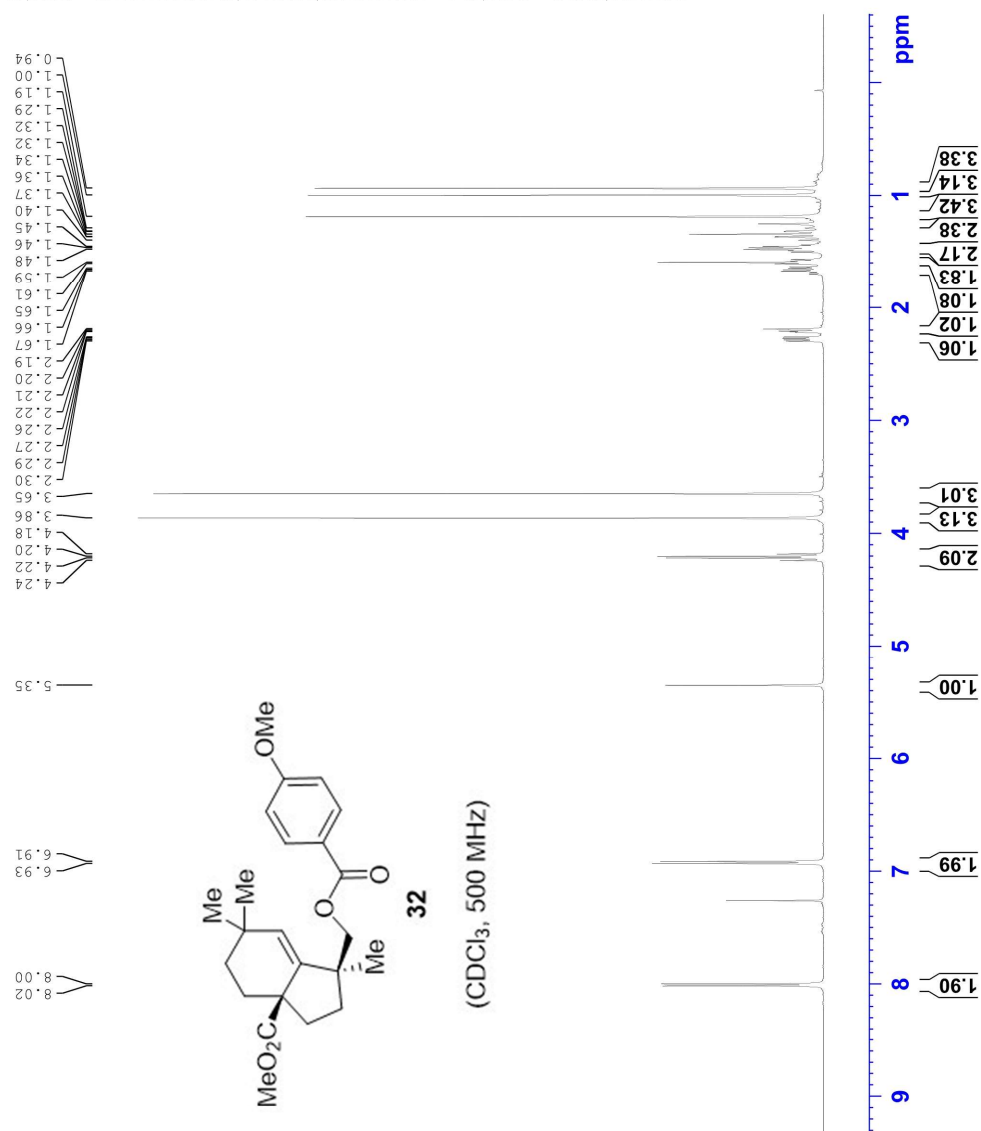


Current Data Parameters
 NAME yh-2-31-1-f
 EXPNO 1
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20171114
 Time_ 17.29
 INSTRUM spect
 PROBHD 5 mm PATXI 1H/
 PULPROG zg
 TD 5998
 SOLVENT CDCl3
 NS 8
 DS 0
 SWH 10000.000 Hz
 FIDRES 0.166672 Hz
 AQ 2.9999001 sec
 RG 79.04
 DW 50.000 usec
 DE 10.00 usec
 TE 294.6 K
 DL 2.0000000 sec
 TD0 1

===== CHANNEL f1 =====
 SFO1 500.1330885 MHz
 NUC1 1H
 PL 8.00 usec
 PLW1 12.19999981 W

F2 - Processing parameters
 SI 65536
 SF 500.1300132 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00



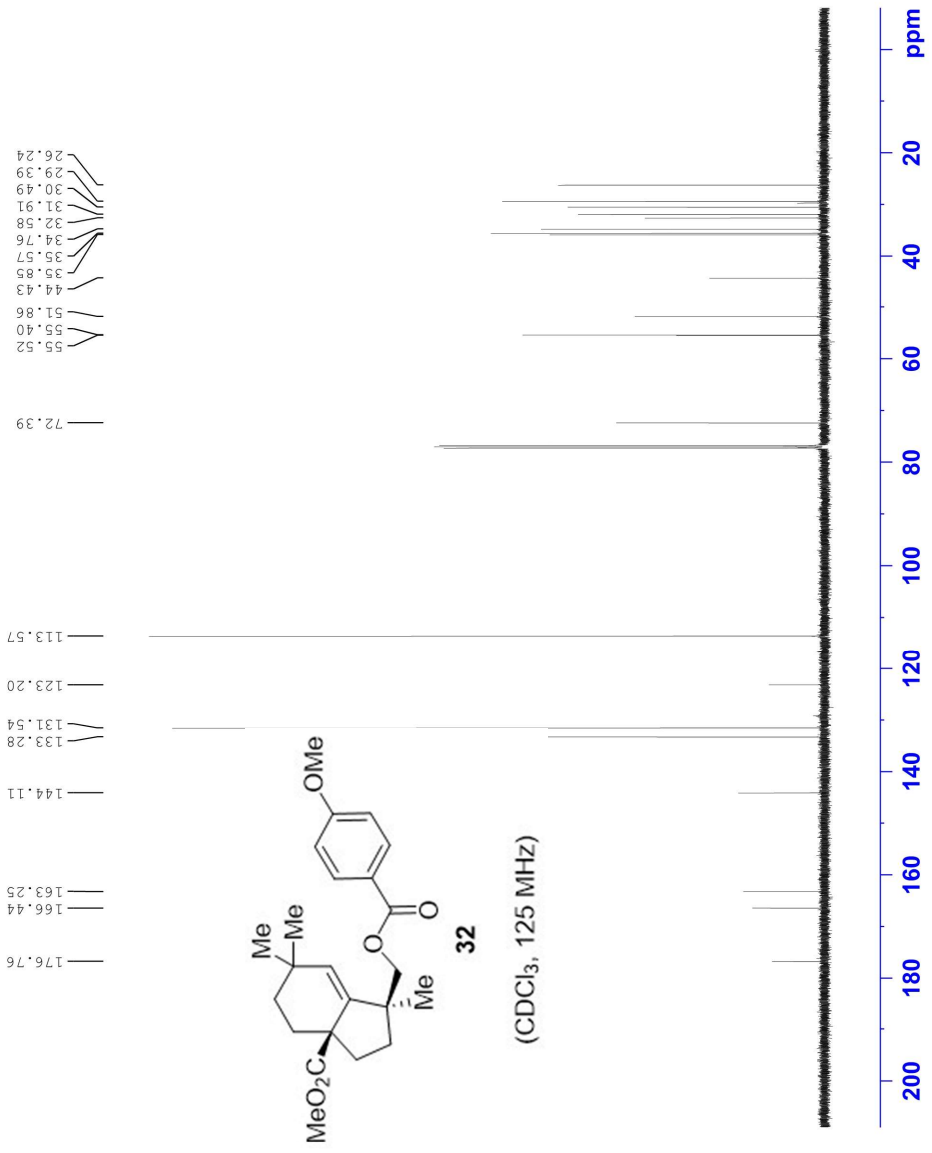
Current Data Parameters
 NAME Yh-2-31-f
 EXPNO 2
 PROCNO 2

F2 - Acquisition Parameters
 Date_ 20171122
 Time_ 21.57
 INSTRUM spect
 PROBHD 5 mm PABBO BB/
 PULPROG zgpg
 TD 187496
 SOLVENT CDCl3
 NS 559
 DS 0
 SWH 31250.000 Hz
 FIDRES 0.166670 Hz
 AQC 2.9999361 sec
 RG 2050
 DW 16.000 usec
 DE 6.50 usec
 TE 298.8 K
 D1 3.0000000 sec
 D11 0.0300000 sec
 TDD 1

==== CHANNEL f1 =====
 SFO1 125.7049802 MHz
 NUC1 13C
 P1 10.00 usec
 PLW1 72.83999634 W

==== CHANNEL f2 =====
 SFO2 499.8724993 MHz
 NUC2 1H
 CPDPRG12 waitz16
 PCPD2 80.00 usec
 PLW2 19.0000000 W
 PLWI2 0.29688001 W

F2 - Processing parameters
 SI 1048576
 SF 125.6924115 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.40

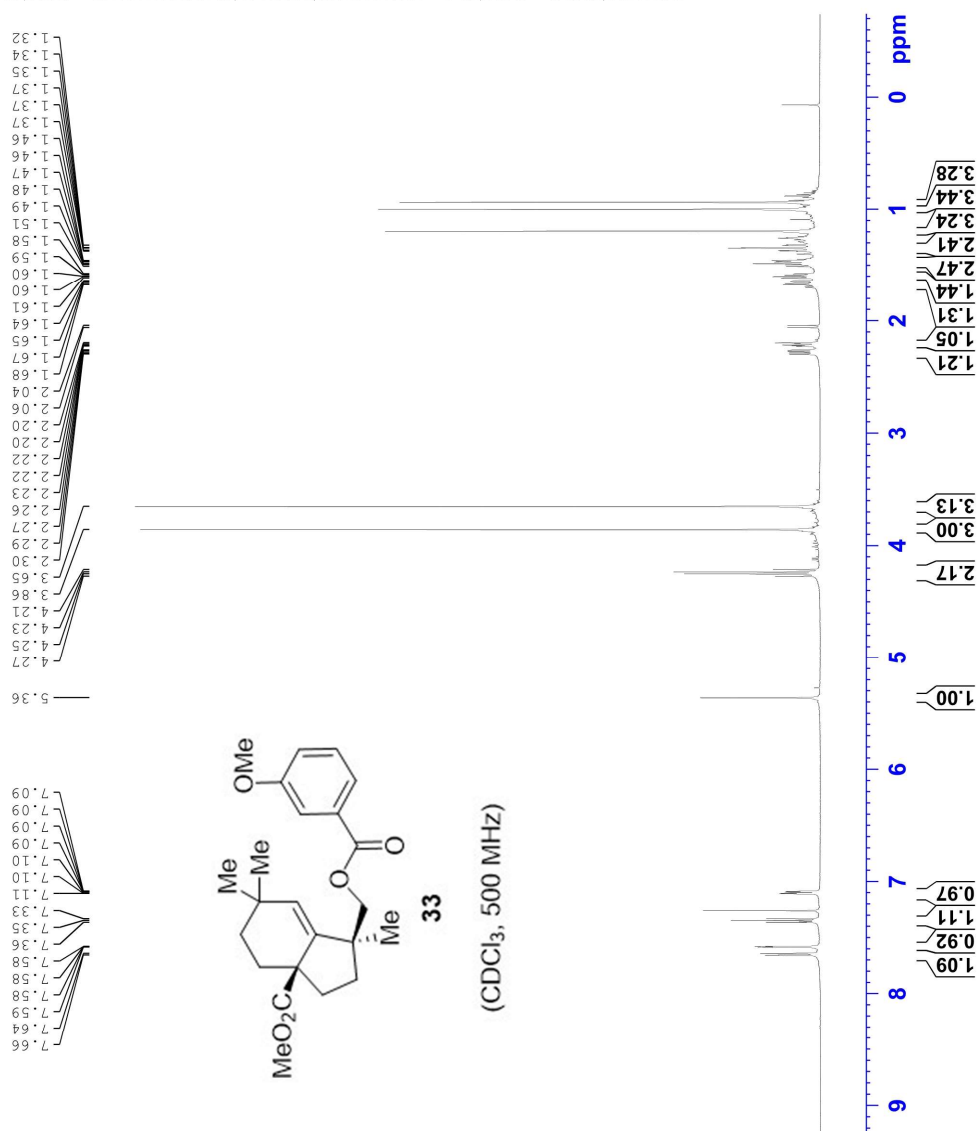


Current Data Parameters
 NAME Yh-2-62-a
 EXPNO 1
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20180104
 Time_ 22.12
 INSTRUM spect
 PROBHD 5 mm PABBO BB/
 PULPROG zg
 TD 5998
 SOLVENT CDCl3
 NS 2
 DS 0
 SWH 10000.000 Hz
 FIDRES 0.166672 Hz
 AQC 2.9999001 sec
 RG 37.92
 DW 50.000 usec
 DE 6.50 usec
 TE 296.0 K
 DL 3.0000000 sec
 TDO 1

===== CHANNEL f1 =====
 SFO1 499.8730869 MHz
 NUC1 1H
 P1 10.75 usec
 PLW1 18.25000000 W

F2 - Processing parameters
 SI 65536
 SF 499.8700123 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00



```

Current Data Parameters
NAME      Yh-2-62-a
EXPNO    4
PROCNO   4

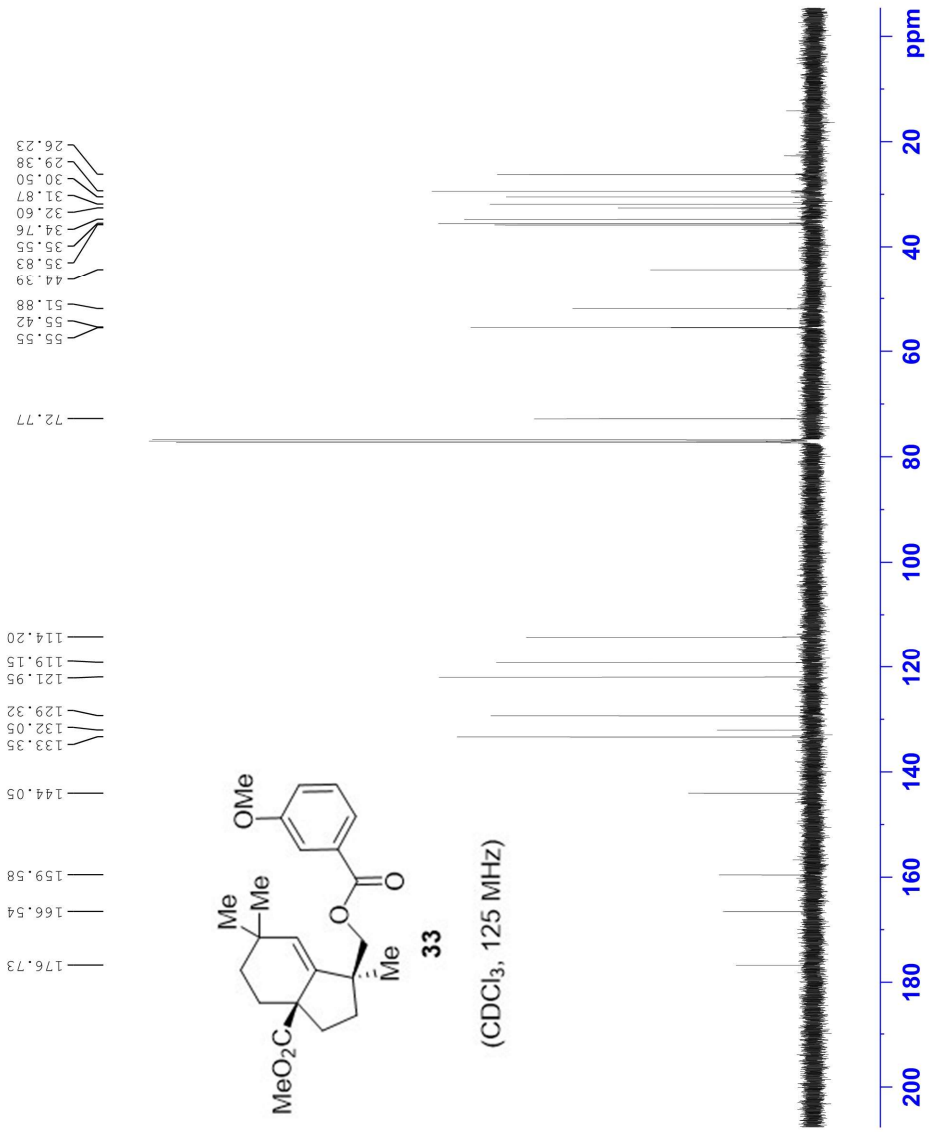
F2 - Acquisition Parameters
Date_    20180104
Time     22.55
INSTRUM  spect
PROBHD   5 mm PABBO BB/
PULPROG  zgpg
TD       187496
SOLVENT  CDCl3
NS       366
DS       0
SWH      31250.000 Hz
FIDRES   0.166670 Hz
AQ       2.9999361 sec
RG       2050
DW       16.000 usec
DE       6.50 usec
TE       297.7 K
D1       3.0000000 sec
D11      0.03000000 sec
TD0      1

===== CHANNEL f1 =====
SFO1    125.7049802 MHz
NUC1    13C
P1      10.00 usec
PLW1    72.83999634 W

===== CHANNEL f2 =====
SFO2    499.8724993 MHz
NUC2    1H
CPDPRG2 waitz16
PCPD2   80.00 usec
PLW2    19.0000000 W
PLW12   0.29688001 W

F2 - Processing parameters
SI      1048576
SF      125.6924115 MHz
WDW     EM
SSB     0
LB      0.30 Hz
GB      0
PC      1.40

```

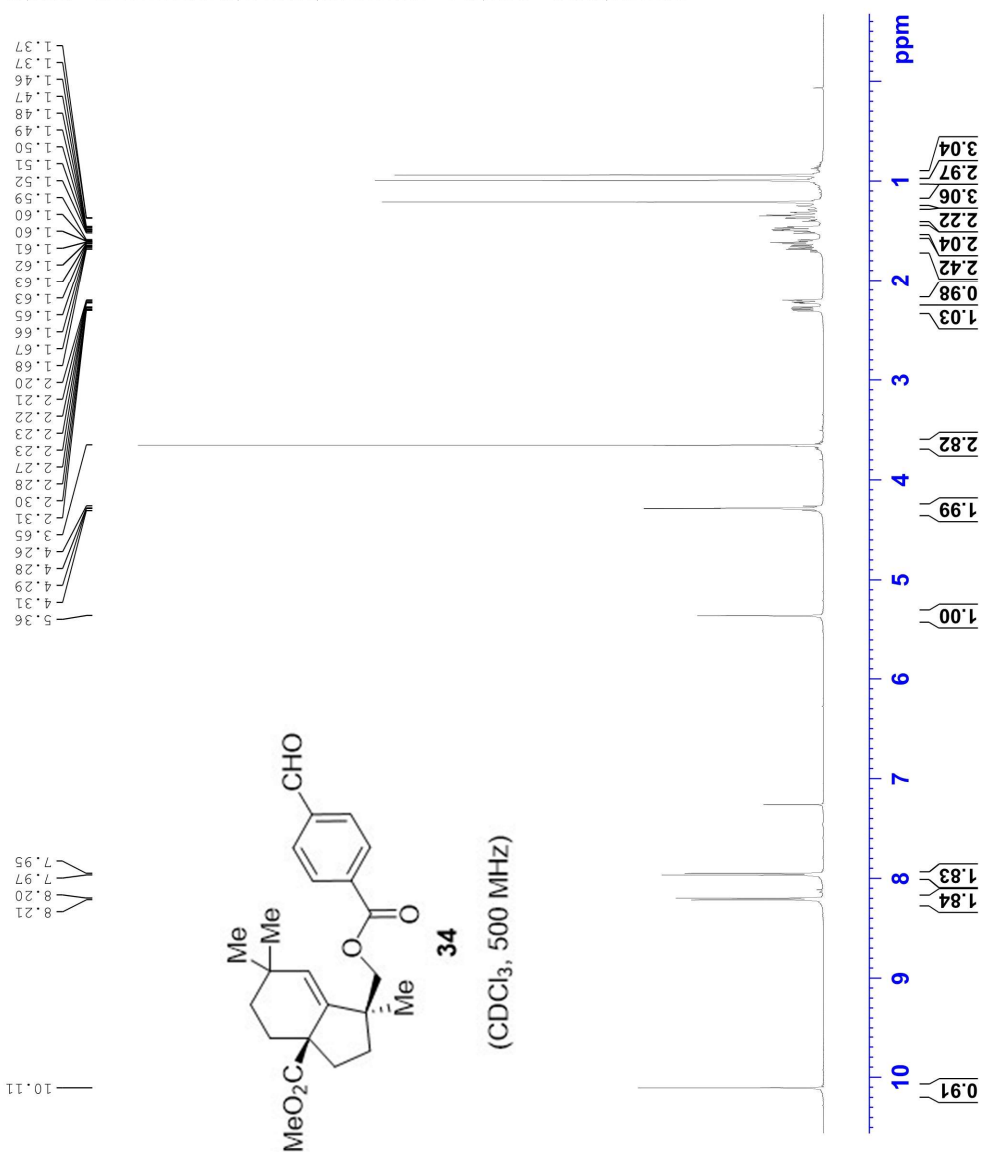


Current Data Parameters
 NAME Yh-2-77-a
 EXPNO 1
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20180124
 Time_ 22.13
 INSTRUM spect
 PROBHD 5 mm PABBO BB/
 PULPROG zg
 TD 5998
 SOLVENT CDCl3
 NS 2
 DS 0
 SWH 10000.000 Hz
 FIDRES 0.166672 Hz
 AQ 2.9999001 sec
 RG 35.92
 DW 50.000 usec
 DE 6.50 usec
 TE 296.9 K
 DL 3.0000000 sec
 TDO 1

===== CHANNEL f1 =====
 SFO1 499.8730869 MHz
 NUC1 1H
 PL 10.75 usec
 PLW1 18.25000000 W

F2 - Processing parameters
 SI 65536
 SF 499.8700124 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00



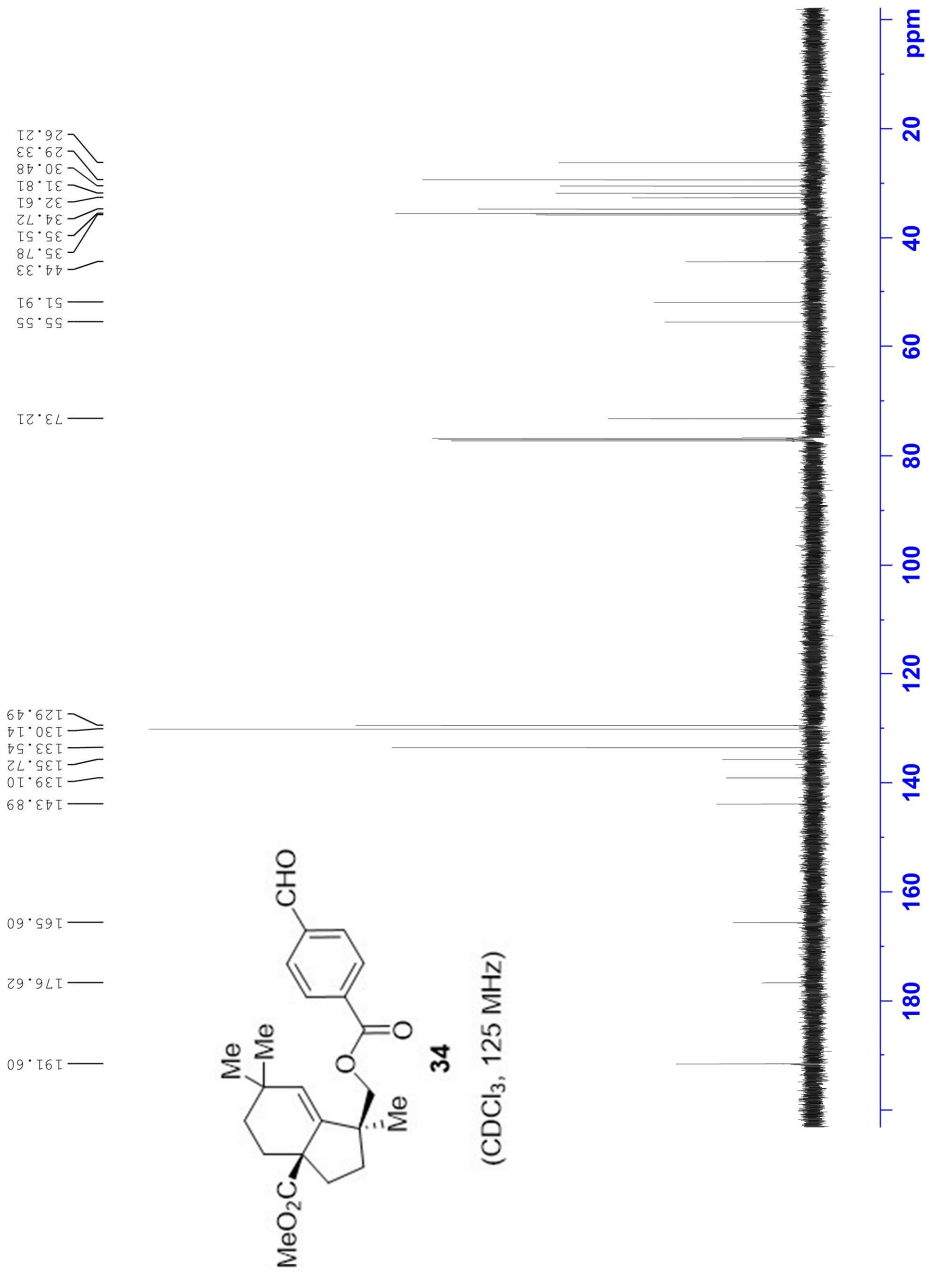
Current Data Parameters
 NAME Yh-2-77-a
 EXPNO 3
 PROCNO 3

F2 - Acquisition Parameters
 Date_ 20180124
 Time_ 22.35
 INSTRUM spect
 PROBHD 5 mm PABBO BB/
 PULPROG zgpg
 TD 187496
 SOLVENT CDCl3
 NS 135
 DS 0
 SWH 31250.000 Hz
 FIDRES 0.166670 Hz
 AQ 2.9999361 sec
 RG 2050
 DW 16.000 usec
 DE 6.50 usec
 TE 298.2 K
 D1 3.00000000 sec
 D11 0.03000000 sec
 TDO 1

==== CHANNEL f1 =====
 SFO1 125.7049802 MHz
 NUC1 13C
 P1 10.00 usec
 PLW1 72.83999634 W

==== CHANNEL f2 =====
 SFO2 499.8724993 MHz
 NUC2 1H
 CPDPRG12 waitz16
 FCPD2 80.00 usec
 PLW2 19.00000000 W
 PLW12 0.29688001 W

F2 - Processing parameters
 SI 1048576
 SF 125.6924115 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.40

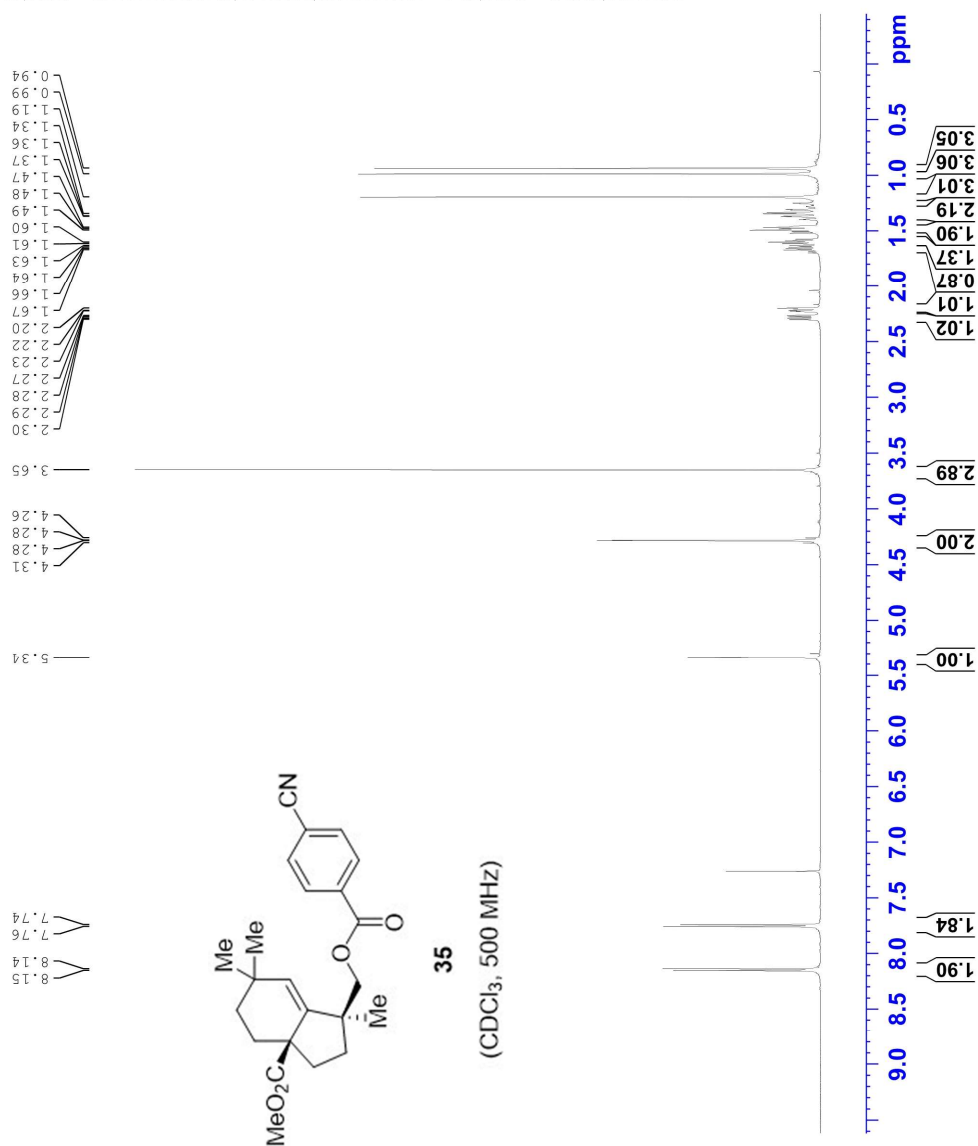


Current Data Parameters
 NAME Yh-2-53-a
 EXPNO 1
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20180102
 Time_ 21:59
 INSTRUM spect
 PROBHD 5 mm PABBO BB/
 PULPROG zg
 TD 5998
 SOLVENT CDCl3
 NS 4
 DS 0
 SWH 10000.000 Hz
 FIDRES 0.166672 Hz
 AQ 2.9999001 sec
 RG 35.92
 DW 50.000 usec
 DE 6.50 usec
 TE 296.1 K
 DL 3.0000000 sec
 TDL 1

===== CHANNEL f1 =====
 SFO1 499.8730869 MHz
 NUC1 1H
 PL 10.75 usec
 PLW1 18.25000000 W

F2 - Processing parameters
 SI 65536
 SF 499.8700122 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00



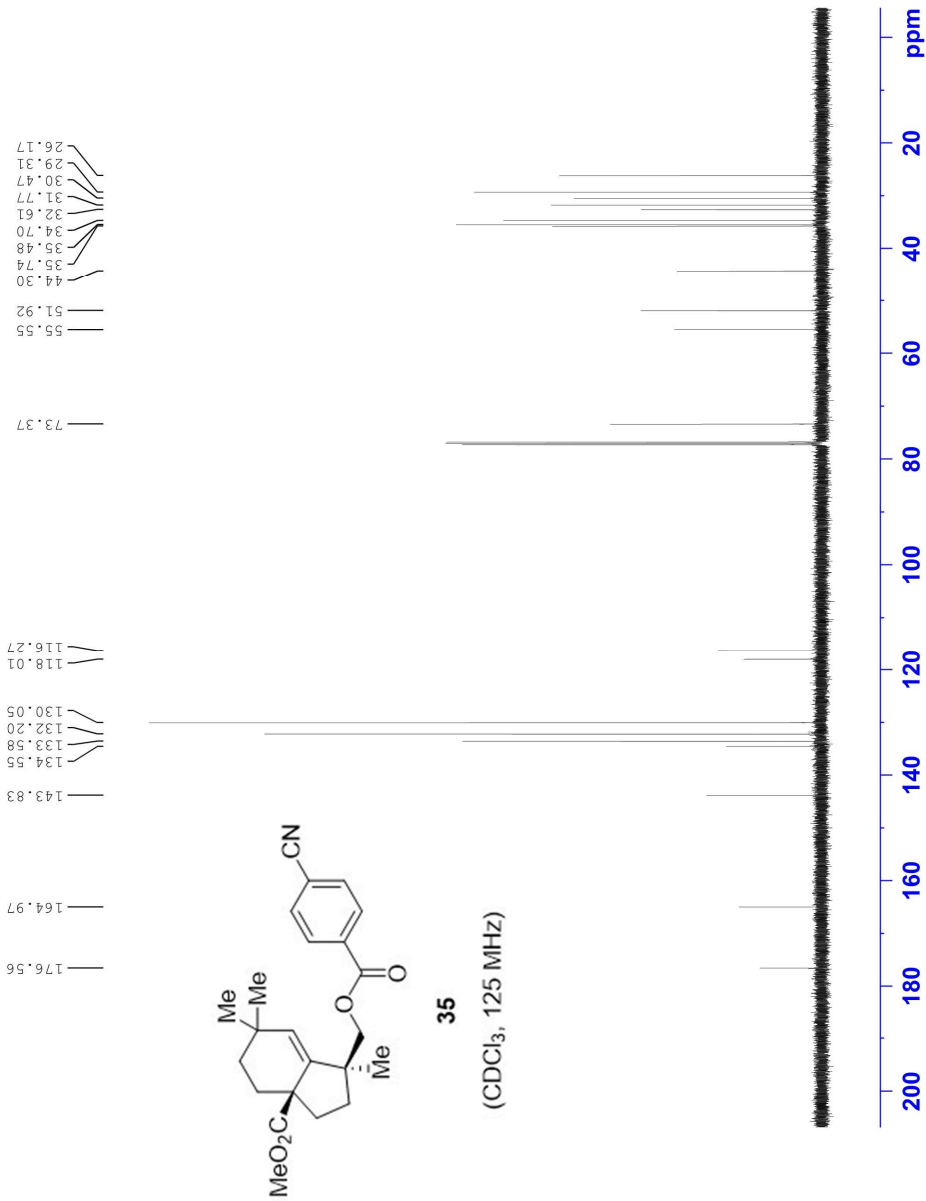
Current Data Parameters
 NAME Yh-2-53-a
 EXPNO 2
 PROCNO 2

F2 - Acquisition Parameters
 Date_ 20180102
 Time_ 22.29
 INSTRUM spect
 PROBHD 5 mm PABBO BB/
 PULPROG zgpg30
 TD 187496
 SOLVENT CDC13
 NS 273
 DS 0
 SWH 31250.000 Hz
 FIDRES 0.166670 Hz
 AQC 2.9999361 sec
 RG 2050
 DW 16.000 usec
 DE 6.50 usec
 TE 297.5 K
 D1 3.0000000 sec
 D11 0.0300000 sec
 TDO 1

===== CHANNEL f1 =====
 SFO1 125.7049802 MHz
 NUC1 13C
 P1 10.00 usec
 PLW1 72.83999634 W

===== CHANNEL f2 =====
 SFO2 499.8724993 MHz
 NUC2 1H
 CPDPRG12 waitz16
 PCPD2 80.00 usec
 PLW2 19.0000000 W
 PLW12 0.29688001 W

F2 - Processing parameters
 SI 1048576
 SF 125.6924115 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.40

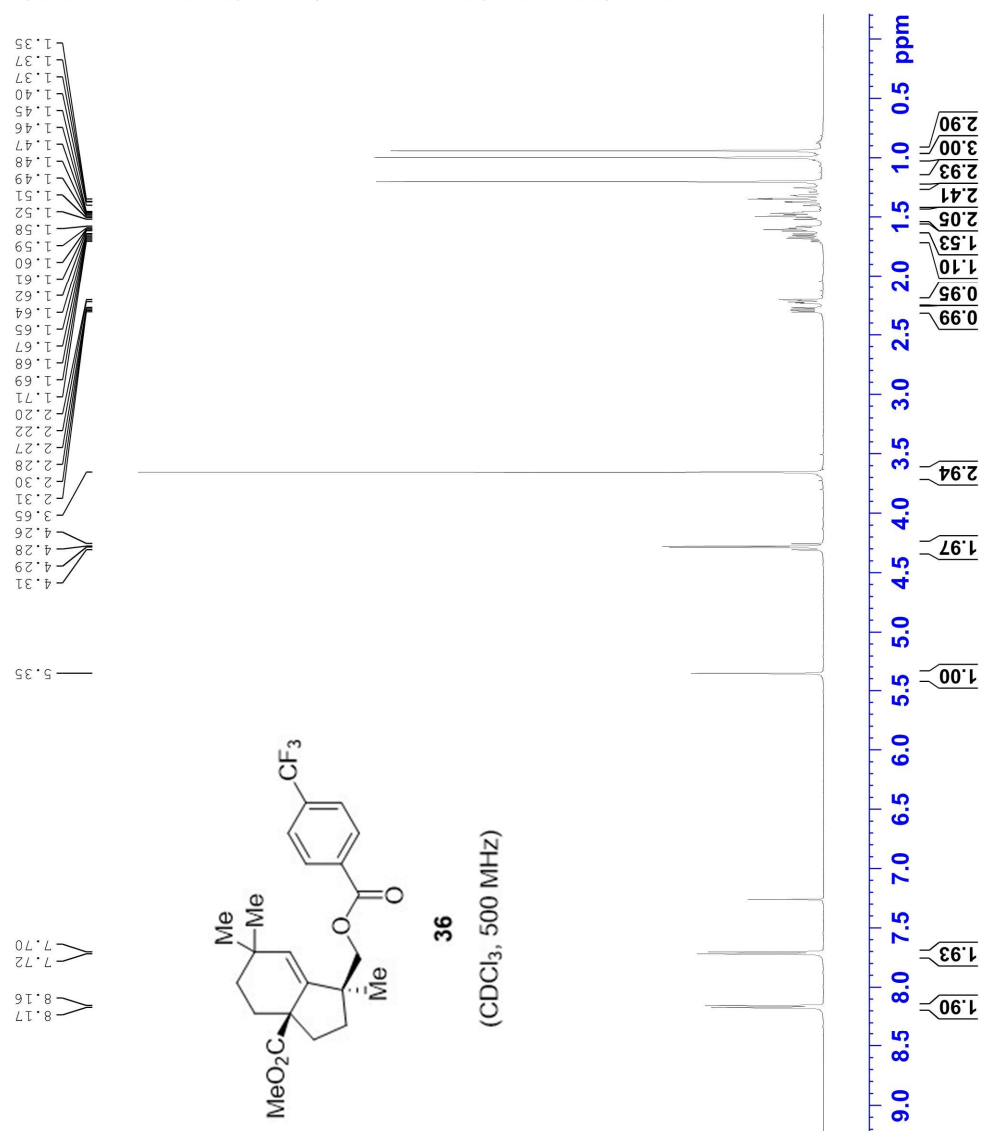


Current Data Parameters
 NAME Yh-2-43-a
 EXPNO 1
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20180812
 Time_ 22.11
 INSTRUM spect
 PROBHD 5 mm PABBO BB/
 PULPROG zg
 TD 5998
 SOLVENT CDCl3
 NS 5
 DS 0
 SWH 10000.000 Hz
 FIDRES 0.166672 Hz
 AQ 2.9999001 sec
 RG 62.78
 DW 50.000 usec
 DE 6.50 usec
 TE 296.8 K
 DL 4.0000000 sec
 TDO 1

===== CHANNEL f1 =====
 SFO1 499.8730869 MHz
 NUC1 1H
 P1 3.58 usec
 PLW1 18.25000000 W

F2 - Processing parameters
 SI 65536
 SF 499.8700121 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00



```

Current Data Parameters
NAME      Yh-2-43-a
EXPNO    2
PROCNO   2

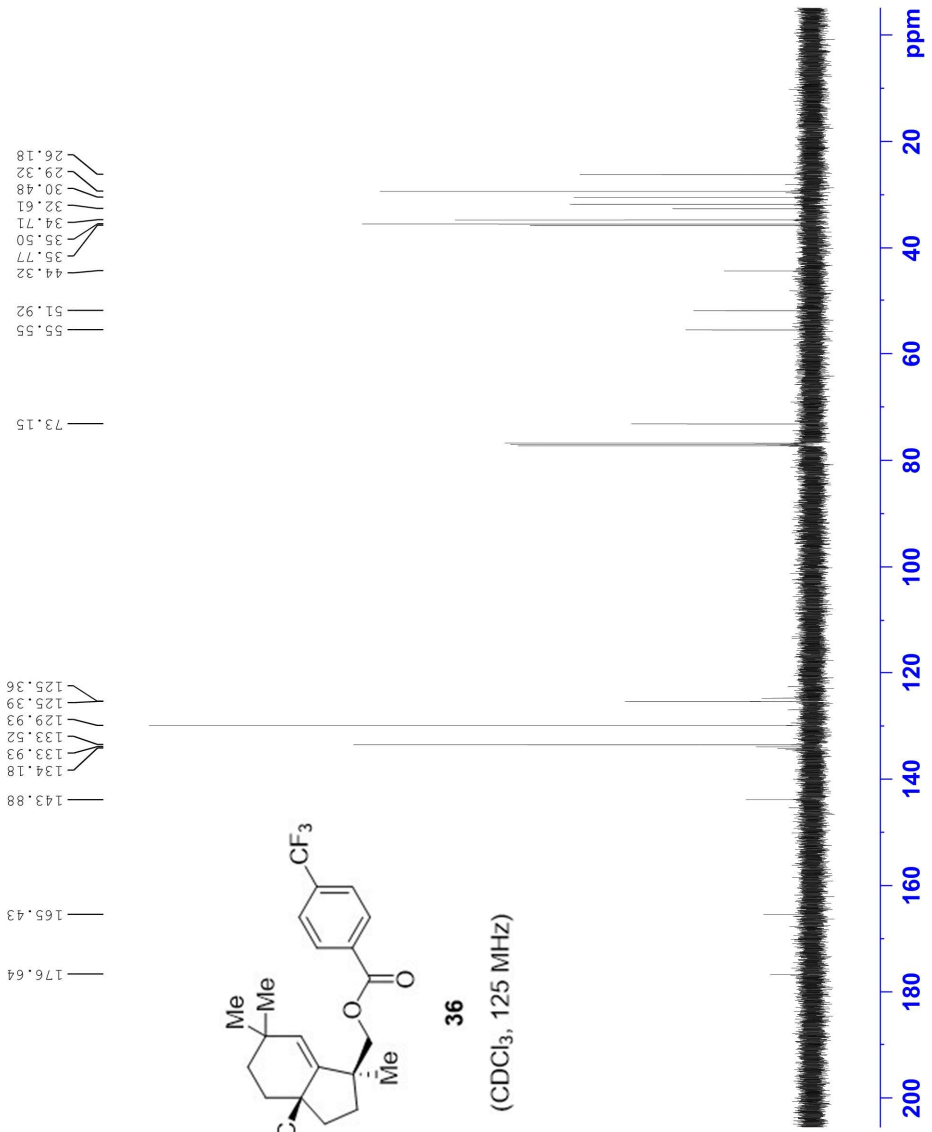
F2 - Acquisition Parameters
Date_     20180812
Time      22.23
INSTRUM   spect
PROBHD    5 mm PABBO BB/
PULPROG   zgpg
TD         187496
SOLVENT   CDCl3
NS         64
DS         0
SWH       31250.000 Hz
FIDRES    0.166670 Hz
AQ         2.9999361 sec
RG         2050
DW         16.000 usec
DE         6.50 usec
TE         297.2 K
D1         3.0000000 sec
D11        0.0300000 sec
TD0        1

===== CHANNEL f1 =====
SFO1      125.7049802 MHz
NUC1       13C
P1         10.00 usec
PLW1      72.83999634 W

===== CHANNEL f2 =====
SFO2      499.8724993 MHz
NUC2       1H
CFPRG12   waitz16
PCPD2     80.00 usec
PLW2      19.0000000 W
PLW12     0.29688001 W

F2 - Processing parameters
SI         1048576
SF         125.6924115 MHz
WDW        EM
SSB        0
LB         0.30 Hz
GB         0
PC         1.40

```

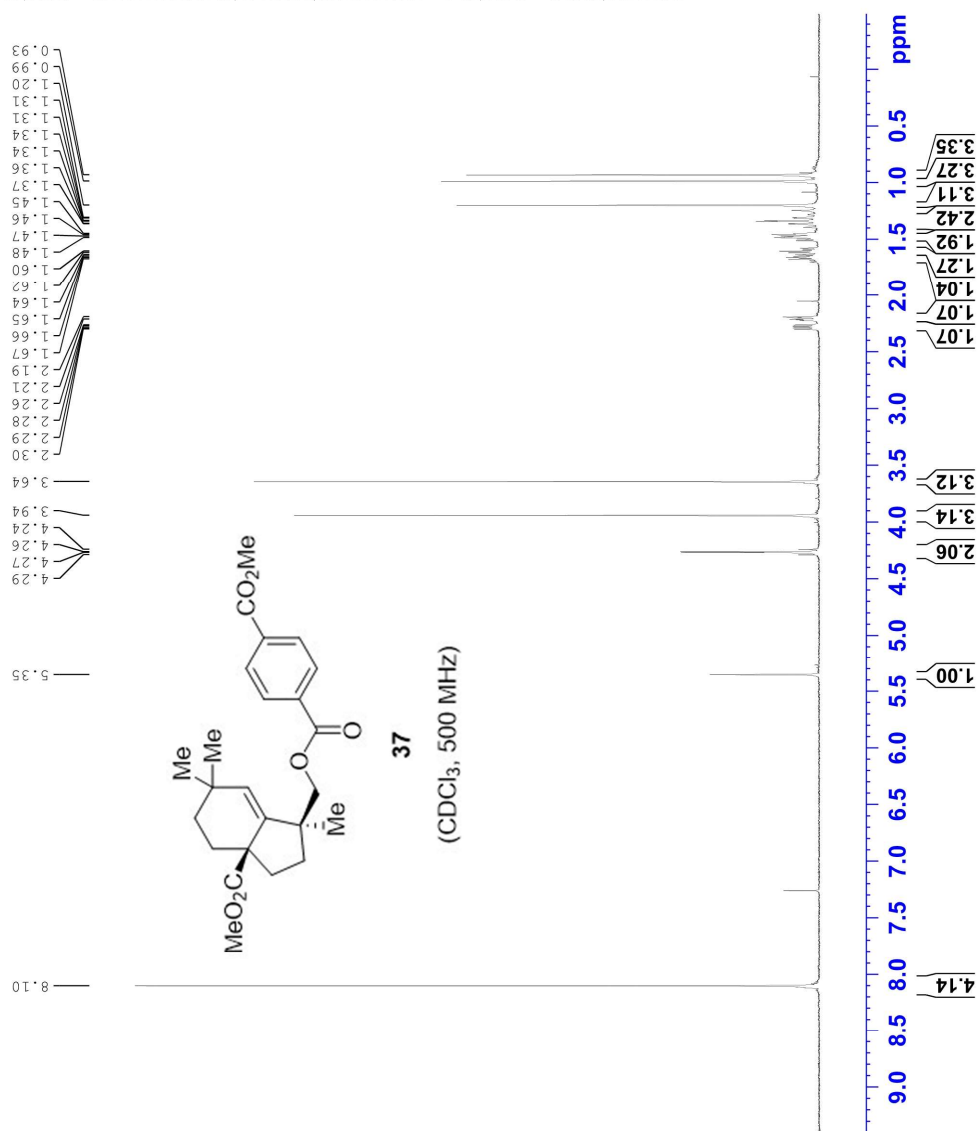


Current Data Parameters
 NAME Yh-2-69-a
 EXPNO 1
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20180119
 Time_ 21.15
 INSTRUM spect
 PROBHD 5 mm PABBO BB/
 PULPROG zg
 TD 5998
 SOLVENT CDCl3
 NS 3
 DS 0
 SWH 10000.000 Hz
 FIDRES 0.166672 Hz
 AQ 2.9999001 sec
 RG 893.6
 DW 50.000 usec
 DE 6.50 usec
 TE 297.2 K
 DL 3.0000000 sec
 TDO 1

===== CHANNEL f1 =====
 SFO1 499.8730869 MHz
 NUC1 1H
 P1 10.75 usec
 PLW1 18.25000000 W

F2 - Processing parameters
 SI 65536
 SF 499.8700123 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00



```

Current Data Parameters
NAME      Yh-2-69-a
EXPNO    2
PROCNO    2

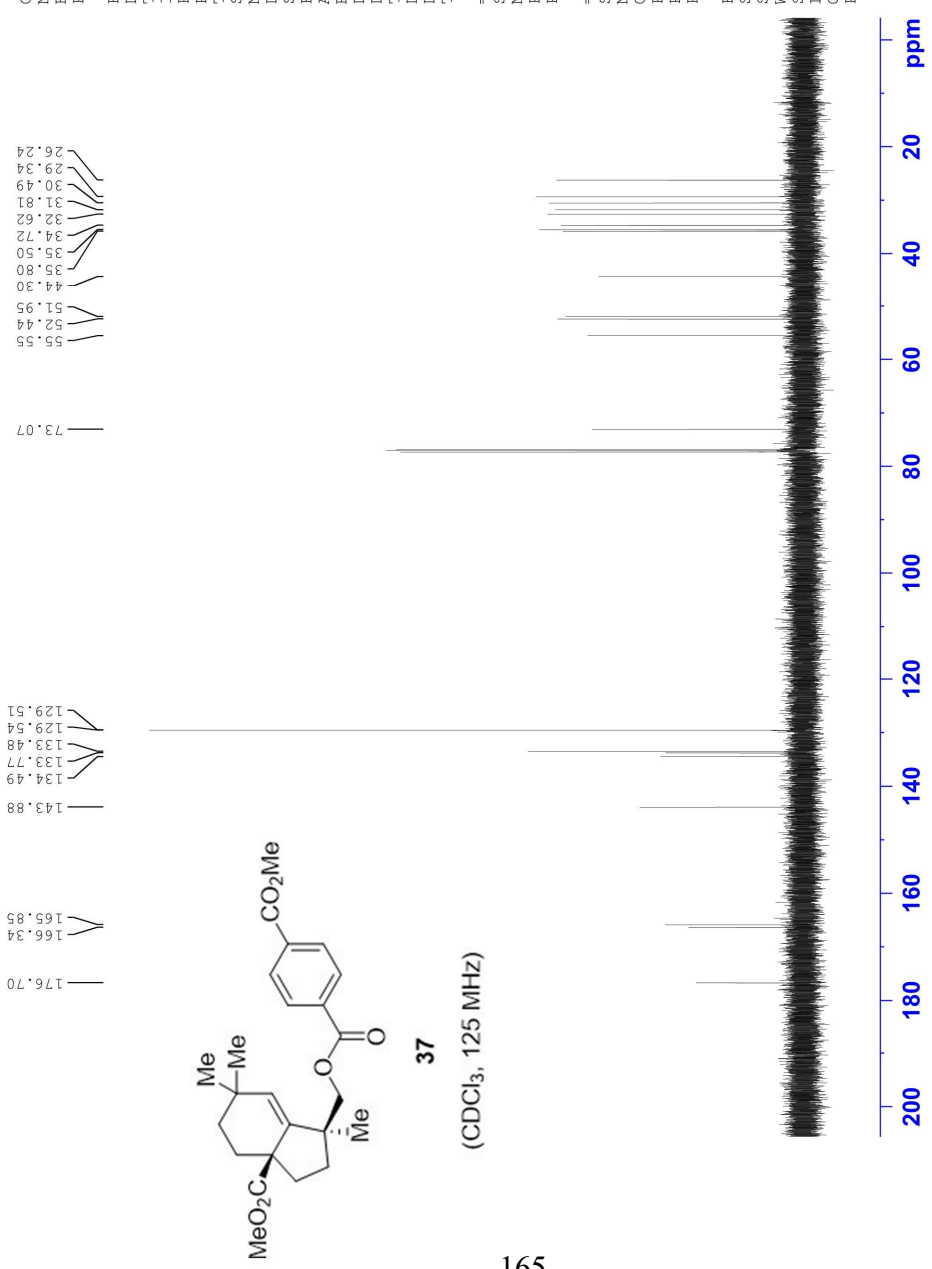
F2 - Acquisition Parameters
Date_     20180119
Time      21.22
INSTRUM   spect
PROBHD    5 mm PABBO BB/
PULPROG   zgpg
TD         187496
SOLVENT   CDC13
NS         6
DS         0
SWH        31250.000 Hz
FIDRES     0.166670 Hz
AQ         2.9999361 sec
RG         2050
DE         16.000 usec
WE         6.50 usec
TE         297.2 K
D1         3.00000000 sec
D11        0.03000000 sec
TD0        1

===== CHANNEL f1 =====
SFO1      125.7049802 MHz
NUC1       13C
P1         10.00 usec
PLW1      72.83999634 W

===== CHANNEL f2 =====
SFO2      499.8724993 MHz
NUC2       1H
PCPDPRG[2] waltz16
PCPD2     80.00 usec
PLW2      19.00000000 W
PLW12     0.29688001 W

F2 - Processing parameters
SI         1048576
SF         125.6924115 MHz
WDW        EM
SSB        0
LB         0.30 Hz
GB         0
PC         1.40

```

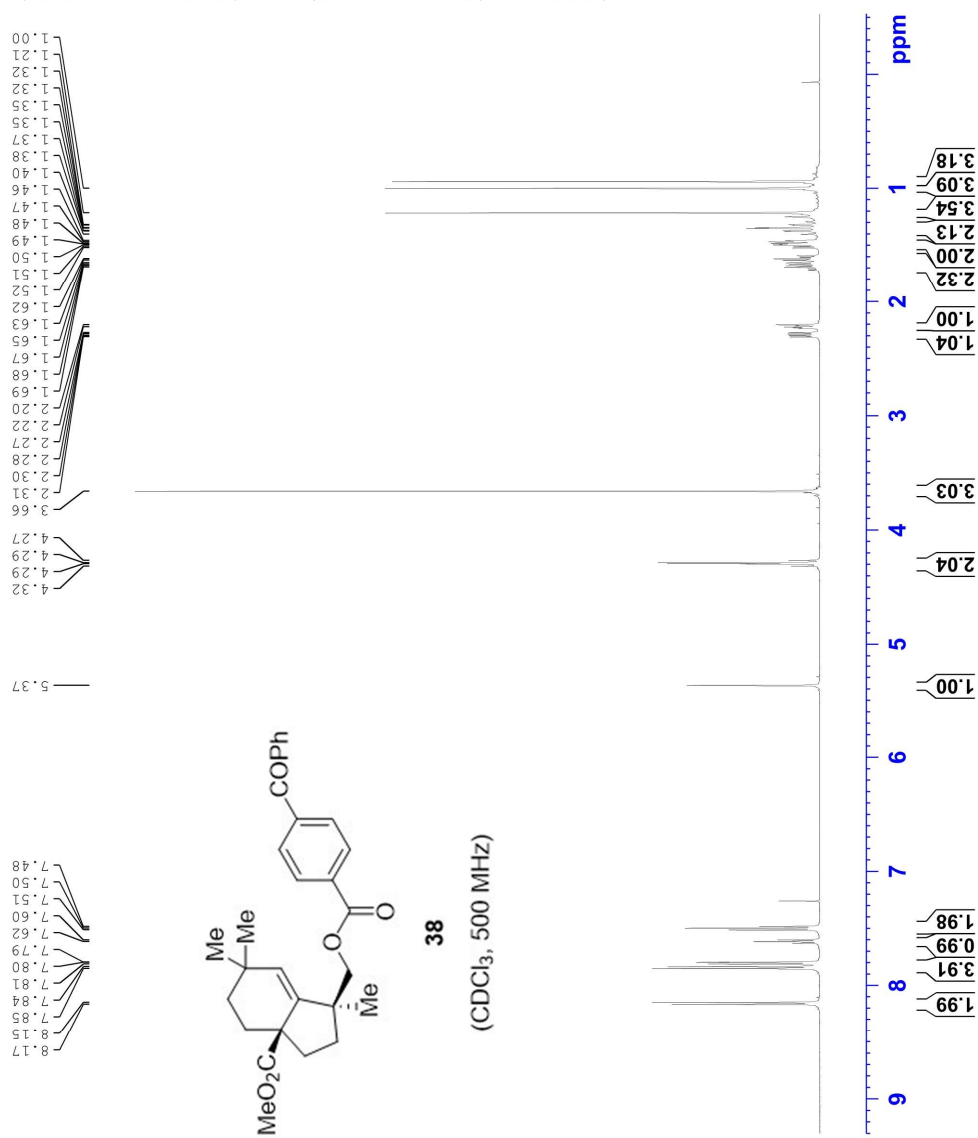


Current Data Parameters
 NAME Yh-2-72-a
 EXPNO 1
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20180111
 Time_ 22.37
 INSTRUM spect
 PROBHD 5 mm PABBO BB/
 PULPROG zg
 TD 5998
 SOLVENT CDCl3
 NS 2
 DS 0
 SWH 10000.000 Hz
 FIDRES 0.166672 Hz
 AQ 2.9999001 sec
 RG 28.76
 DW 50.000 usec
 DE 6.50 usec
 TE 296.7 K
 DL 3.00000000 sec
 TDO 1

===== CHANNEL f1 =====
 SFO1 499.8730869 MHz
 NUC1 1H
 P1 10.75 usec
 PLW1 18.25000000 W

F2 - Processing parameters
 SI 65536
 SF 499.8700124 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00



```

Current Data Parameters
NAME      Yh-2-72-a
EXPNO    2
PROCNO   2

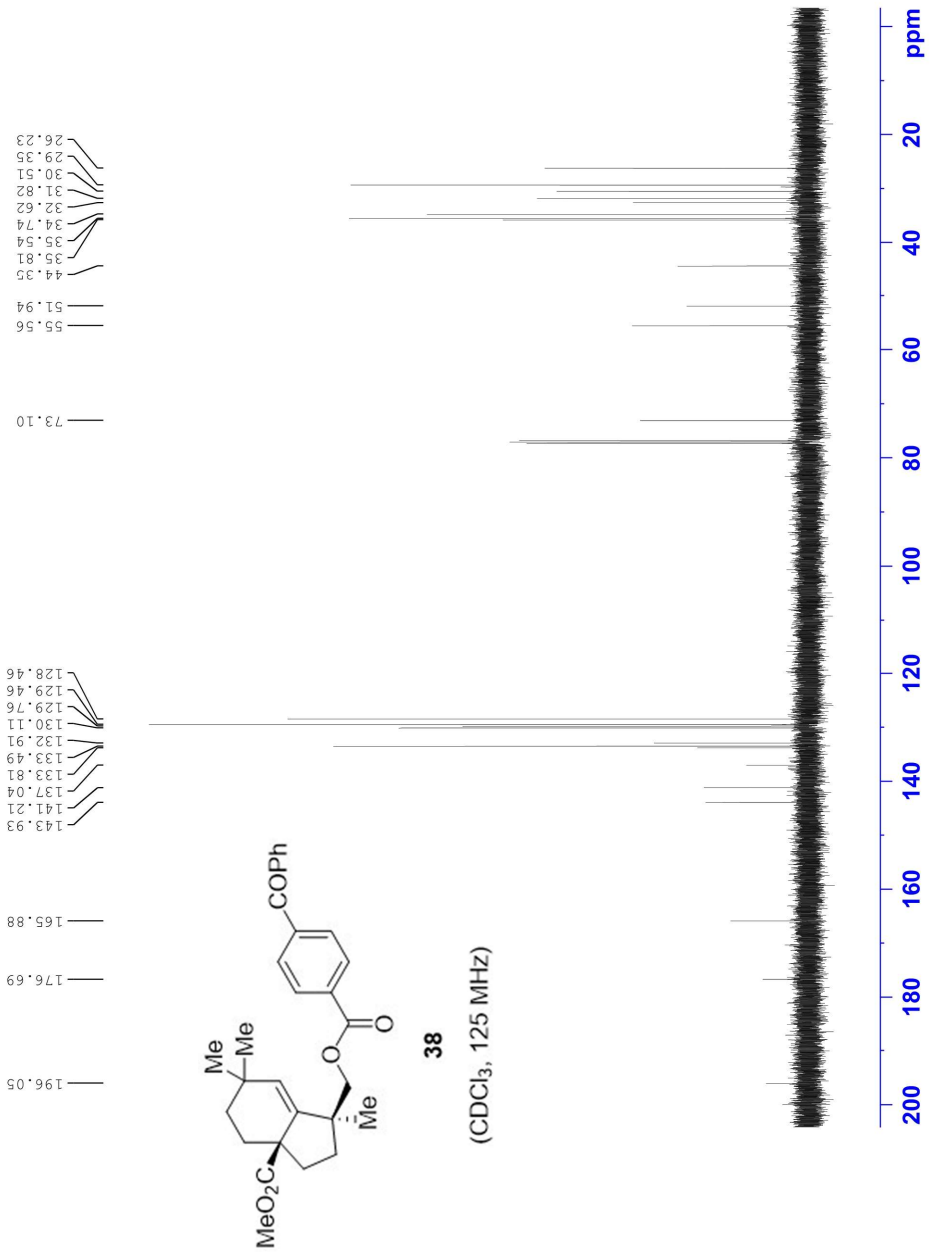
F2 - Acquisition Parameters
Date_     20180111
Time      22.44
INSTRUM   spect
PROBHD    5 mm PABBO BB/
PULPROG   zgpg
TD         187496
SOLVENT   CDCl3
NS         43
DS         0
SWH        31250.000 Hz
FIDRES     0.166670 Hz
AQ         2.9999361 sec
RG         2050
DW         16.000 usec
DE         6.50 usec
TE         297.4 K
D1         3.0000000 sec
D11        0.0300000 sec
TDO        1

===== CHANNEL f1 =====
SFO1      125.7049802 MHz
NUC1       13C
P1         10.00 usec
PLW1       72.83999634 W

===== CHANNEL f2 =====
SFO2      499.8724993 MHz
NUC2       1H
CFPRG12   waitz16
PCPD2     80.00 usec
PLW2      19.0000000 W
PLW12     0.29688001 W

F2 - Processing parameters
SI         1048576
SF         125.6924115 MHz
WDW        EM
SSB        0
LB         0.30 Hz
GB         0
PC         1.40

```

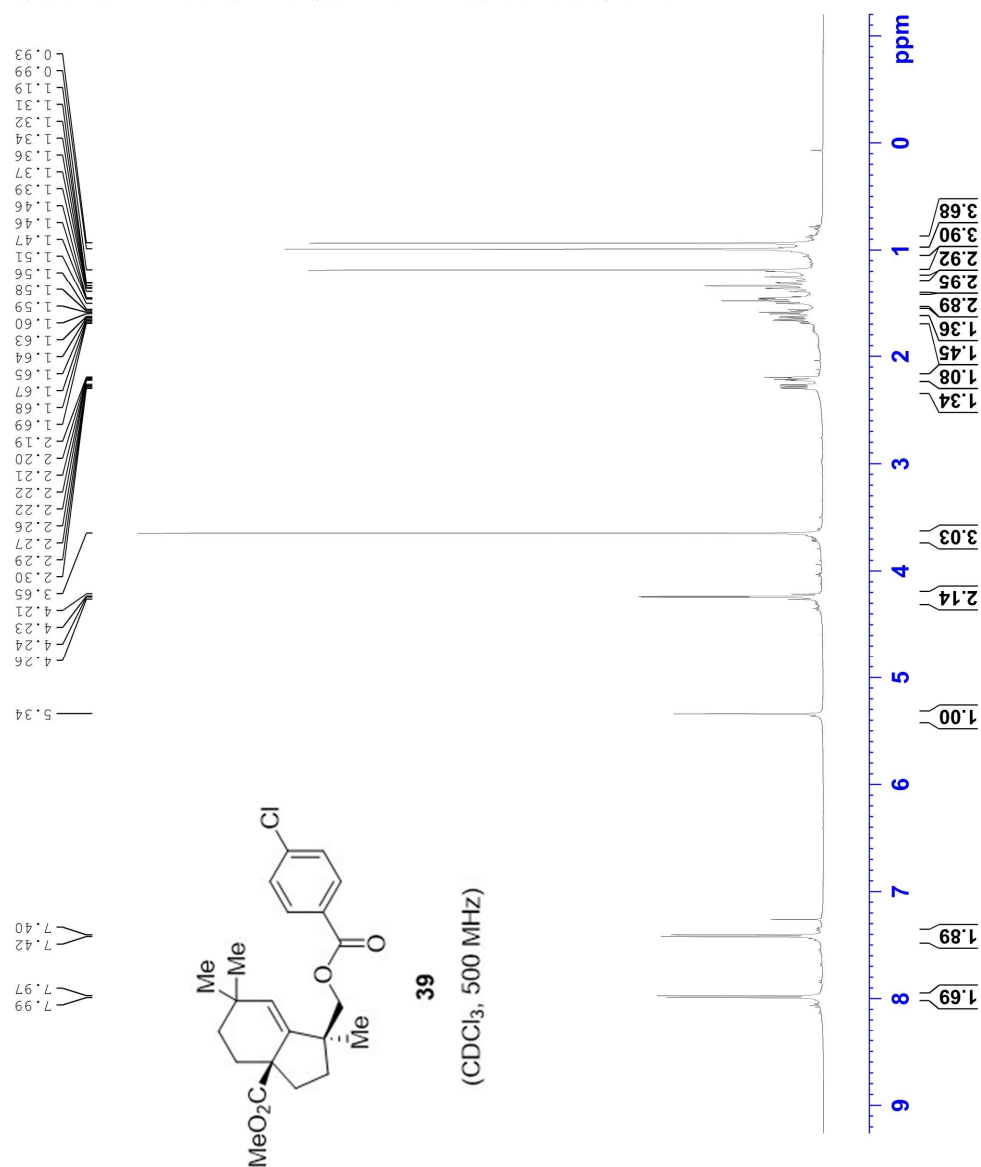


Current Data Parameters
 NAME Yh-2-47-a
 EXPNO 1
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20180101
 Time_ 21:59
 INSTRUM spect
 PROBHD 5 mm PABBO BB/
 PULPROG zg
 TD 5998
 SOLVENT CDCl3
 NS 16
 DS 0
 SWH 10000.000 Hz
 FIDRES 0.166672 Hz
 AQ 2.9999001 sec
 RG 28.76
 DW 50.000 usec
 DE 6.50 usec
 TE 296.7 K
 DL 3.0000000 sec
 TDO 1

===== CHANNEL f1 =====
 SFO1 499.8730869 MHz
 NUC1 1H
 PL 10.75 usec
 PLW1 18.25000000 W

F2 - Processing parameters
 SI 65536
 SF 499.8700122 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00



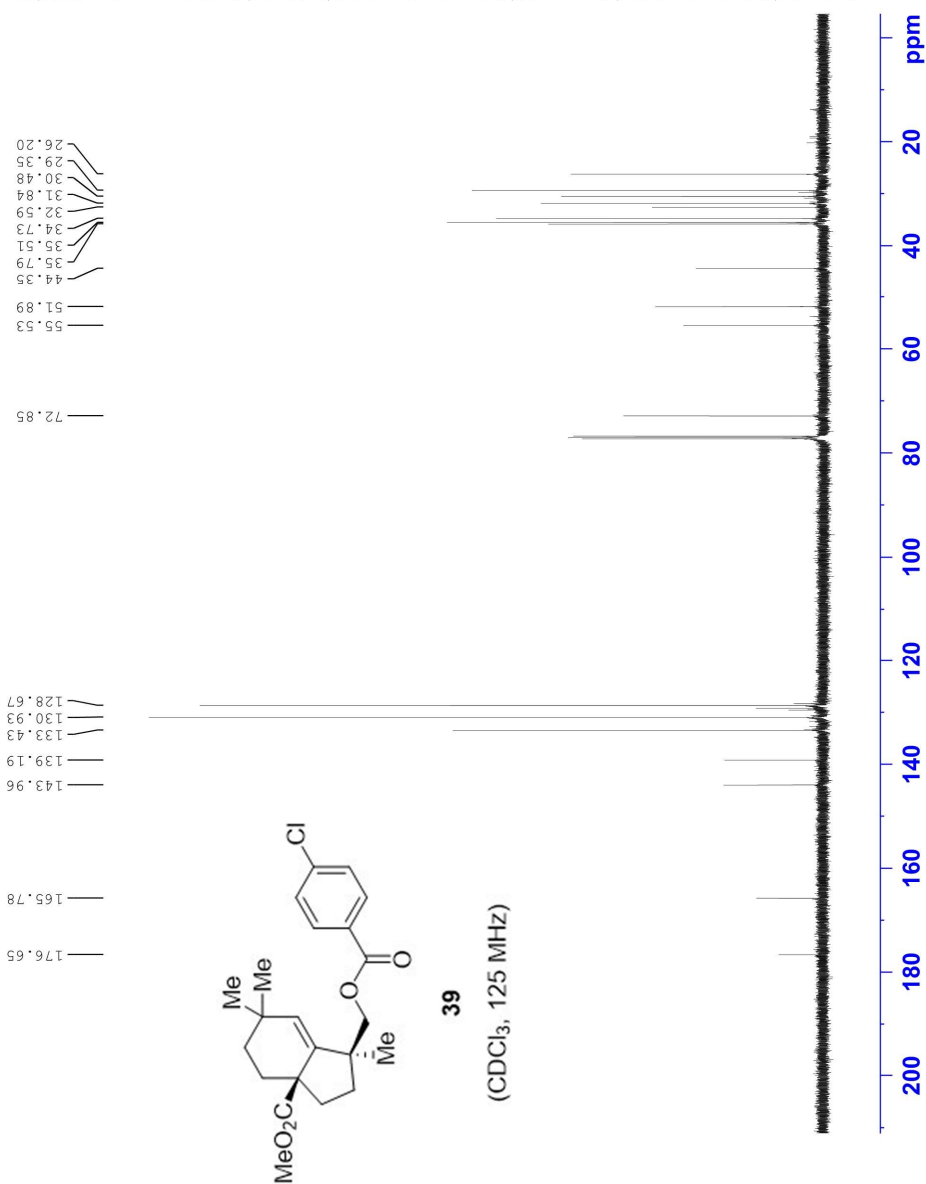
Current Data Parameters
 NAME Yh-2-47-a
 EXPNO 2
 PROCNO 2

F2 - Acquisition Parameters
 Date_ 20180101
 Time_ 22.30
 INSTRUM spect
 PROBHD 5 mm PABBO BB/
 PULPROG zgpg30
 TD 187496
 SOLVENT CDC13
 NS 261
 DS 0
 SWH 31250.000 Hz
 FIDRES 0.166670 Hz
 AQ 2.9999361 sec
 RG 2050
 DW 16.000 usec
 DE 6.50 usec
 TE 297.7 K
 D1 3.0000000 sec
 D11 0.0300000 sec
 TDO 1

==== CHANNEL f1 =====
 SFO1 125.7049802 MHz
 NUC1 13C
 P1 10.00 usec
 PLW1 72.83999634 W

==== CHANNEL f2 =====
 SFO2 499.8724993 MHz
 NUC2 1H
 CPDPRG12 waitz16
 PCPD2 80.00 usec
 PLW2 19.0000000 W
 PLW12 0.29688001 W

F2 - Processing parameters
 SI 1048576
 SF 125.6924115 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.40

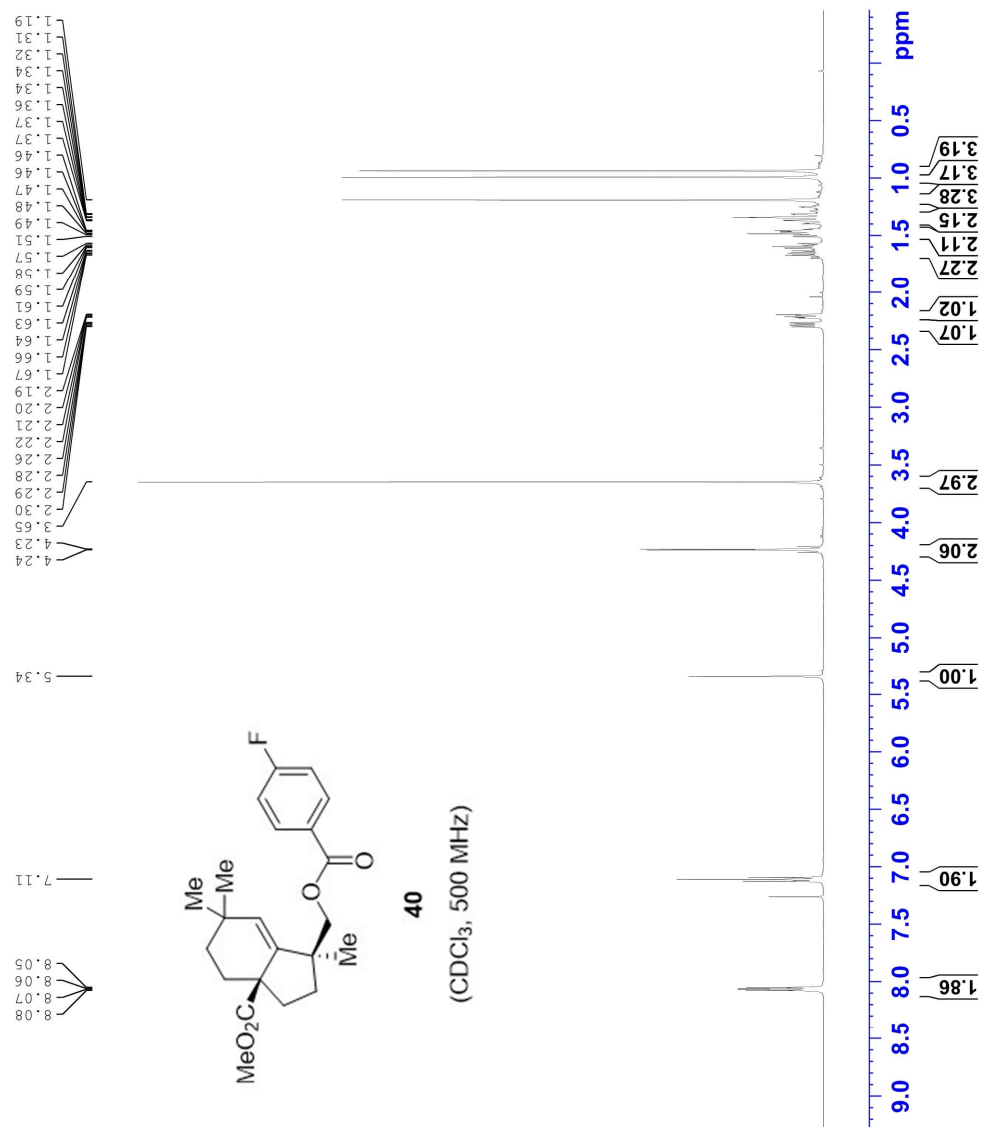


Current Data Parameters
 NAME Yh-2-55-a
 EXPNO 1
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20180103
 Time_ 21.58
 INSTRUM spect
 PROBHD 5 mm PABBO BB/
 PULPROG zg
 TD 5998
 SOLVENT CDCl3
 NS 4
 DS 0
 SWH 10000.000 Hz
 FIDRES 0.166672 Hz
 AQ 2.9999001 sec
 RG 29.95
 DW 50.000 usec
 DE 6.50 usec
 TE 296.2 K
 DL 3.0000000 sec
 TD0 1

===== CHANNEL f1 =====
 SFO1 499.8730869 MHz
 NUC1 1H
 PL 10.75 usec
 PLW1 18.25000000 W

F2 - Processing parameters
 SI 65536
 SF 499.8700123 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00



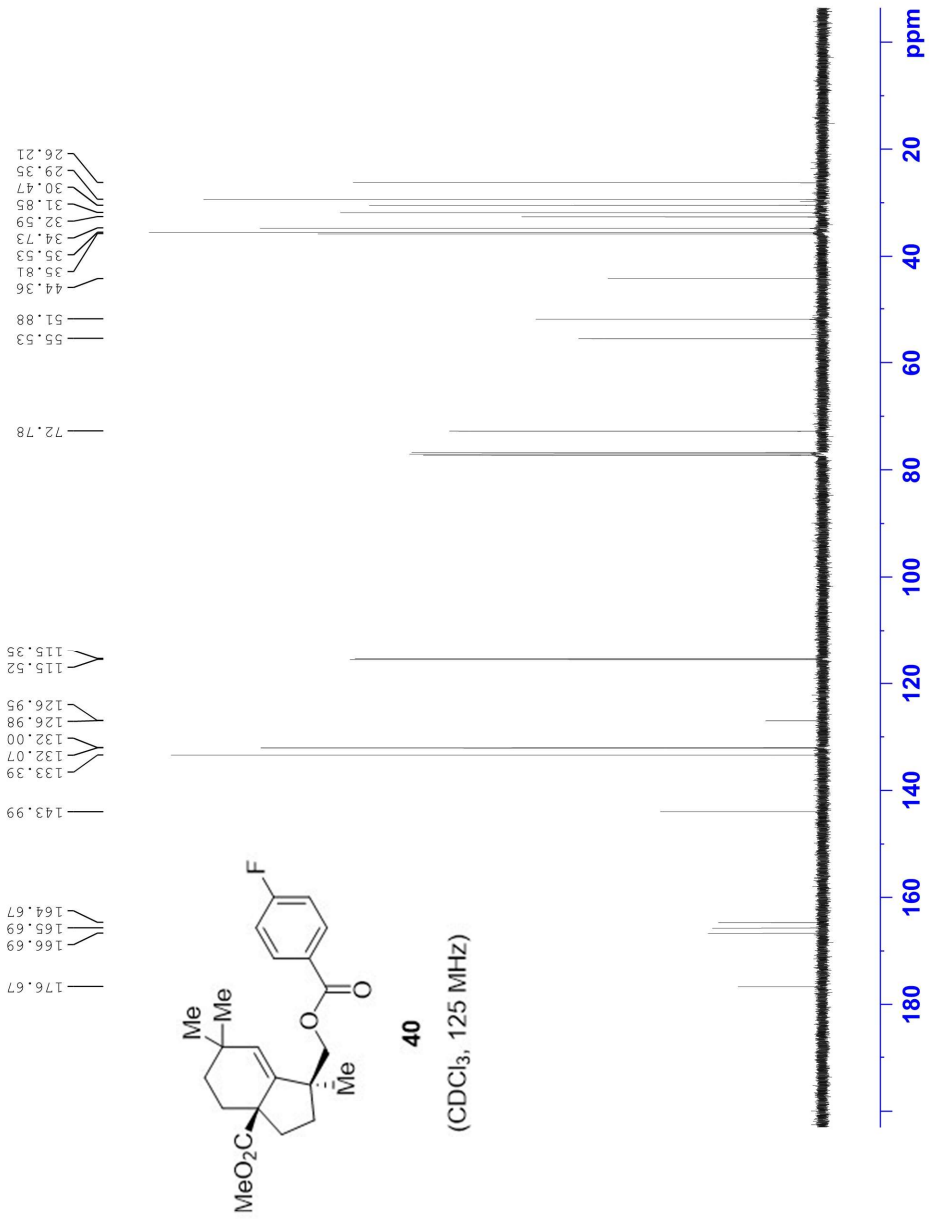
Current Data Parameters
 NAME Yh-2-55-a
 EXPNO 2
 PROCNO 2

F2 - Acquisition Parameters
 Date_ 20180103
 Time_ 22.36
 INSTRUM spect
 PROBHD 5 mm PABBO BB/
 PULPROG zgpg
 TD 187496
 SOLVENT CDCl3
 NS 354
 DS 0
 SWH 31250.000 Hz
 FIDRES 0.166670 Hz
 AQ 2.9999361 sec
 RG 2050
 DW 16.000 usec
 DE 6.50 usec
 TE 297.9 K
 D1 3.0000000 sec
 D11 0.0300000 sec
 TDO 1

==== CHANNEL f1 =====
 SFO1 125.7049802 MHz
 NUC1 13C
 P1 10.00 usec
 PLW1 72.83999634 W

==== CHANNEL f2 =====
 SFO2 499.8724993 MHz
 NUC2 1H
 CPDPRG12 waitz16
 FCPD2 80.00 usec
 PLW2 19.0000000 W
 PLW12 0.29688001 W

F2 - Processing parameters
 SI 1048576
 SF 125.6924115 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.40

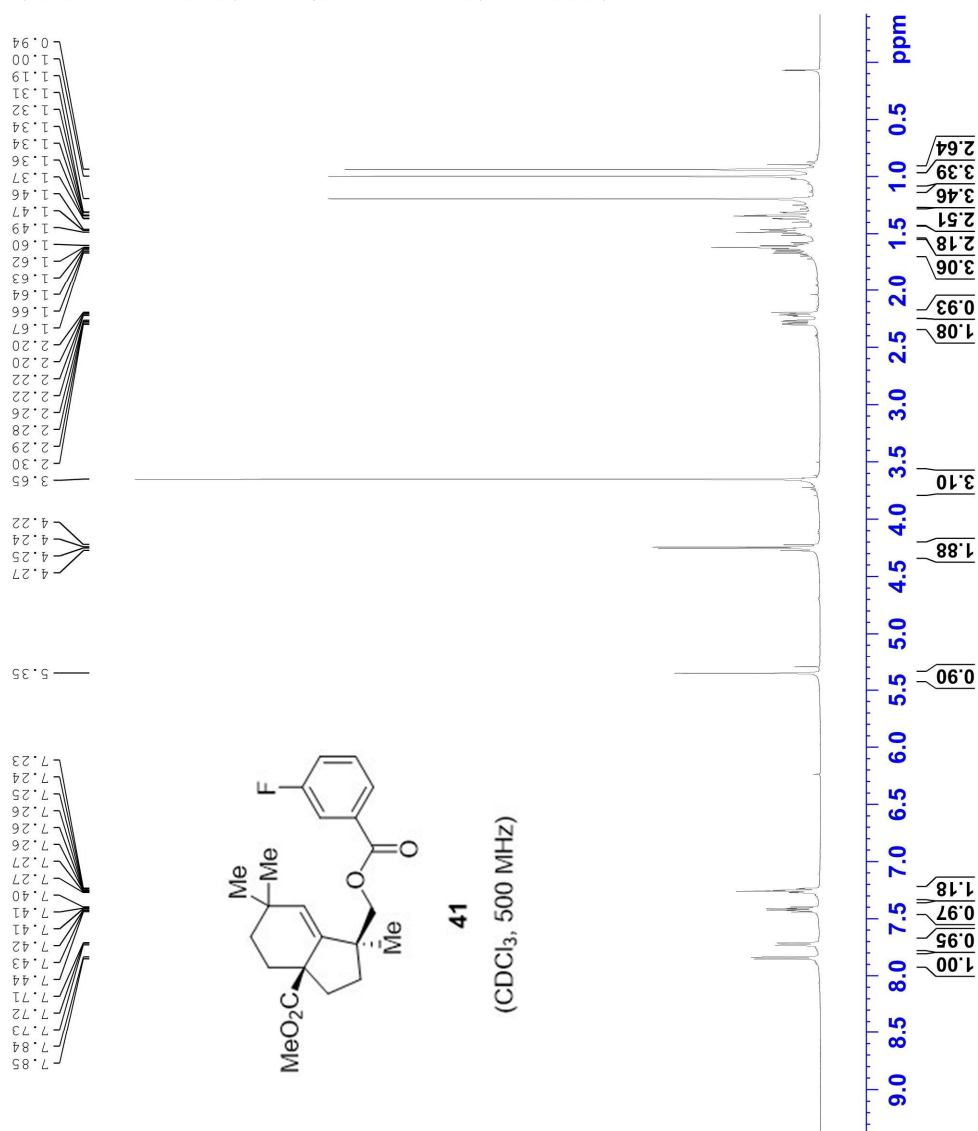


Current Data Parameters
 NAME Yh-2-68-a
 EXPNO 1
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20180821
 Time_ 16.51
 INSTRUM spect
 PROBHD 5 mm PABBO BB/
 PULPROG zg
 TD 5998
 SOLVENT CDCl3
 NS 5
 DS 0
 SWH 10000.000 Hz
 FIDRES 0.166672 Hz
 AQ 2.9999001 sec
 RG 62.78
 DW 50.000 usec
 DE 6.50 usec
 TE 295.7 K
 DL 4.0000000 sec
 TDO 1

===== CHANNEL f1 =====
 SFO1 499.8730869 MHz
 NUC1 1H
 PL 3.58 usec
 PLW1 18.25000000 W

F2 - Processing parameters
 SI 65536
 SF 499.8700123 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00



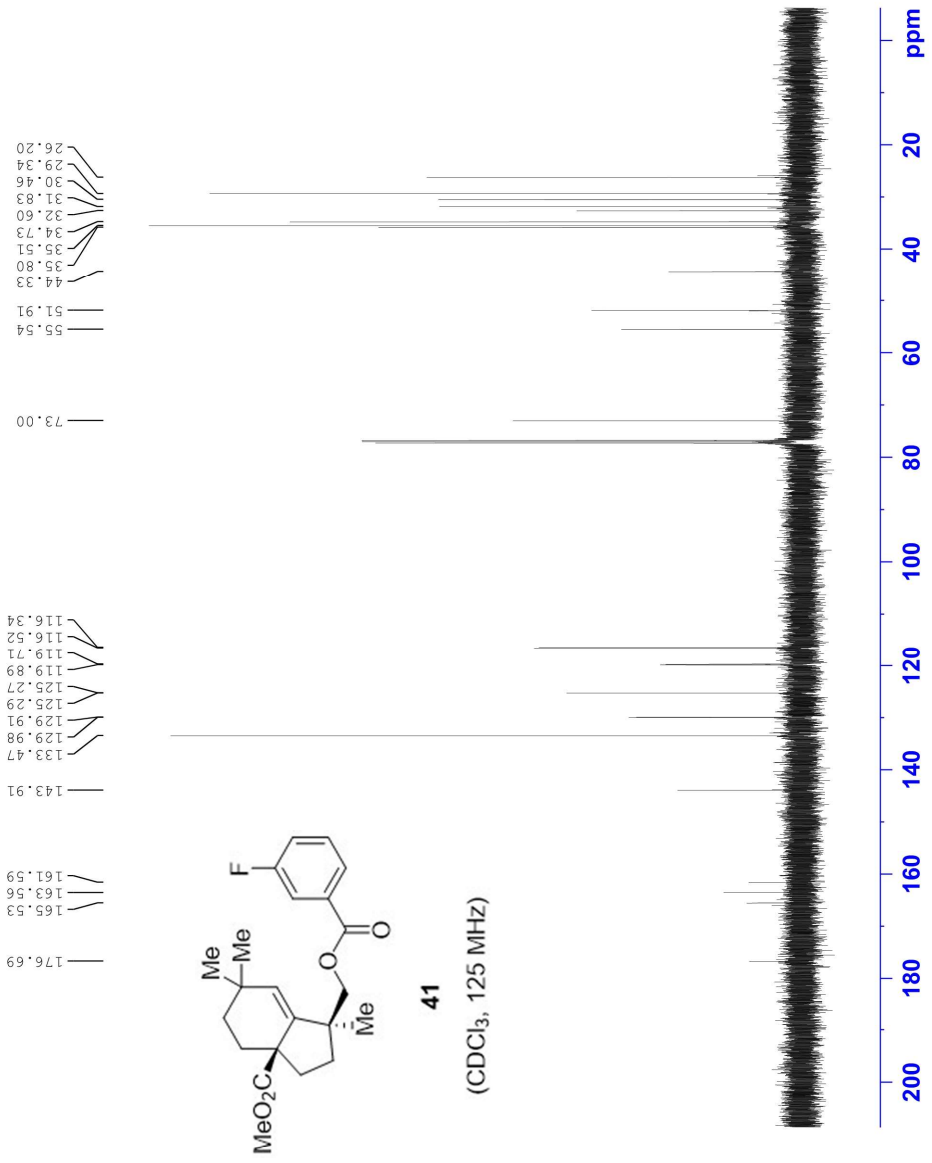
Current Data Parameters
 NAME Yh-2-68-a
 EXPNO 2
 PROCNO 2

F2 - Acquisition Parameters
 Date_ 20180821
 Time_ 17.07
 INSTRUM spect
 PROBHD 5 mm PABBO BB/
 PULPROG zgpg30
 TD 187496
 SOLVENT CDC13
 NS 115
 DS 0
 SWH 31250.000 Hz
 FIDRES 0.166670 Hz
 AQC 2.9999361 sec
 RG 2050
 DW 16.000 usec
 DE 6.50 usec
 TE 297.0 K
 D1 3.0000000 sec
 D11 0.0300000 sec
 TDO 1

==== CHANNEL f1 =====
 SFO1 125.7049802 MHz
 NUC1 13C
 P1 10.00 usec
 PLW1 72.83999634 W

==== CHANNEL f2 =====
 SFO2 499.8724993 MHz
 NUC2 1H
 CPDPRG12 waitz16
 FCPD2 80.00 usec
 PLW2 19.0000000 W
 PLW12 0.29688001 W

F2 - Processing parameters
 SI 1048576
 SF 125.6924115 MHz
 EM
 WDW 0
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.40

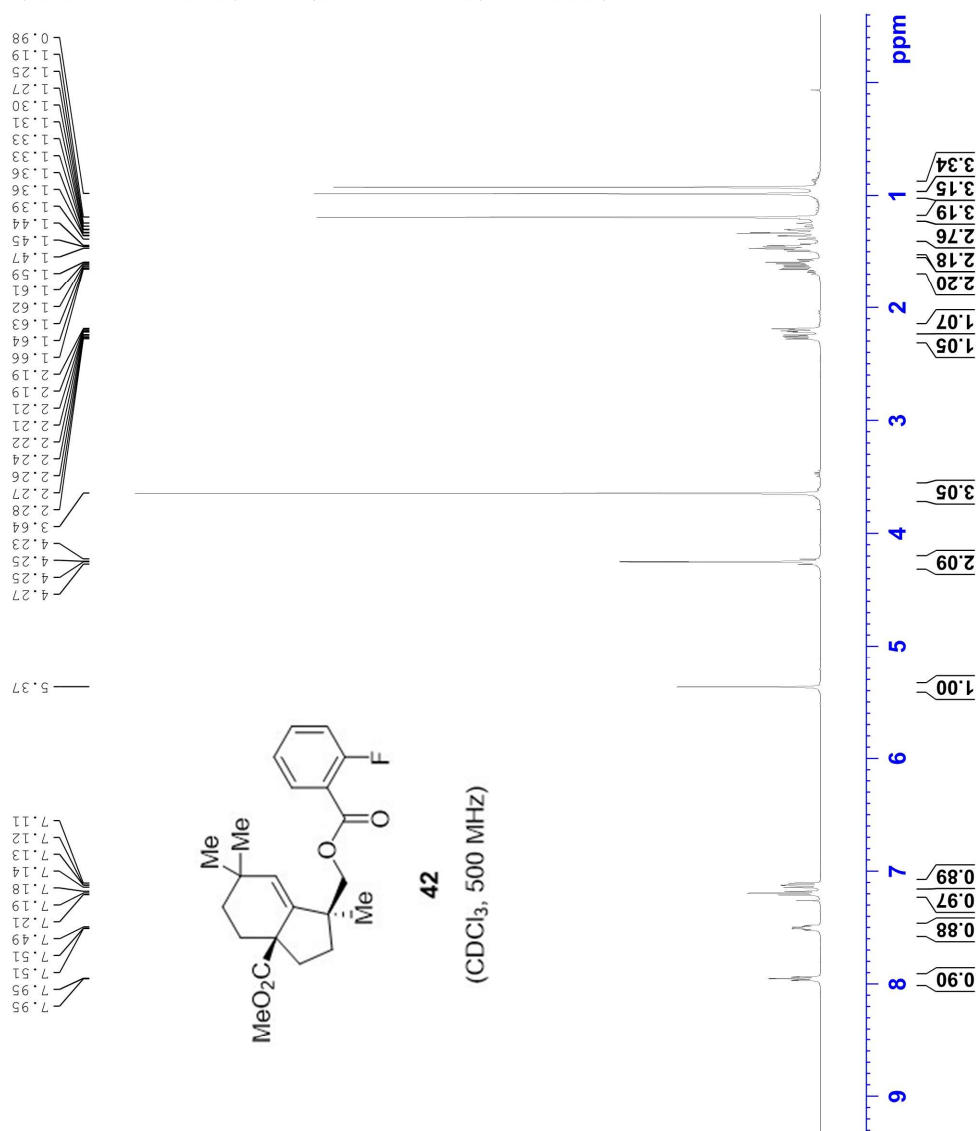


Current Data Parameters
 NAME Yh-2-34-a
 EXPNO 1
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20171218
 Time_ 21:22
 INSTRUM spect
 PROBHD 5 mm PABBO BB/
 PULPROG zg
 TD 5998
 SOLVENT CDCl3
 NS 16
 DS 0
 SWH 10000.000 Hz
 FIDRES 0.166672 Hz
 AQ 2.9999001 sec
 RG 16.71
 DW 50.000 usec
 DE 6.50 usec
 TE 296.8 K
 DL 3.0000000 sec
 TDO 1

===== CHANNEL f1 =====
 SFO1 499.8730869 MHz
 NUC1 1H
 P1 10.75 usec
 PLW1 18.25000000 W

F2 - Processing parameters
 SI 65536
 SF 499.8700122 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00



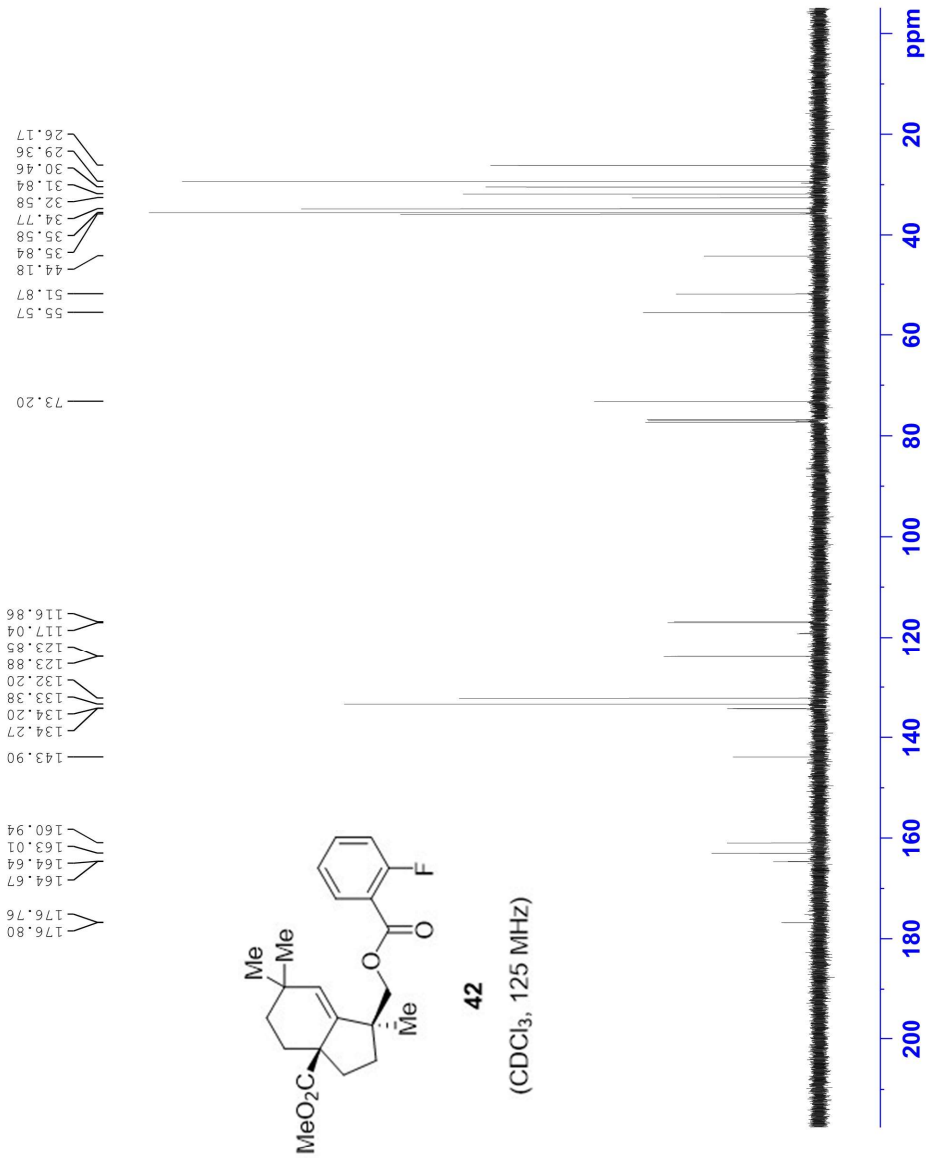
Current Data Parameters
 NAME Yh-2-34-a
 EXPNO 2
 PROCNO 2

F2 - Acquisition Parameters
 Date_ 20171218
 Time_ 21.35
 INSTRUM spect
 PROBHD 5 mm PABBO BB/
 PULPROG zgpg30
 TD 187496
 SOLVENT CDC13
 NS 52
 DS 0
 SWH 31250.000 Hz
 FIDRES 0.166670 Hz
 AQ 2.9999361 sec
 RG 2050
 DW 16.000 usec
 DE 6.50 usec
 TE 297.6 K
 D1 3.0000000 sec
 D11 0.0300000 sec
 TDO 1

==== CHANNEL f1 =====
 SFO1 125.7049802 MHz
 NUC1 13C
 P1 10.00 usec
 PLW1 72.83999634 W

==== CHANNEL f2 =====
 SFO2 499.8724993 MHz
 NUC2 1H
 CPDPRG2 waitz16
 PCPD2 80.00 usec
 PLW2 19.0000000 W
 PLWI2 0.29688001 W

F2 - Processing parameters
 SI 1048576
 SF 125.6924115 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.40

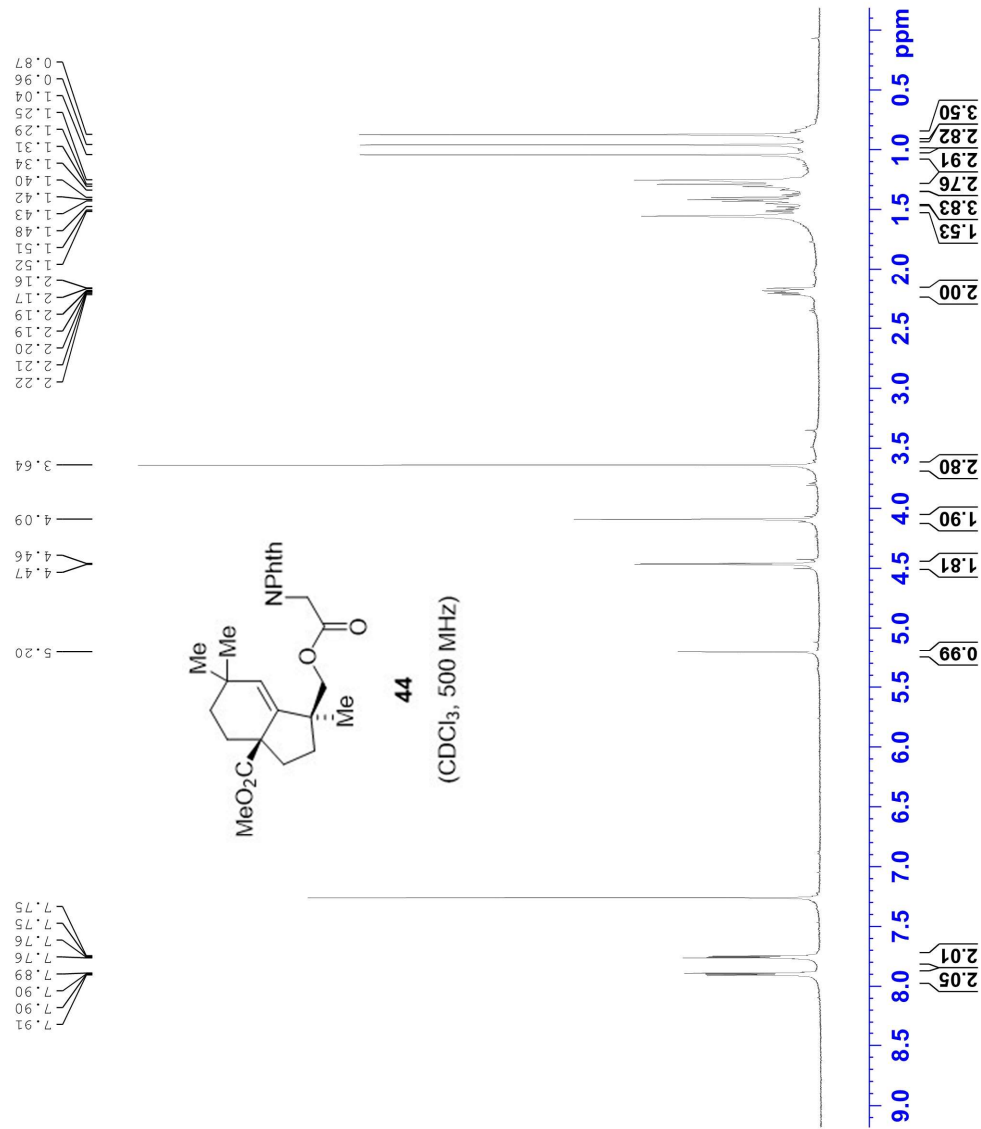


Current Data Parameters
 NAME Yh-5-55-d
 EXPNO 1
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20190712
 Time_ 21.14
 INSTRUM spect
 PROBHD 5 mm PABBO BB/
 PULPROG zg
 TD 5998
 SOLVENT CDCl3
 NS 4
 DS 0
 SWH 10000.000 Hz
 FIDRES 0.166672 Hz
 AQ 2.9999001 sec
 RG 105.66
 DW 50.000 usec
 DE 6.50 usec
 TE 297.5 K
 DL 4.0000000 sec
 TDO 1

===== CHANNEL f1 =====
 SFO1 499.8730869 MHz
 NUC1 1H
 P1 3.58 usec
 PLW1 18.25000000 W

F2 - Processing parameters
 SI 65536
 SF 499.8700124 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00



```

Current Data Parameters
NAME      Yh-5-51-a
EXPNO    2
PROCNO   2

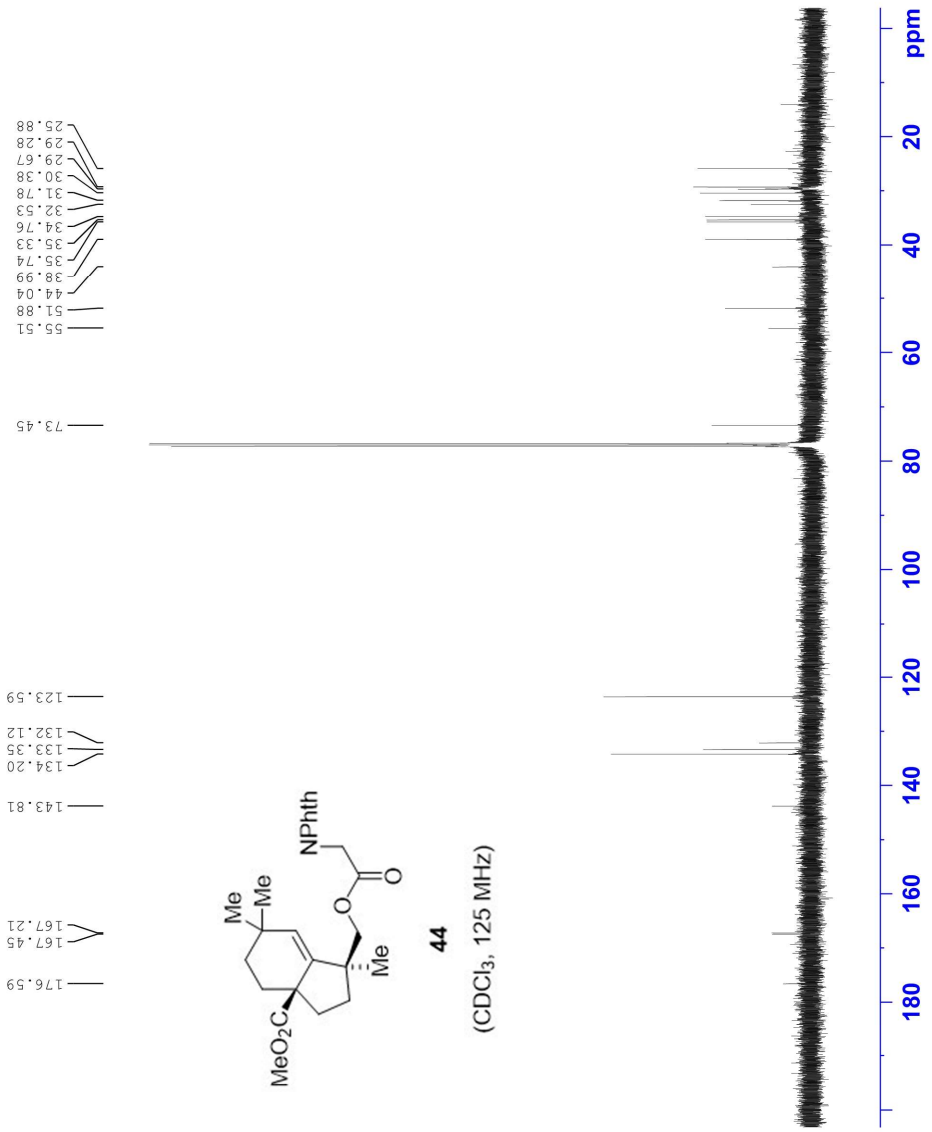
F2 - Acquisition Parameters
Date_     20180926
Time      23.03
INSTRUM   spect
PROBHD    5 mm PABBO BB/
PULPROG   zgpg
TD         187496
SOLVENT   CDCl3
NS         1083
DS         0
SWH        31250.000 Hz
FIDRES     0.166670 Hz
AQ         2.9999361 sec
RG         2050
DW         16.000 usec
DE         6.50 usec
TE         301.2 K
D1         3.00000000 sec
D11        0.03000000 sec
TDO        1

===== CHANNEL f1 =====
SFO1      125.7049802 MHz
NUC1       13C
P1         10.00 usec
PLW1      72.83999634 W

===== CHANNEL f2 =====
SFO2      499.8724993 MHz
NUC2       1H
CFPRG12   waitz16
PCPD2     80.00 usec
PLW2      19.00000000 W
PLW12     0.29688001 W

F2 - Processing parameters
SI         1048576
SF         125.6924115 MHz
WDW        EM
SSB        0
LB         0.30 Hz
GB         0
PC         1.40

```

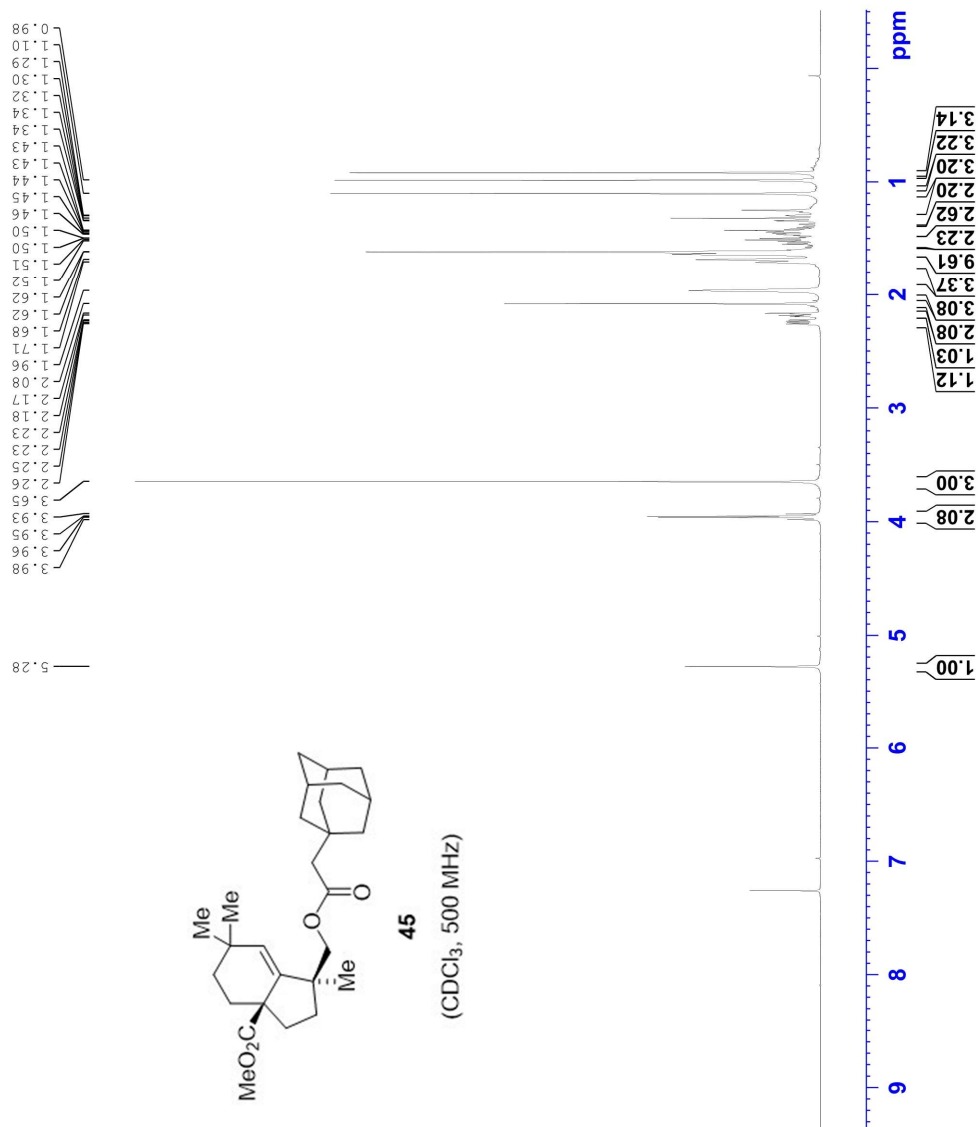


Current Data Parameters
 NAME Yh-2-25-a
 EXPNO 1
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20171214
 Time_ 21.05
 INSTRUM spect
 PROBHD 5 mm PABBO BB/
 PULPROG zg
 TD 5998
 SOLVENT CDCl3
 NS 16
 DS 0
 SWH 10000.000 Hz
 FIDRES 0.166672 Hz
 AQ 2.9999001 sec
 RG 28.76
 DW 50.000 usec
 DE 6.50 usec
 TE 296.9 K
 DL 3.00000000 sec
 TDO 1

===== CHANNEL f1 =====
 SFO1 499.8730869 MHz
 NUC1 1H
 PL 10.75 usec
 PLW1 18.25000000 W

F2 - Processing parameters
 SI 65536
 SF 499.8700123 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00



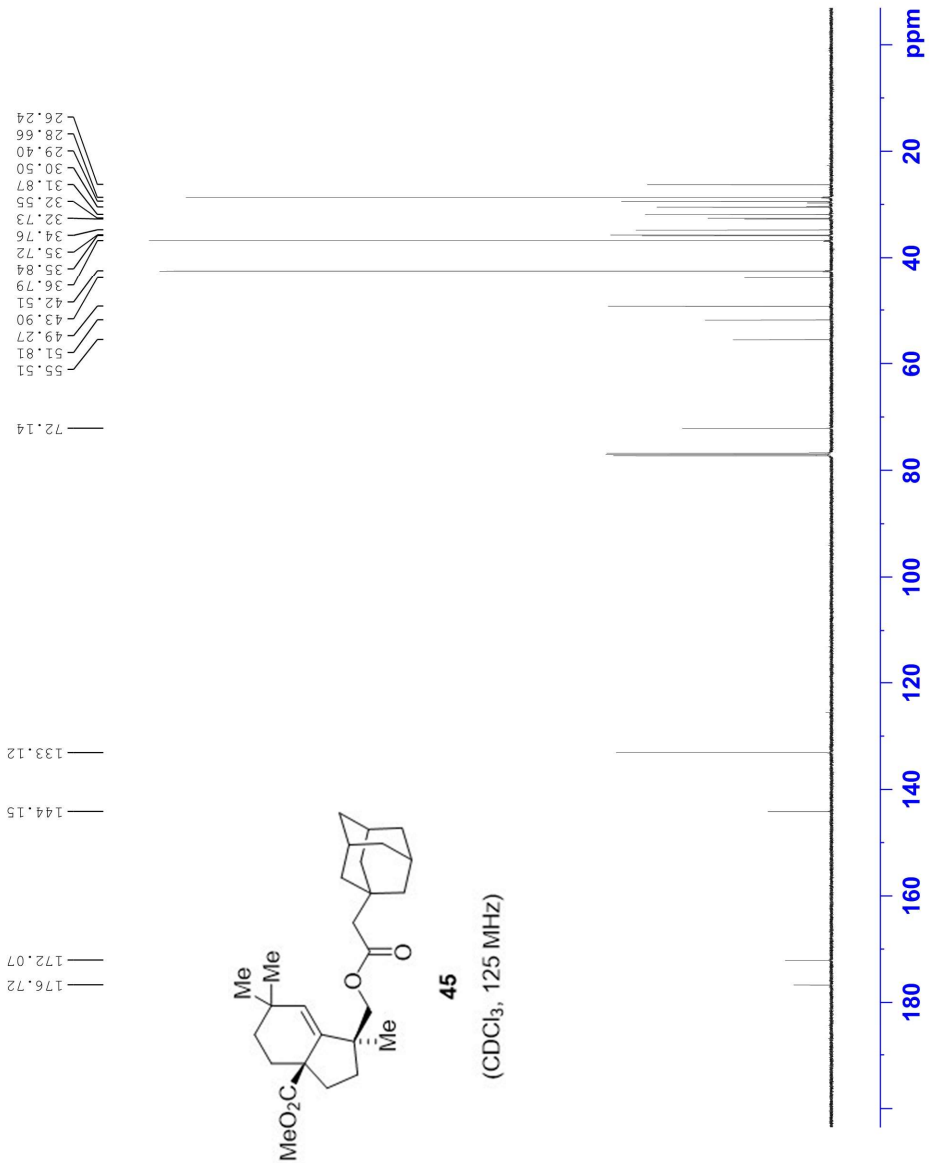
Current Data Parameters
 NAME Yh-2-25-a
 EXPNO 2
 PROCNO 2

F2 - Acquisition Parameters
 Date_ 20171214
 Time_ 22.48
 INSTRUM spect
 PROBHD 5 mm PABBO BB/
 PULPROG zgpgc
 TD 187496
 SOLVENT CDCl3
 NS 978
 DS 0
 SWH 31250.000 Hz
 FIDRES 0.166670 Hz
 AQC 2.9999361 sec
 RG 2050
 DW 16.000 usec
 DE 6.50 usec
 TE 298.9 K
 D1 3.0000000 sec
 D11 0.0300000 sec
 TDO 1

==== CHANNEL f1 =====
 SFO1 125.7049802 MHz
 NUC1 13C
 P1 10.00 usec
 PLW1 72.83999634 W

==== CHANNEL f2 =====
 SFO2 499.8724993 MHz
 NUC2 1H
 CPDPRG2 waitz16
 PCPD2 80.00 usec
 PLW2 19.0000000 W
 PLW12 0.29688001 W

F2 - Processing parameters
 SI 1048576
 SF 125.6924115 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.40

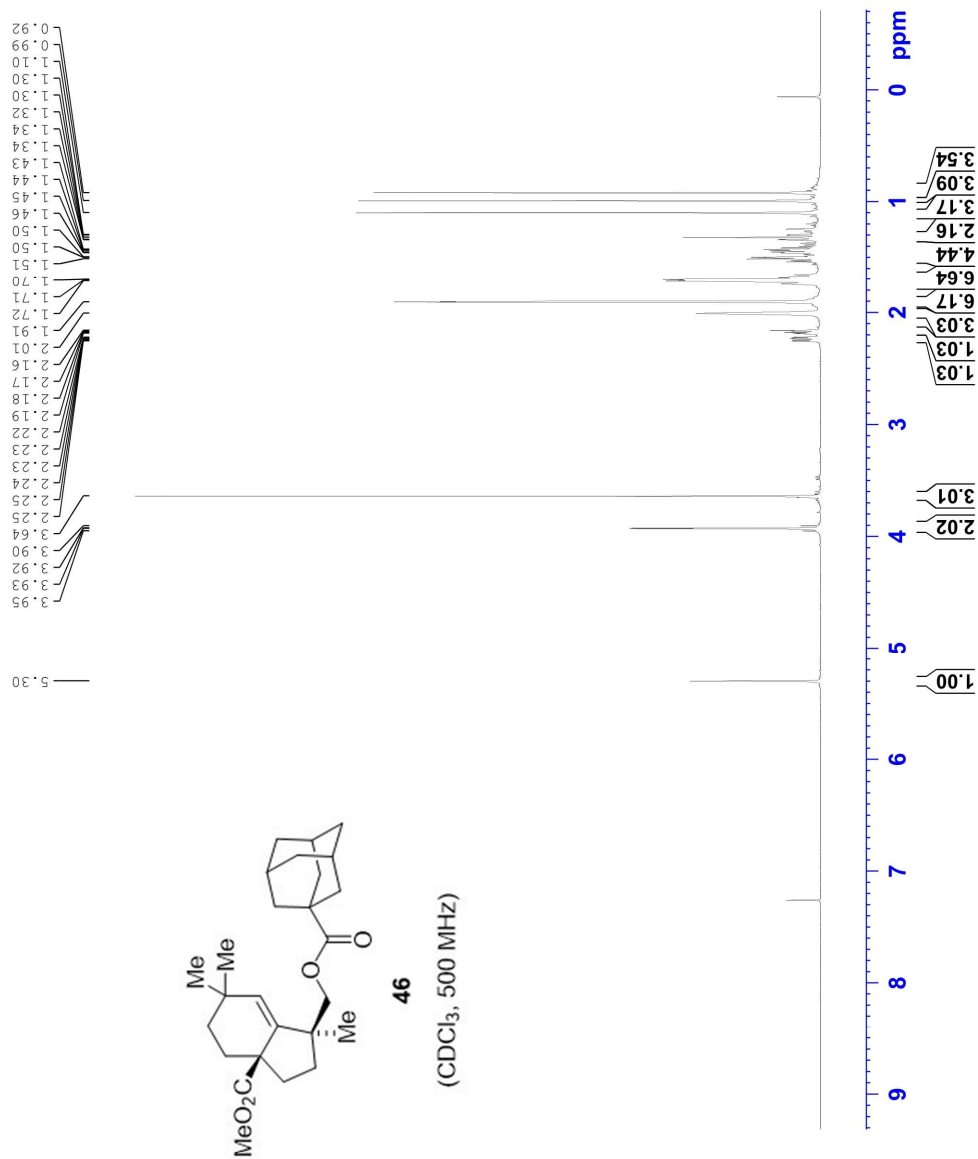


Current Data Parameters
 NAME Yh-2-73-a
 EXPNO 1
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20180121
 Time_ 22.32
 INSTRUM spect
 PROBHD 5 mm PABBO BB/
 PULPROG zg
 TD 5998
 SOLVENT CDCl3
 NS 2
 DS 0
 SWH 10000.000 Hz
 FIDRES 0.166672 Hz
 AQ 2.9999001 sec
 RG 19.64
 DW 50.000 usec
 DE 6.50 usec
 TE 296.8 K
 DL 3.0000000 sec
 TDO 1

===== CHANNEL f1 =====
 SFO1 499.8730869 MHz
 NUC1 1H
 PL 10.75 usec
 PLW1 18.25000000 W

F2 - Processing parameters
 SI 65536
 SF 499.8700124 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00



```

Current Data Parameters
NAME      Yh-2-73-a
EXPNO    2
PROCNO   2

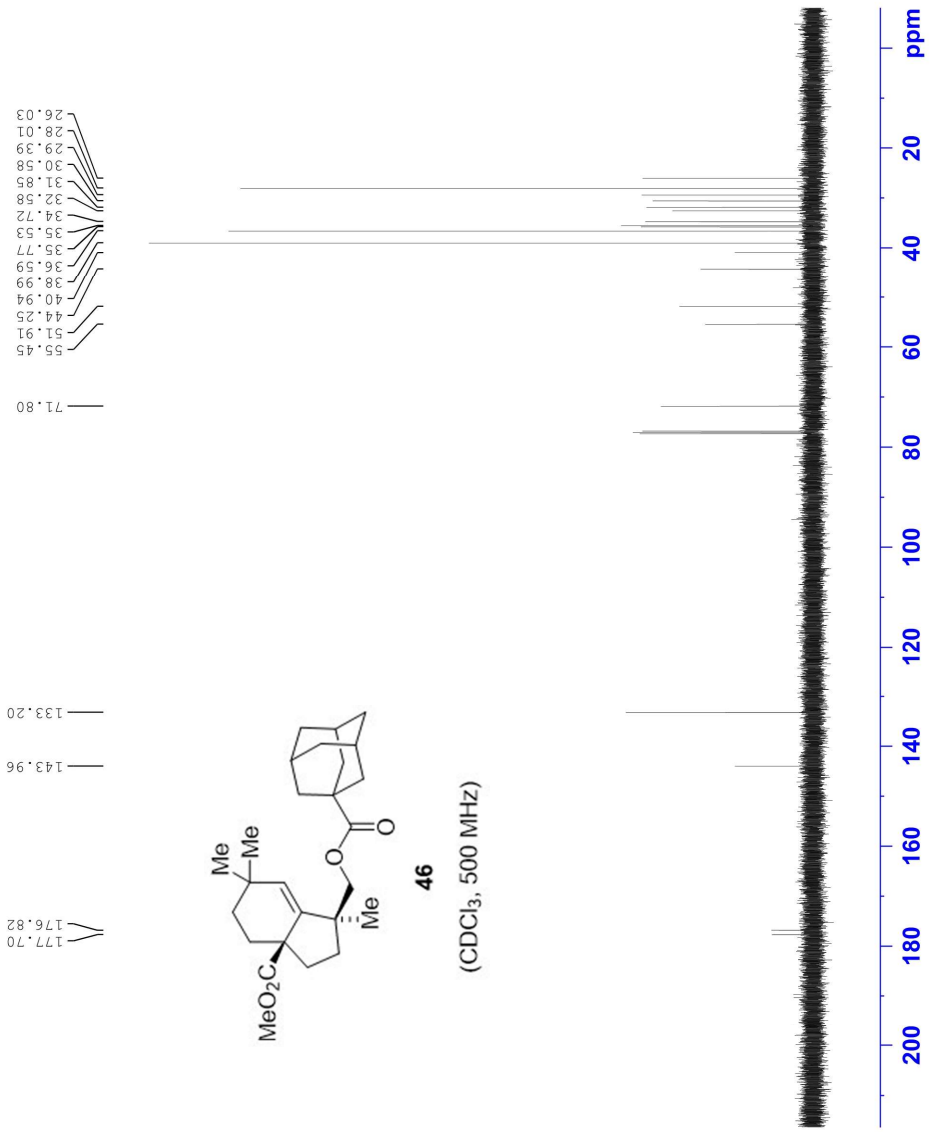
F2 - Acquisition Parameters
Date_     20180121
Time      22.35
INSTRUM   spect
PROBHD    5 mm PABBO BB/
PULPROG   zgpg
TD         187496
SOLVENT   CDCl3
NS         7
DS         0
SWH        31250.000 Hz
FIDRES     0.166670 Hz
AQ         2.9999361 sec
RG         2050
DW         16.000 usec
DE         6.50 usec
TE         296.9 K
D1         3.0000000 sec
D11        0.0300000 sec
TDO        1

===== CHANNEL f1 =====
SFO1      125.7049802 MHz
NUC1      13C
P1         10.00 usec
PLW1      72.83999634 W

===== CHANNEL f2 =====
SFO2      499.8724993 MHz
NUC2      1H
CFDPRG12 waitz16
PCPD2     80.00 usec
PLW2      19.0000000 W
PLW12     0.29688001 W

F2 - Processing parameters
SI         1048576
SF         125.6924115 MHz
WDW        EM
SSB        0
LB         0.30 Hz
GB         0
PC         1.40

```

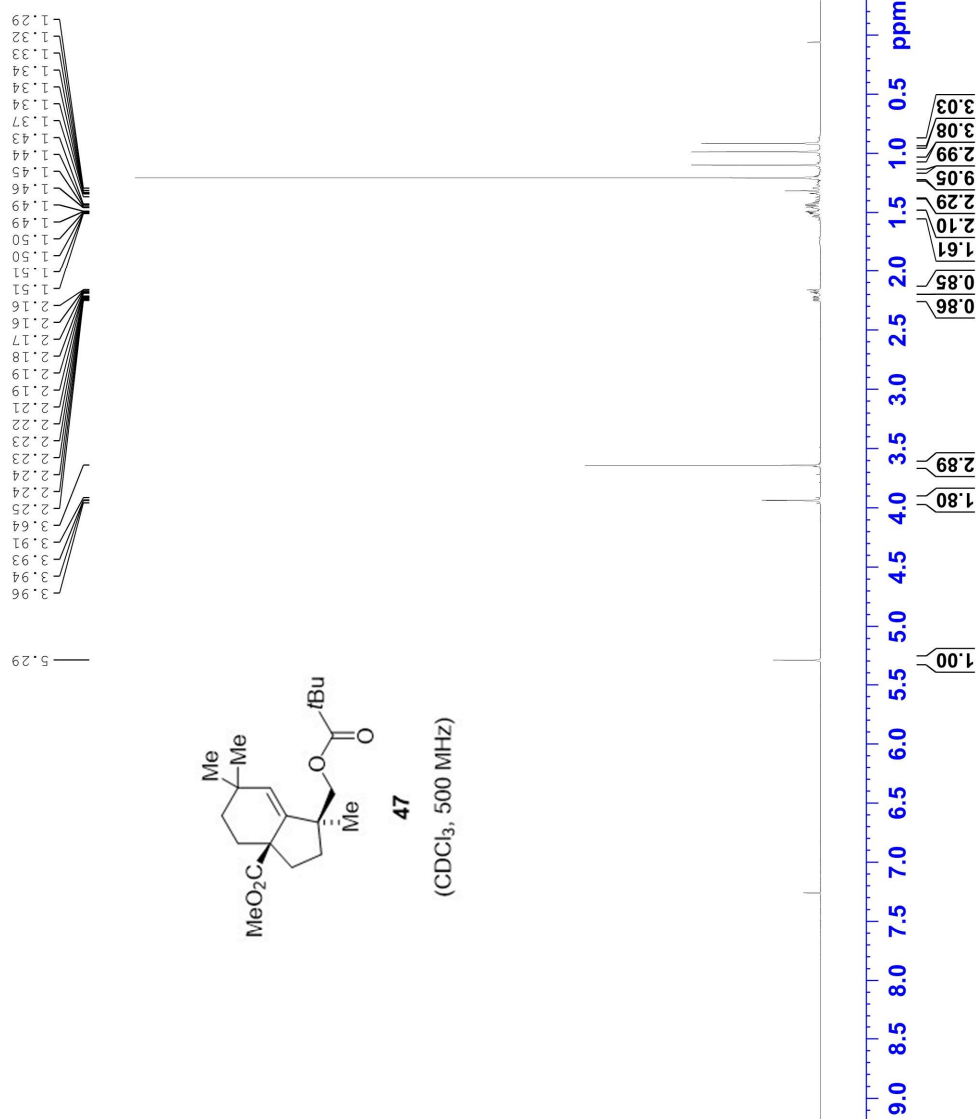


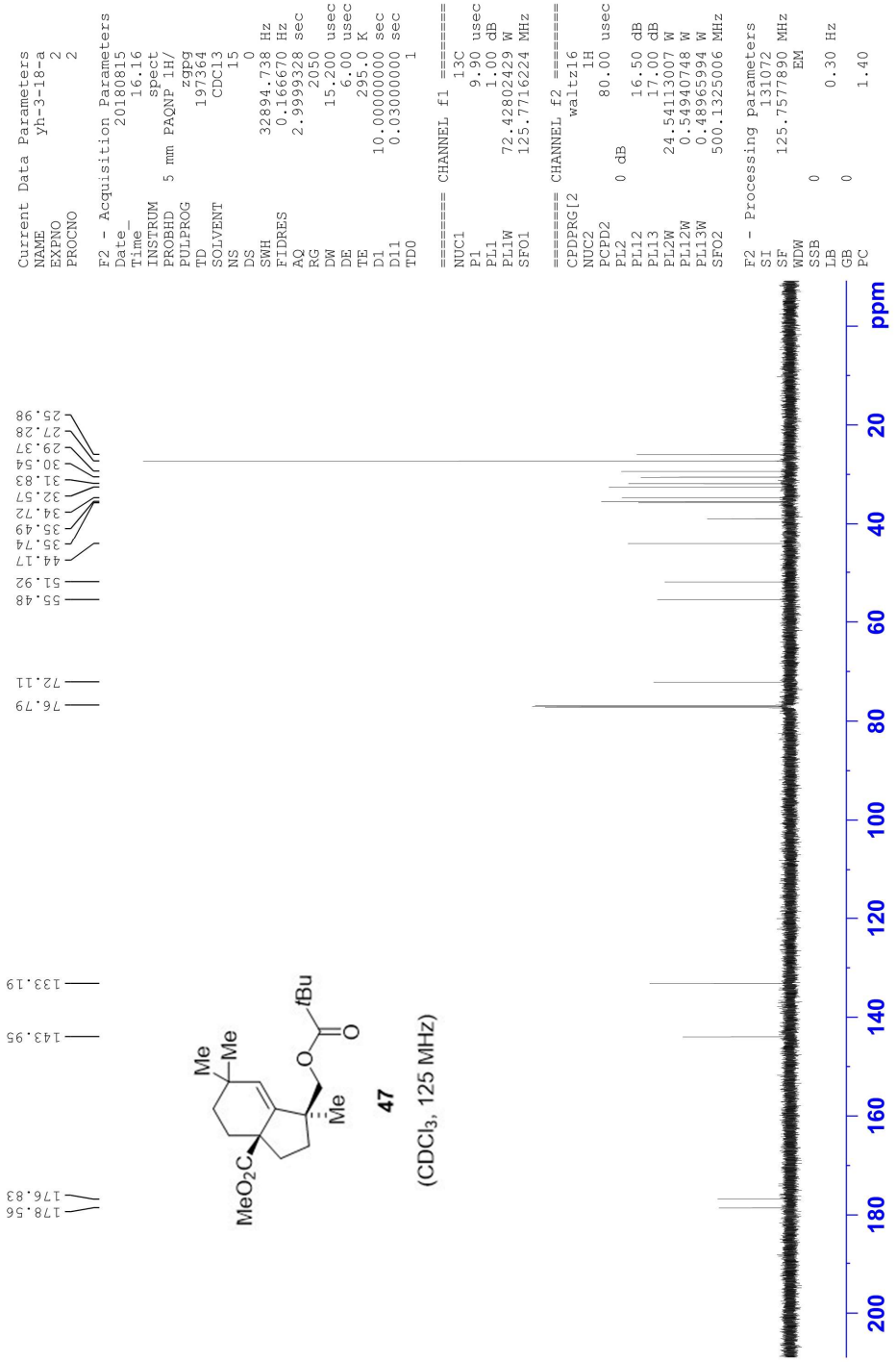
Current Data Parameters
 NAME Yh-3-18-a
 EXPNO 1
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20180815
 Time_ 16.03
 INSTRUM spect
 PROBHD 5 mm PAQNP 1H/
 PULPROG zg
 TD 4498
 SOLVENT CDCl3
 NS 1
 DS 0
 SWH 7500.000 Hz
 FIDRES 0.166674 Hz
 AQ 2.9998667 sec
 RG 50.8
 DW 66.667 usec
 DE 71.43 usec
 TE 294.5 K
 DL 3.0000000 sec
 TDO 1

===== CHANNEL f1 =====
 NUC1 1H
 P1 14.50 usec
 PL1 0 dB
 PL1W 24.54113007 W
 SF01 500.1330008 MHz

F2 - Processing parameters
 SI 32768
 SF 500.1300138 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00





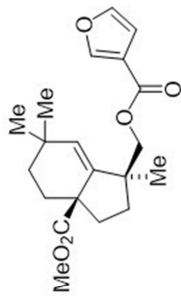
Current Data Parameters
 NAME yh-2-143-a
 EXPNO 1
 PROCNO 1

F2 - Acquisition Parameters
 Date 20180419
 Time 20.41
 INSTRUM spect
 PROBHD 5 mm PAQNP 1H/
 PULPROG zg
 TD 4498
 SOLVENT CDCl3
 NS 8
 DS 0
 SWH 7500.000 Hz
 FIDRES 0.166674 Hz
 AQ 2.9998667 sec
 RG 181
 DW 66.667 usec
 DE 71.43 usec
 TE 295.1 K
 DL 3.0000000 sec
 TDO 1

===== CHANNEL f1 =====
 NUC1 1H
 P1 12.00 usec
 PL1 0 dB
 PL1W 24.54113007 W
 SFO1 500.1330008 MHz

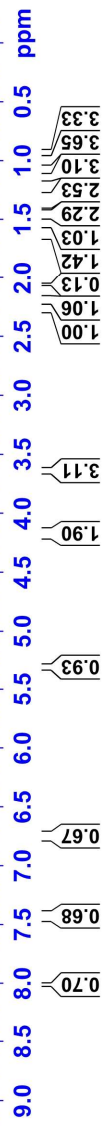
F2 - Processing parameters
 SI 16384
 SF 500.1300133 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00

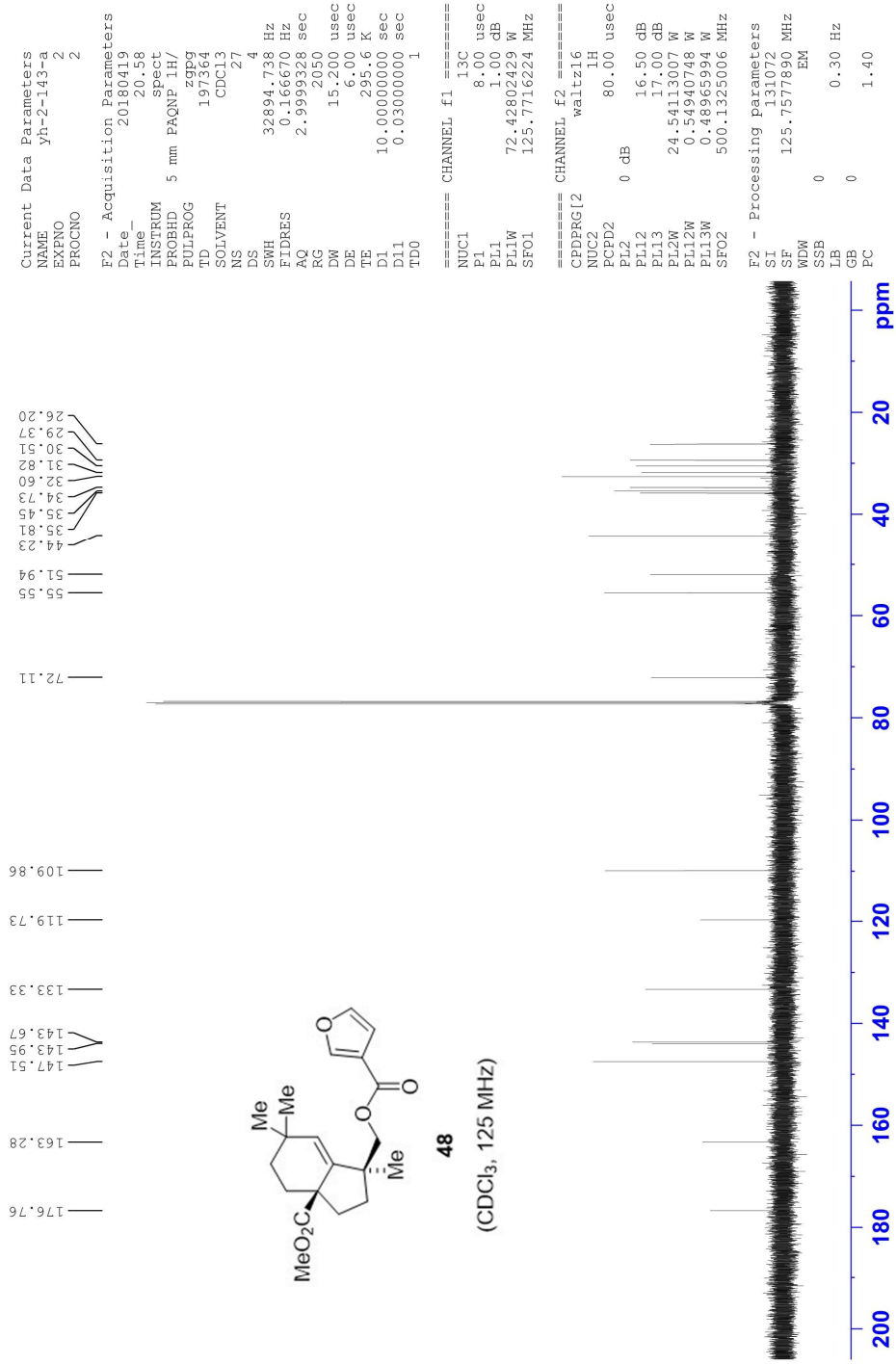
8.01
7.43
6.75
5.32
4.20
4.18
4.17
4.15
3.65
3.28
2.27
2.26
2.25
2.21
2.21
2.19
1.63
1.62
1.60
1.59
1.56
1.55
1.52
1.49
1.48
1.47
1.46
1.46
1.45
1.45
1.44
1.36
1.36
1.36
1.33
1.31
1.31



48

(CDCl₃, 500 MHz)





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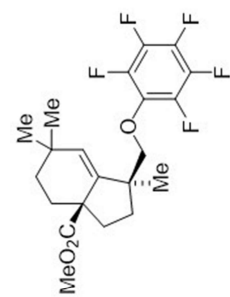
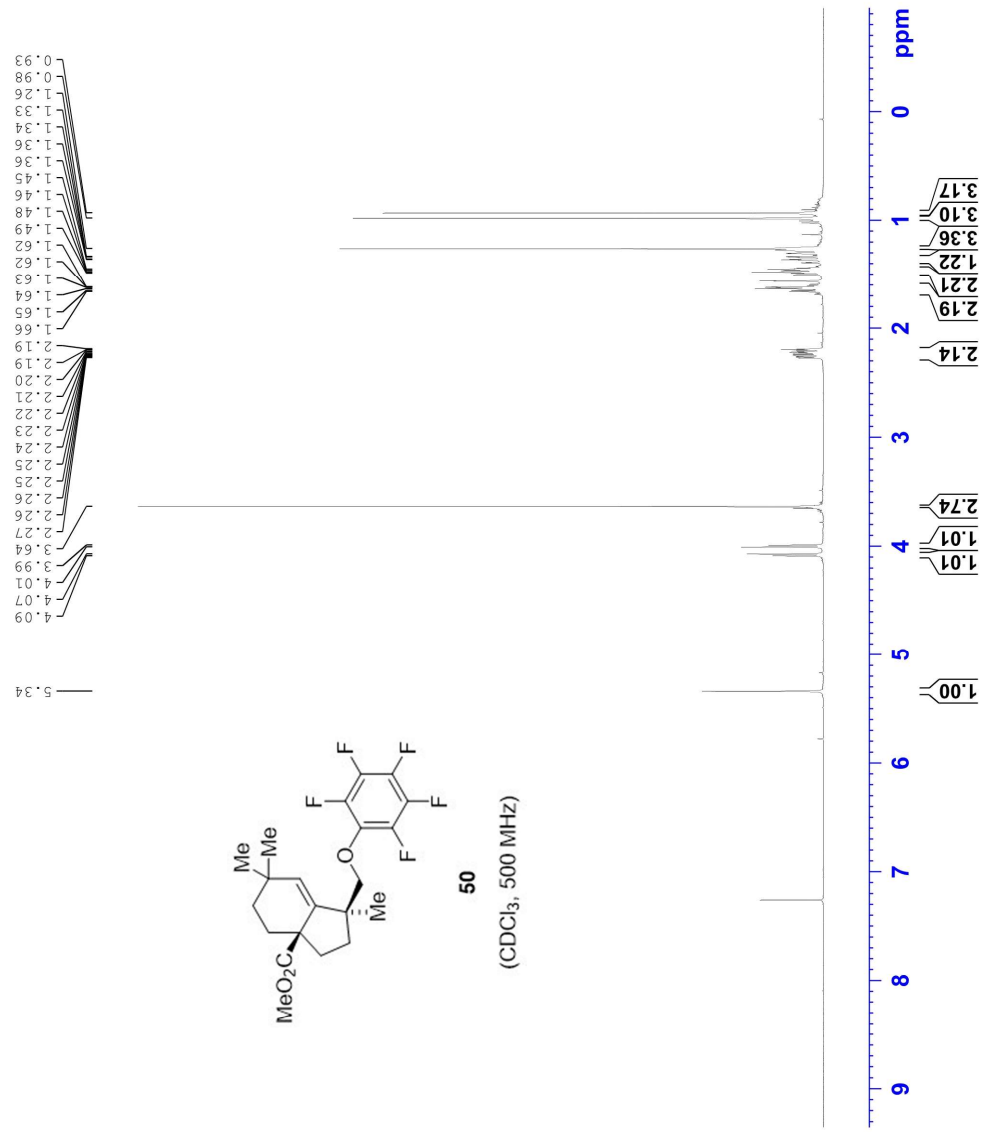
Current Data Parameters
NAME      Yh-2-63-a
EXPNO    1
PROCNO   1

F2 - Acquisition Parameters
Date_    20180110
Time     21.27
INSTRUM  spect
PROBHD   5 mm PABBO BB/
PULPROG  zg
TD        59998
SOLVENT  CDCl3
NS        2
DS        0
SWH       10000.000 Hz
FIDRES    0.166672 Hz
AQ        2.9999001 sec
RG        37.92
DW        50.000 usec
DE        6.50 usec
TE        296.7 K
DL        3.0000000 sec
TD0       1

===== CHANNEL f1 =====
SFO1     499.8730869 MHz
NUC1      1H
P1        10.75 usec
PLW1     18.25000000 W

F2 - Processing parameters
SI        65536
SF        499.8700123 MHz
WDW       EM
SSB       0
LB        0.30 Hz
GB        0
PC        1.00

```



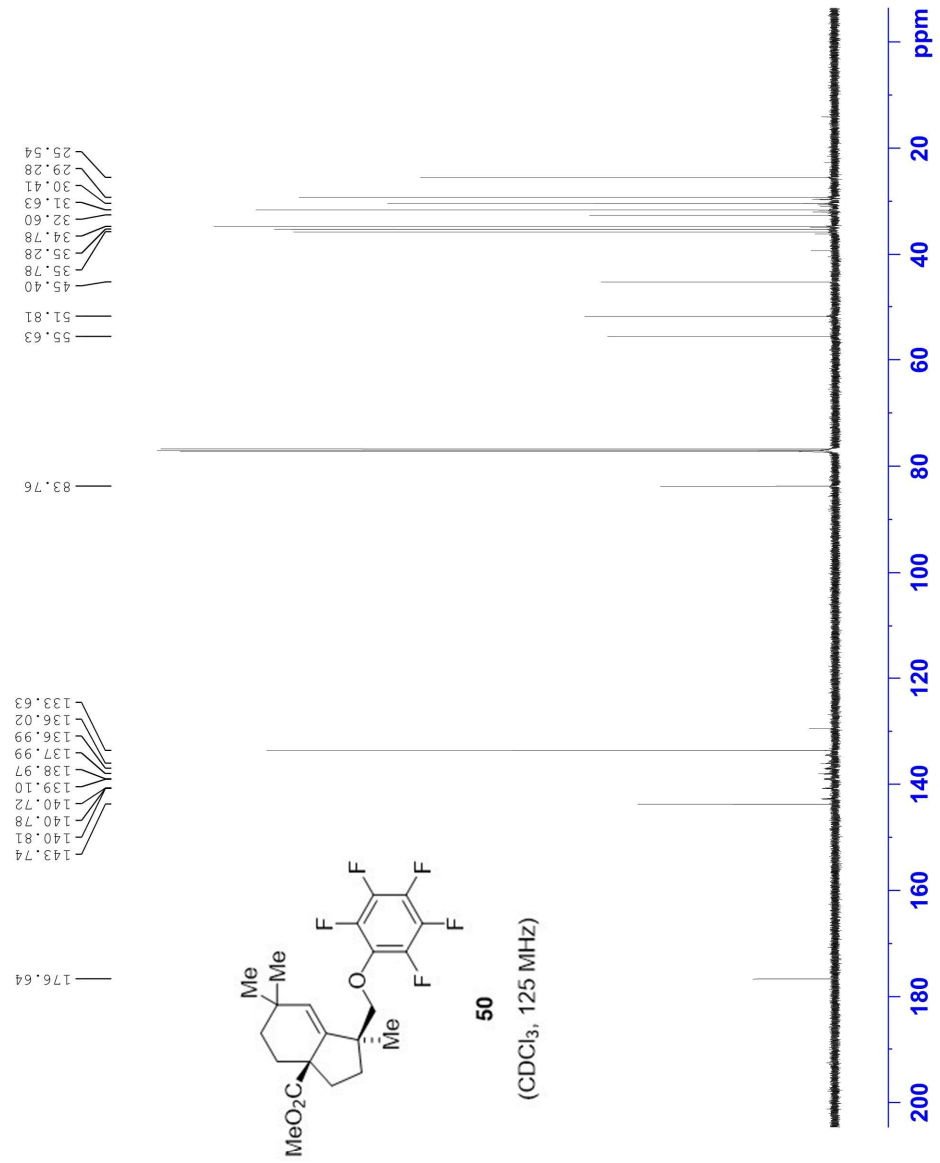
Current Data Parameters
 NAME Yh-2-63-a
 EXPNO 2
 PROCNO 2

F2 - Acquisition Parameters
 Date_ 20180111
 Time_ 0.59
 INSTRUM spect
 PROBHD 5 mm PABBO BB/
 PULPROG zgpg
 TD 187496
 SOLVENT CDC13
 NS 2048
 DS 0
 SWH 31250.000 Hz
 FIDRES 0.166670 Hz
 AQ 2.9999361 sec
 RG 2050
 DW 16.000 usec
 DE 6.50 usec
 TE 298.7 K
 D1 3.0000000 sec
 D11 0.0300000 sec
 TDO 1

==== CHANNEL f1 =====
 SFO1 125.7049802 MHz
 NUC1 13C
 P1 10.00 usec
 PLW1 72.83999634 W

==== CHANNEL f2 =====
 SFO2 499.8724993 MHz
 NUC2 1H
 CPDPRG12 waitz16
 FCPD2 80.00 usec
 PLW2 19.0000000 W
 PLW12 0.29688001 W

F2 - Processing parameters
 SI 1048576
 SF 125.6924115 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.40

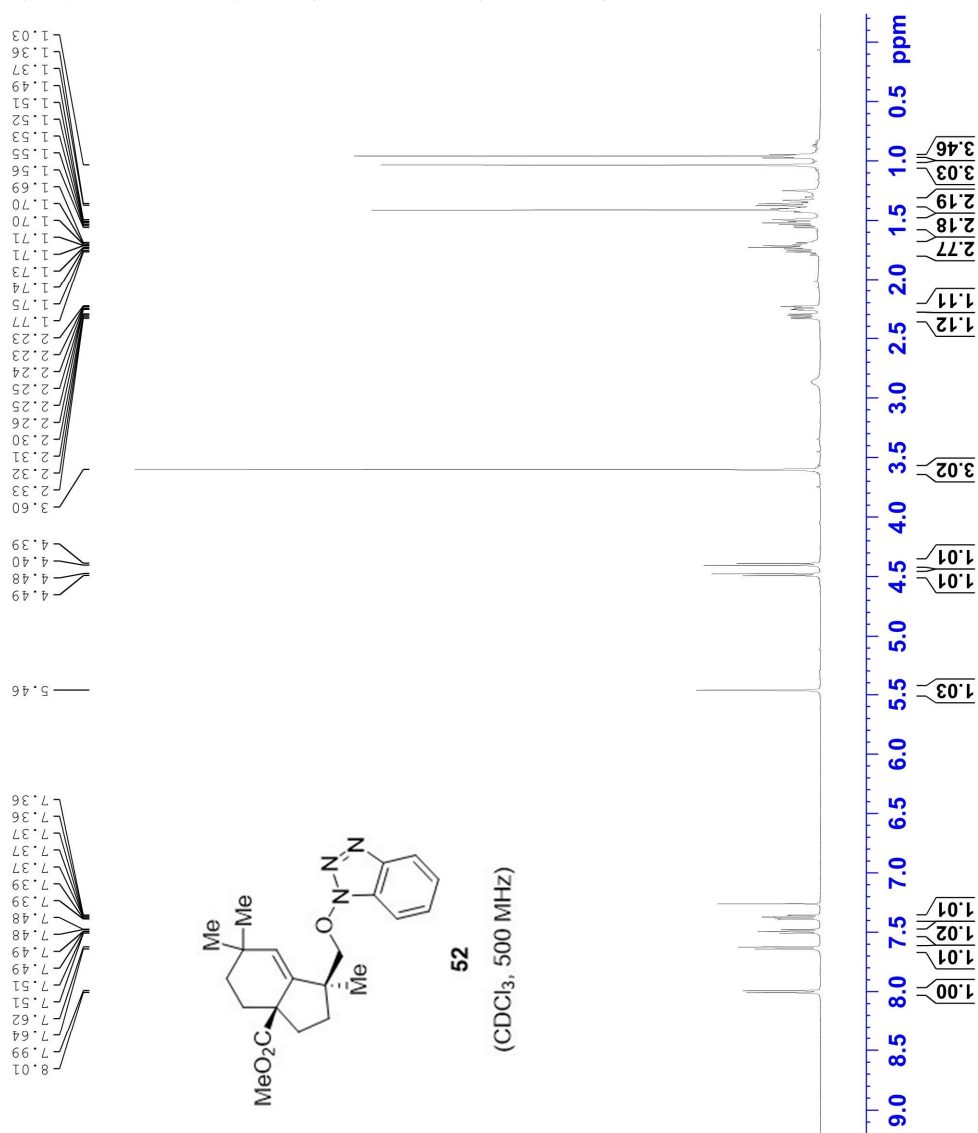


Current Data Parameters
 NAME Yh-7-27-B
 EXPNO 1
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20190930
 Time_ 15.25
 INSTRUM spect
 PROBHD 5 mm PABBO BB/
 PULPROG zg
 TD 5998
 SOLVENT CDCl3
 NS 4
 DS 0
 SWH 10000.000 Hz
 FIDRES 0.166672 Hz
 AQC 2.9999001 sec
 RG 70.49
 DW 50.000 usec
 DE 6.50 usec
 TE 298.0 K
 DL 4.0000000 sec
 TDO 1

===== CHANNEL f1 =====
 SFO1 499.8730869 MHz
 NUC1 1H
 P1 3.58 usec
 PLW1 18.25000000 W

F2 - Processing parameters
 SI 65536
 SF 499.8700109 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00



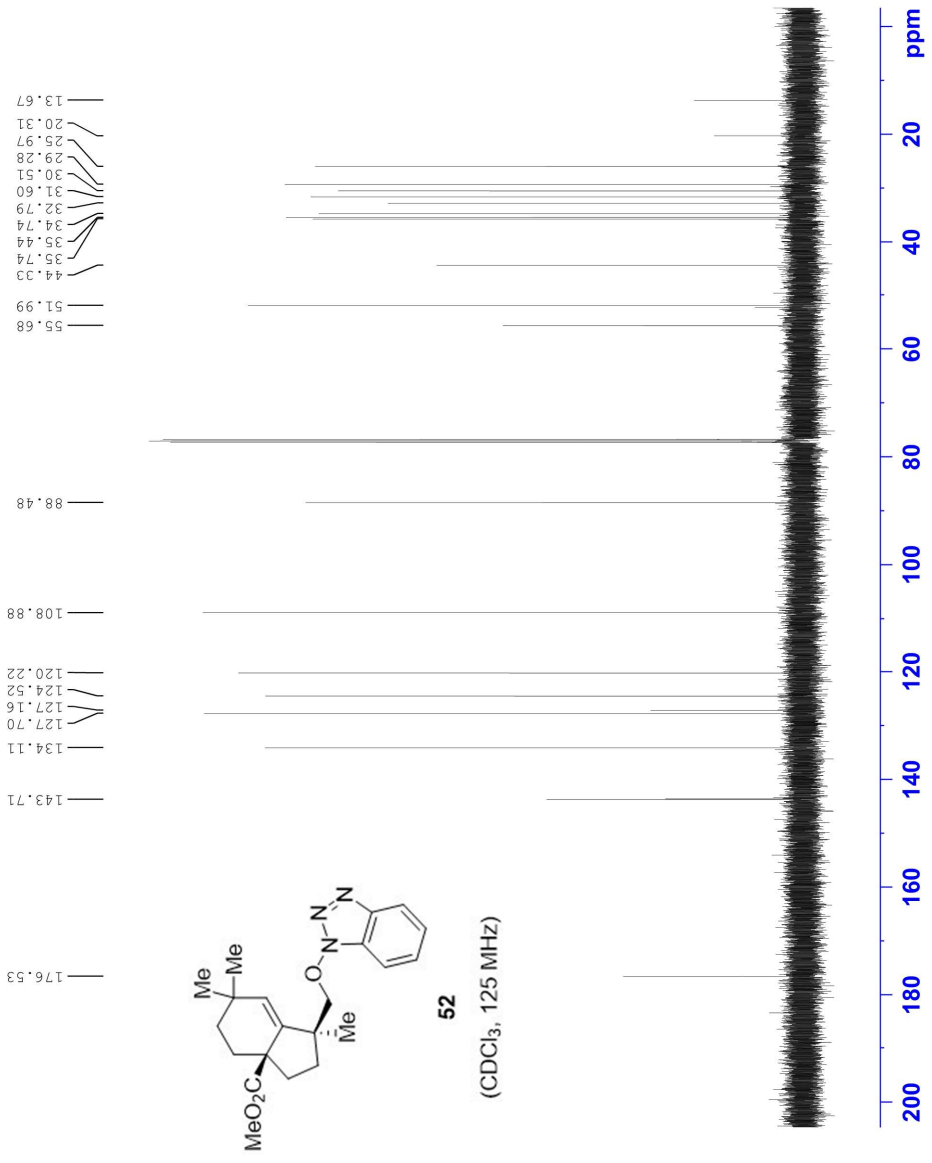
Current Data Parameters
 NAME Yh-7-27-B
 EXPNO 2
 PROCNO 2

F2 - Acquisition Parameters
 Date_ 20190930
 Time_ 16.10
 INSTRUM spect
 PROBHD 5 mm PABBO BB/
 PULPROG zgpg30
 TD 187496
 SOLVENT CDCl3
 NS 67
 DS 0
 SWH 31250.000 Hz
 FIDRES 0.166670 Hz
 AQ 2.9999361 sec
 RG 2050
 DW 16.000 usec
 DE 6.50 usec
 TE 298.0 K
 D1 3.0000000 sec
 D11 0.0300000 sec
 TDO 1

==== CHANNEL f1 =====
 SFO1 125.7049802 MHz
 NUC1 13C
 P1 10.00 usec
 PLW1 72.83999634 W

==== CHANNEL f2 =====
 SFO2 499.8724993 MHz
 NUC2 1H
 CPDPRG2 waitz16
 FCPD2 80.00 usec
 PLW2 19.0000000 W
 PLW12 0.29688001 W

F2 - Processing parameters
 SI 1048576
 SF 125.6924115 MHz
 EM
 WDW 0
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.40

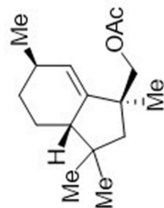
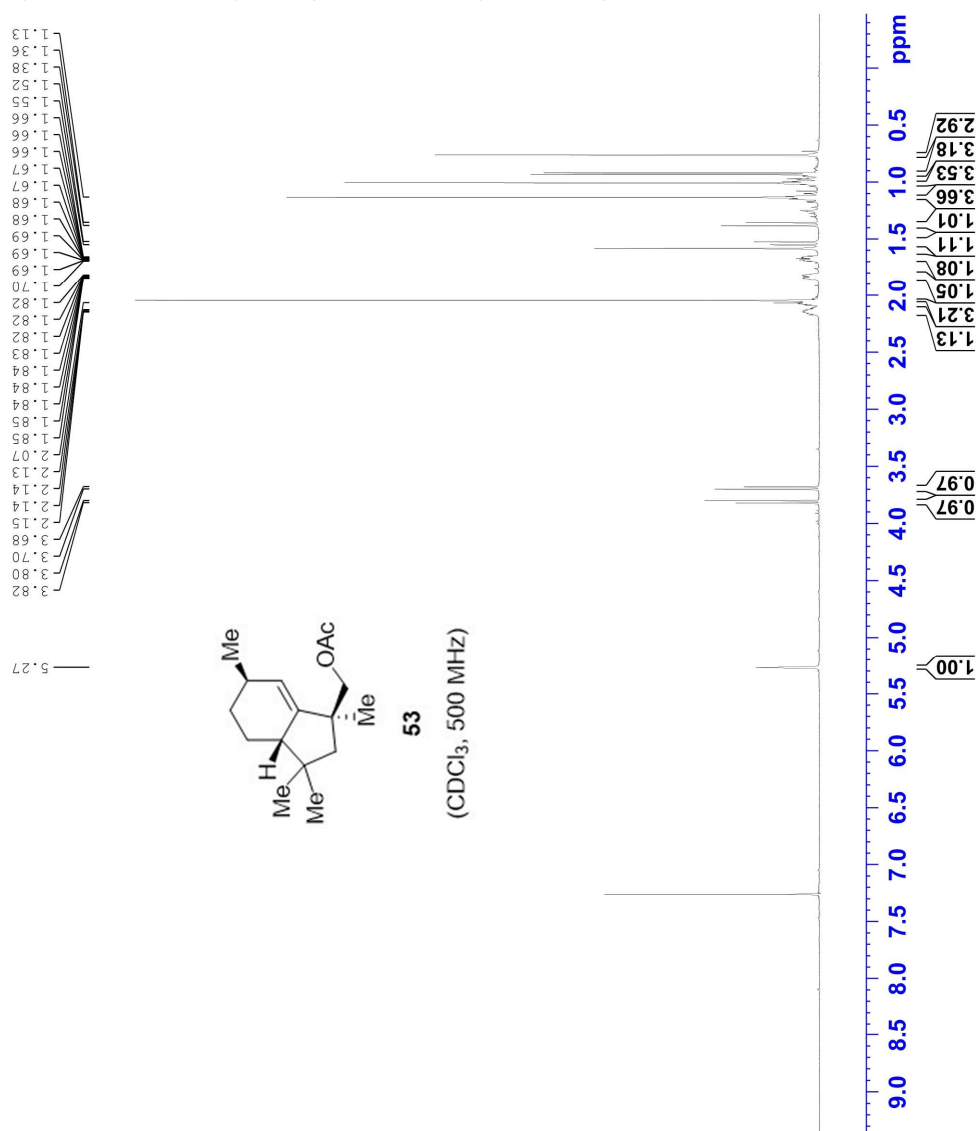


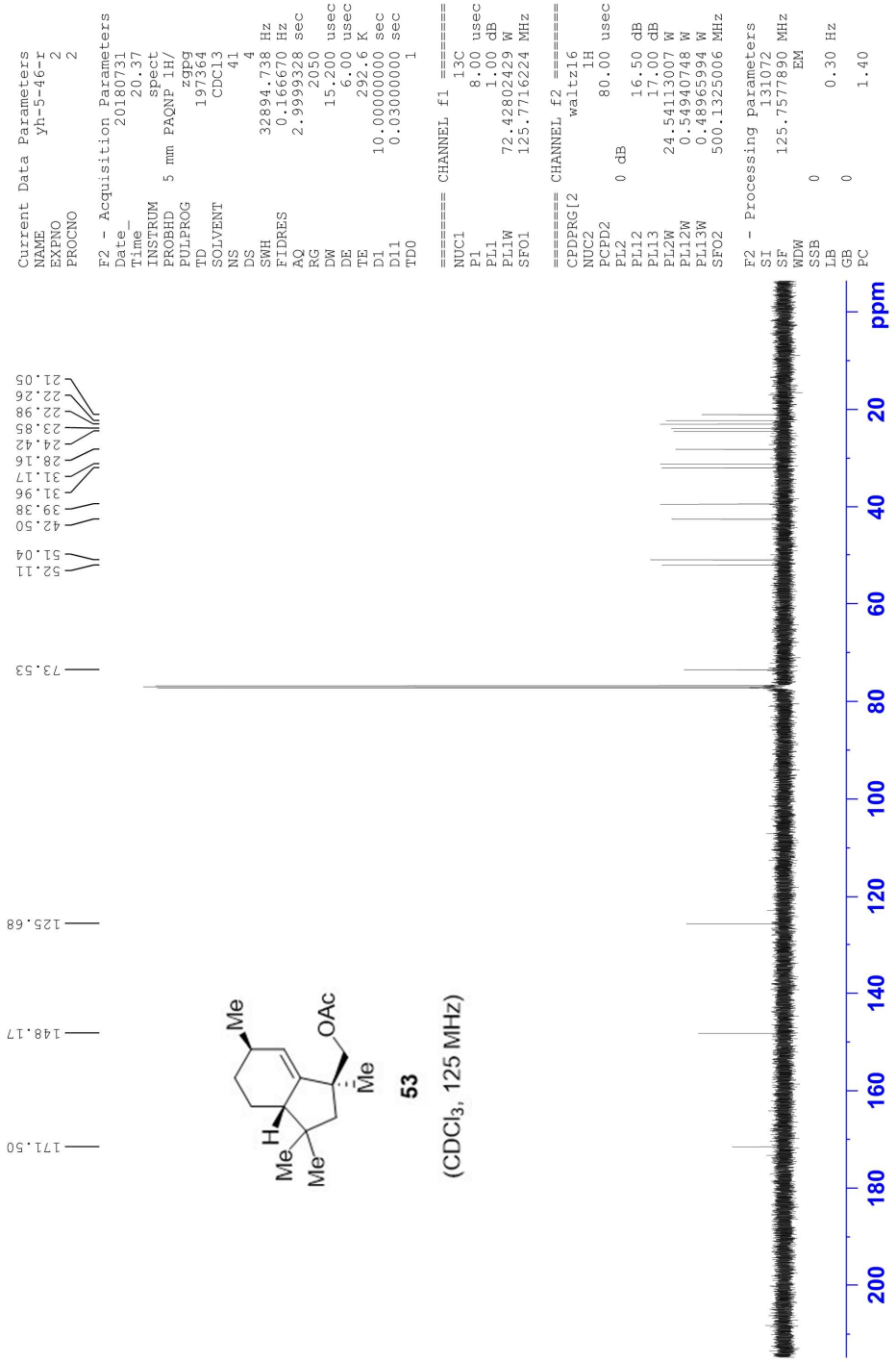
Current Data Parameters
 NAME Yh-5-46-f
 EXPNO 1
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20180731
 Time_ 20.07
 INSTRUM spect
 PROBHD 5 mm PATXI 1H/
 PULPROG zg
 TD 5998
 SOLVENT CDCl3
 NS 3
 DS 0
 SWH 10000.000 Hz
 FIDRES 0.166672 Hz
 AQ 2.9999001 sec
 RG 196.79
 DW 50.000 usec
 DE 10.00 usec
 TE 294.1 K
 DL 3.0000000 sec
 TDO 1

===== CHANNEL f1 =====
 SFO1 500.1330885 MHz
 NUC1 1H
 PL1 2.67 usec
 PLW1 12.19999981 W

F2 - Processing parameters
 SI 65536
 SF 500.1300138 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00



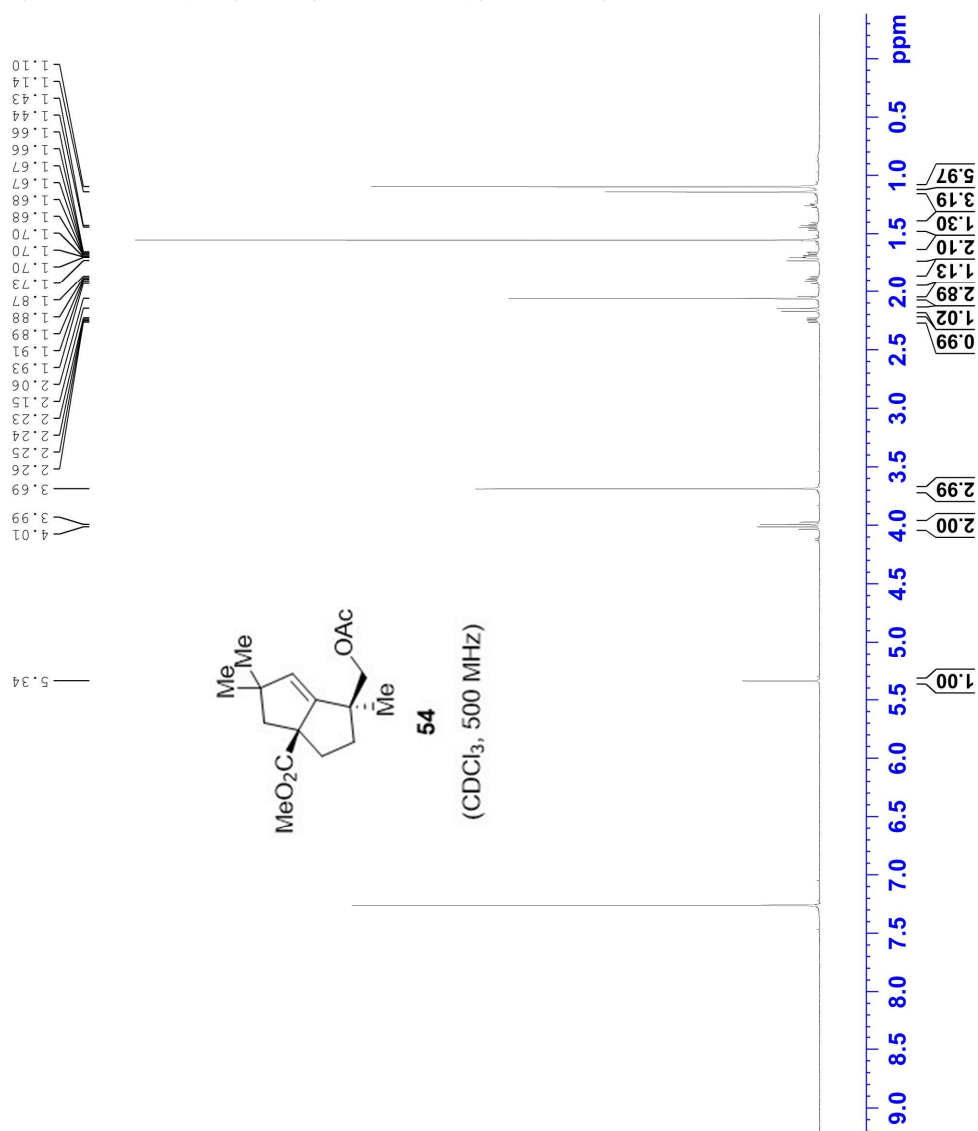


Current Data Parameters
 NAME 5-member-ring
 EXPNO 1
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20190812
 Time_ 11.28
 INSTRUM spect
 PROBHD 5 mm PATXI 1H/
 PULPROG zg
 TD 5998
 SOLVENT CDCl3
 NS 6
 DS 0
 SWH 10000.000 Hz
 FIDRES 0.166672 Hz
 AQ 2.9999001 sec
 RG 196.79
 DW 50.000 usec
 DE 10.00 usec
 TE 294.4 K
 DL 4.0000000 sec
 TDO 1

===== CHANNEL f1 =====
 SFO1 500.1330885 MHz
 NUC1 1H
 PL1 3.30 usec
 PLW1 12.19999981 W

F2 - Processing parameters
 SI 65536
 SF 500.1300138 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00



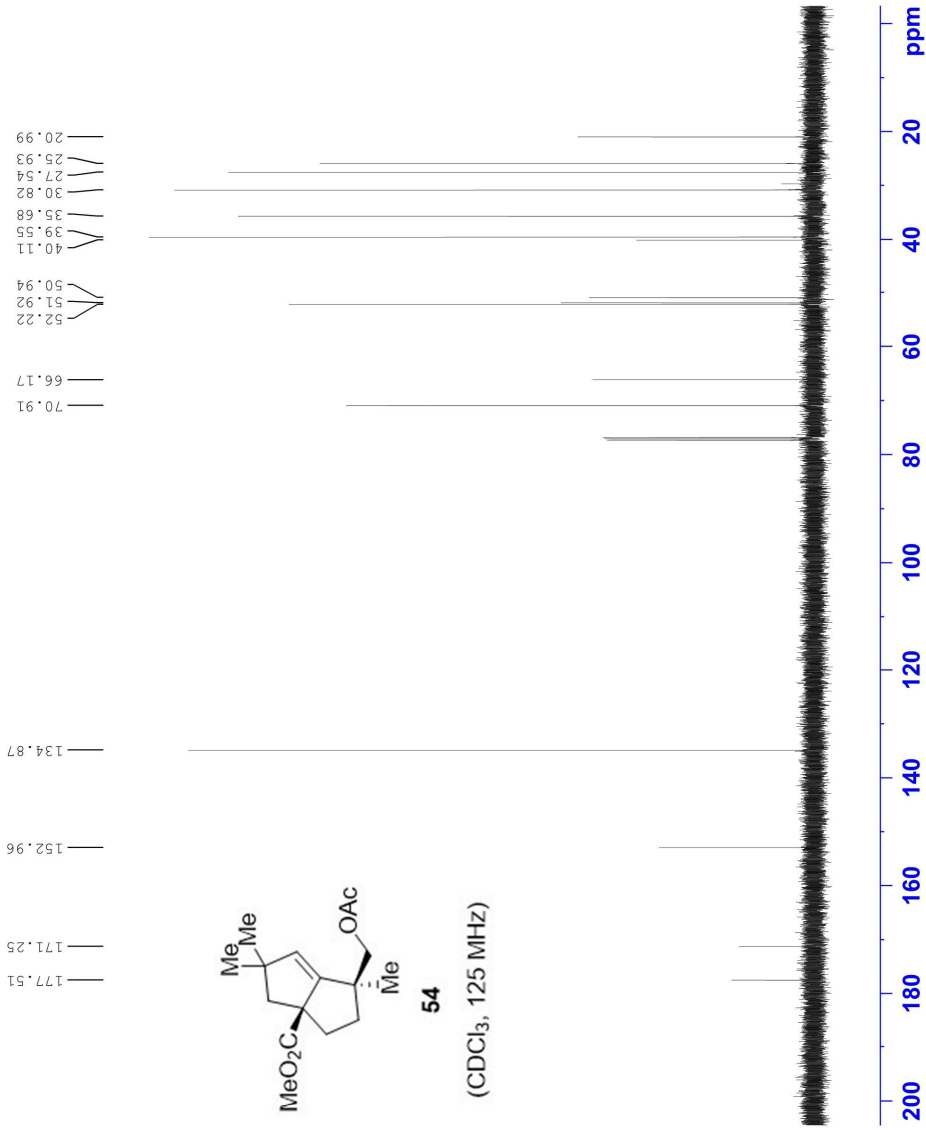
Current Data Parameters
 NAME 5-member-ring
 EXPNO 2
 PROCNO 2

F2 - Acquisition Parameters
 Date_ 20190812
 Time_ 21.07
 INSTRUM spect
 PROBHD 5 mm PABBO BB/
 PULPROG zgpg30
 TD 187496
 SOLVENT CDCl3
 NS 15
 DS 0
 SWH 31250.000 Hz
 FIDRES 0.166670 Hz
 AQ 2.9999361 sec
 RG 2050
 DW 16.000 usec
 DE 6.50 usec
 TE 294.5 K
 D1 3.0000000 sec
 D11 0.0300000 sec
 TDO 1

==== CHANNEL f1 =====
 SFO1 125.7049802 MHz
 NUC1 13C
 P1 10.00 usec
 PLW1 72.83999634 W

==== CHANNEL f2 =====
 SFO2 499.8724993 MHz
 NUC2 1H
 CPDPRG12 waltz16
 PCPD2 80.00 usec
 PLW2 19.0000000 W
 PLW12 0.29688001 W

F2 - Processing parameters
 SI 1048576
 SF 125.6924115 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.40

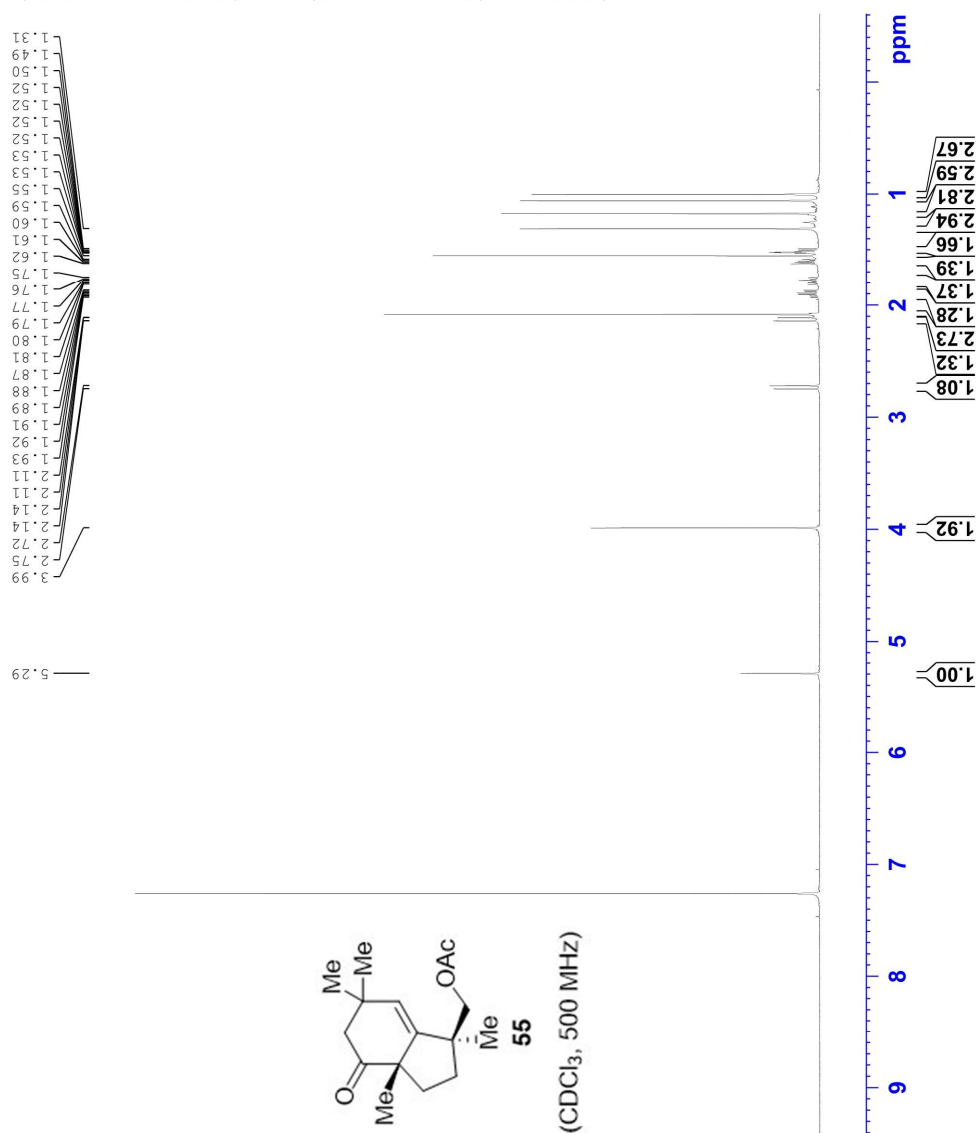


Current Data Parameters
 NAME yh-5-39
 EXPNO 1
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20180724
 Time_ 15.25
 INSTRUM spect
 PROBHD 5 mm PABBO BB/
 PULPROG zg
 TD 5998
 SOLVENT CDCl3
 NS 16
 DS 0
 SWH 10000.000 Hz
 FIDRES 0.166672 Hz
 AQ 2.9999001 sec
 RG 118.99
 DW 50.000 usec
 DE 6.50 usec
 TE 296.2 K
 DL 4.00000000 sec
 TDL 1

===== CHANNEL f1 =====
 SFO1 499.8730869 MHz
 NUC1 1H
 PL 3.58 usec
 PLW1 18.25000000 W

F2 - Processing parameters
 SI 65536
 SF 499.8700122 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00



```

Current Data Parameters
NAME      yh-5-39
EXPNO    3
PROCNO   3

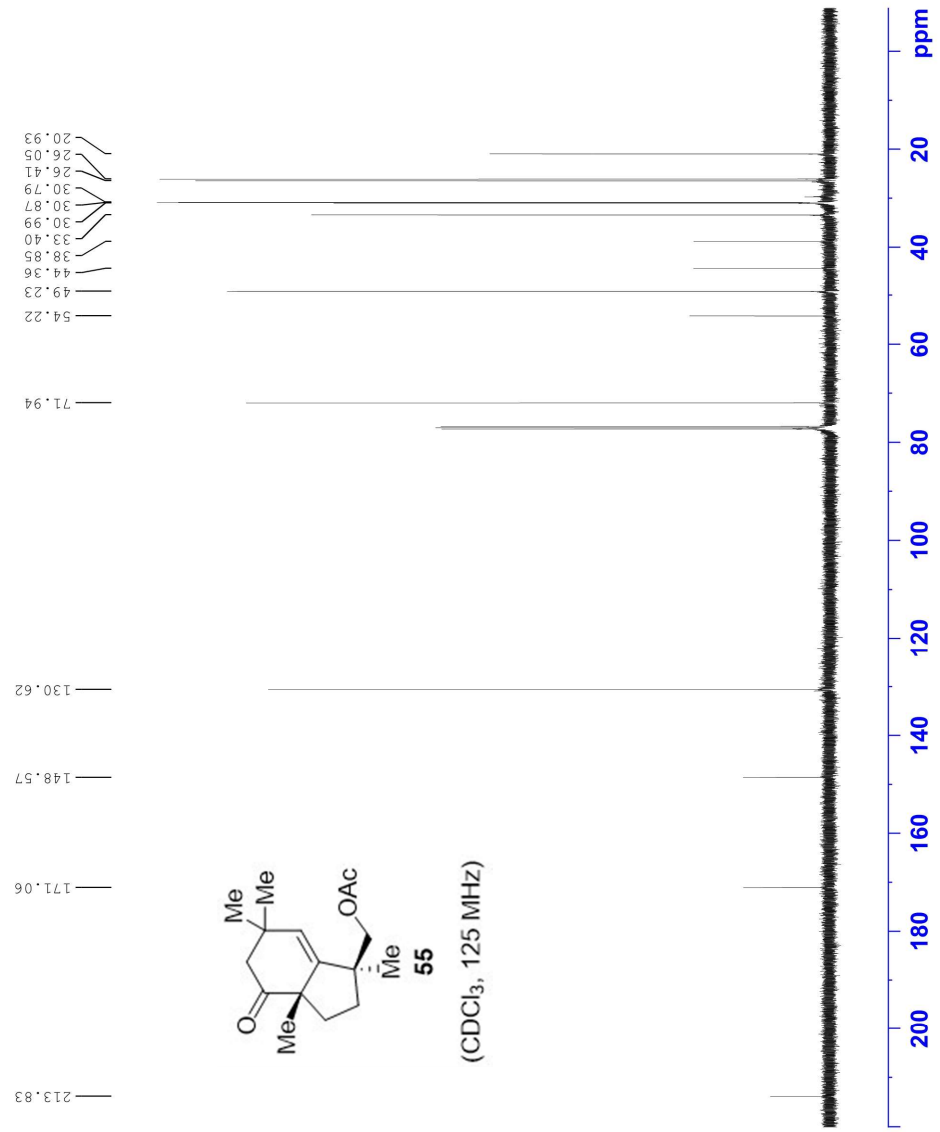
F2 - Acquisition Parameters
Date_     20180725
Time      22.44
INSTRUM   spect
PROBHD    5 mm PABBO BB/
PULPROG   zgpg
TD         187496
SOLVENT   CDCl3
NS         507
DS         0
SWH        31250.000 Hz
FIDRES     0.166670 Hz
AQ         2.9999361 sec
RG         2050
DM         16.000 usec
DE         6.50 usec
TE         298.2 K
D1         3.00000000 sec
D11        0.03000000 sec
TD0        1

===== CHANNEL f1 =====
SFO1      125.7049802 MHz
NUC1       13C
P1         10.00 usec
PLW1      72.83999634 W

===== CHANNEL f2 =====
SFO2      499.8724993 MHz
NUC2       1H
CFPRG2    waitz16
PCPD2     80.00 usec
PLW2      19.00000000 W
PLW12     0.29688001 W

F2 - Processing parameters
SI         1048576
SF         125.6924115 MHz
WDW        EM
SSB        0
LB         0.30 Hz
GB         0
PC         1.40

```

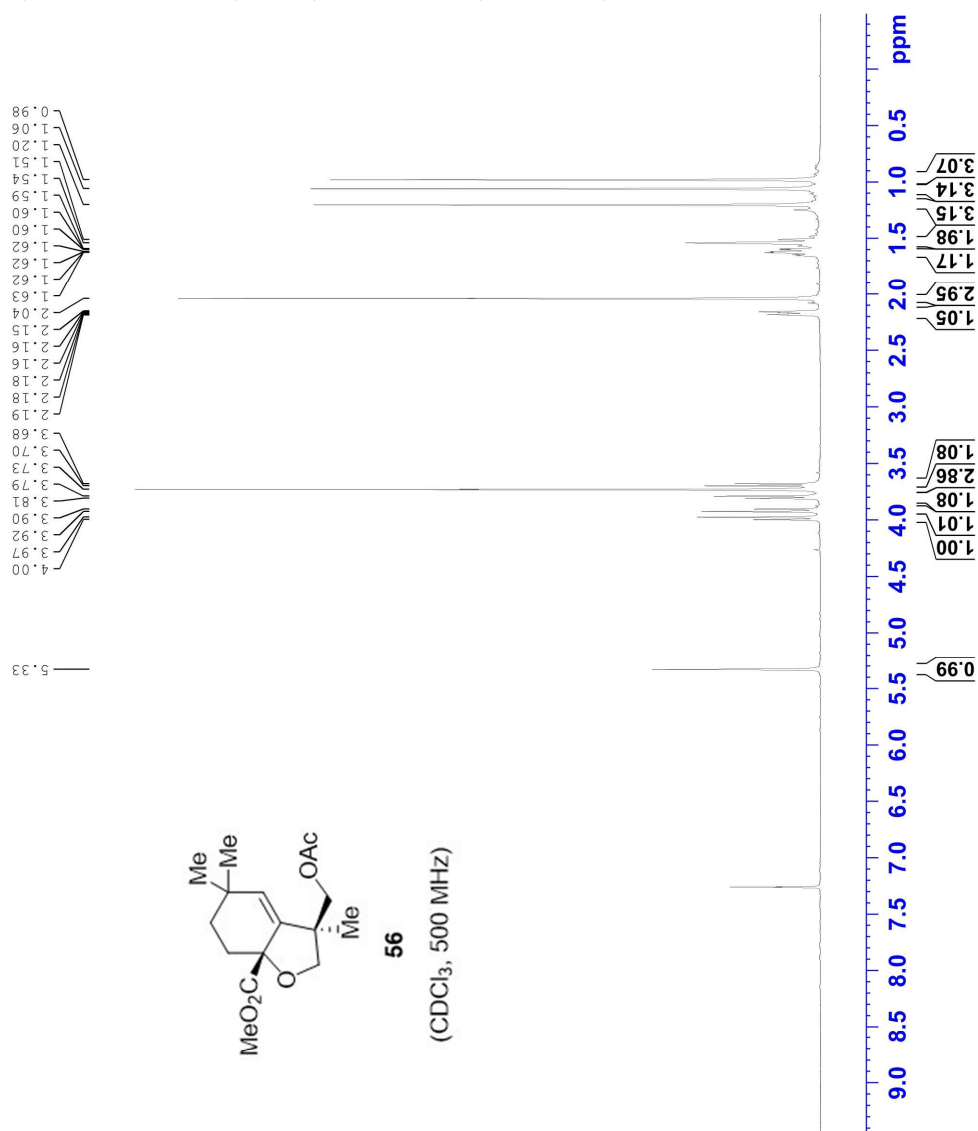


Current Data Parameters
 NAME Yh-5-88-a
 EXPNO 1
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20181022
 Time_ 11.53
 INSTRUM spect
 PROBHD 5 mm PABBO BB/
 PULPROG zg
 TD 5998
 SOLVENT CDCl3
 NS 8
 DS 0
 SWH 10000.000 Hz
 FIDRES 0.166672 Hz
 AQ 2.9999001 sec
 RG 56.75
 DW 50.000 usec
 DE 6.50 usec
 TE 297.2 K
 DL 4.0000000 sec
 TDO 1

===== CHANNEL f1 =====
 SFO1 499.8730869 MHz
 NUC1 1H
 P1 3.58 usec
 PLW1 18.25000000 W

F2 - Processing parameters
 SI 65536
 SF 499.8700120 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00



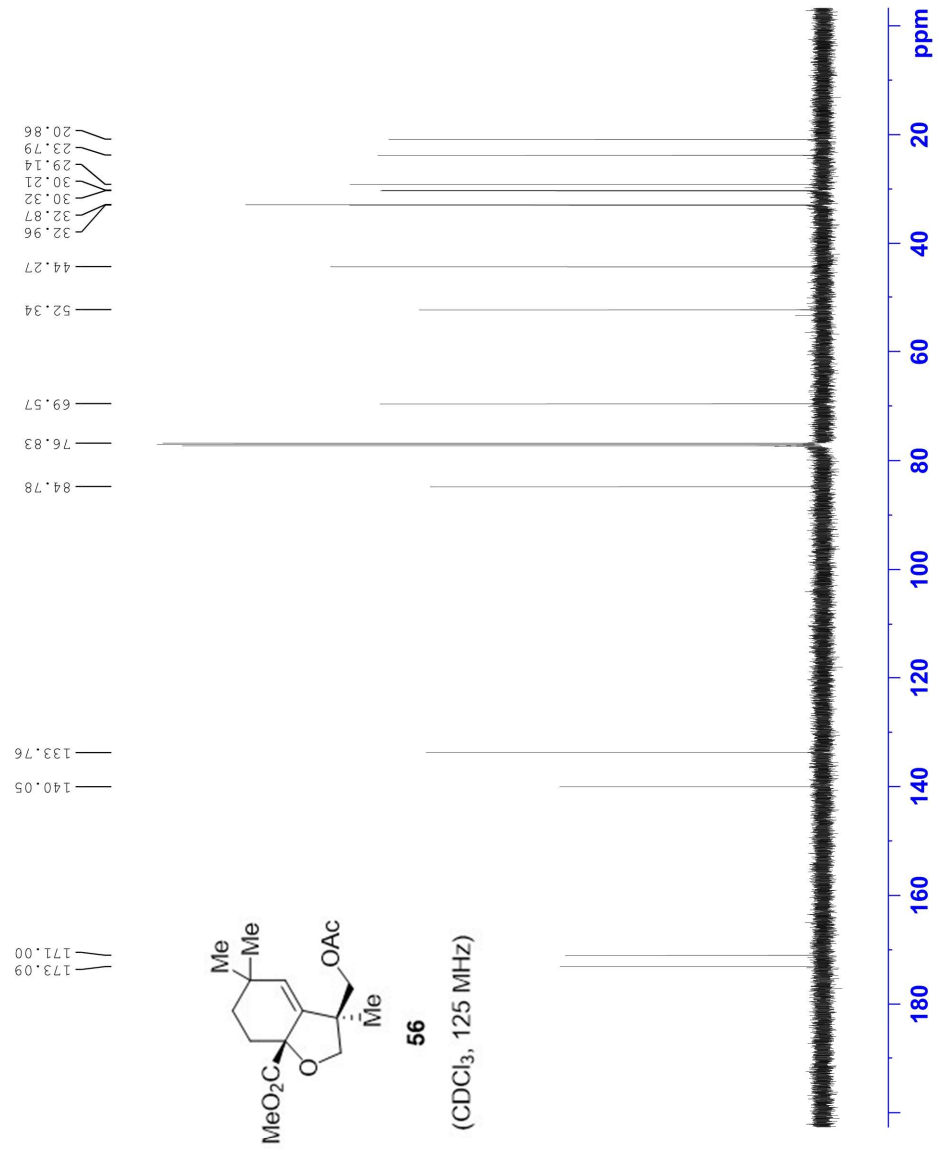
Current Data Parameters
 NAME Yh-5-88-a
 EXPNO 2
 PROCNO 2

F2 - Acquisition Parameters
 Date_ 20181022
 Time 13.52
 INSTRUM spect
 PROBHD 5 mm PAQNP 1H/
 PULPROG zgpg30
 TD 197364
 SOLVENT CDC13
 NS 64
 DS 0
 SWH 32894.738 Hz
 FIDRES 0.166670 Hz
 AQ 2.9999328 sec
 RG 2050
 DM 15.200 usec
 DE 6.00 usec
 TE 295.5 K
 D1 10.00000000 sec
 D11 0.03000000 sec
 TDO 1

==== CHANNEL f1 =====
 NUC1 13C
 P1 9.90 usec
 PL1 1.00 dB
 EL1W 72.42802429 W
 SFO1 125.7716224 MHz

==== CHANNEL f2 =====
 CPDPRG12 waltz16
 NUC2 1H
 FCPD2 80.00 usec
 PL2 0 dB
 PL12 16.50 dB
 PL13 17.00 dB
 PL2W 24.54113007 W
 PL12W 0.54940748 W
 PL13W 0.48965994 W
 SFO2 500.1325006 MHz

F2 - Processing parameters
 SI 131072
 SF 125.7577890 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 FC 1.40

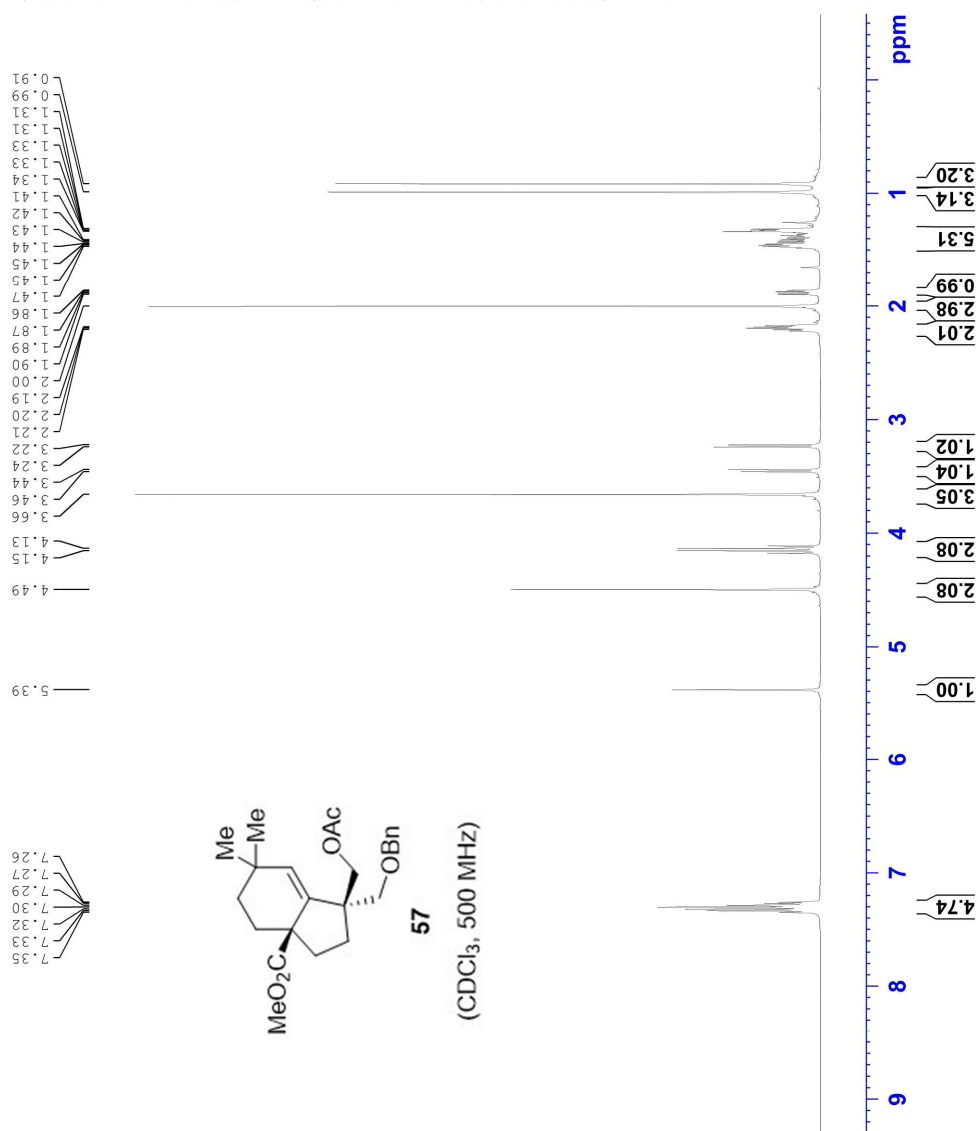


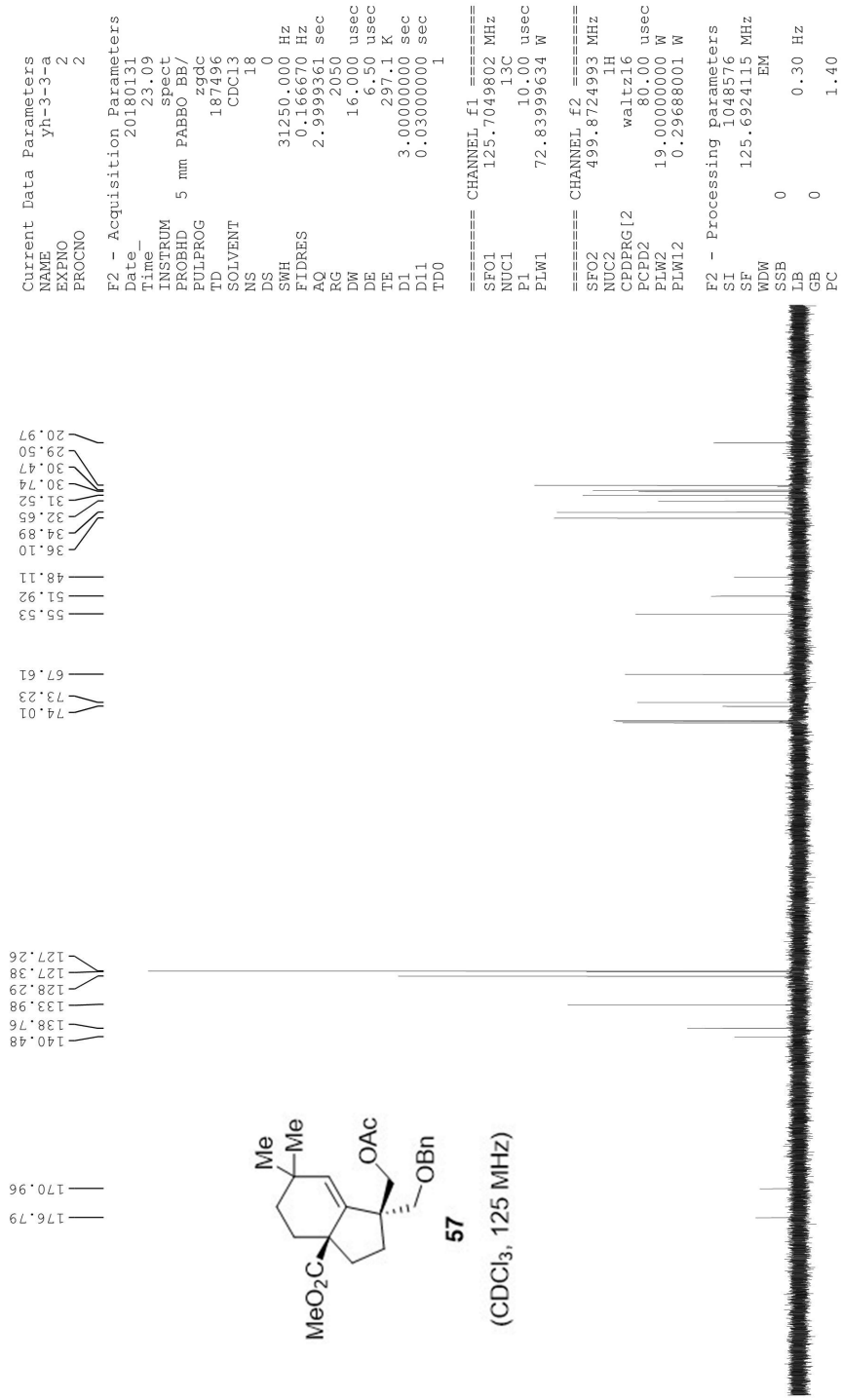
Current Data Parameters
 NAME yn-3-3-a
 EXPNO 1
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20180131
 Time_ 22.59
 INSTRUM spect
 PROBHD 5 mm PABBO BB/
 PULPROG zg
 TD 5998
 SOLVENT CDCl3
 NS 4
 DS 0
 SWH 10000.000 Hz
 FIDRES 0.166672 Hz
 AQ 2.9999001 sec
 RG 22.37
 DW 50.000 usec
 DE 6.50 usec
 TE 296.8 K
 DL 3.0000000 sec
 TDO 1

===== CHANNEL f1 =====
 SFO1 499.8730869 MHz
 NUC1 1H
 PL 10.75 usec
 PLW1 18.25000000 W

F2 - Processing parameters
 SI 65536
 SF 499.8700120 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00





```

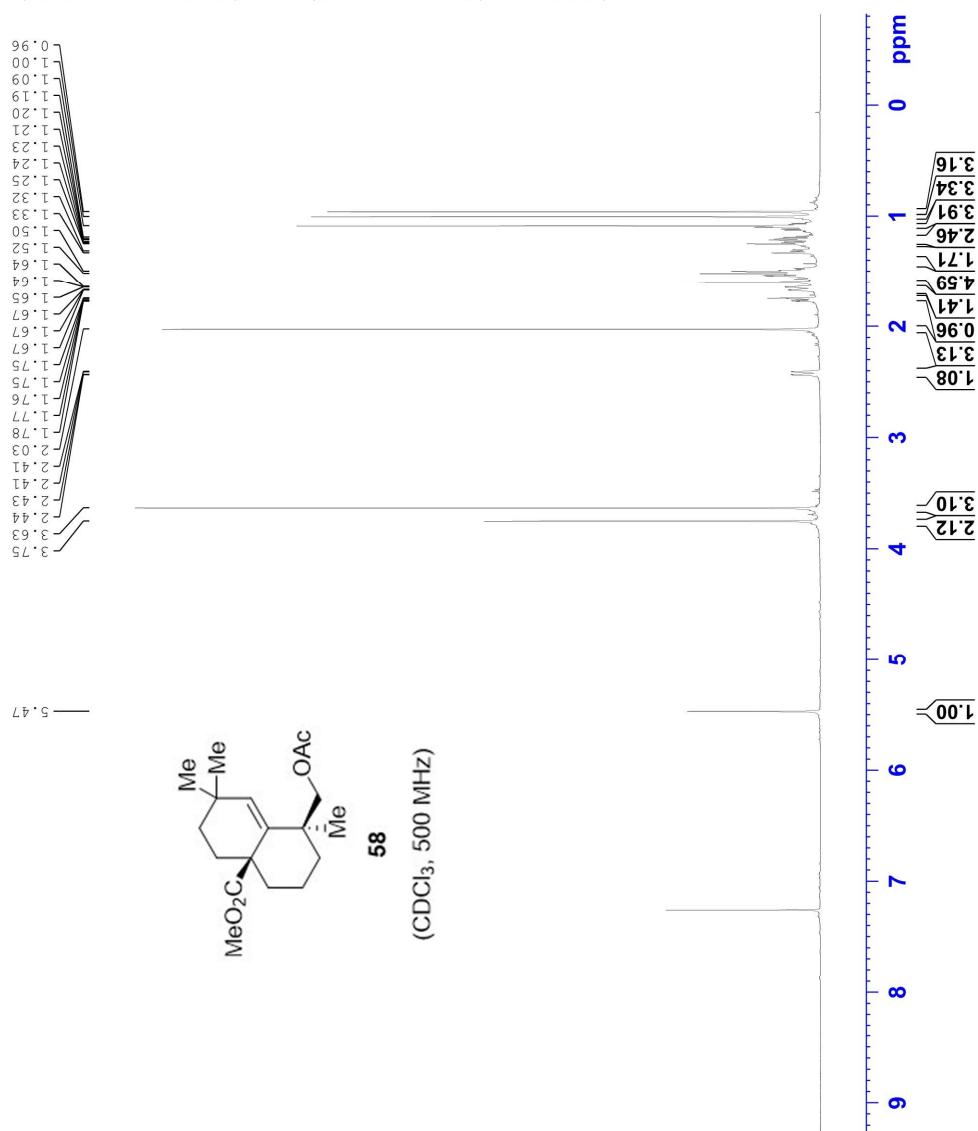
Current Data Parameters
NAME      yh-5-21
EXPNO    1
PROCNO   1

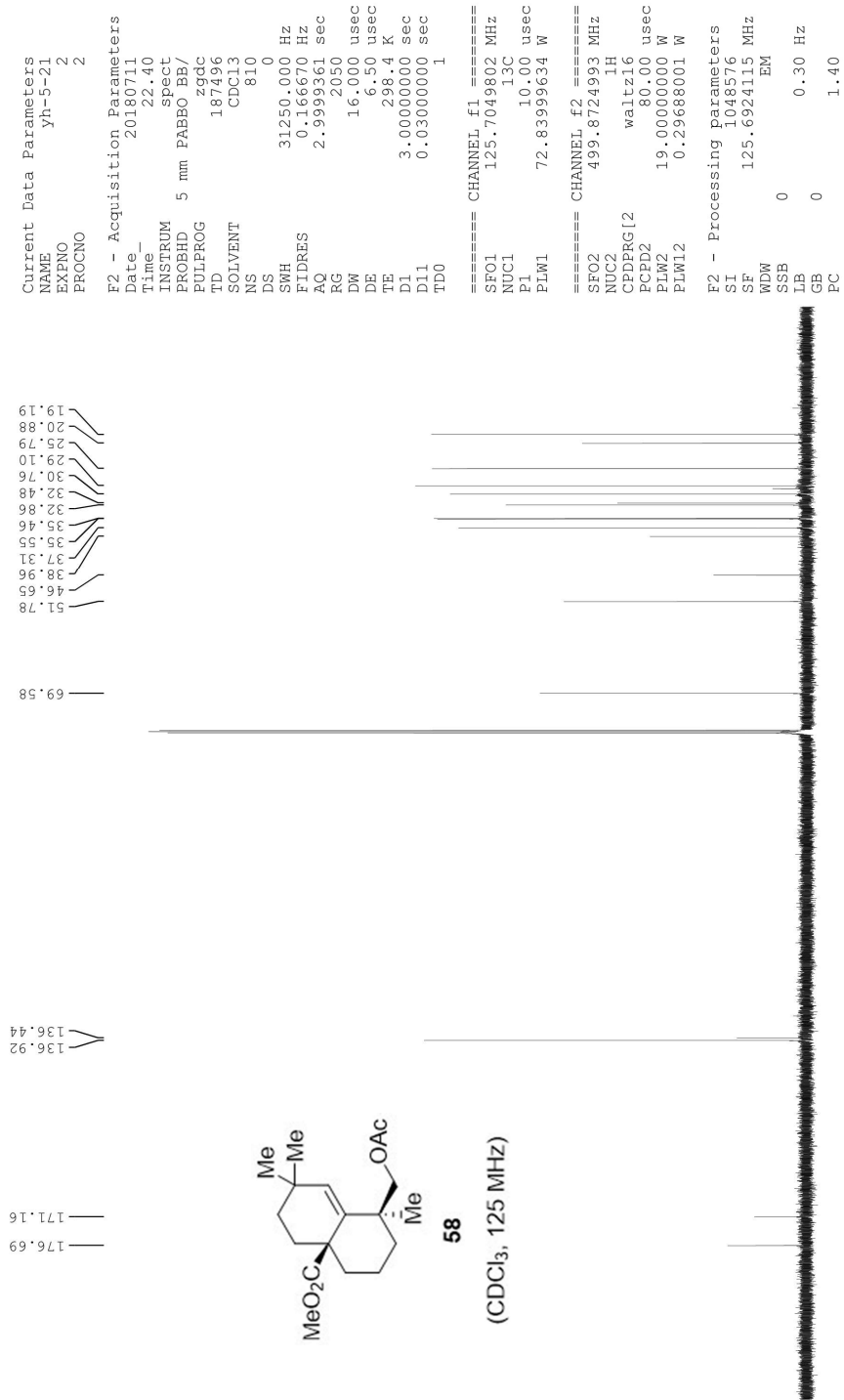
F2 - Acquisition Parameters
Date_    20180711
Time     21.11
INSTRUM spect
PROBHD   5 mm PABBO BB/
PULPROG zg
TD       59998
SOLVENT  CDCl3
NS       16
DS       0
SWH      10000.000 Hz
FIDRES   0.166672 Hz
AQ       2.9999001 sec
RG       77.07
DW       50.000 usec
DE       6.50 usec
TE       296.7 K
DL       4.0000000 sec
TD0      1

===== CHANNEL f1 =====
SFO1    499.8730869 MHz
NUC1     1H
P1       3.58 usec
PLW1    18.25000000 W

F2 - Processing parameters
SI      65536
SF      499.8700122 MHz
WDW     EM
SSB     0
LB      0.30 Hz
GB      0
PC      1.00

```

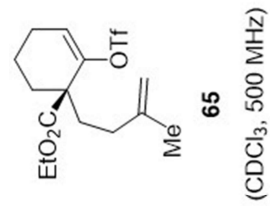




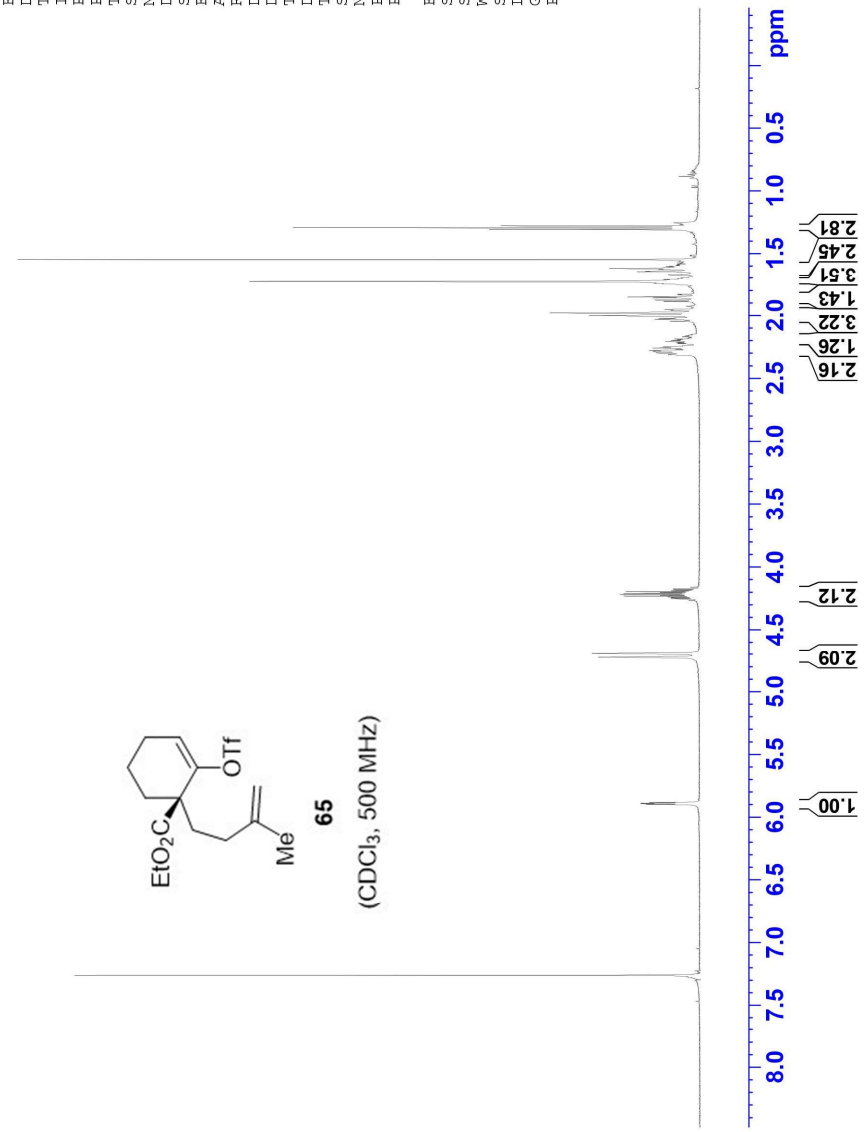
5.90
5.89
5.88
4.72
4.69
4.25
4.24
4.24
4.22
4.20
4.19
4.18
4.17
4.17
2.31
2.30
2.28
2.28
2.27
2.26
2.25
2.22
2.22
2.21
2.20
2.20
2.19
2.19
2.18
2.17
2.17
2.04
2.03
2.02
1.96
1.95
1.89
1.88
1.86
1.85
1.83
1.83
1.73
1.67
1.62
1.60
1.59
1.58

Current Data Parameters
 NAME Yh-7-74
 EXPNO 1
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20191106
 Time 22.58 h
 INSTRUM spect
 PROBHD zll3652_0187 (
 PULPROG zg
 TD 59998
 SOLVENT CDCl3
 NS 4
 DS 0
 SWH 10000.000 Hz
 FIDRES 0.333344 Hz
 AQ 2.9999001 sec
 RG 77.07
 DW 50.000 usec
 DE 6.50 usec
 TE 295.3 K
 DL 3.00000000 sec
 TD0 1
 SFO1 499.8730869 MHz
 NUC1 1H
 PI 10.75 usec
 PLW1 18.25000000 W



F2 - Processing parameters
 SI 65536
 SF 499.8700121 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00



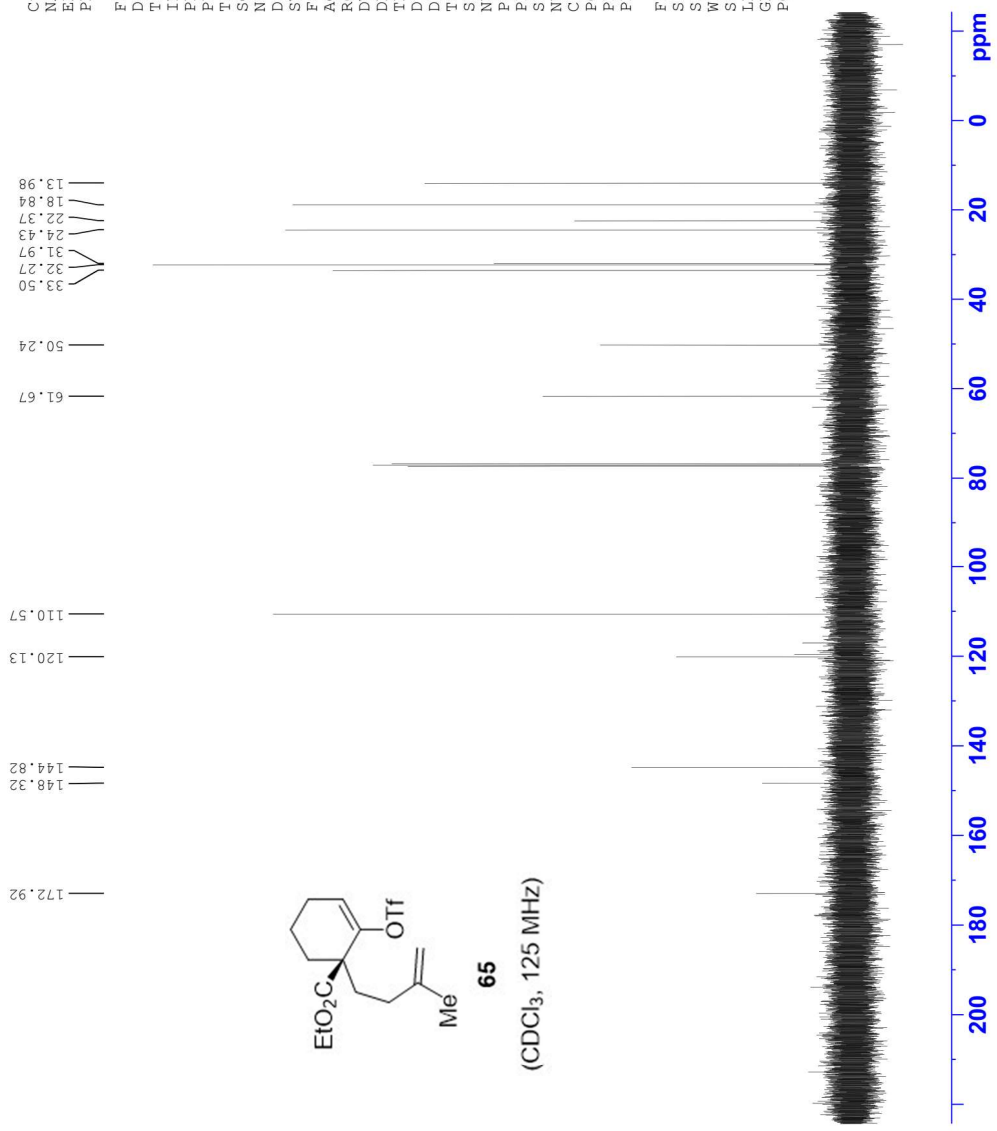
```

Current Data Parameters
NAME      yh-7-74
EXPNO    2
PROCNO   1

F2 - Acquisition Parameters
Date_    20191107
Time     22.20 h
INSTRUM  spect
PROBHD   Z113652_0187 (
PULPROG  zgdc
TD       187496
SOLVENT  CDCl3
NS       13
DS       0
SWH      31250.000 Hz
FIDRES   0.333340 Hz
AQ       2.9993361 sec
RG       2050
DW       16.000 usec
DE       6.50 usec
TE       295.7 K
D1       3.00000000 sec
D11      0.03000000 sec
TD0      1
SF01     125.7049802 MHz
NUC1     13C
P1       10.00 usec
PLW1     72.83999634 W
SE02     499.8724993 MHz
NUC2     1H
CPDPRG[2] waltz16
PCPD2    80.00 usec
PLW2     19.00000000 W
PLW12    0.29688001 W

F2 - Processing parameters
SI       1048576
SF       125.6924115 MHz
WDW      EM
SSB      0
LB       0.30 Hz
GB       0
PC       1.40

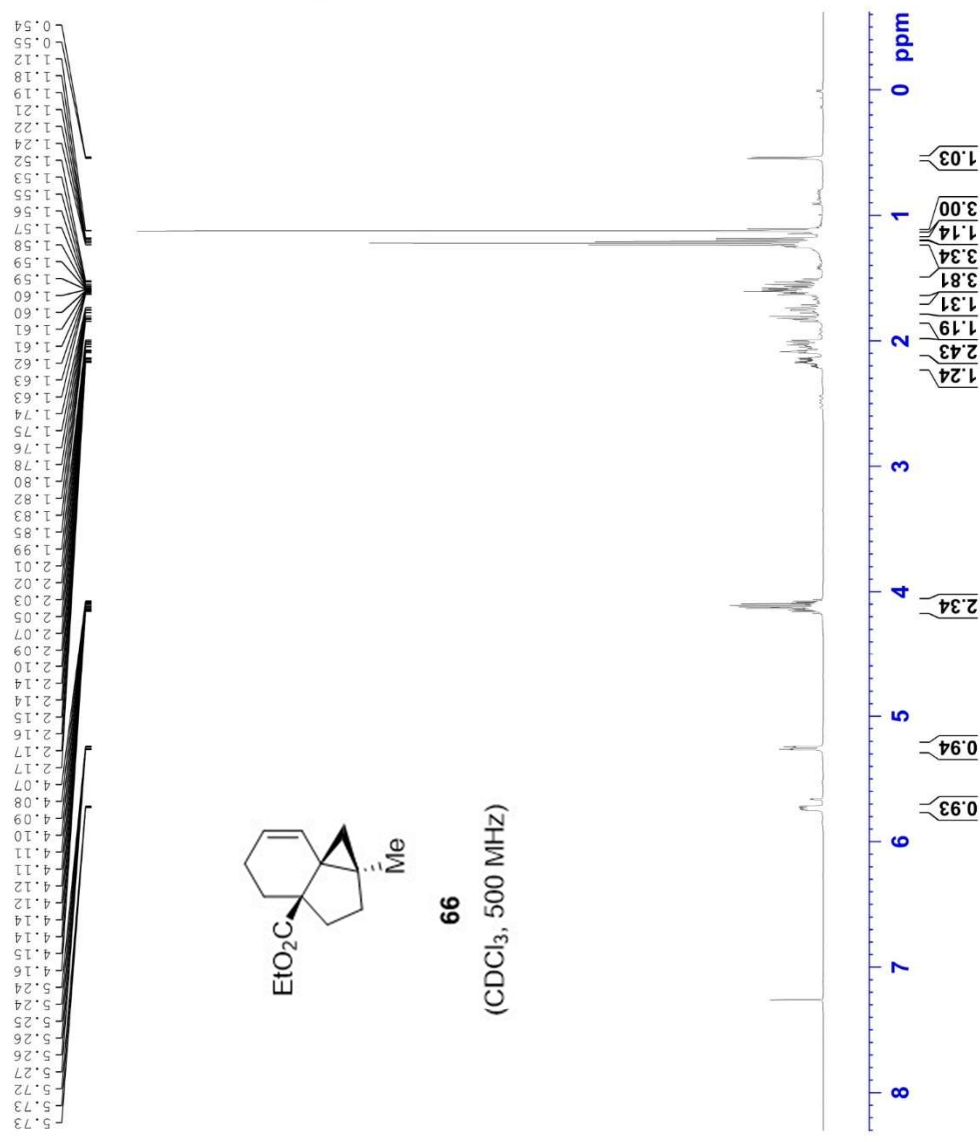
```



Current Data Parameters
 NAME yh-7-76-a
 EXPNO 1
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20191108
 Time 0.02 h
 INSTRUM spect
 PROBHD z113652_0187 (zg)
 PULPROG 59998
 TD CDC13
 NS 4
 DS 0
 SWH 10000.000 Hz
 FIDRES 0.333344 Hz
 AQ 2.9999001 sec
 RG 29.95
 DW 50.000 usec
 DE 6.50 usec
 TE 295.4 K
 D1 3.0000000 sec
 TD0 1
 SFO1 499.8730869 MHz
 NUC1 1H
 P1 10.75 usec
 PLW1 18.25000000 W

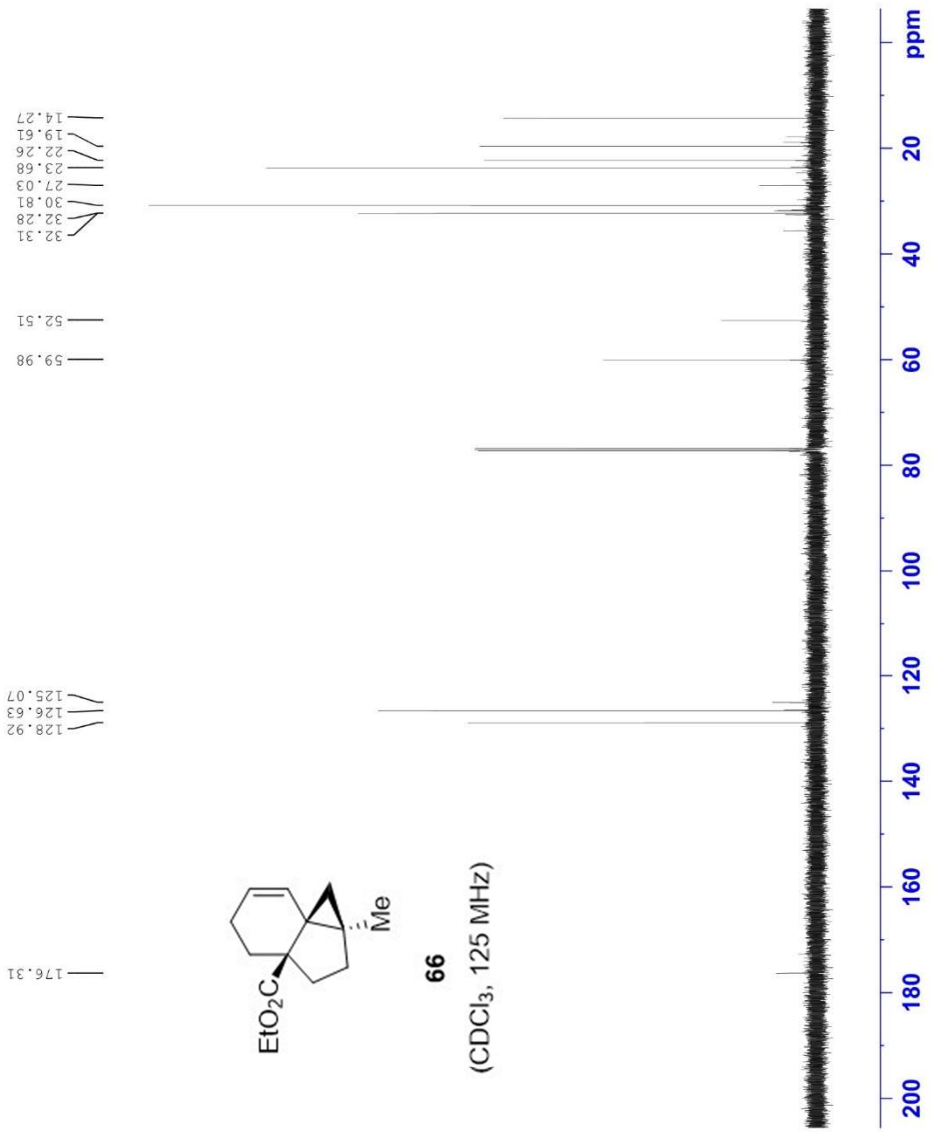
F2 - Processing parameters
 SI 65536
 SF 499.8700125 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00



Current Data Parameters
 NAME Yh-7-76-a
 EXPNO 2
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20191108
 Time_ 0.24 h
 INSTRUM spect
 PROBHD Z113652_0187 (zdc
 PULPROG zgpg30
 TD 65536
 SOLVENT CDCl3
 NS 1024
 DS 4
 SWH 31250.000 Hz
 FIDRES 0.333340 Hz
 AQ 2.9999361 sec
 RG 2050
 DW 16.000 usec
 DE 6.50 usec
 TE 296.6 K
 D1 3.0000000 sec
 D11 0.0300000 sec
 TDO 1
 SFO1 125.7049802 MHz
 NUC1 13C
 P1 10.00 usec
 PLW1 72.83999634 W
 SFO2 499.8724993 MHz
 NUC2 1H
 CEDPRG12 waltz16
 PCPD2 80.00 usec
 PLW2 19.0000000 W
 PLW12 0.29688001 W

F2 - Processing parameters
 SI 1048576
 SF 125.6924115 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 FC 1.40



```

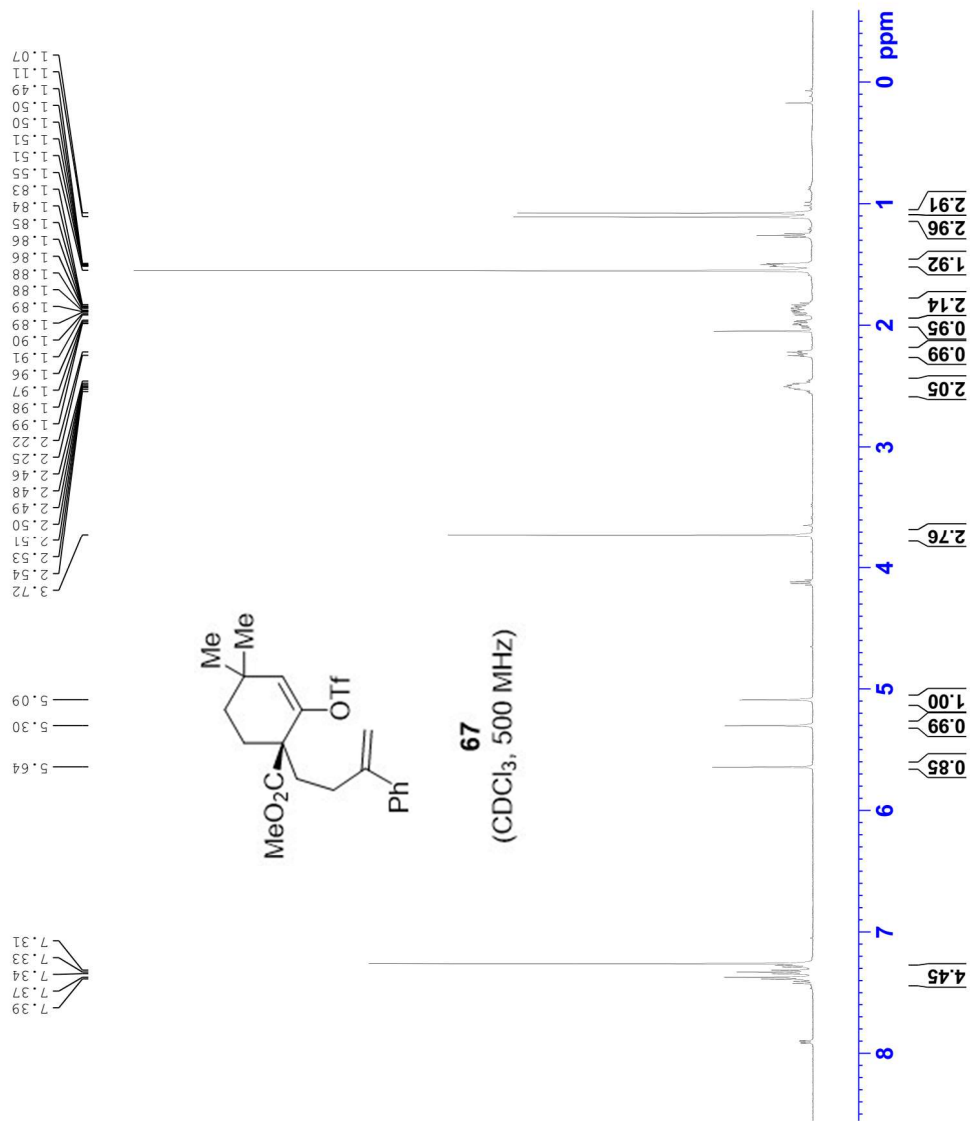
Current Data Parameters
NAME      yh-2-149
EXPNO    1
PROCNO   1

F2 - Acquisition Parameters
Date_    20170906
Time_    23.05
INSTRUM  spect
PROBHD   5 mm PAXI 1H/
PULPROG  zg
TD        59998
SOLVENT  CDCl3
NS        8
DS        0
SWH       10000.000 Hz
FIDRES   0.166672 Hz
AQ        2.9999001 sec
RG        196.79
DW        50.000 usec
DE        10.00 usec
TE        295.0 K
D1        2.00000000 sec
TDO       1

===== CHANNEL f1 =====
SFO1     500.1330885 MHz
NUC1     1H
P1       8.00 usec
PLW1     12.19999981 W

F2 - Processing parameters
SI        65536
SF        500.1300134 MHz
WDW       EM
SSB       0
LB        0.30 Hz
GB        0
PC        1.00

```

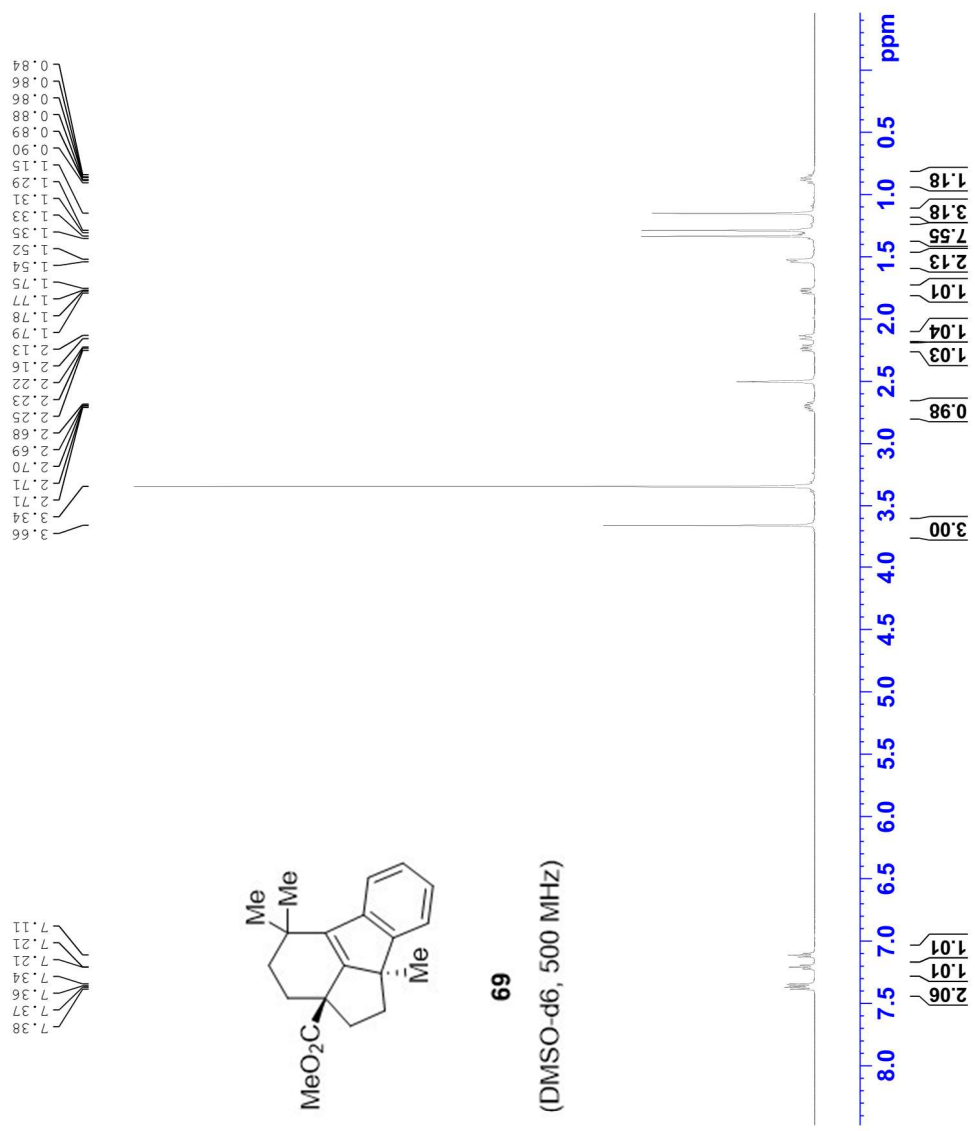


Current Data Parameters
 NAME Yh-2-150-1-r
 EXPNO 1
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20170908
 Time_ 15.30
 INSTRUM spect
 PROBHD 5 mm PAXI 1H/
 PULPROG zg
 TD 59998
 SOLVENT DMSO
 NS 8
 DS 0
 SWH 10000.000 Hz
 FIDRES 0.166672 Hz
 AQ 2.999001 sec
 RG 110.37
 DW 50.000 usec
 DE 10.000 usec
 TE 293.4 K
 D1 2.00000000 sec
 TDO 1

===== CHANNEL f1 =====
 SF01 500.1330885 MHz
 NUC1 1H
 P1 8.00 usec
 PLW1 12.1999981 W

F2 - Processing parameters
 SI 65536
 SF 500.1300049 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00



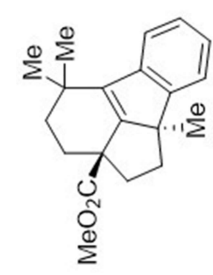
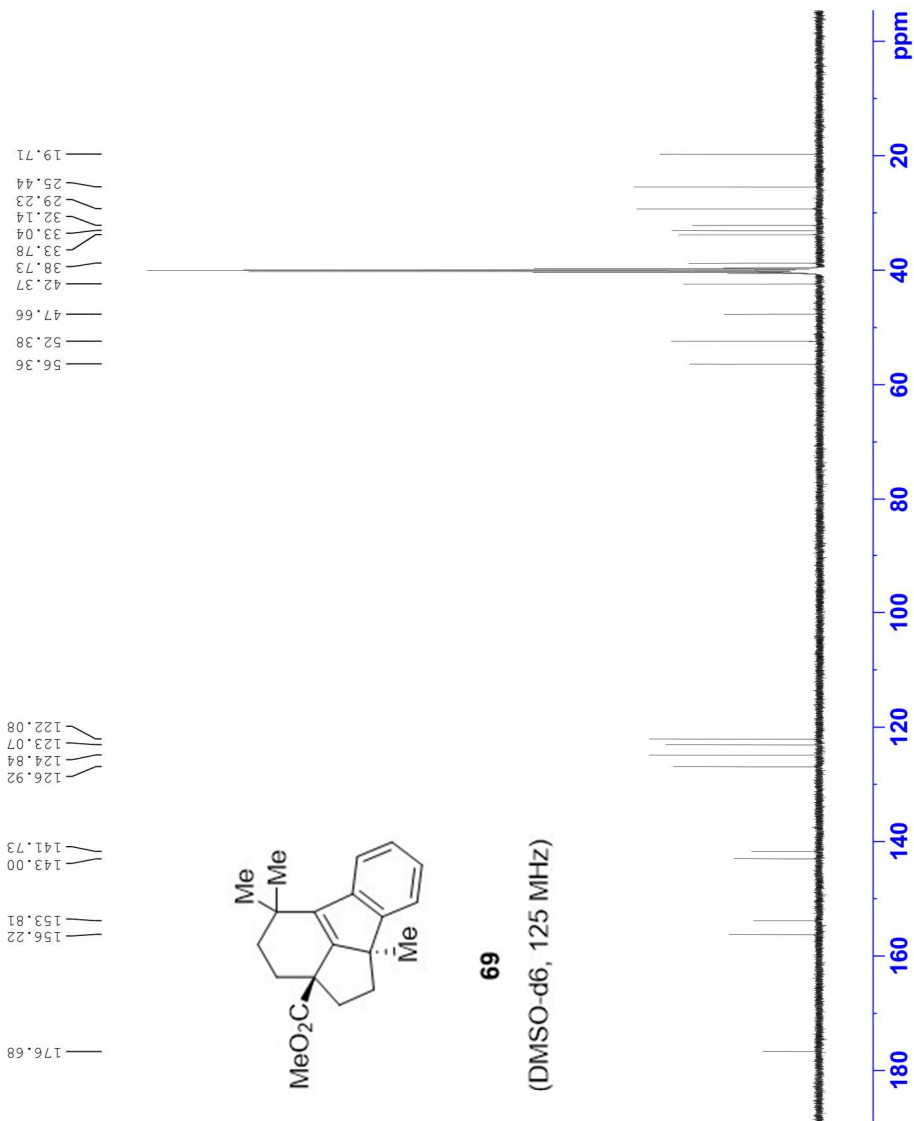
Current Data Parameters
 NAME Yh-2-150-1-rc
 EXPNO 1
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20170917
 Time_ 21.21
 INSTRUM spect
 PROBHD 5 mm PABBO BB/
 PULPROG zgdc
 TD 187496
 SOLVENT DMSO
 NS 530
 DS 0
 SWH 31250.000 Hz
 FIDRES 0.166670 Hz
 AQ 2.9999361 sec
 RG 2050
 DM 16.000 usec
 DE 6.50 usec
 TE 297.3 K
 D1 3.00000000 sec
 D11 0.03000000 sec
 TDO 1

=====
 CHANNEL f1
 SFO1 125.7049802 MHz
 NUC1 13C
 P1 10.00 usec
 PLW1 72.83999634 W

=====
 CHANNEL f2
 SFO2 499.8724993 MHz
 NUC2 1H
 CPDPRG2 waltz16
 FCPD2 80.00 usec
 PLW2 19.00000000 W
 PLWI2 0.29688001 W

F2 - Processing parameters
 SI 1048576
 SF 125.6924115 MHz
 WDM EM
 SSB 0
 LB 0.30 Hz
 GB 0
 FC 1.40



69
 (DMSO-d₆, 125 MHz)

```

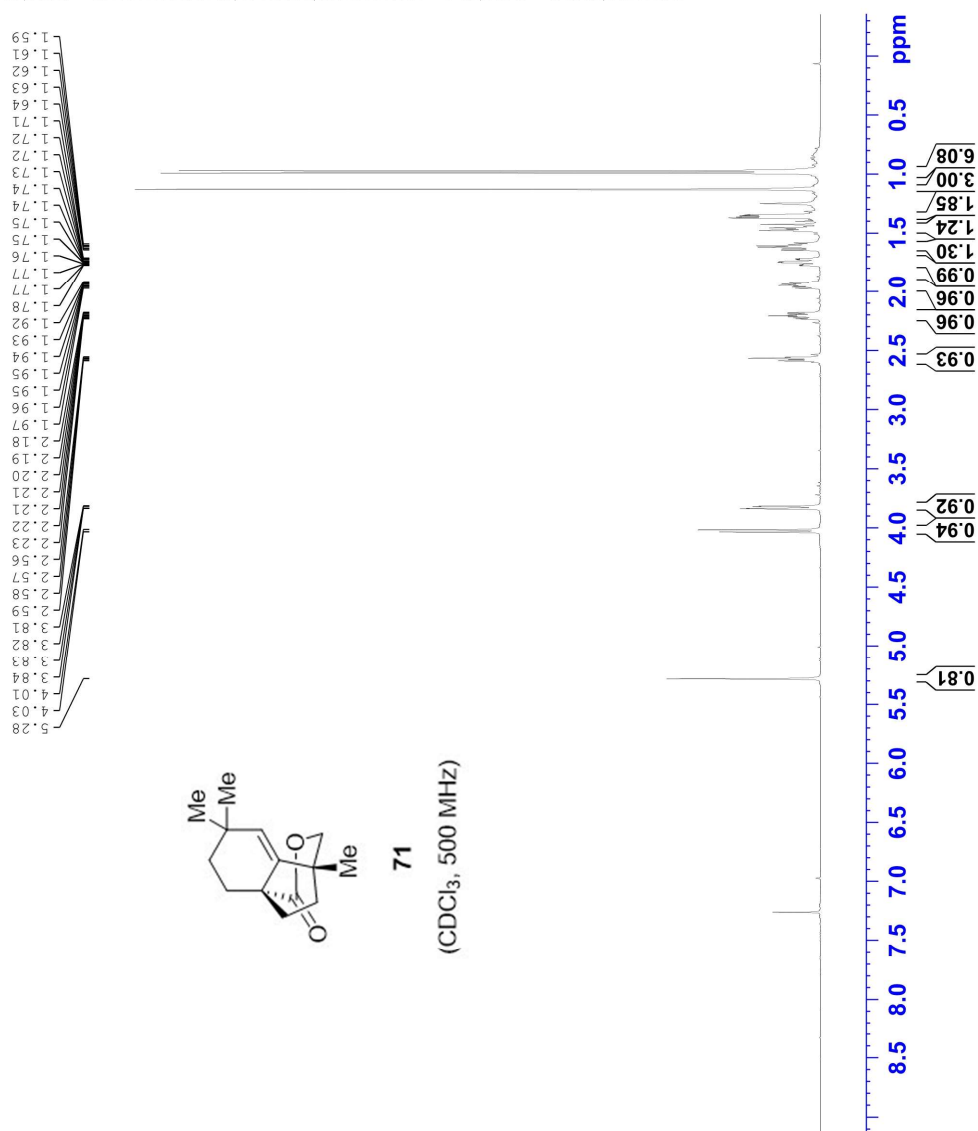
Current Data Parameters
NAME      yh-3-36-1H
EXPNO     1
PROCNO    1

F2 - Acquisition Parameters
Date_     20171104
Time      21.34
INSTRUM   spect
PROBHD    5 mm PATXI 1H/
PULPROG   zg
TD         59998
SOLVENT   CDCl3
NS         8
DS         0
SWH        10000.000 Hz
FIDRES     0.166672 Hz
AQ         2.9999001 sec
RG         55.8
DW         50.000 usec
DE         10.00 usec
TE         294.7 K
DL         2.0000000 sec
TDO        1

===== CHANNEL f1 =====
SFO1      500.1330885 MHz
NUC1      1H
P1         8.00 usec
PLW1      12.19999981 W

F2 - Processing parameters
SI         65536
SF         500.1300131 MHz
WDW        EM
SSB        0
LB         0.30 Hz
GB         0
PC         1.00

```



```

Current Data Parameters
NAME      Yh-3-36-1C
EXENO    1
PROCNO   1

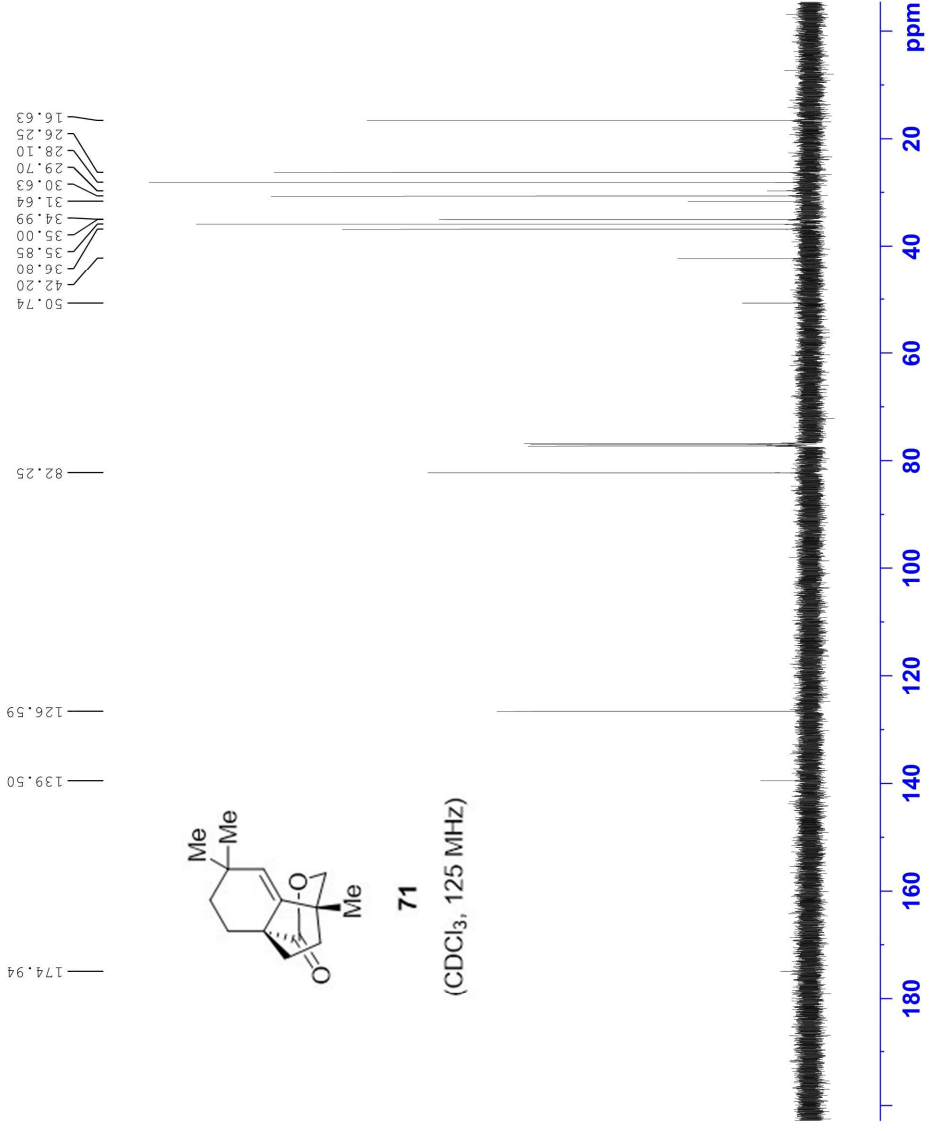
F2 - Acquisition Parameters
Date     20180822
Time    14.15
INSTRUM spect
PROBHD  5 mm PABBO BB/
PULPROG zgdc
TD      187496
SOLVENT CDCl3
NS      147
DS      0
SWH     31250.000 Hz
FIDRES  0.166670 Hz
AQ      2.9999361 sec
RG      2050
DM      16.000 usec
DE      6.50 usec
TE      296.7 K
D1      3.00000000 sec
D11     0.03000000 sec
TD0     1

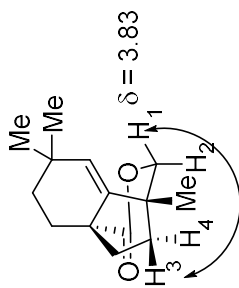
===== CHANNEL f1 =====
SF01    125.7049802 MHz
NUC1    13C
P1      10.00 usec
PLW1    72.83999634 W

===== CHANNEL f2 =====
SF02    499.8724993 MHz
NUC2    1H
CPDPRG2 waltz16
PCPD2   80.00 usec
PLW2    19.00000000 W
PLW12   0.29688001 W

F2 - Processing parameters
SI      1048576
SF      125.6924115 MHz
WDW     EM
SSB     0
LB      0.30 Hz
GB      0
PC      1.40

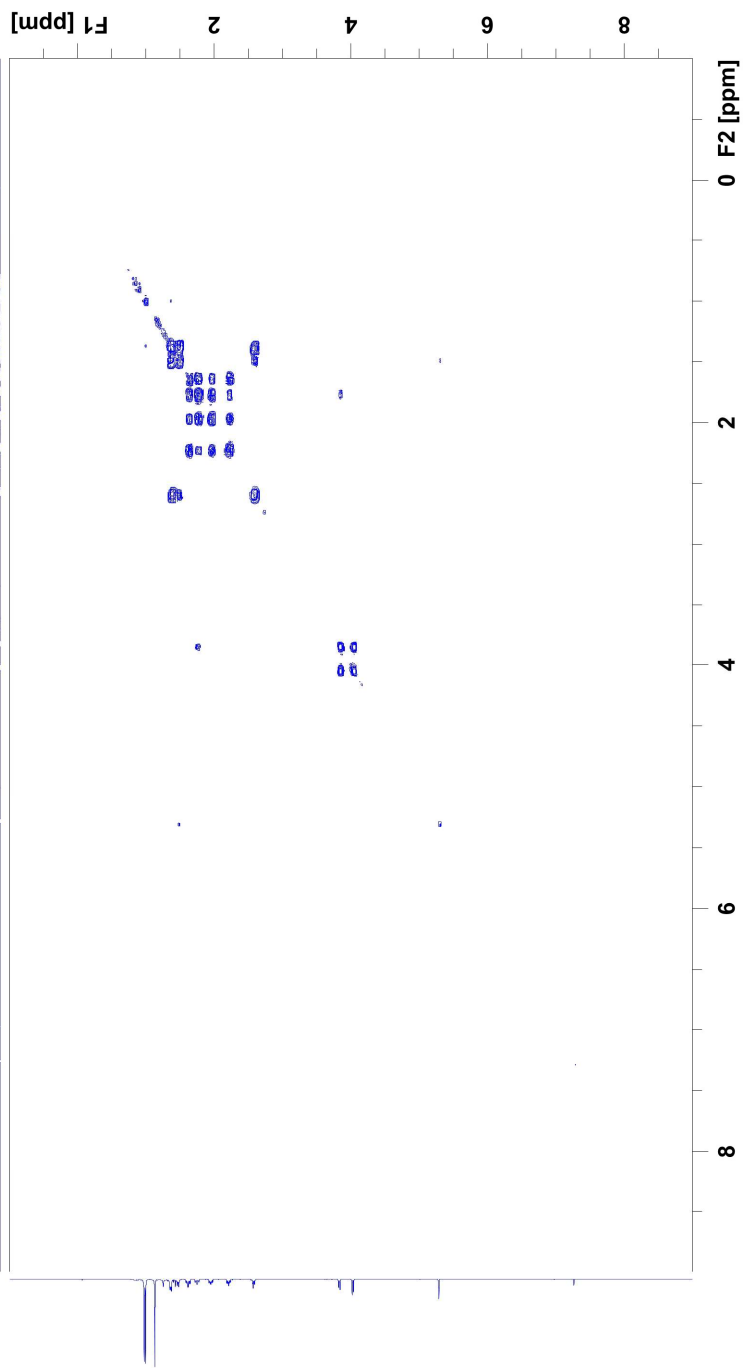
```

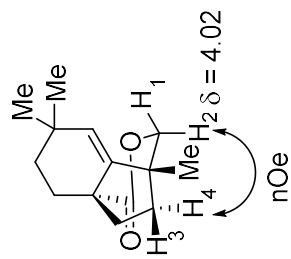




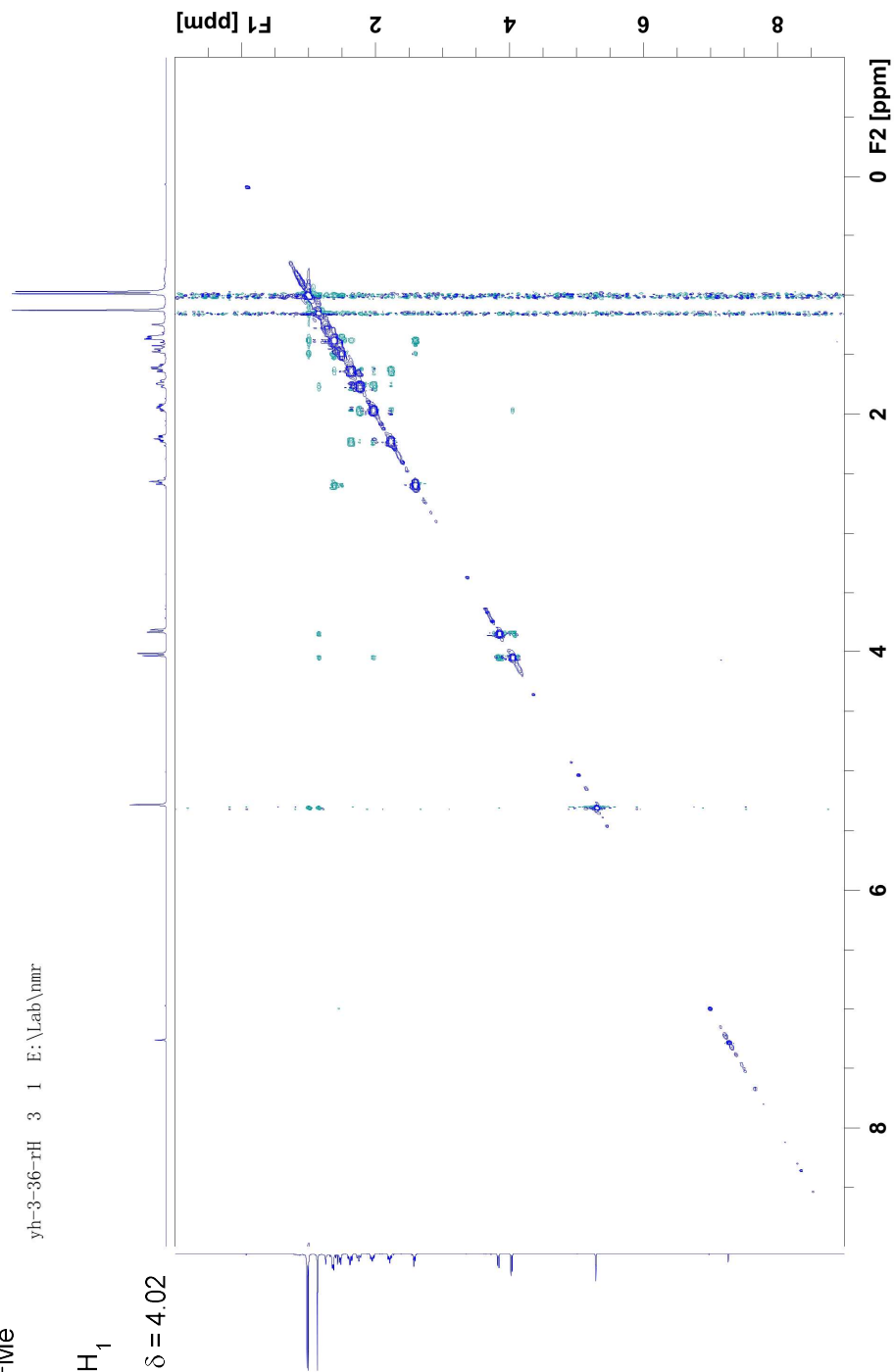
cosey (W coupling)

yh-3-36-rfl 2 1 E:\Lab\nmr





yh-3-36-rH 3 1 E:\Lab\nmr

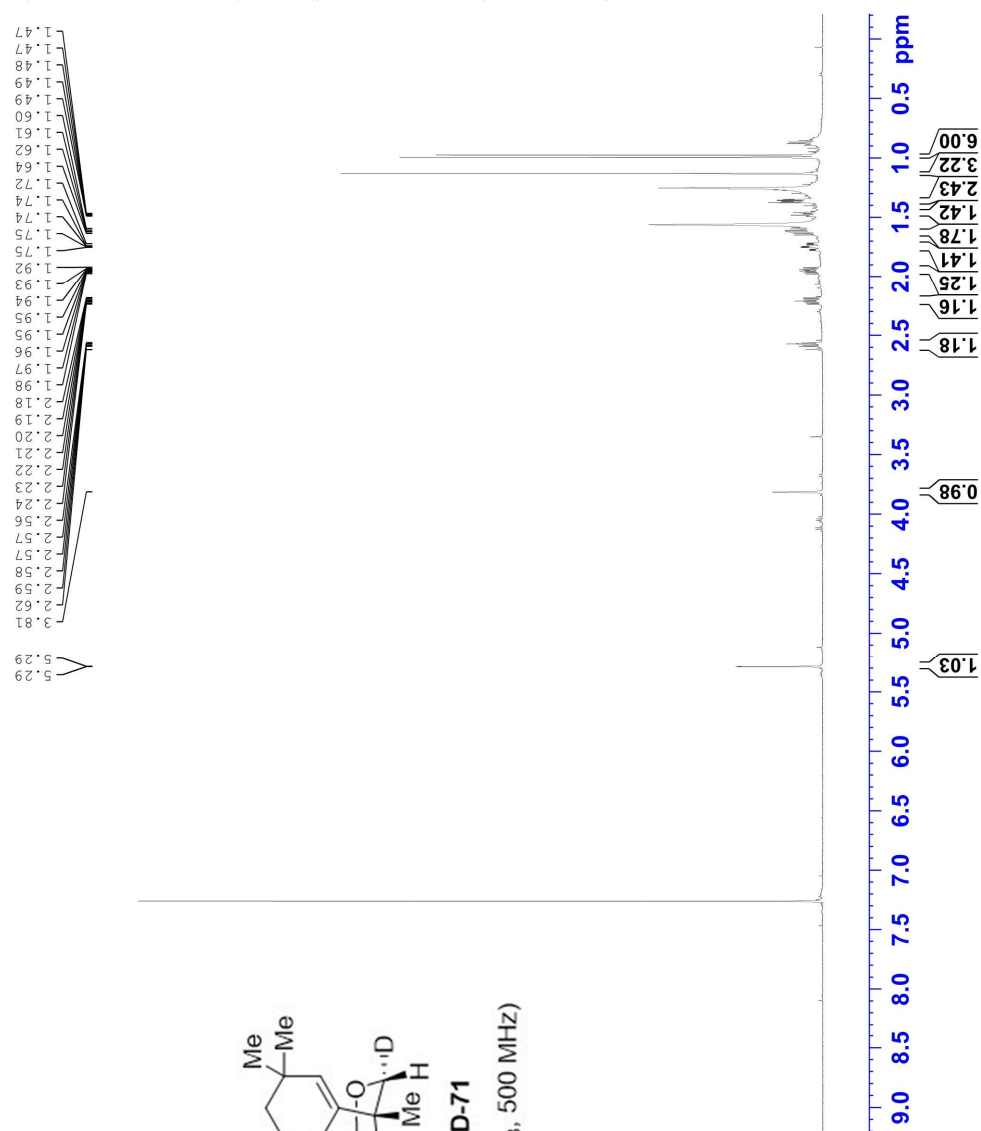


Current Data Parameters
 NAME Yh-6-146-3-a
 EXPNO 1
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20190811
 Time_ 21.15
 INSTRUM spect
 PROBHD 5 mm PATXI 1H/
 PULPROG zg
 TD 5998
 SOLVENT CDCl3
 NS 8
 DS 0
 SWH 10000.000 Hz
 FIDRES 0.166672 Hz
 AQ 2.9999001 sec
 RG 196.79
 DW 50.000 usec
 DE 10.00 usec
 TE 294.8 K
 DL 4.0000000 sec
 TDO 1

===== CHANNEL f1 =====
 SFO1 500.1330885 MHz
 NUC1 1H
 P1 3.30 usec
 PLW1 12.19999981 W

F2 - Processing parameters
 SI 65536
 SF 500.1300138 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00



```

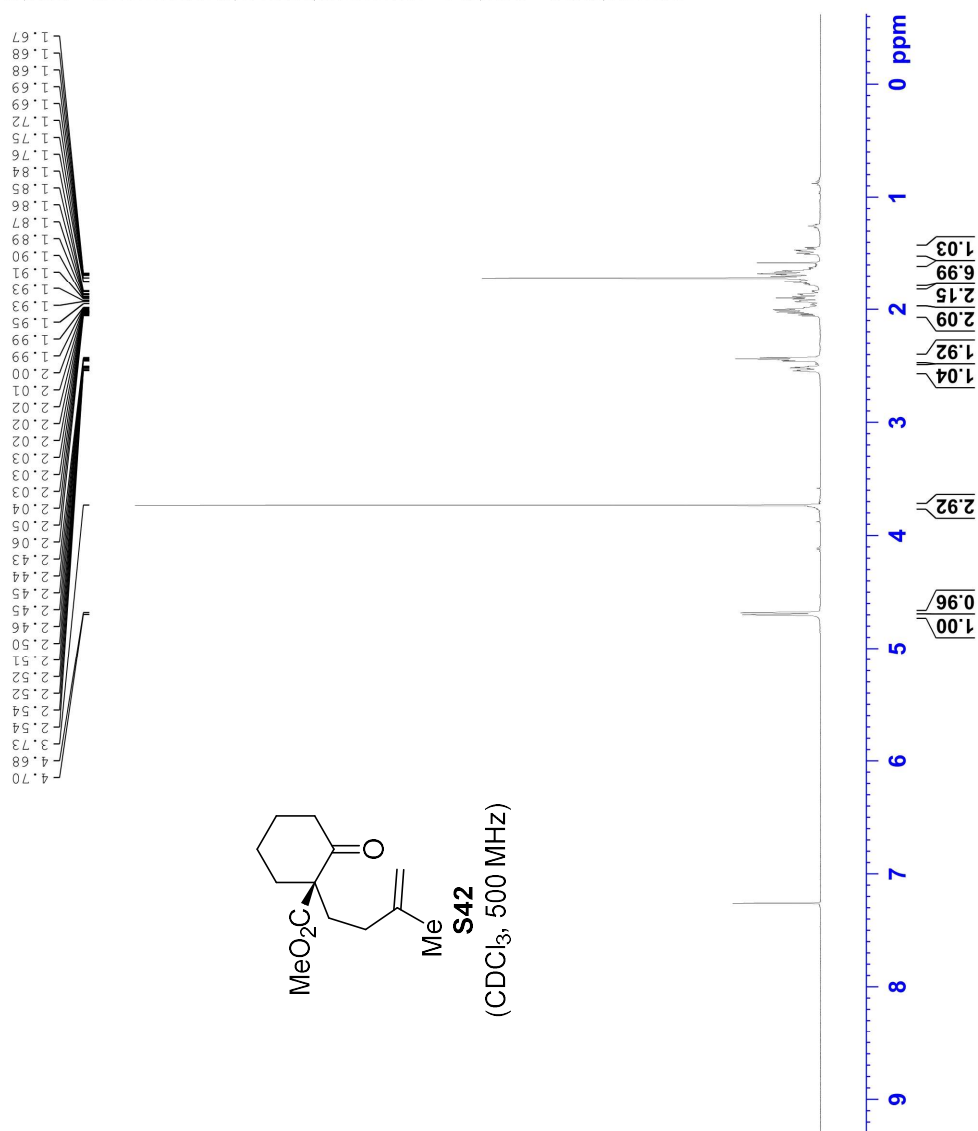
Current Data Parameters
NAME      yh-5-49
EXPNO    1
PROCNO   1

F2 - Acquisition Parameters
Date_    20180813
Time     15.54
INSTRUM spect
PROBHD   5 mm PABBO BB/
PULPROG zg
TD       5998
SOLVENT  CDCl3
NS       2
DS       0
SWH      10000.000 Hz
FIDRES   0.166672 Hz
AQ       2.9999001 sec
RG       77.07
DW       50.000 usec
DE       6.50 usec
TE       295.7 K
DL       4.0000000 sec
TD0      1

===== CHANNEL f1 =====
SFO1    499.8730869 MHz
NUC1     1H
P1       3.58 usec
PLW1    18.25000000 W

F2 - Processing parameters
SI      65536
SF      499.8700122 MHz
WDW     EM
SSB     0
LB      0.30 Hz
GB      0
PC      1.00

```



```

Current Data Parameters
NAME      yh-5-49
EXPNO    2
PROCNO   2

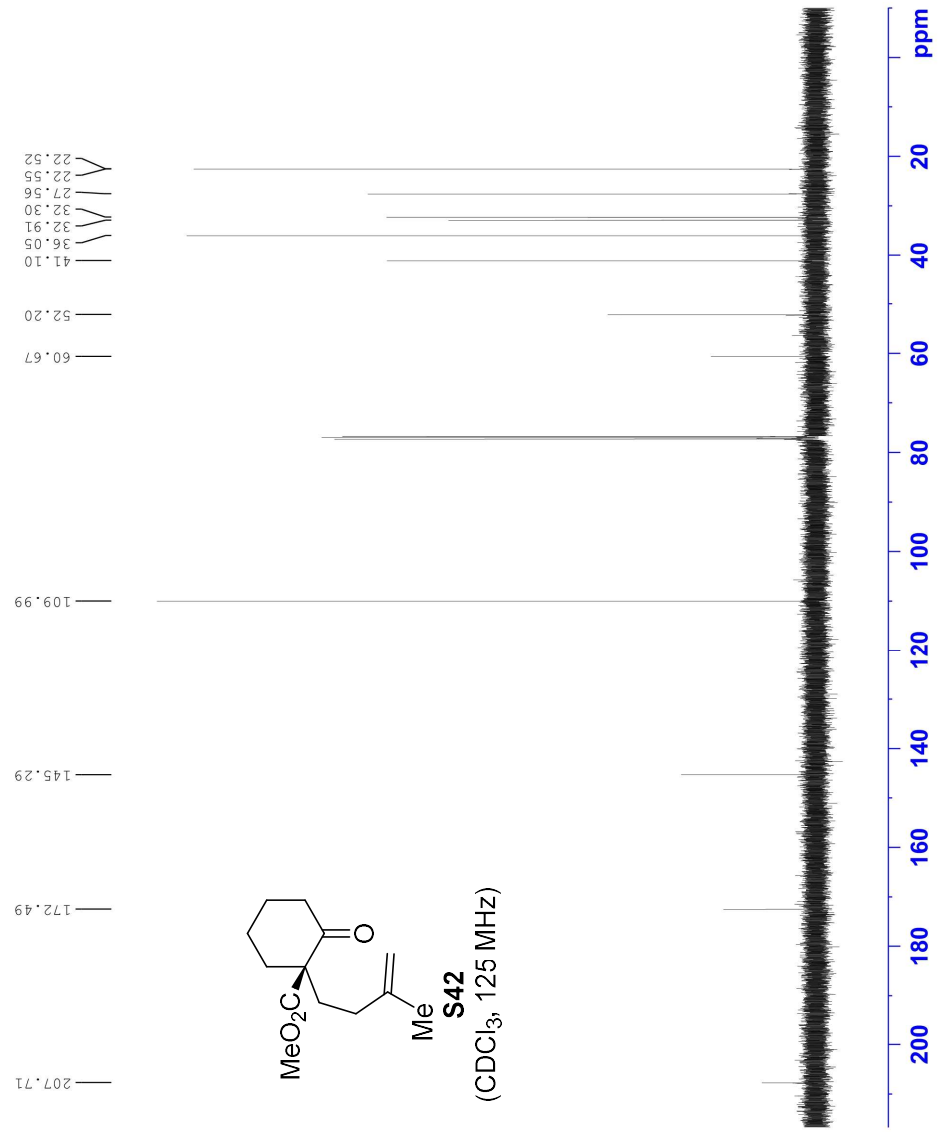
F2 - Acquisition Parameters
Date_    20180813
Time     20.19
INSTRUM spect
PROBHD   5 mm PABBO BB/
PULPROG zgpg
TD       187496
SOLVENT  CDCl3
NS       165
DS       0
SWH      31250.000 Hz
FIDRES   0.166670 Hz
AQ       2.9999361 sec
RG       2050
DW       16.000 usec
DE       6.50 usec
TE       297.3 K
D1       3.0000000 sec
D11      0.0300000 sec
TDO      1

===== CHANNEL f1 =====
SFO1    125.7049802 MHz
NUC1    13C
P1      10.00 usec
PLW1    72.83999634 W

===== CHANNEL f2 =====
SFO2    499.8724993 MHz
NUC2    1H
CFPRG12 waitz16
PCPD2   80.00 usec
PLW2    19.0000000 W
PLW12   0.29688001 W

F2 - Processing parameters
SI      1048576
SF      125.6924115 MHz
WDW     EM
SSB     0
LB      0.30 Hz
GB      0
PC      1.40

```

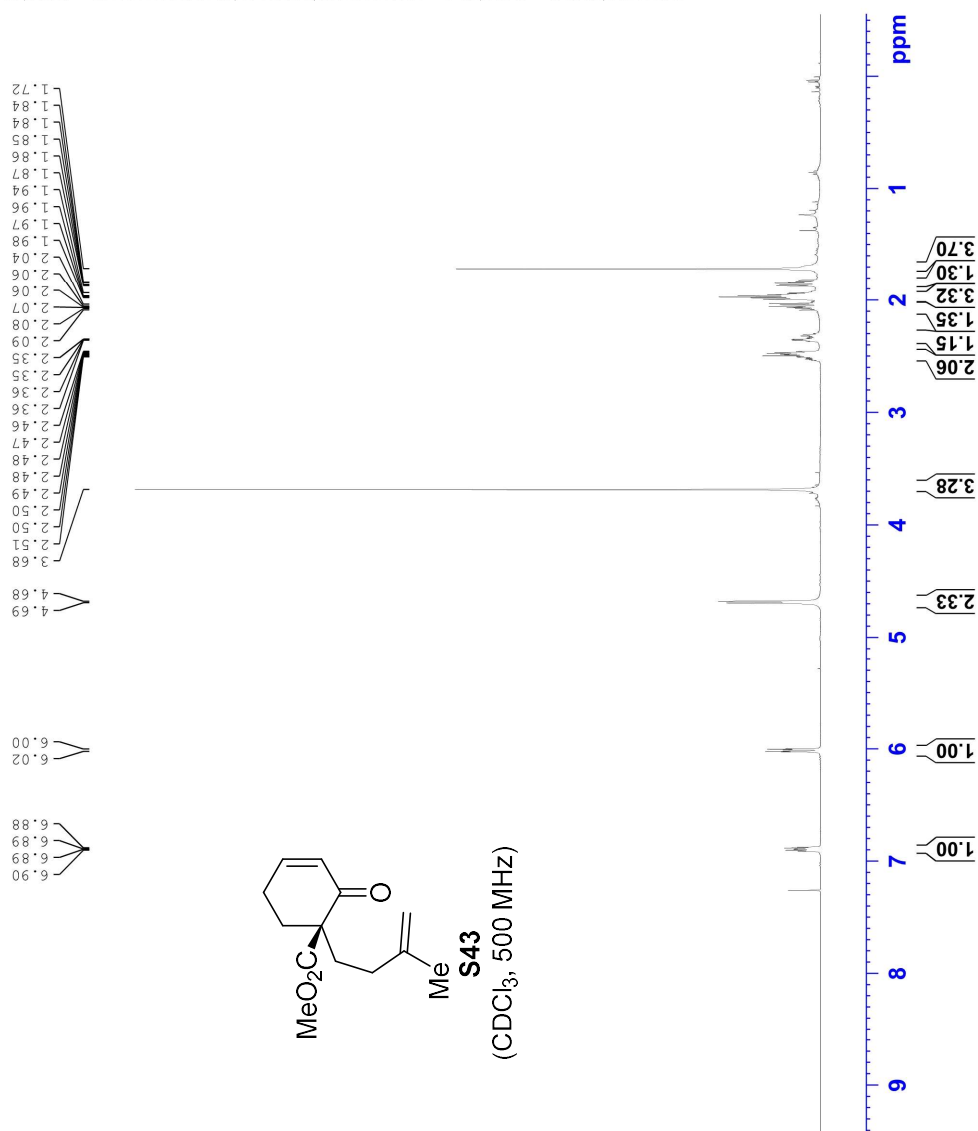


Current Data Parameters
 NAME Yh-4-22-a
 EXPNO 1
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20180812
 Time_ 20.18
 INSTRUM spect
 PROBHD 5 mm PATXI 1H/
 PULPROG zg
 TD 5998
 SOLVENT CDCl3
 NS 8
 DS 0
 SWH 1000.000 Hz
 FIDRES 0.166672 Hz
 AQ 2.9999001 sec
 RG 52.86
 DW 50.000 usec
 DE 10.00 usec
 TE 294.2 K
 DL 3.0000000 sec
 TDO 1

===== CHANNEL f1 =====
 SFO1 500.1330885 MHz
 NUC1 1H
 P1 2.67 usec
 PLW1 12.19999981 W

F2 - Processing parameters
 SI 65536
 SF 500.1300136 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00



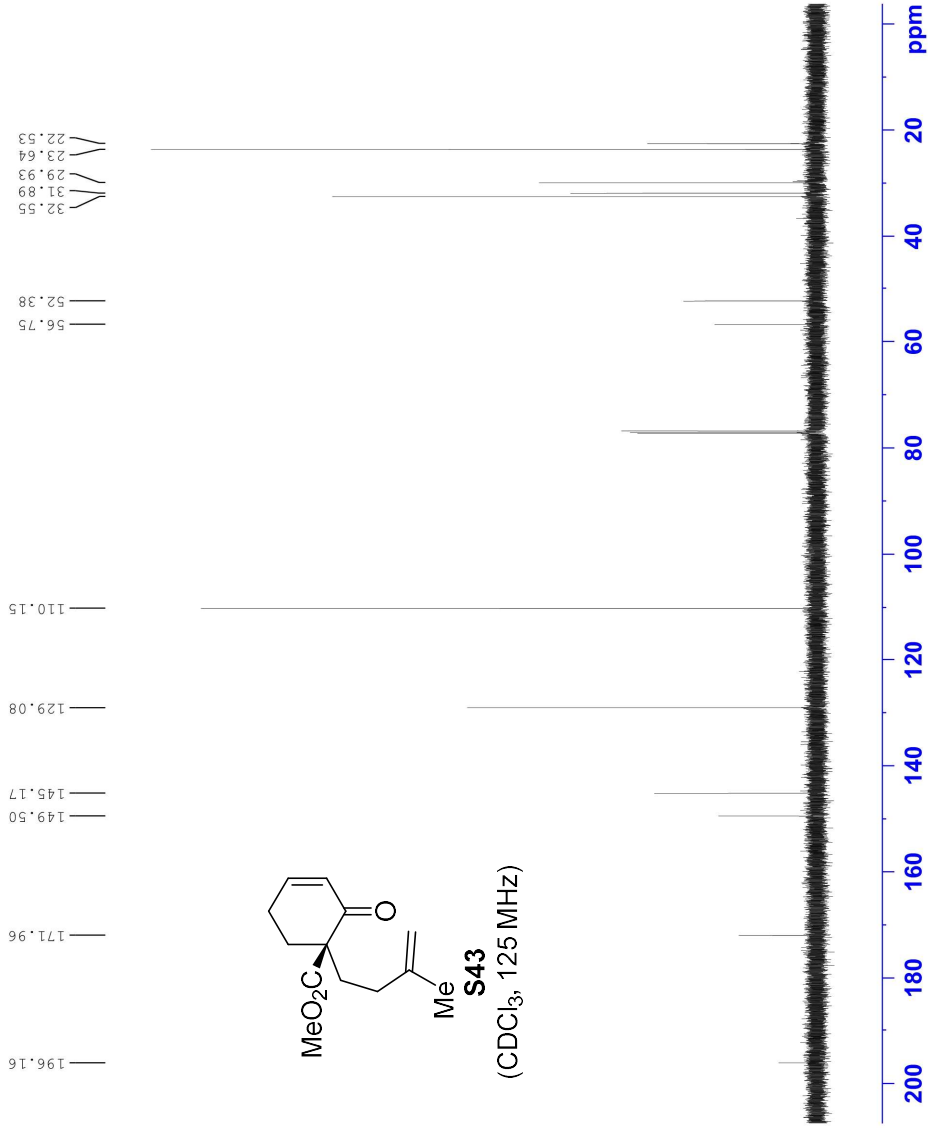
Current Data Parameters
 NAME Yh-4-22-a
 EXPNO 2
 PROCNO 2

F2 - Acquisition Parameters
 Date_ 20180812
 Time_ 20.53
 INSTRUM spect
 PROBHD 5 mm PABBO BB/
 PULPROG zgpg30
 TD 187496
 SOLVENT CDC13
 NS 23
 DS 0
 SWH 31250.000 Hz
 FIDRES 0.166670 Hz
 AQ 2.9999361 sec
 RG 2050
 DW 16.000 usec
 DE 6.50 usec
 TE 296.3 K
 D1 3.0000000 sec
 D11 0.0300000 sec
 TDO 1

==== CHANNEL f1 =====
 SFO1 125.7049802 MHz
 NUC1 13C
 P1 10.00 usec
 PLW1 72.83999634 W

==== CHANNEL f2 =====
 SFO2 499.8724993 MHz
 NUC2 1H
 CPDPRG12 waitz16
 FCPD2 80.00 usec
 PLW2 19.0000000 W
 PLW12 0.29688001 W

F2 - Processing parameters
 SI 1048576
 SF 125.6924115 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.40

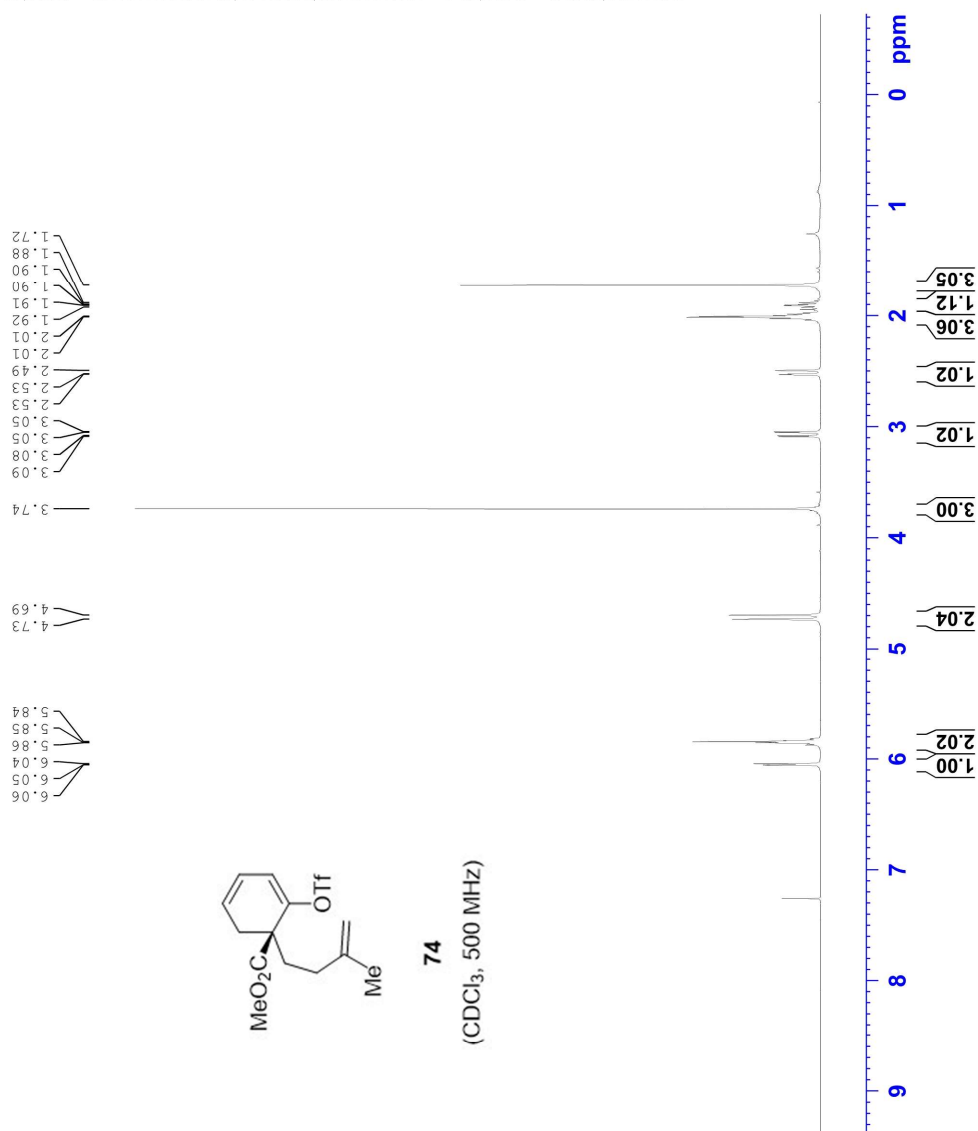


Current Data Parameters
 NAME Yh-4-24-a
 EXPNO 1
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20180314
 Time_ 22.08
 INSTRUM spect
 PROBHD 5 mm PABBO BB/
 PULPROG zg
 TD 5998
 SOLVENT CDCl3
 NS 8
 DS 0
 SWH 10000.000 Hz
 FIDRES 0.166672 Hz
 AQ 2.9999001 sec
 RG 29.95
 DW 50.000 usec
 DE 6.50 usec
 TE 296.7 K
 DL 4.0000000 sec
 TDO 1

===== CHANNEL f1 =====
 SFO1 499.8730869 MHz
 NUC1 1H
 PL 3.58 usec
 PLW1 18.25000000 W

F2 - Processing parameters
 SI 32768
 SF 499.8700123 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00



```

Current Data Parameters
NAME      Yh-4-24-a
EXPNO    2
PROCNO   2

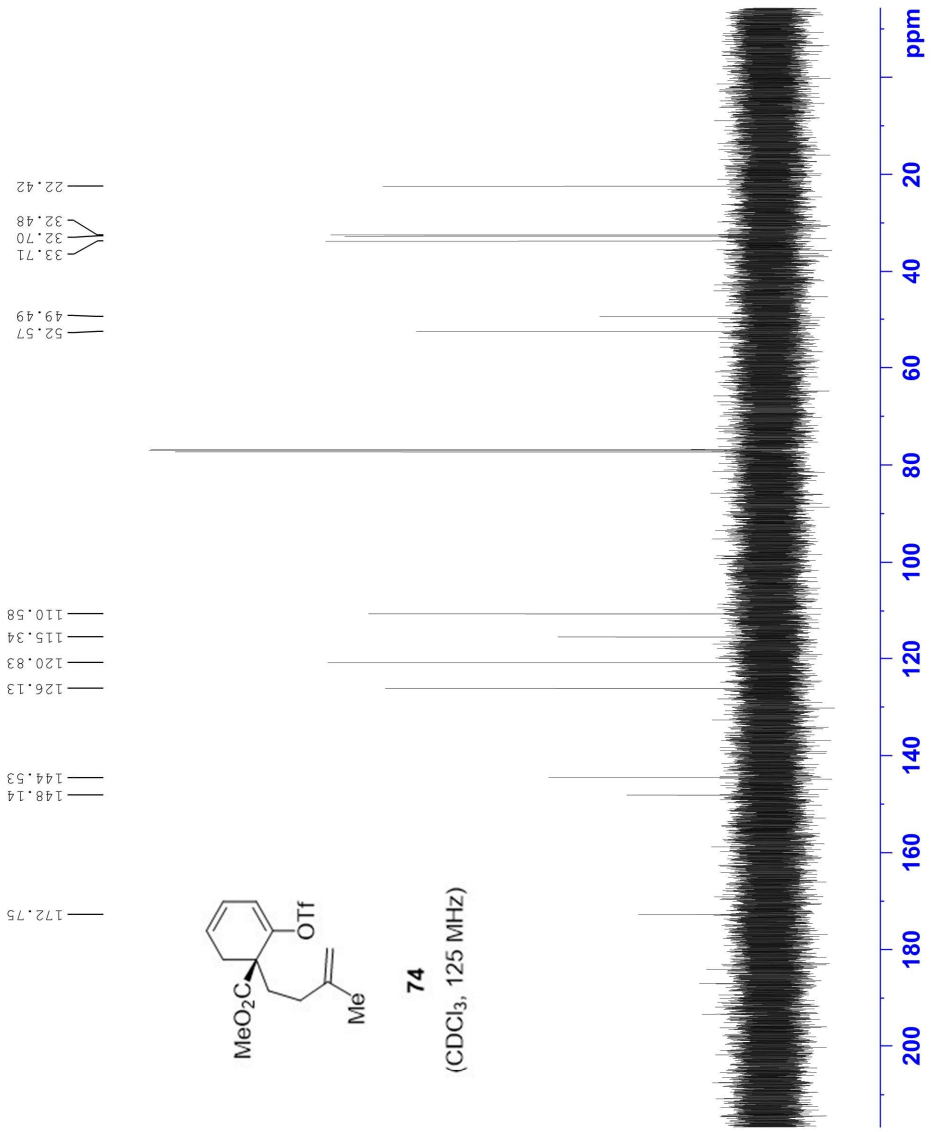
F2 - Acquisition Parameters
Date_    20180314
Time     22.16
INSTRUM  spect
PROBHD   5 mm PABBO BB/
PULPROG  zgpg
TD        187496
SOLVENT  CDCl3
NS        2
DS        0
SWH       31250.000 Hz
FIDRES    0.166670 Hz
AQ         2.9999361 sec
RG         2050
DW         16.000 usec
DE         6.50 usec
TE         296.9 K
D1         3.0000000 sec
D11        0.0300000 sec
TD0        1

===== CHANNEL f1 =====
SFO1      125.7049802 MHz
NUC1      13C
P1         10.00 usec
PLW1      72.83999634 W

===== CHANNEL f2 =====
SFO2      499.8724993 MHz
NUC2      1H
CPDPRG12 waitz16
PCPD2     80.00 usec
PLW2      19.0000000 W
PLW12     0.29688001 W

F2 - Processing parameters
SI         1048576
SF         125.6924115 MHz
WDW        EM
SSB        0
LB         0.30 Hz
GB         0
PC         1.40

```

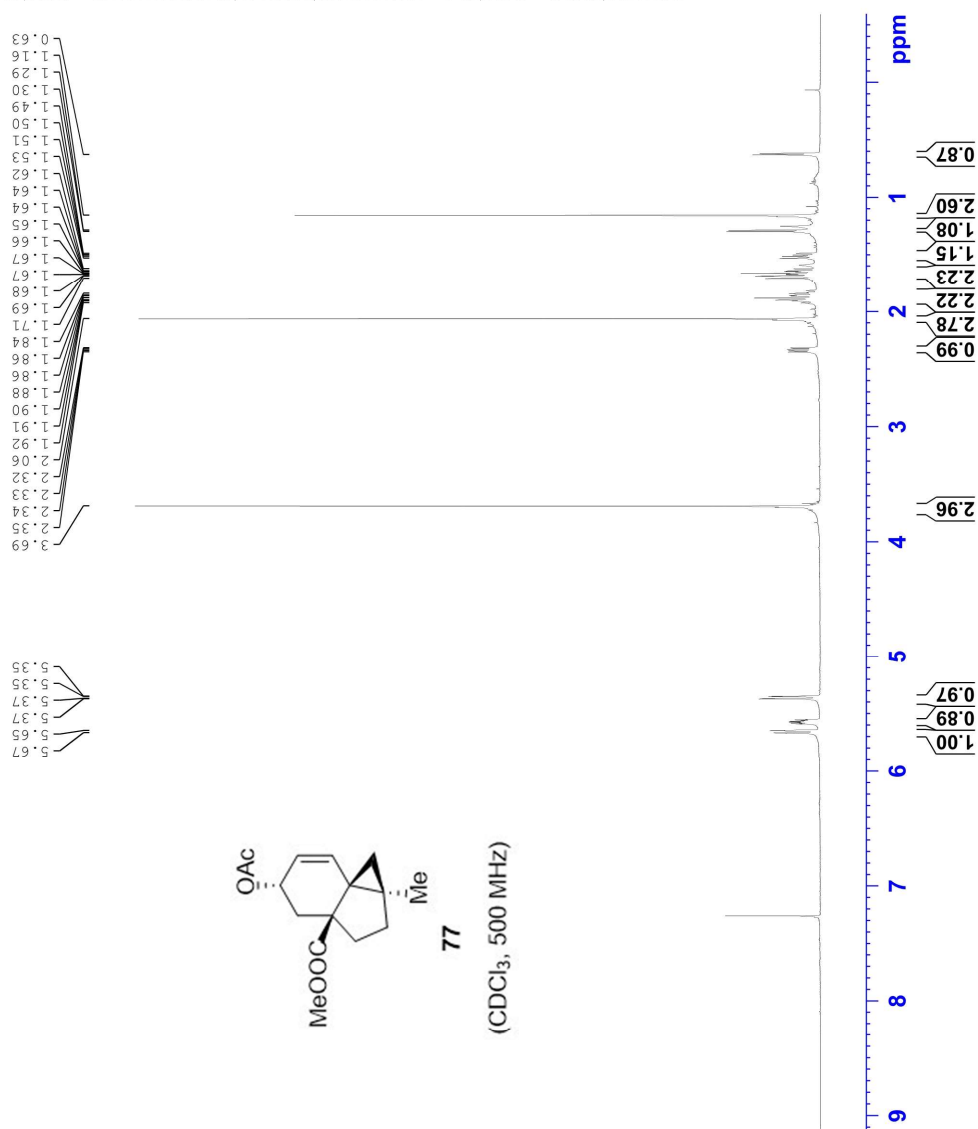


Current Data Parameters
 NAME yh-4-25-2-a
 EXPNO 2
 PROCNO 2

F2 - Acquisition Parameters
 Date_ 20180313
 Time_ 21.11
 INSTRUM spect
 PROBHD 5 mm PABBO BB/
 PULPROG zg
 TD 5998
 SOLVENT CDCl3
 NS 8
 DS 0
 SWH 10000.000 Hz
 FIDRES 0.166672 Hz
 AQ 2.9999001 sec
 RG 51.11
 DW 50.000 usec
 DE 6.50 usec
 TE 296.9 K
 DL 4.0000000 sec
 TDO 1

===== CHANNEL f1 =====
 SFO1 499.8730869 MHz
 NUC1 1H
 P1 3.58 usec
 PLW1 18.25000000 W

F2 - Processing parameters
 SI 65536
 SF 499.8700123 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00



```

Current Data Parameters
NAME      yh-4-25-2-a
EXPNO    3
PROCNO   3

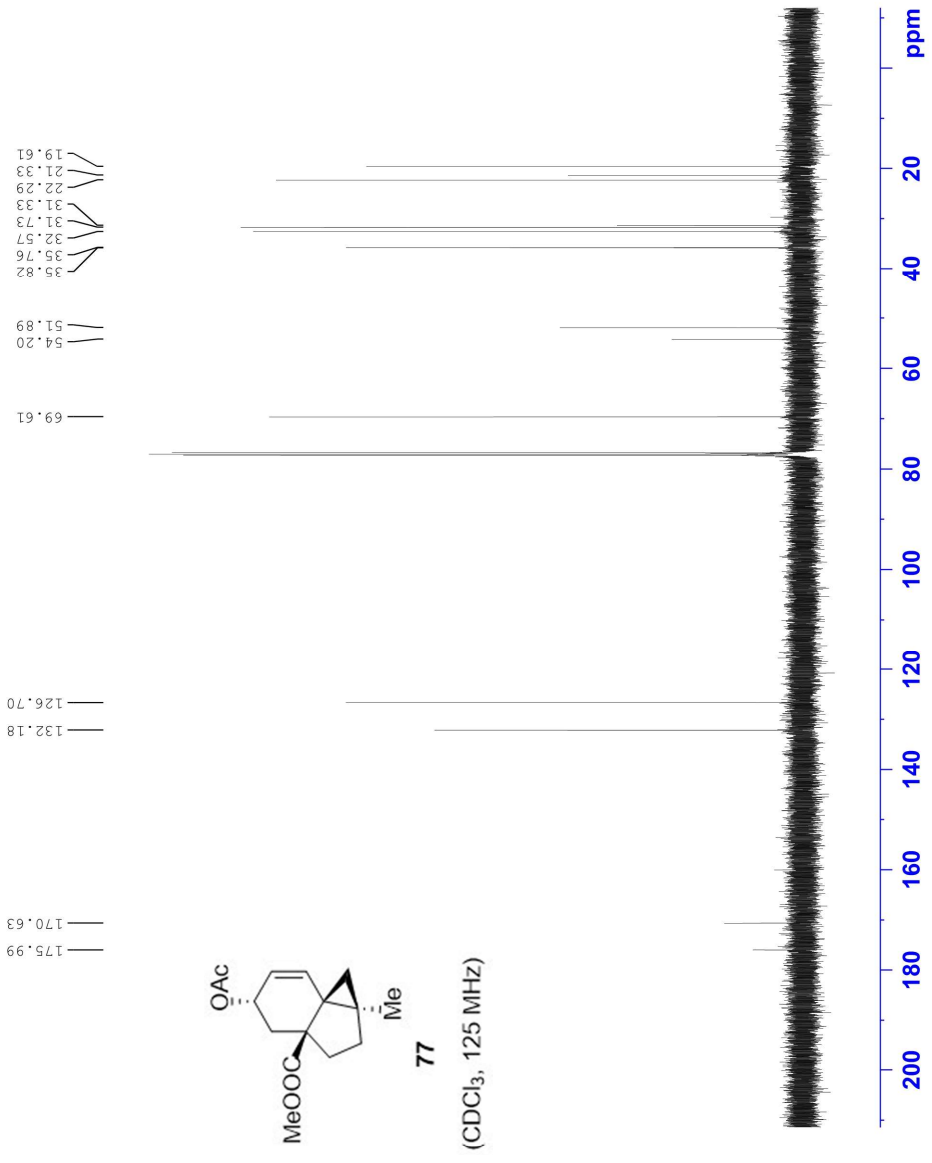
F2 - Acquisition Parameters
Date_    20180313
Time     21.47
INSTRUM  spect
PROBHD   5 mm PABBO BB/
PULPROG  zgpg
TD        187496
SOLVENT  CDCl3
NS        329
DS        0
SWH       31250.000 Hz
FIDRES    0.166670 Hz
AQ         2.9999361 sec
RG         2050
DW         16.000 usec
DE         6.50 usec
TE         298.5 K
D1         3.0000000 sec
D11        0.0300000 sec
TDO       1

===== CHANNEL f1 =====
SFO1      125.7049802 MHz
NUC1       13C
P1         10.00 usec
PLW1      72.83999634 W

===== CHANNEL f2 =====
SFO2      499.8724993 MHz
NUC2       1H
CFPRG12   waitz16
PCPD2     80.00 usec
PLW2      19.0000000 W
PLW12     0.29688001 W

F2 - Processing parameters
SI         1048576
SF         125.6924115 MHz
WDW        EM
SSB        0
LB         0.30 Hz
GB         0
PC         1.40

```

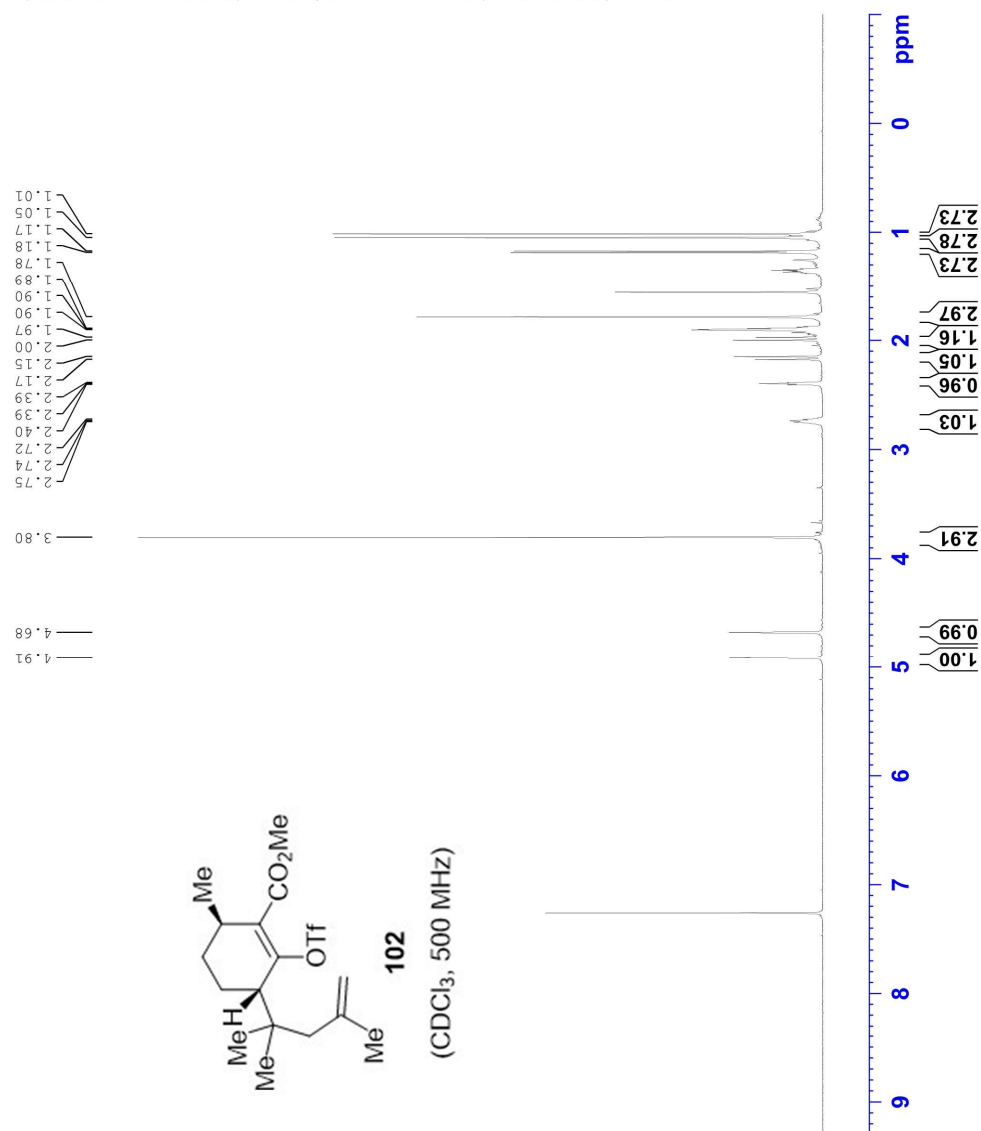


Current Data Parameters
 NAME yn-4-121
 EXPNO 1
 PROCNO 1

F2 - Acquisition Parameters
 Date 20180725
 Time 21.16
 INSTRUM spect
 PROBHD 5 mm PABBO BB/
 PULPROG zg
 TD 5998
 SOLVENT CDCl3
 NS 3
 DS 0
 SWH 10000.000 Hz
 FIDRES 0.166672 Hz
 AQ 2.9999001 sec
 RG 105.66
 DW 50.000 usec
 DE 6.50 usec
 TE 296.5 K
 DL 4.0000000 sec
 TDO 1

===== CHANNEL f1 =====
 SFO1 499.8730869 MHz
 NUC1 1H
 P1 3.58 usec
 PLW1 18.25000000 W

F2 - Processing parameters
 SI 65536
 SF 499.8700122 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00



```

Current Data Parameters
NAME      yn-4-121
EXPNO    2
PROCNO   2

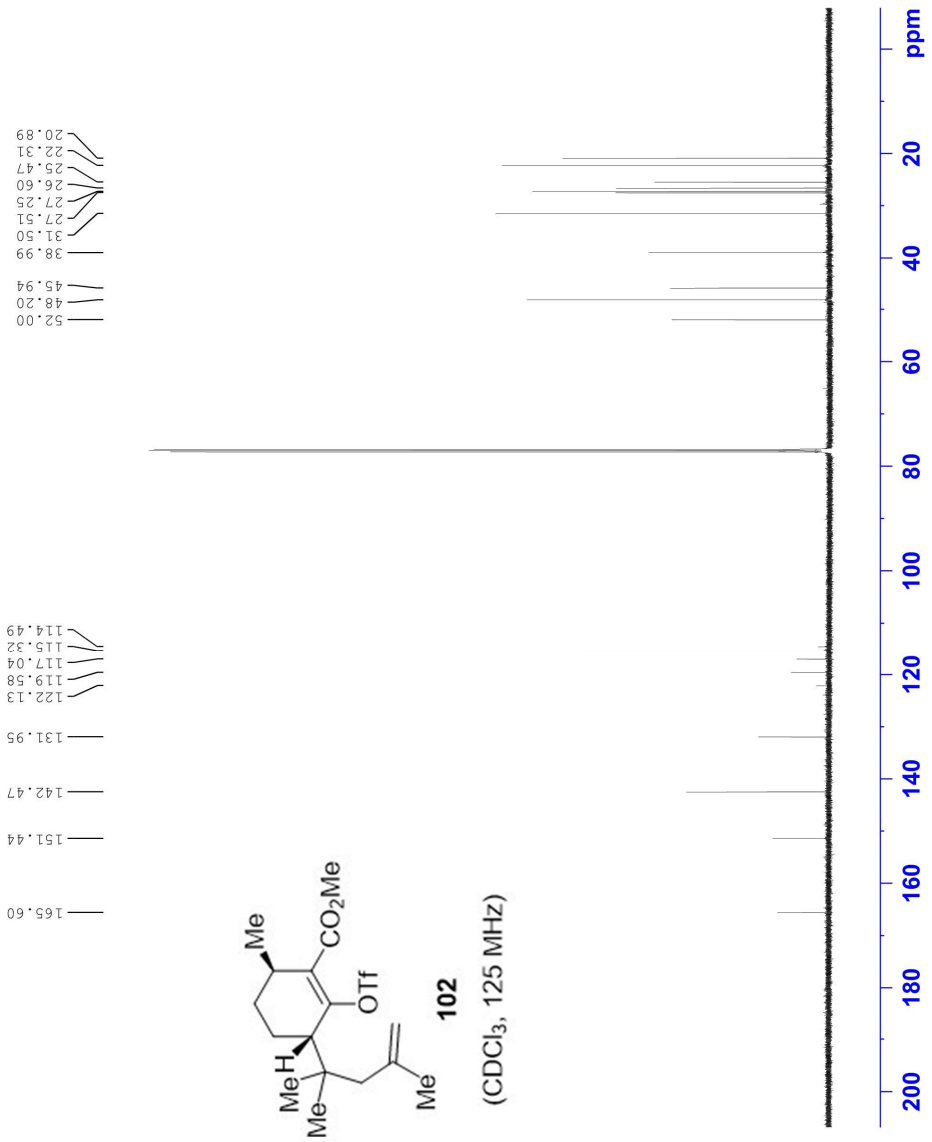
F2 - Acquisition Parameters
Date_    20180726
Time     7.14
INSTRUM  spect
PROBHD   5 mm PABBO BB/
PULPROG  zgpg
TD        187496
SOLVENT  CDCl3
NS        5000
DS        0
SWH       31250.000 Hz
FIDRES    0.166670 Hz
AQ         2.9999361 sec
RG         2050
DW         16.000 usec
DE         6.50 usec
TE         298.4 K
D1         3.0000000 sec
D11        0.0300000 sec
TD0        1

===== CHANNEL f1 =====
SFO1      125.7049802 MHz
NUC1       13C
P1         10.00 usec
PLW1       72.83999634 W

===== CHANNEL f2 =====
SFO2      499.8724993 MHz
NUC2       1H
CFPRG12   waitz16
PCPD2     80.00 usec
PLW2      19.0000000 W
PLW12     0.29688001 W

F2 - Processing parameters
SI         1048576
SF         125.6924115 MHz
WDW        EM
SSB        0
LB         0.30 Hz
GB         0
PC         1.40

```



```

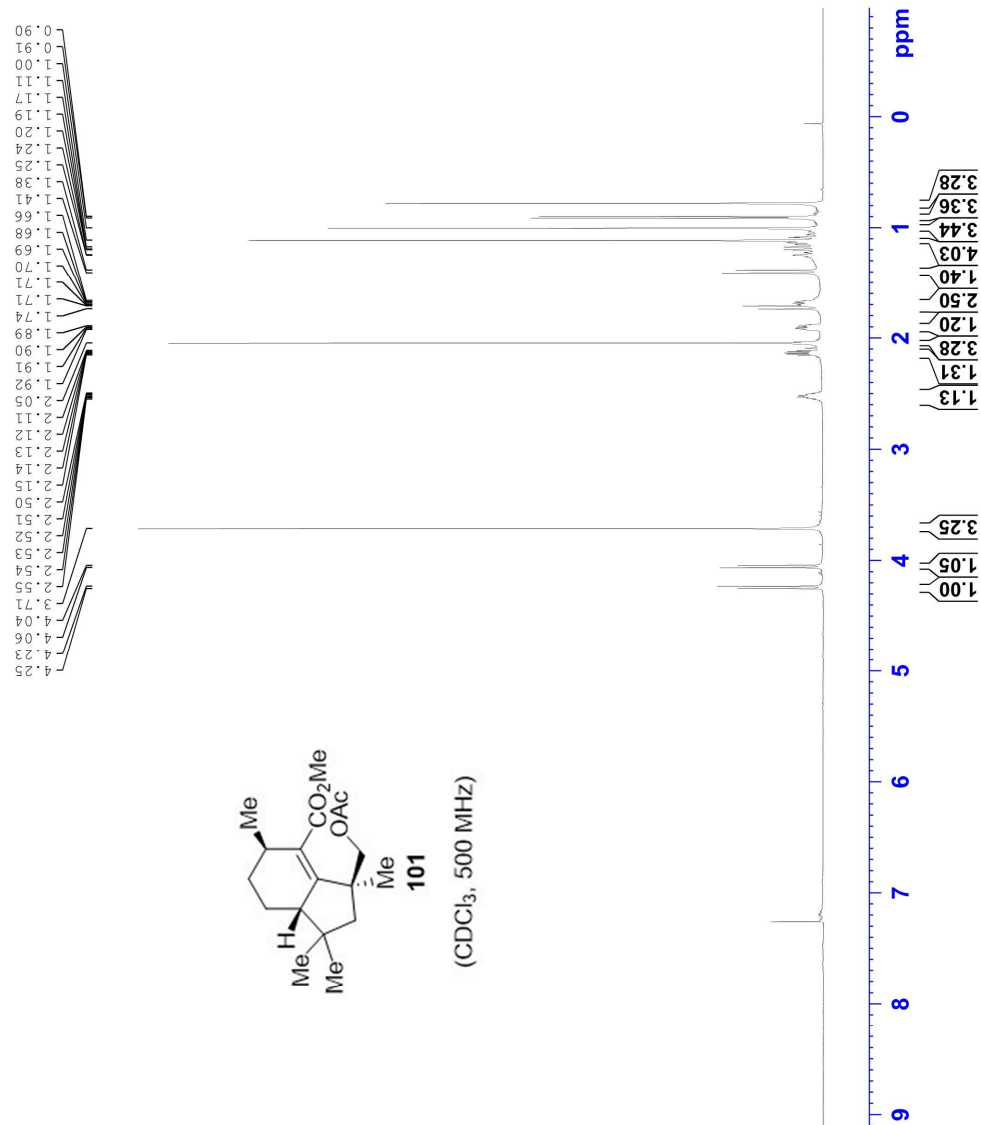
Current Data Parameters
NAME      yh-5-48
EXPNO     1
PROCNO    1

F2 - Acquisition Parameters
Date_     20180809
Time      21.01
INSTRUM   spect
PROBHD    5 mm PABBO BB/
PULPROG   zg
TD         59998
SOLVENT   CDCl3
NS         3
DS         0
SWH        10000.000 Hz
FIDRES     0.166672 Hz
AQ         2.9999001 sec
RG         51.11
DW         50.000 usec
DE         6.50 usec
TE         296.3 K
DL         4.0000000 sec
TDO        1

===== CHANNEL f1 =====
SFO1      499.8730869 MHz
NUC1       1H
P1         3.58 usec
PLW1      18.25000000 W

F2 - Processing parameters
SI         65536
SF         499.8700123 MHz
WDW        EM
SSB        0
LB         0.30 Hz
GB         0
PC         1.00

```



```

Current Data Parameters
NAME      yh-5-48
EXPNO    2
PROCNO   2

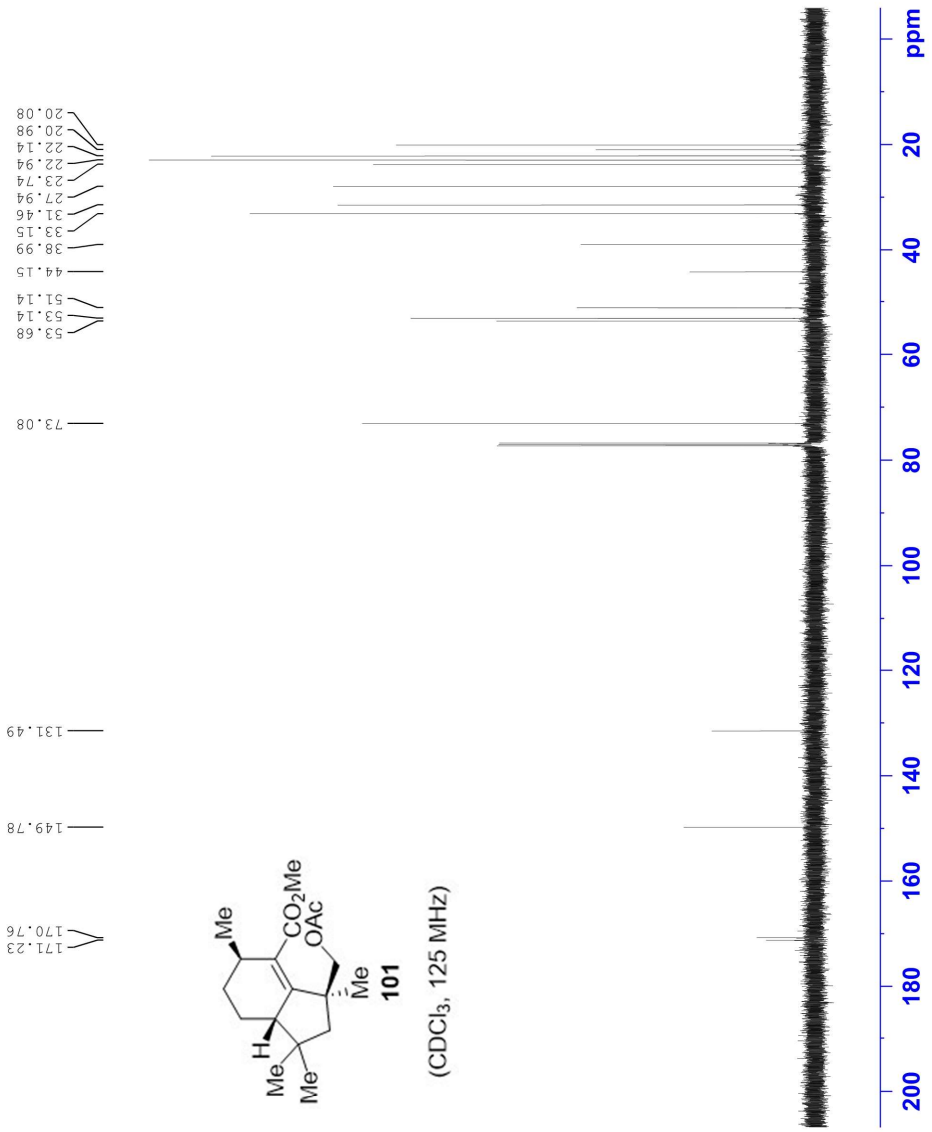
F2 - Acquisition Parameters
Date_    20180809
Time     21.22
INSTRUM spect
PROBHD   5 mm PABBO BB/
PULPROG zgpg
TD       187496
SOLVENT  CDC13
NS       149
DS       0
SWH      31250.000 Hz
FIDRES   0.166670 Hz
AQ       2.9999361 sec
RG       2050
DM       16.000 usec
DE       6.50 usec
TE       297.3 K
D1       3.0000000 sec
D11      0.03000000 sec
TD0      1

===== CHANNEL f1 =====
SFO1    125.7049802 MHz
NUC1    13C
P1      10.00 usec
PLW1    72.83999634 W

===== CHANNEL f2 =====
SFO2    499.8724993 MHz
NUC2    1H
PCPDG12 waitz16
PCPD2   80.00 usec
PLW2    19.0000000 W
PLW12   0.29688001 W

F2 - Processing parameters
SI      1048576
SF      125.6924115 MHz
WDW     EM
SSB     0
LB      0.30 Hz
GB      0
PC      1.40

```



```

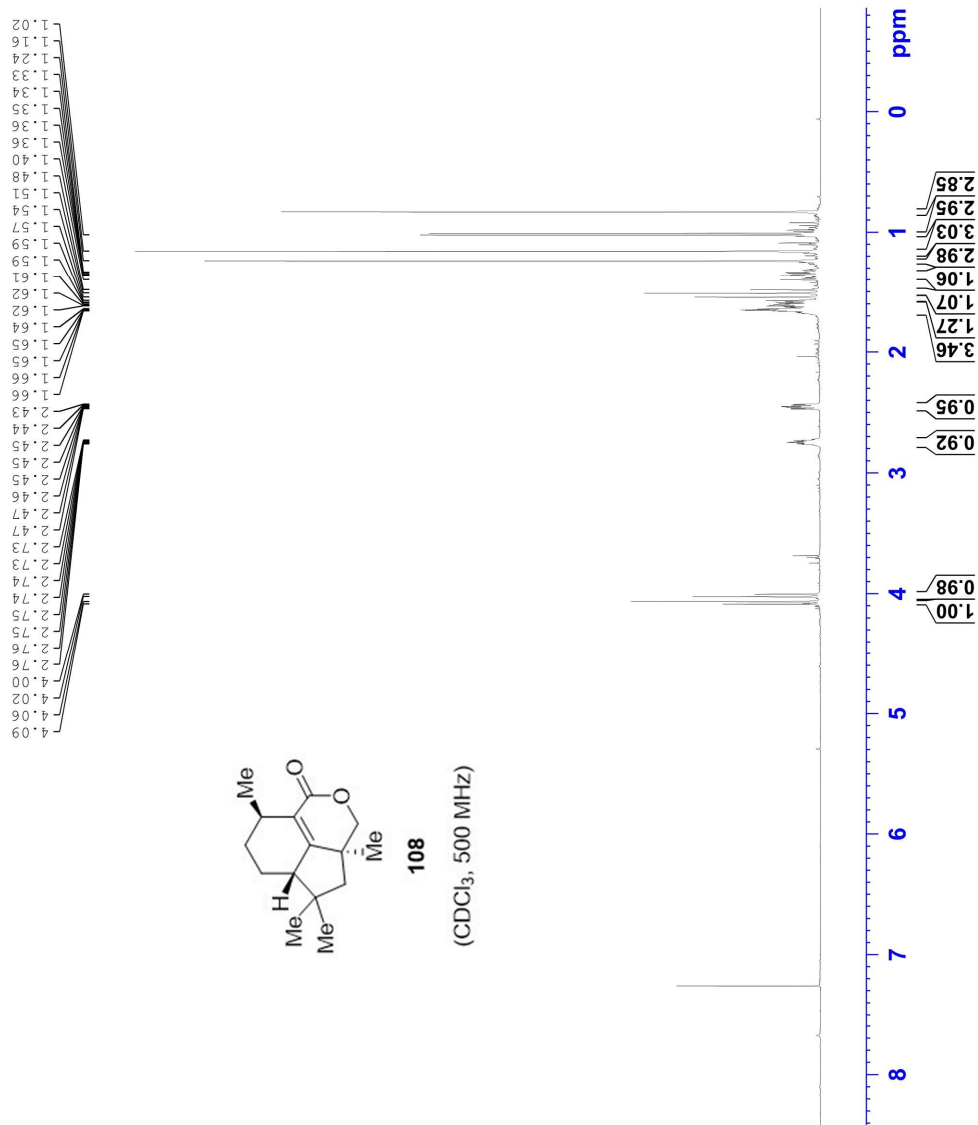
Current Data Parameters
NAME      Yh-4-57-B
EXPNO    1
PROCNO   1

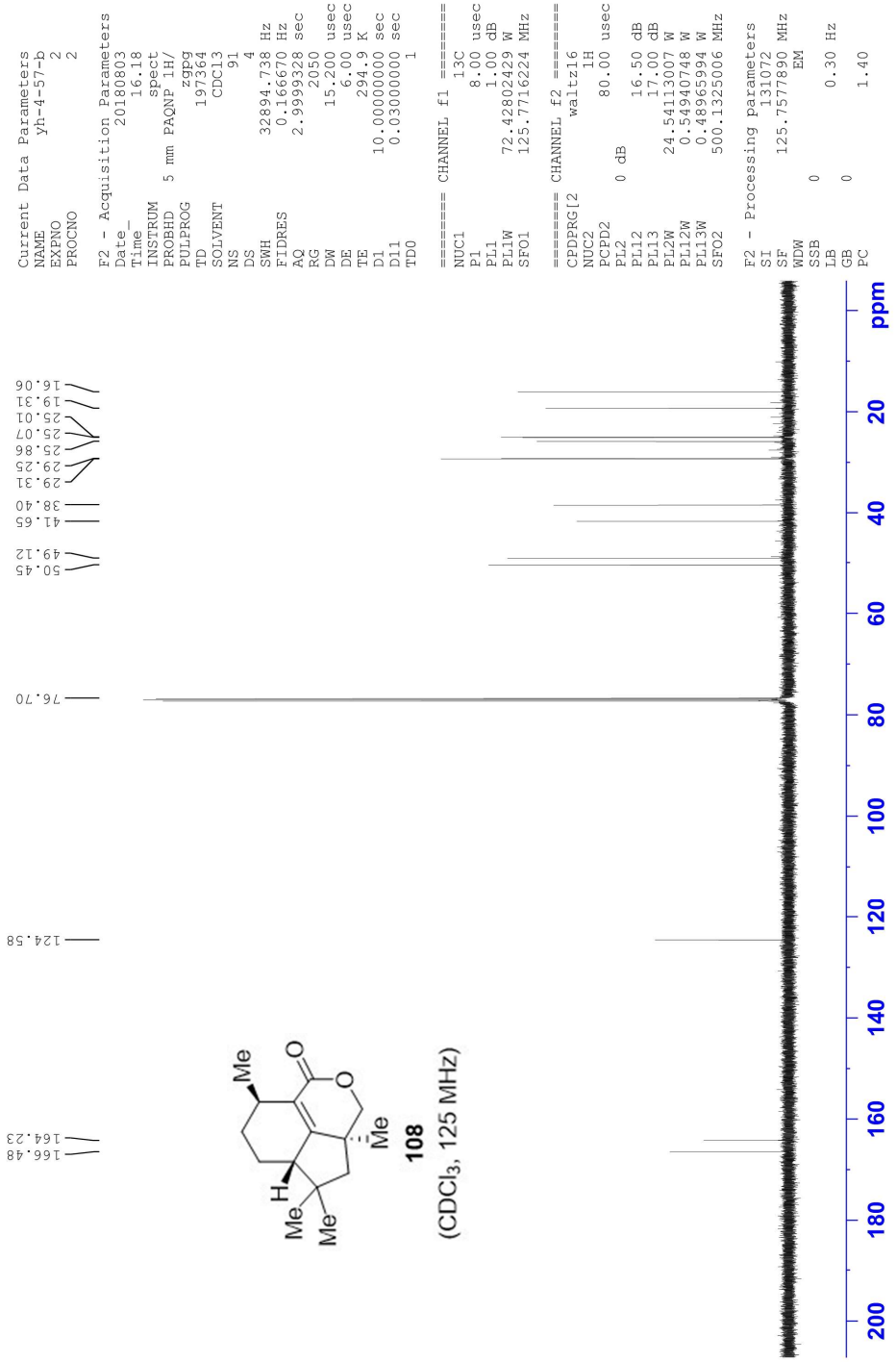
F2 - Acquisition Parameters
Date_    20180803
Time     11.05
INSTRUM spect
PROBHD   5 mm PATXI 1H/
PULPROG zg
TD       59998
SOLVENT  CDCl3
NS       8
DS       0
SWH      10000.000 Hz
FIDRES   0.166672 Hz
AQ       2.9999001 sec
RG       110.37
DW       50.000 usec
DE       10.00 usec
TE       294.1 K
DL       3.0000000 sec
TD0      1

===== CHANNEL f1 =====
SFO1    500.1330885 MHz
NUC1    1H
P1       2.67 usec
PLW1    12.19999981 W

F2 - Processing parameters
SI      65536
SF      500.1300137 MHz
WDW     EM
SSB     0
LB      0.30 Hz
GB      0
PC      1.00

```



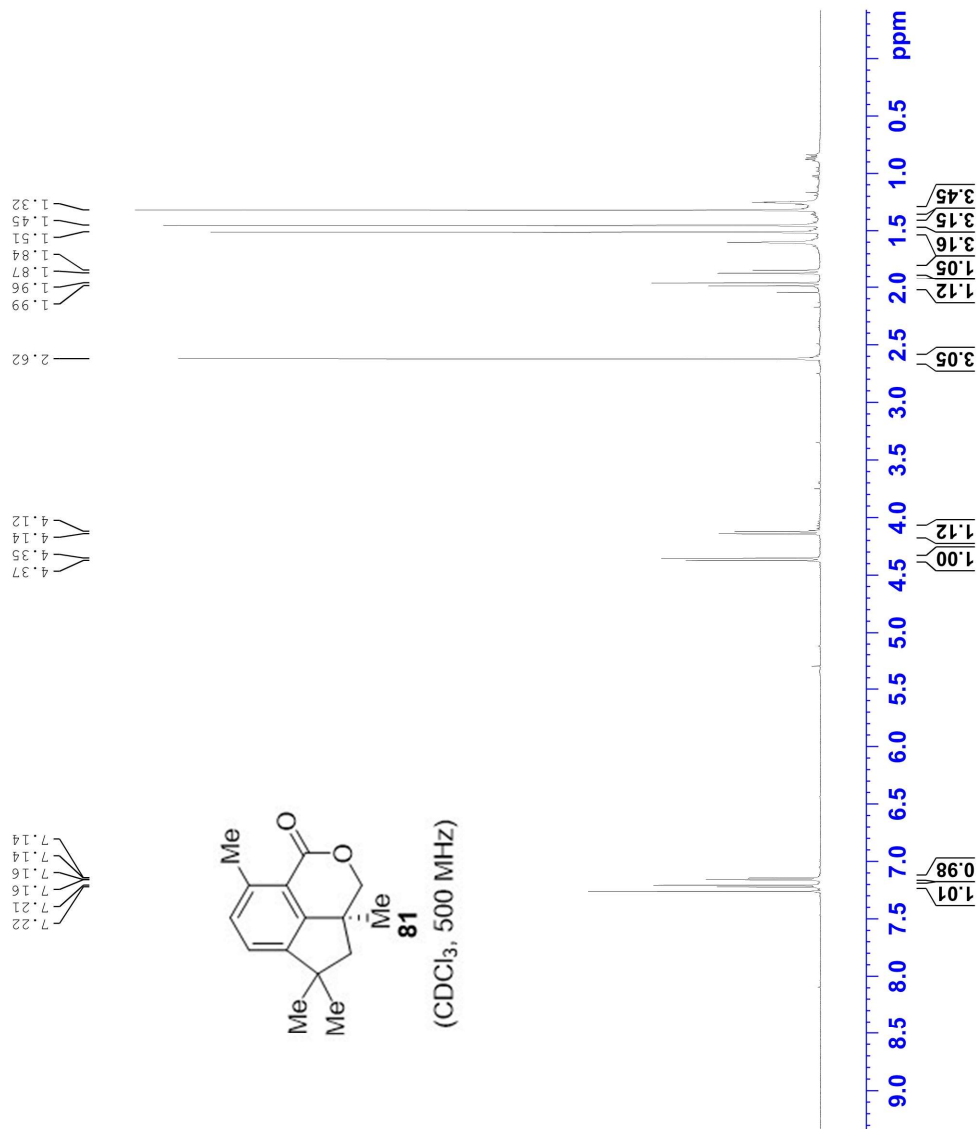


Current Data Parameters
 NAME yh-5-47
 EXPNO 1
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20180804
 Time_ 22.32
 INSTRUM spect
 PROBHD 5 mm PATXI 1H/
 PULPROG zg
 TD 5998
 SOLVENT CDCl3
 NS 2
 DS 0
 SWH 10000.000 Hz
 FIDRES 0.166672 Hz
 AQ 2.9999001 sec
 RG 196.79
 DW 50.000 usec
 DE 10.00 usec
 TE 294.4 K
 DL 3.0000000 sec
 TDO 1

===== CHANNEL f1 =====
 SFO1 500.1330885 MHz
 NUC1 1H
 PL 2.67 usec
 PLW1 12.19999981 W

F2 - Processing parameters
 SI 65536
 SF 500.1300138 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00



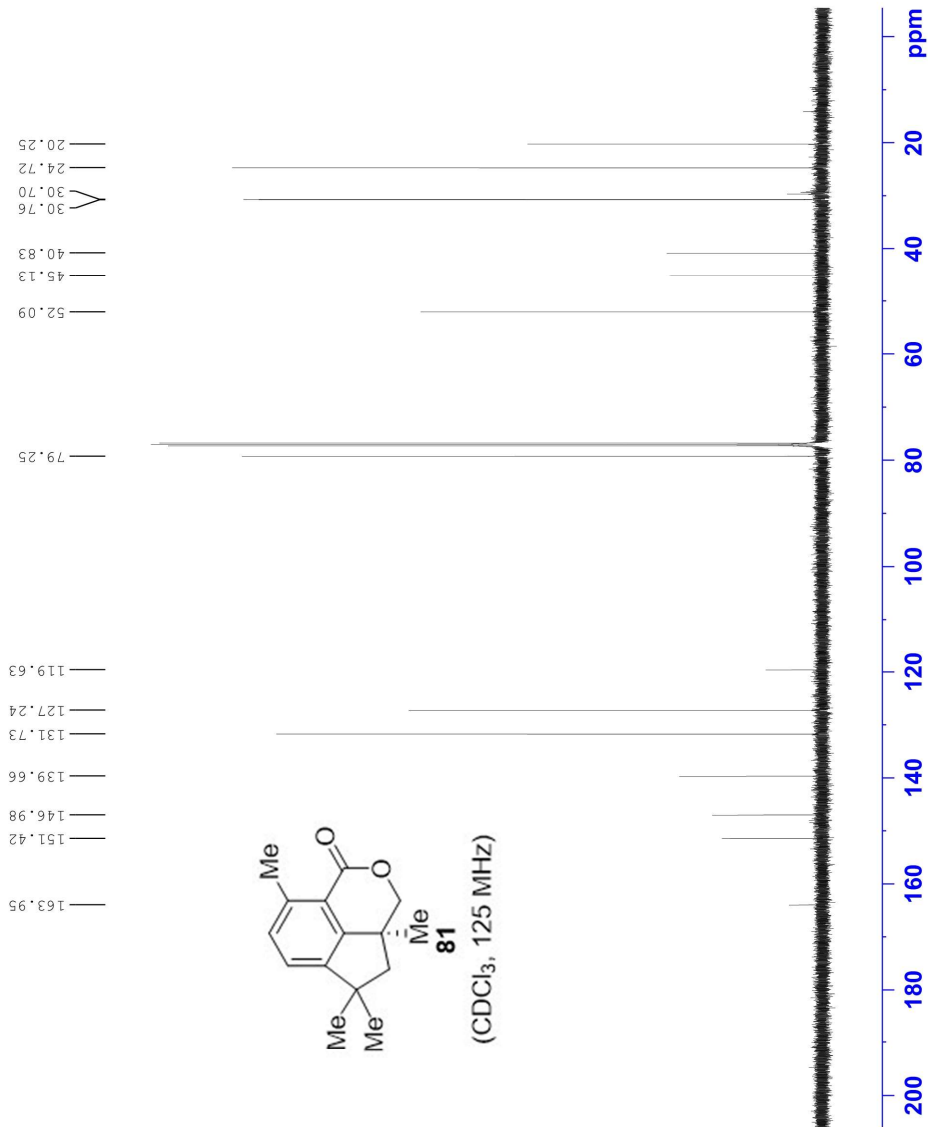
Current Data Parameters
 NAME Yh-5-47-a
 EXPNO 2
 PROCNO 2

F2 - Acquisition Parameters
 Date_ 20180805
 Time_ 20.54
 INSTRUM spect
 PROBHD 5 mm PABBO BB/
 PULPROG zgpgc
 TD 187496
 SOLVENT CDC13
 NS 872
 DS 0
 SWH 31250.000 Hz
 FIDRES 0.166670 Hz
 AQ 2.9999361 sec
 RG 2050
 DW 16.000 usec
 DE 6.50 usec
 TE 297.9 K
 D1 3.0000000 sec
 D11 0.0300000 sec
 TDO 1

==== CHANNEL f1 =====
 SFO1 125.7049802 MHz
 NUC1 13C
 P1 10.00 usec
 PLW1 72.83999634 W

==== CHANNEL f2 =====
 SFO2 499.8724993 MHz
 NUC2 1H
 CPDPRG12 waitz16
 PCPD2 80.00 usec
 PLW2 19.0000000 W
 PLW12 0.29688001 W

F2 - Processing parameters
 SI 1048576
 SF 125.6924115 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.40



```

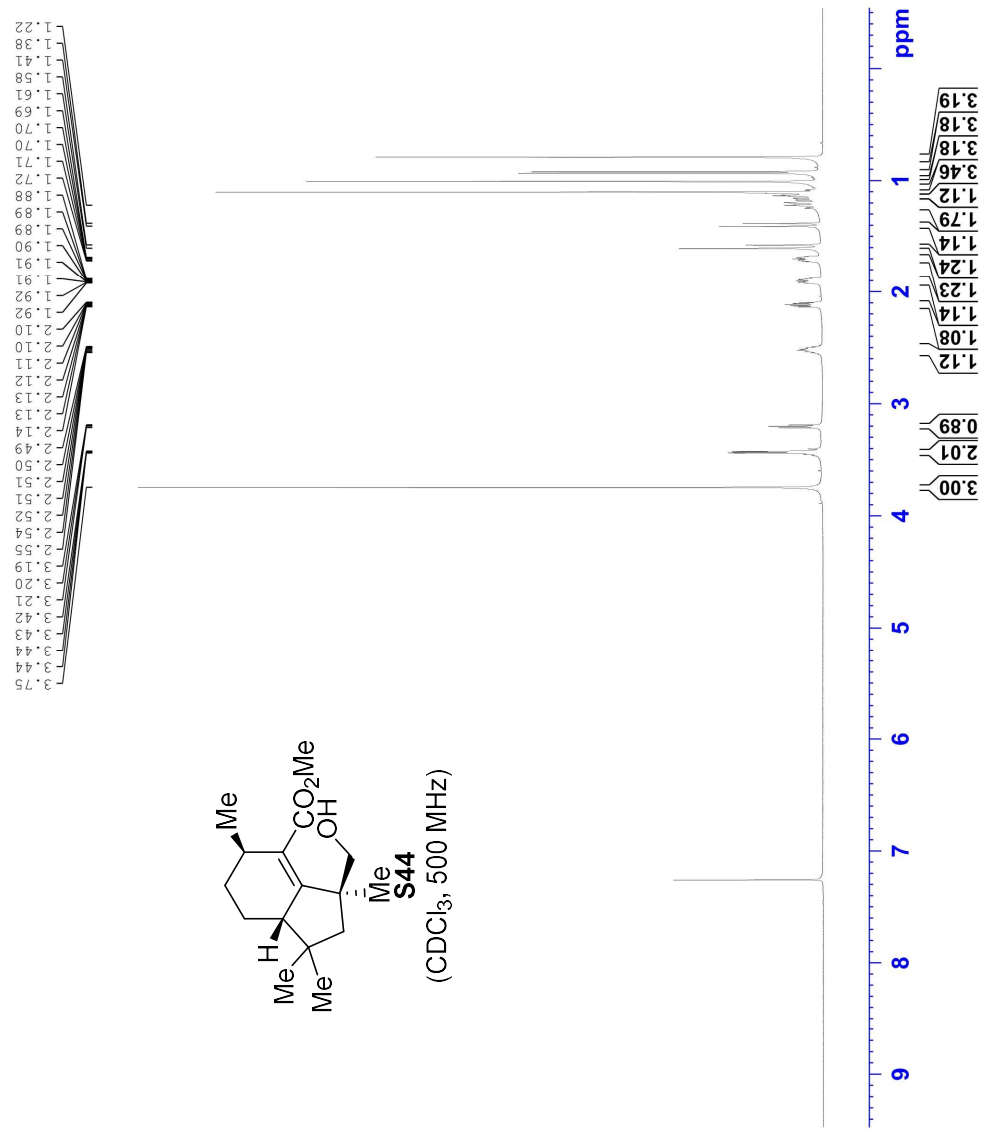
Current Data Parameters
NAME      yn-4-107
EXPNO    1
PROCNO   1

F2 - Acquisition Parameters
Date_    20180525
Time     21.45
INSTRUM spect
PROBHD   5 mm PABBO BB/
PULPROG zg
TD       5998
SOLVENT  CDCl3
NS       7
DS       0
SWH      10000.000 Hz
FIDRES   0.166672 Hz
AQ       2.9999001 sec
RG       77.07
DW       50.000 usec
DE       6.50 usec
TE       296.3 K
DL       4.0000000 sec
TD0      1

===== CHANNEL f1 =====
SFO1    499.8730869 MHz
NUC1    1H
P1      3.58 usec
PLW1    18.25000000 W

F2 - Processing parameters
SI      65536
SF      499.8700122 MHz
WDW     EM
SSB     0
LB      0.30 Hz
GB      0
PC      1.00

```



```

Current Data Parameters
NAME      yn-4-107
EXPNO    2
PROCNO   2

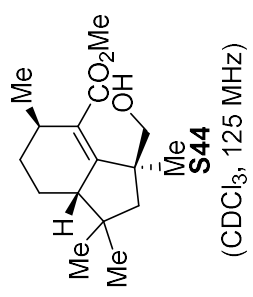
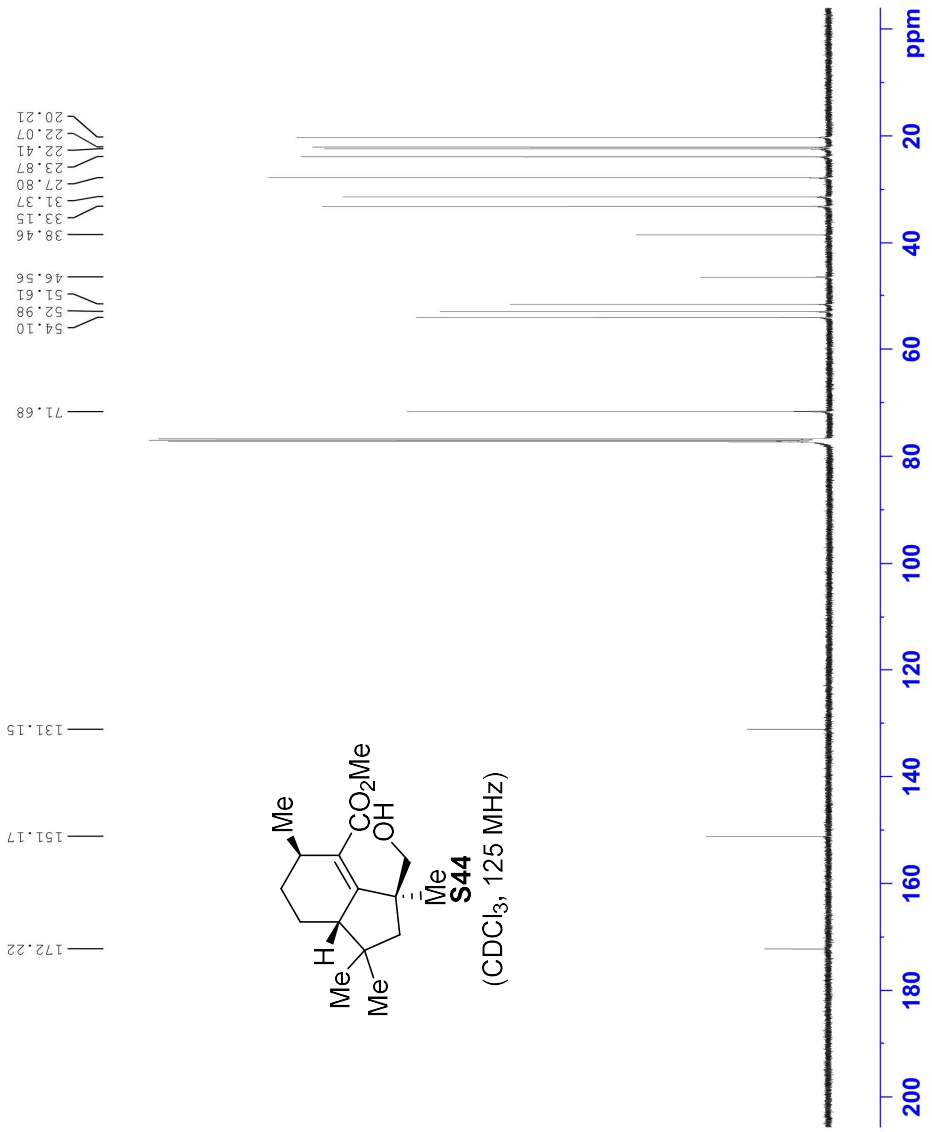
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TE        297.7 K
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PLW12     0.29688001 W

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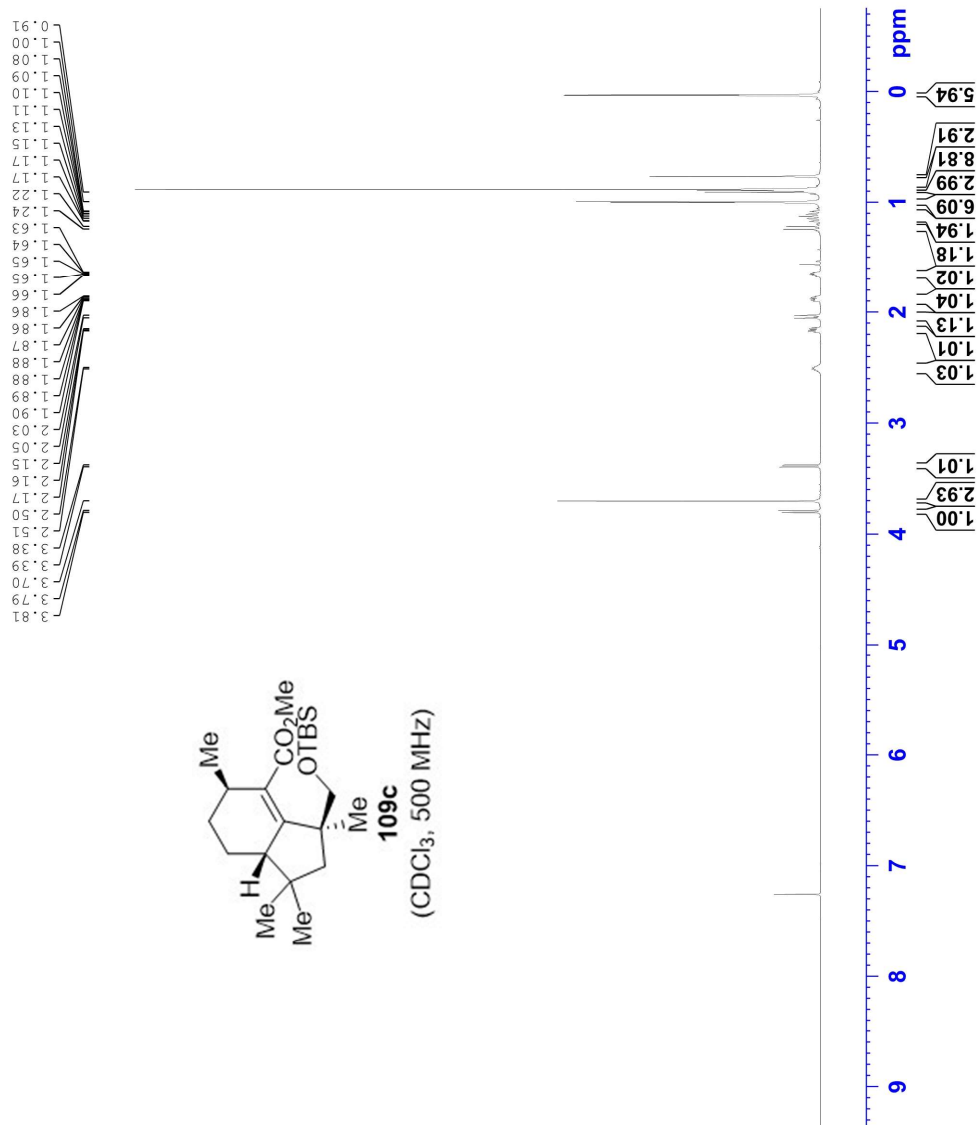


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 TE 295.6 K
 DL 4.0000000 sec
 TDO 1

===== CHANNEL f1 =====
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 NUC1 1H
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F2 - Processing parameters
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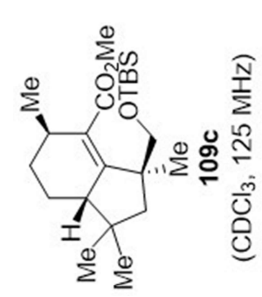
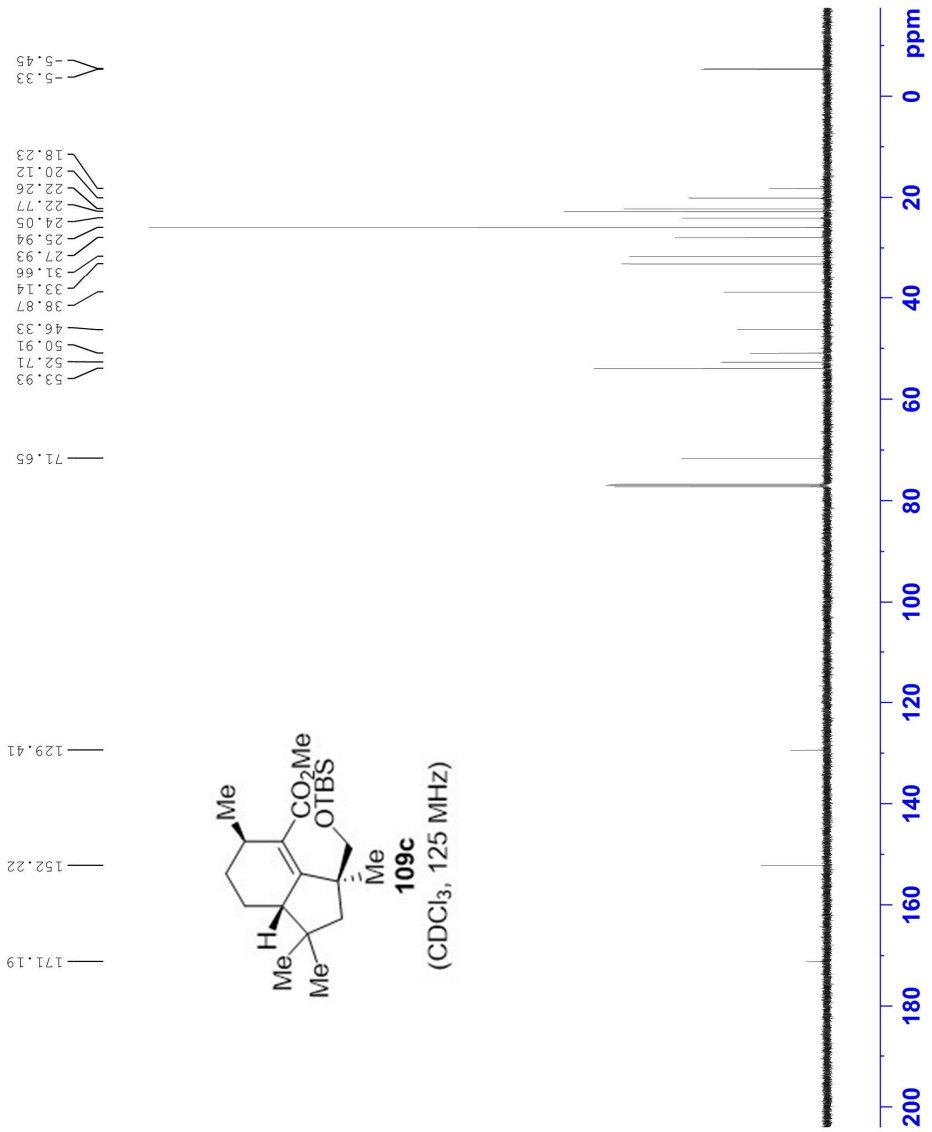
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P1         10.00 usec
PLW1      72.83999634 W

===== CHANNEL f2 =====
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NUC2       1H
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PCPD2     80.00 usec
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PLW12     0.29688001 W

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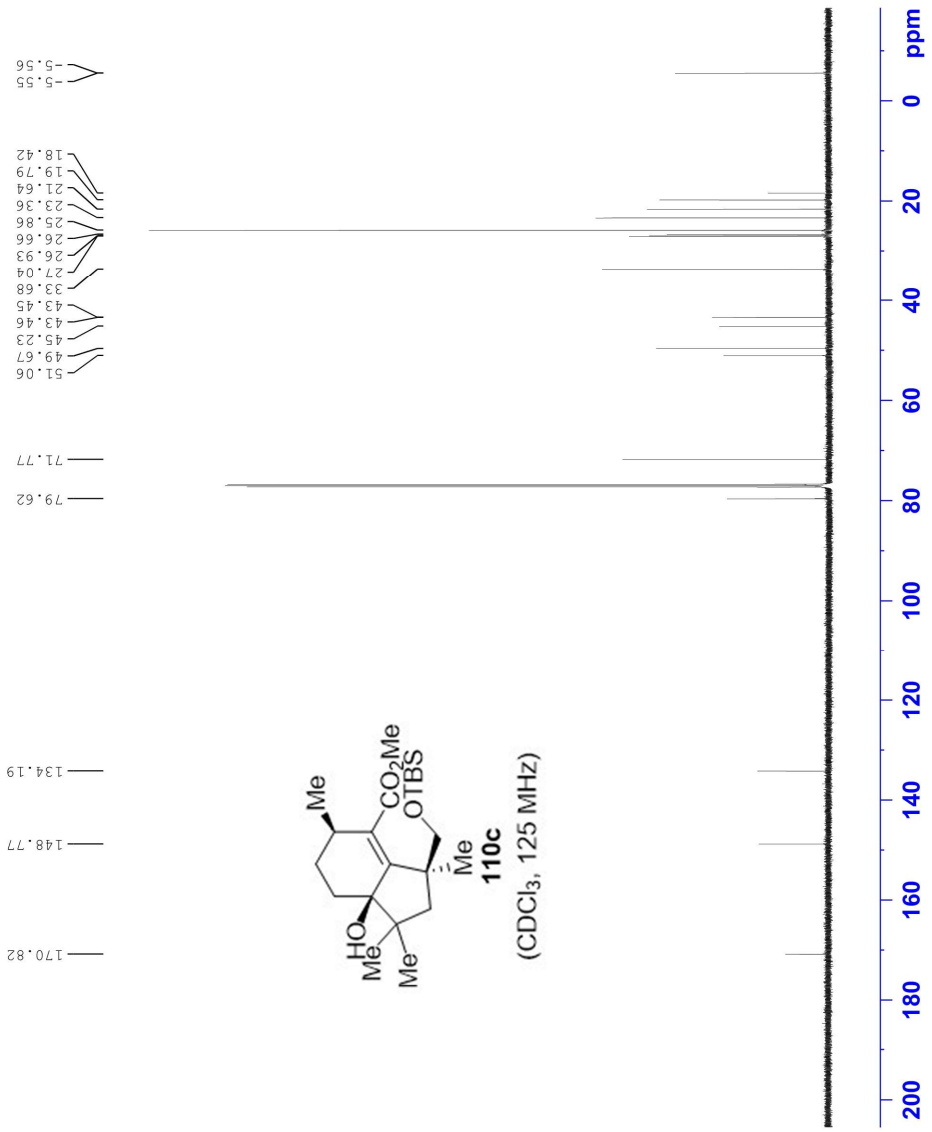
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 DE 6.50 usec
 TE 297.6 K
 D1 3.0000000 sec
 D11 0.0300000 sec
 TDO 1

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 NUC1 13C
 P1 10.00 usec
 PLW1 72.83999634 W

==== CHANNEL f2 =====
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 NUC2 1H
 CPDPRG2 waitz16
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F2 - Processing parameters
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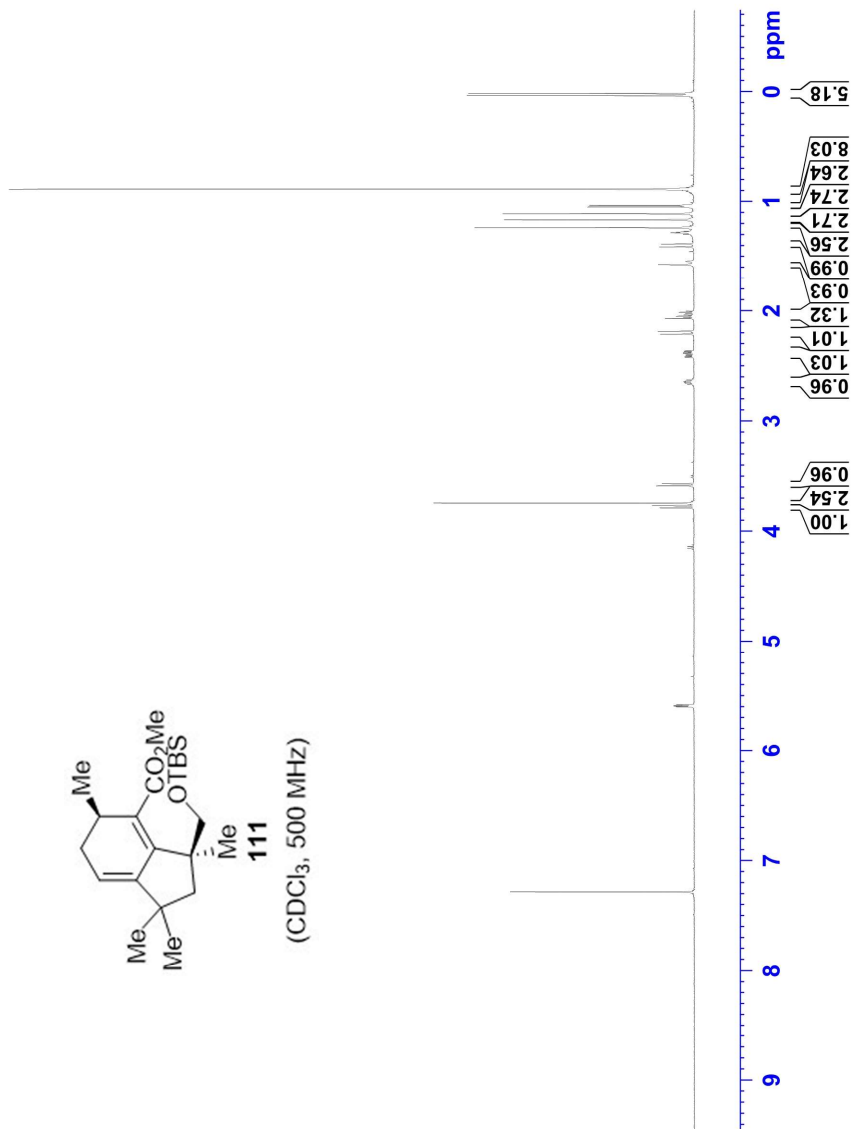
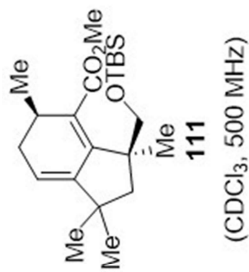
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===== CHANNEL f1 =====
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 NUC1 1H
 P1 3.58 usec
 PLW1 18.25000000 W

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3.74
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3.57
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2.66
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2.64
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2.41
2.40
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2.38
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2.36
2.21
2.19
2.07
2.06
2.05
2.04
2.02
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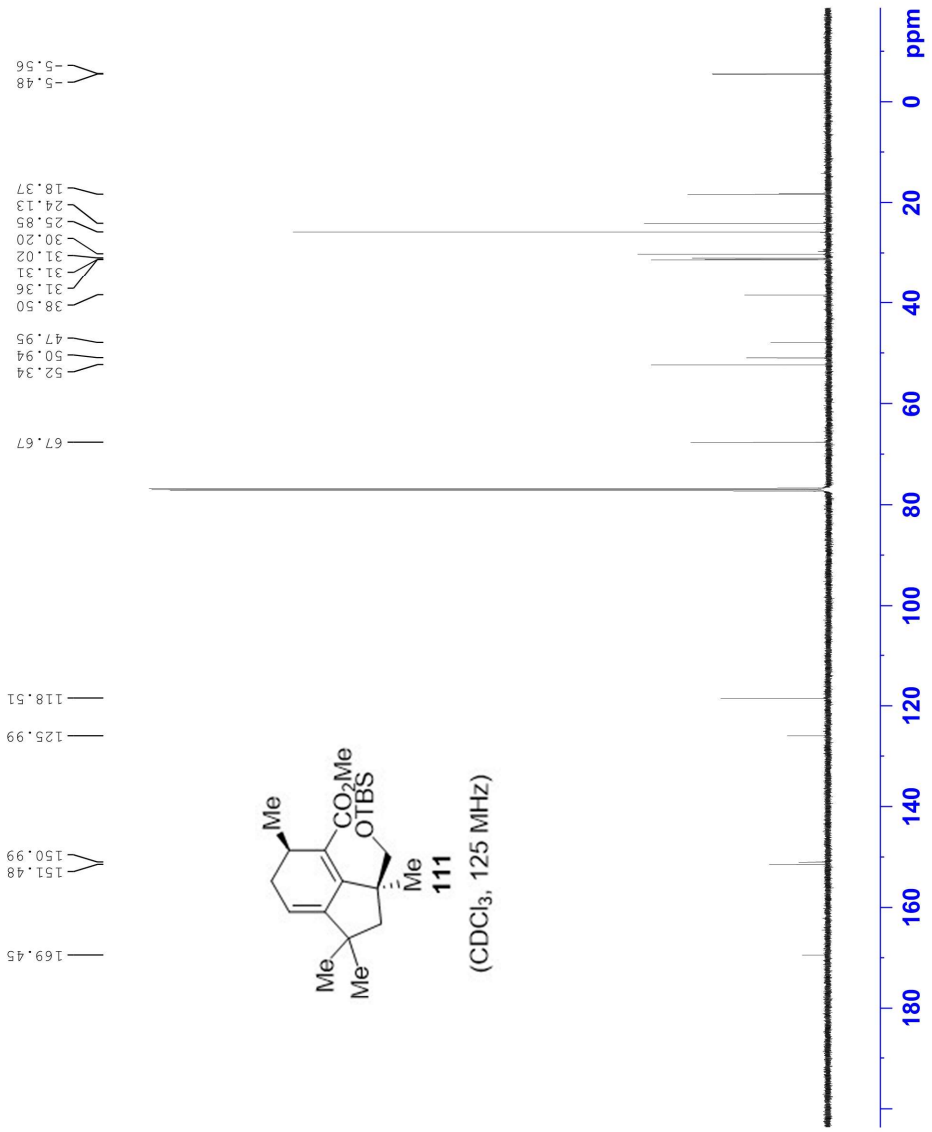
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 SOLVENT CDC13
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 SWH 31250.000 Hz
 FIDRES 0.166670 Hz
 AQ 2.9999361 sec
 RG 2050
 DW 16.000 usec
 DE 6.50 usec
 TE 297.7 K
 D1 3.0000000 sec
 D11 0.0300000 sec
 TDO 1

==== CHANNEL f1 =====
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 NUC1 13C
 P1 10.00 usec
 PLW1 72.83999634 W

==== CHANNEL f2 =====
 SFO2 499.8724993 MHz
 NUC2 1H
 CPDPRG12 waitz16
 PCPD2 80.00 usec
 PLW2 19.0000000 W
 PLW12 0.29688001 W

F2 - Processing parameters
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111
 (CDCl₃, 125 MHz)

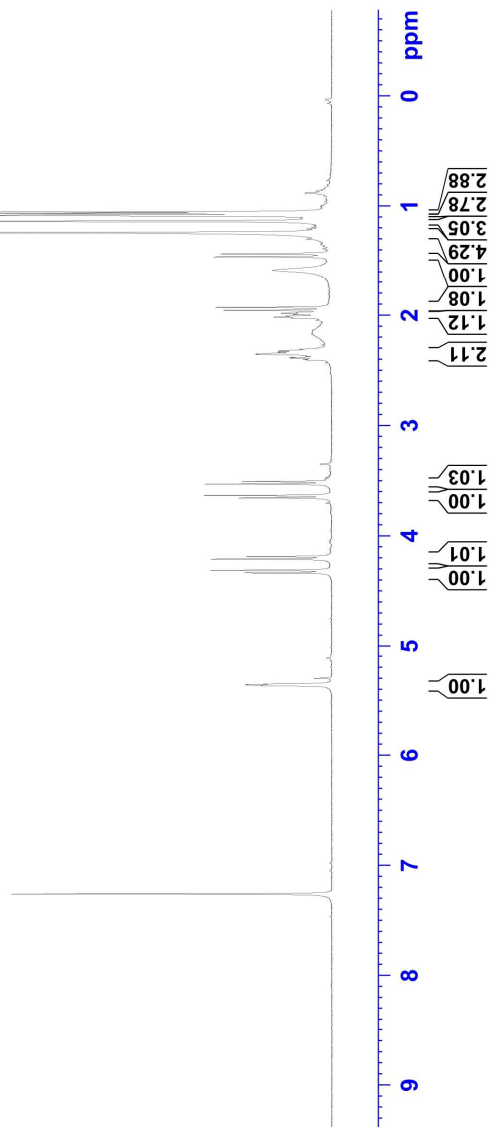
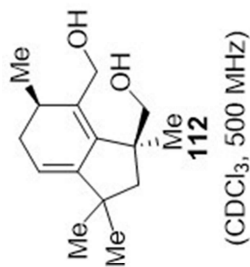
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 PULPROG zg
 TD 5998
 SOLVENT CDCl3
 NS 8
 DS 0
 SWH 10000.000 Hz
 FIDRES 0.166672 Hz
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 RG 196.79
 DW 50.000 usec
 DE 10.00 usec
 TE 293.8 K
 DL 3.0000000 sec
 TDO 1

===== CHANNEL f1 =====
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F2 - Processing parameters
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 WDW EM
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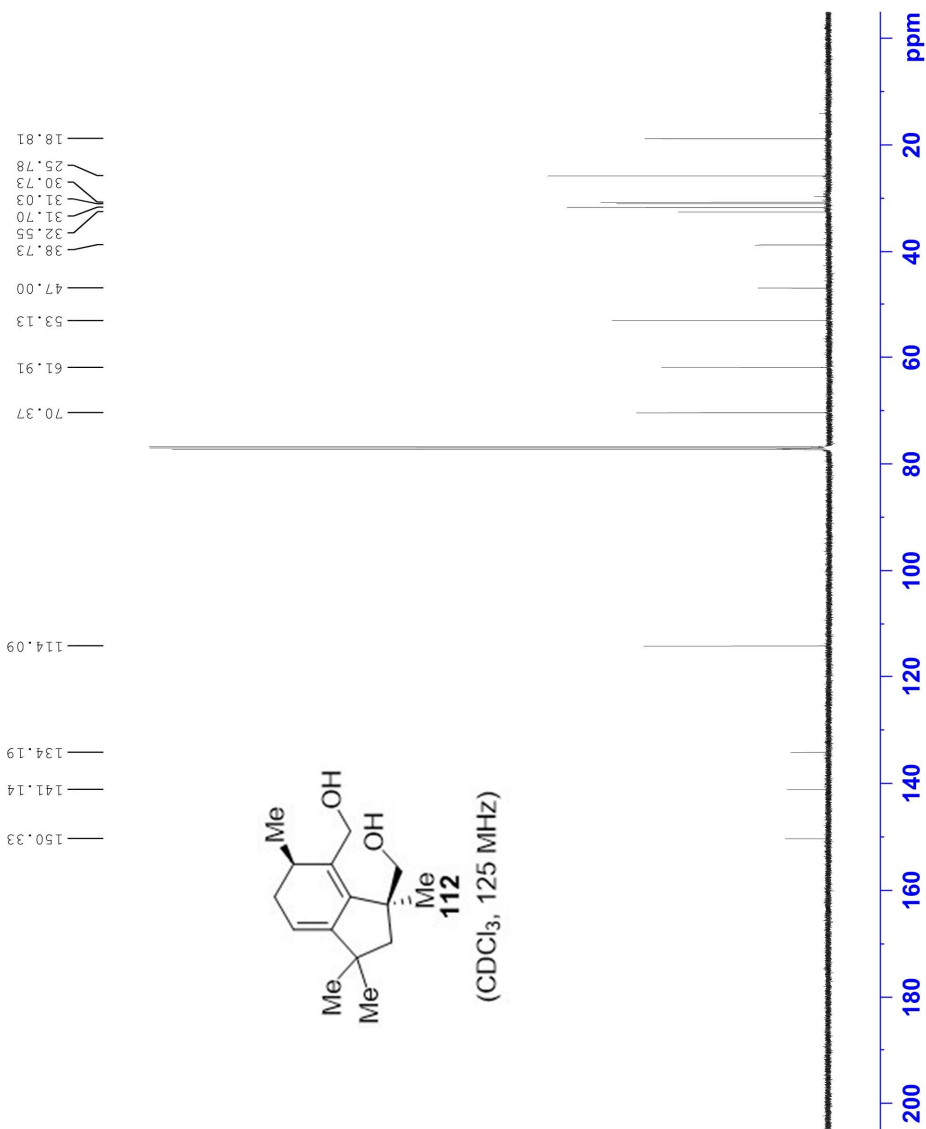
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PULPROG   zgpg
TD         187496
SOLVENT   CDCl3
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FIDRES    0.166670 Hz
AQ         2.9999361 sec
RG         2050
DE         16.000 usec
TE         297.7 K
D1         3.00000000 sec
D11        0.03000000 sec
TD0        1

===== CHANNEL f1 =====
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NUC1       13C
P1         10.00 usec
PLW1      72.83999634 W

===== CHANNEL f2 =====
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NUC2       1H
PCPDPRG[2] waltz16
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PLW2      19.00000000 W
PLW12     0.296888001 W

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PC         1.40

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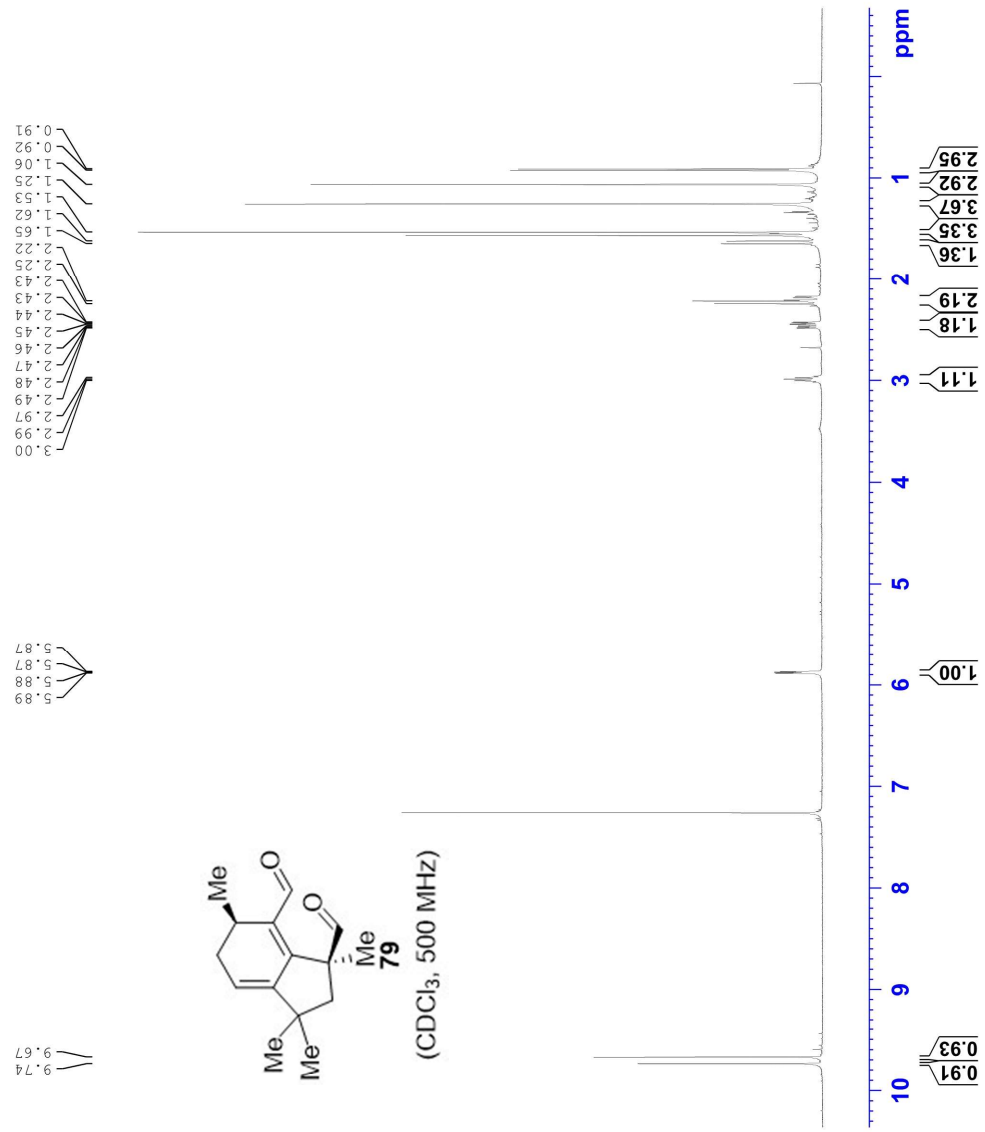


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 SOLVENT CDCl3
 NS 4
 DS 0
 SWH 10000.000 Hz
 FIDRES 0.166672 Hz
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 RG 133.95
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 DE 6.50 usec
 TE 295.5 K
 DL 4.0000000 sec
 TDO 1

===== CHANNEL f1 =====
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 NUC1 1H
 PL 3.58 usec
 PLW1 18.25000000 W

F2 - Processing parameters
 SI 65536
 SF 499.8700123 MHz
 WDW EM
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 LB 0.30 Hz
 GB 0
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PROCNO   2

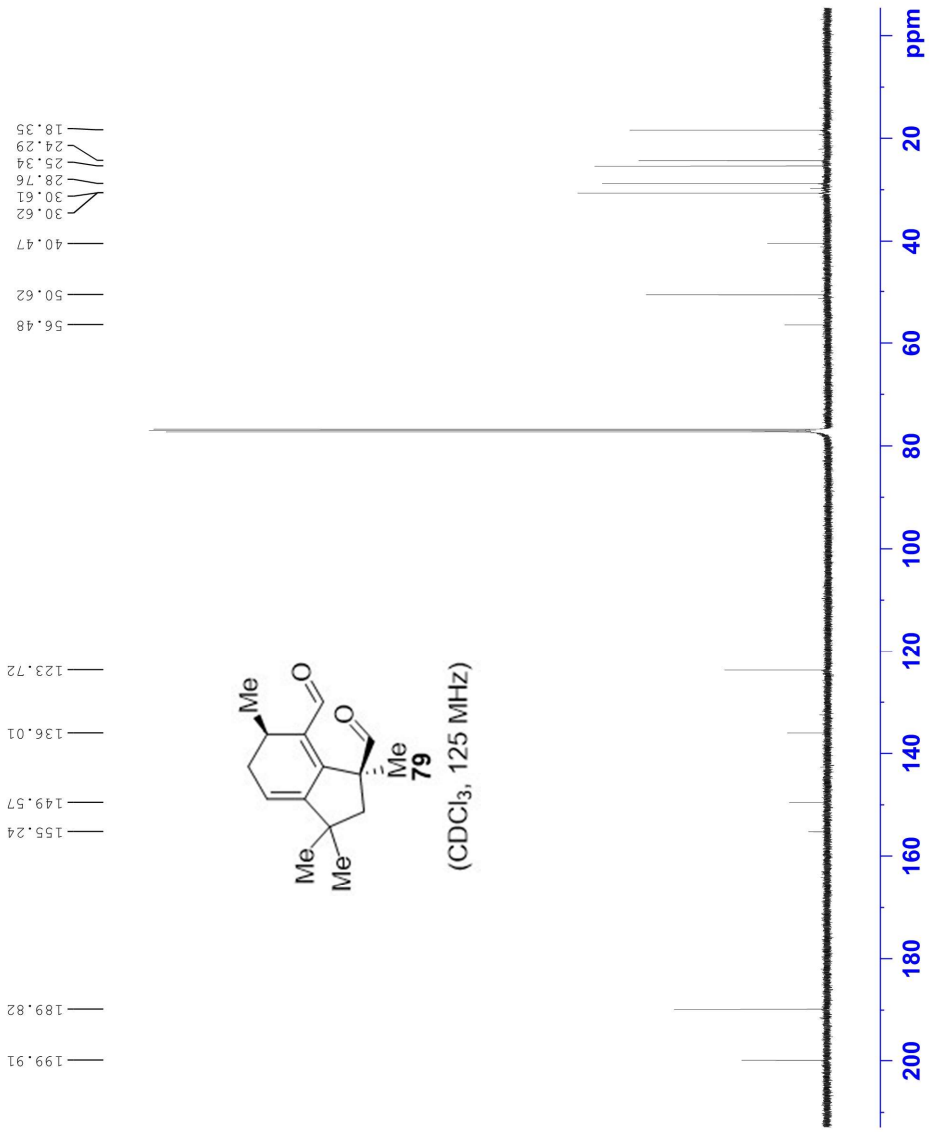
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PULPROG zgpg
TD       187496
SOLVENT  CDC13
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FIDRES   0.166670 Hz
AQ       2.9999361 sec
RG       2050
DM       16.000 usec
DE       6.50 usec
TE       297.5 K
D1       3.0000000 sec
D11      0.03000000 sec
TD0      1

===== CHANNEL f1 =====
SFO1    125.7049802 MHz
NUC1    13C
P1      10.00 usec
PLW1    72.83999634 W

===== CHANNEL f2 =====
SFO2    499.8724993 MHz
NUC2    1H
CPDPRG12 waitz16
PCPD2   80.00 usec
PLW2    19.0000000 W
PLW12   0.29688001 W

F2 - Processing parameters
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PC      1.40

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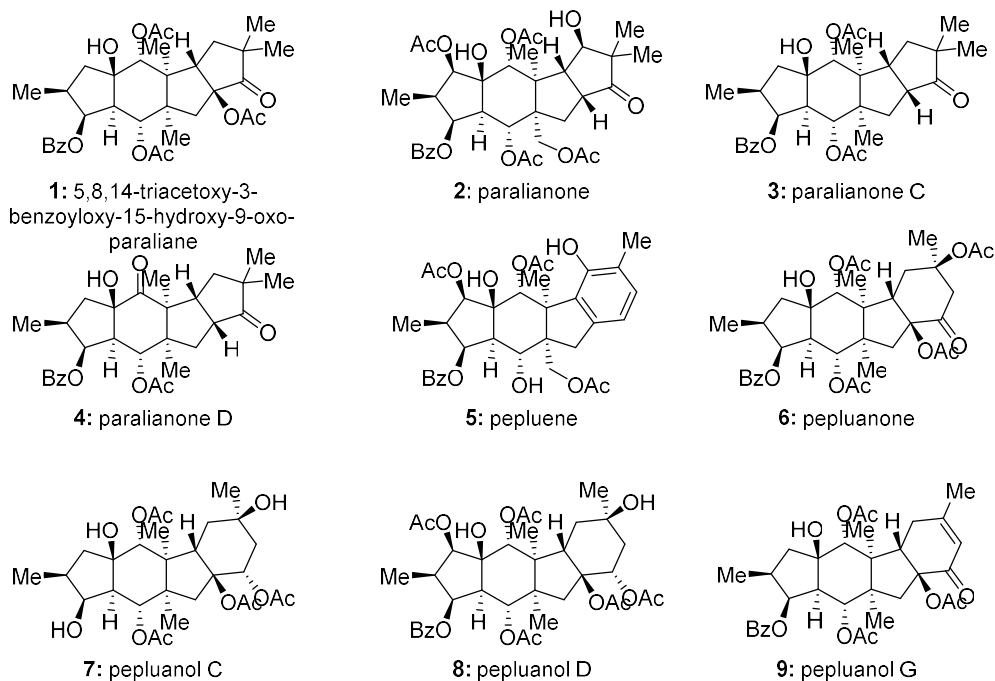
CHAPTER 3

SYNTHETIC STUDY TOWARDS *PARALIANE* FAMILY OF NATURAL PRODUCTS

3.1 Introduction

3.1.1 Structures of selected members of the *paraliane* family of natural products

Figure 3-1: Structure of selected members of *paraliane* and *pepluane* family

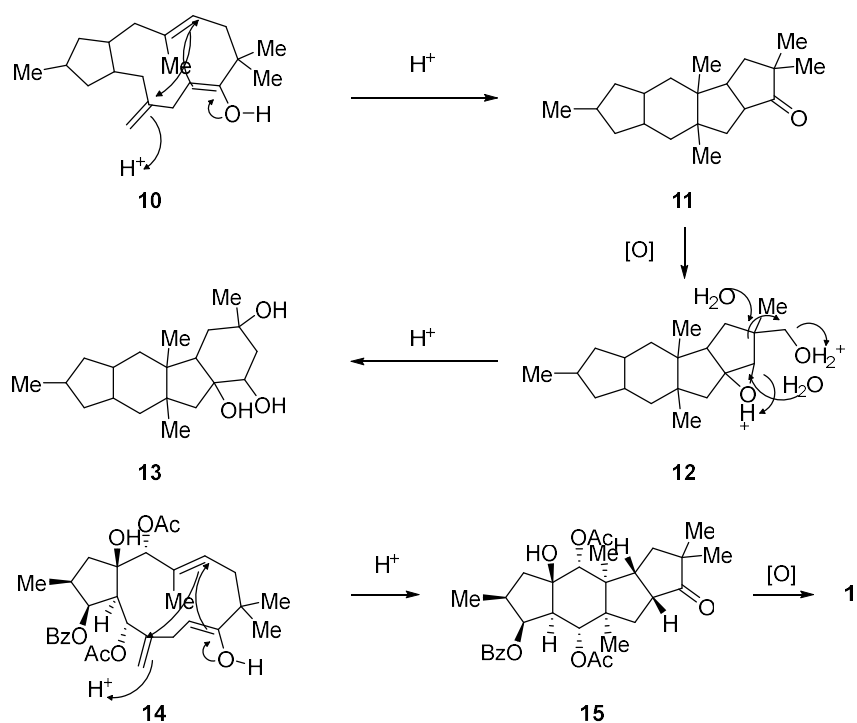


The *paraliane* family of natural products is comprised of complex tetracyclic terpenes. The first member **1** was first isolated in 1998 by the Jakupovic group from the plant *Euphorbia paralias*.^[1] Other members of the family have since been further isolated from *E. paralias* and the related traditional herb *Euphorbia peplus*, together with the similar *pepluane* family.^[2] The *paraliane* family (**1-4**) compounds share a special 5,5,6,5-tetracyclic skeleton while the *pepluane* (**5-9**) skeleton is 6,5,6,5-tetracyclic. All *paraliane* family members (**1-4**) are highly oxidized terpenes, and the density of continuous stereocenters makes them even more challenging targets of organic synthesis. As far as we know, no total synthesis of *paraliane* family members have been reported. (Figure 3-1)

3.1.2 Biochemistry and biosynthesis study on *paraliane* family

E. peplus has been used in the treatment of skin diseases since the ancient times in Europe. The extraction of *E. peplus* was reported to be effective in the treatment against nonmelanoma skin cancer.^[3] Researchers have been working on isolating the active ingredients from those plants.^[4] *Paralianones* were reported to be anti-inflammatorily bioactive, though moderate, with the best IC_{50} equaling $33.7 \mu\text{M}$ for *paralinaone C* (**3**).^[2b,5]

Scheme 3-1: Proposed biosynthesis of *pepluane* and *paraliane* skeleton



As *jatrophanes* were co-isolated with *paralianes* from same plant^[2b], it was proposed that the biosynthesis process of *paralianes* started from relatively simple *jatrophanes*. The acid catalyzed polyene cyclization of *jatrophone* skeleton **10** could directly provide *paraliane* skeleton **11** in one step. Following oxidation and ring expansion would result in *pepluane* skeleton **13**.^[4] Jakupovic et al. proposed that **1** was synthesized by nature from a hypothetical precursor **14**, via

cyclization forming **15** and following α -oxidation of ketone.^[1] However, though **14** is similar to natural product *guyonianin E*,^[2c] it has been neither isolated nor synthesized so far. (Scheme 3-1)

3.2 Synthetic study of *paralianones* via Pd-catalyzed cyclization cascade

3.2.1 Overview of the design

The proposed biosynthesis of the *paraliane* skeleton was based on the acid-catalyzed polyene cyclization. And herein we proposed that our palladium chemistry could be applied to construct the complex ring structures.

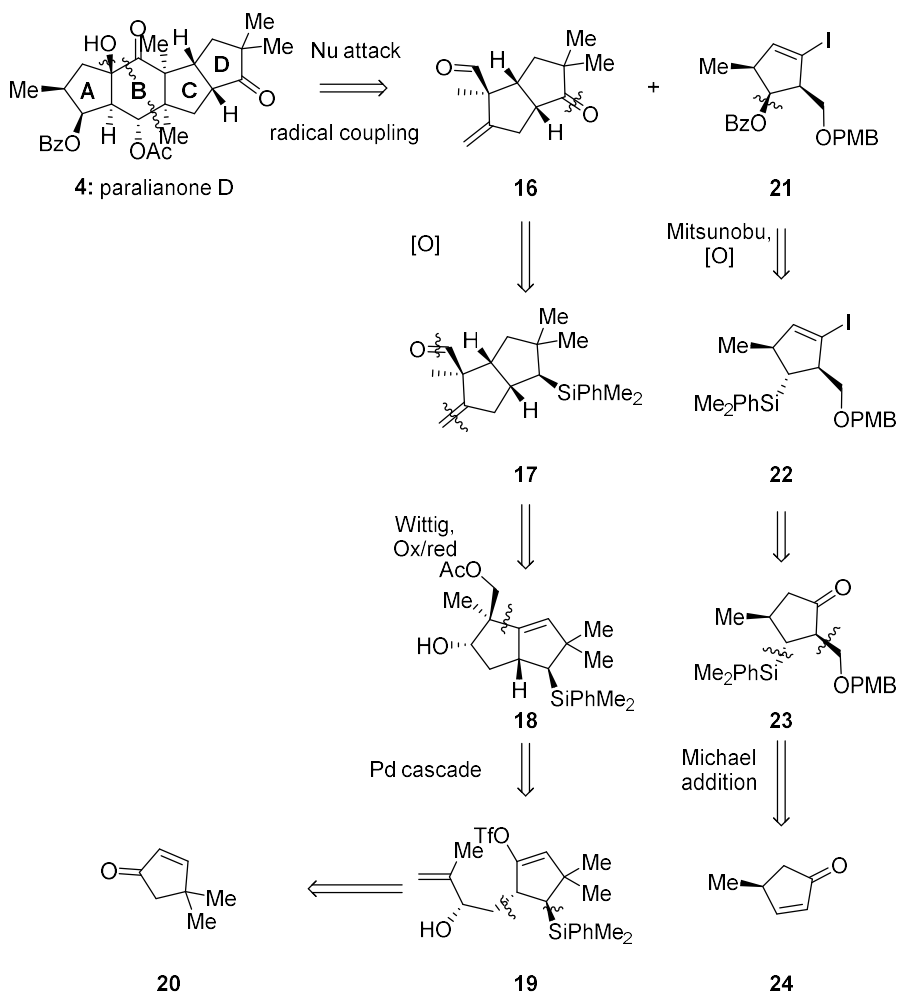
All members of the *paraliane* family have the same stereochemistry in all four rings. We noticed that the 1,3-*trans* stereochemistry and gem-dimethyl quaternary center of the C and D rings, as well as the C–O bond (acetate) on the B ring, could be the target of our cascade chemistry. So, we proposed this stereospecific cascade would be a potential method for the convergent synthesis of the *paraliane* family. (Scheme 3-2)

The retrosynthetic analysis of *paralianone D*, as an example, was shown in Scheme 3-2. The first disconnection focused on the B ring of natural product **4**. This 6-membered ring could be synthesized by a nucleophilic attack to the carbonyl and reductive radical coupling of the alkene and aldehyde, to form the upper and lower bonds, respectively. Then, **4** could be traced back to 2 fragments: east fragment **16** and west fragment **21**.

The ketone of **16** could be introduced by oxidation of a silyl group.^[6] Intermediate **17** could be the product of a Wittig reaction and oxidation/hydrogenation from acetate **18**. The stereochemistry and C–O bond next to quaternary center in **18** indicated that we could synthesize it via our Pd-catalyzed cascade chemistry from triflate **19**. Finally, **19** could be traced back to commercial enone **20** by an asymmetrical Michael addition.

For the west fragment **21**, the benzoate group could be introduced by the Mitsunobu reaction^[7] and oxidation, so it could also be traced back to a silyl intermediate **22**. The vinyl iodide could be readily synthesized from ketone, and ketone **23** would be easily produced by a silyl Michael addition from chiral enone **24**.

Scheme 3-2: Retrosynthetic analysis of *paralianone D*

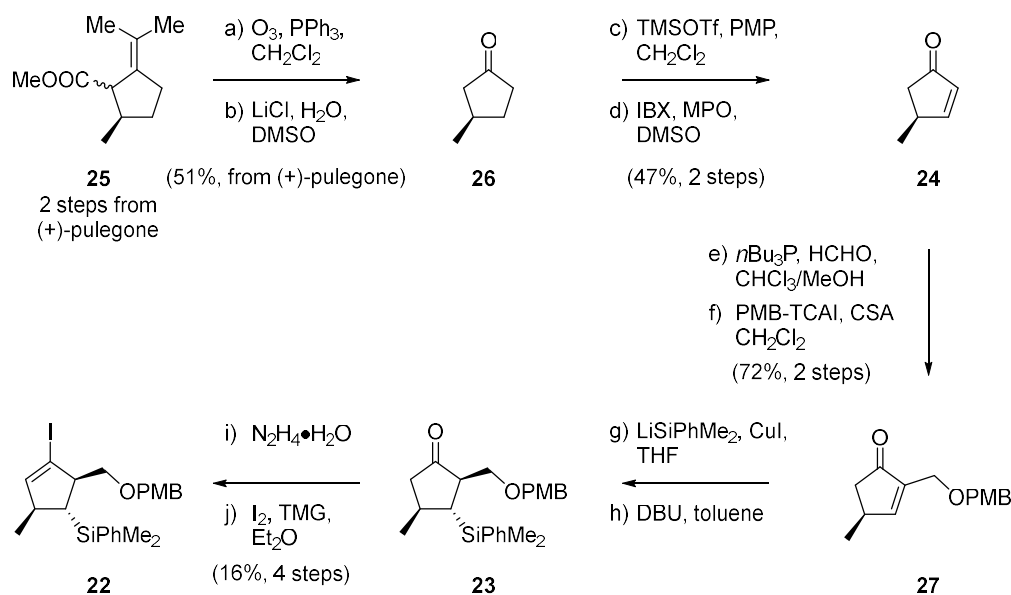


3.2.2 Synthesis of west fragment

We started the synthesis from chiral ketone **26**. Though commercially available, we also developed a synthesis of **26** due to its cost (\$ 96/g from Sigma-Aldrich). The chiral compound **25** was readily prepared on a decagram scale from cheap commercially available natural product, (+)-

pulegone (\$ 1.24/g from Sigma-Aldrich), by bromination then one-pot Favorskii rearrangement and elimination (see experimental section for detailed procedures).^[8] Ozonolysis cleavage of the alkene^[9] followed by decarboxylation of the resulting keto-ester provided chiral ketone **26**. Selective formation of silyl enol ether with hindered base 1,2,2,6,6-pentamethylpiperidine (PMP)^[10] followed by IBX oxidation^[11] would provide enone **24** in moderate yield. (Scheme 3-3)

Scheme 3-3: Synthesis of west fragment^a



a. reaction conditions: (a) O₃, CH₂Cl₂, -78 °C, then PPh₃ (1.1 equiv), -78 °C - 23 °C, 12 h; (b) LiCl (2.0 equiv), DMSO/H₂O (200:1), 120 °C, 7 h; (c) PMP (1,2,2,6,6-pentamethylpiperidine, 1.2 equiv), TMSOTf (1.1 equiv), CH₂Cl₂, -78 °C, 8 h; (d) IBX (2.0 equiv), MPO (1.3 equiv), DMSO, 23 °C, 3 h; (e) *n*Bu₃P (10 mol%), HCHO (37% in H₂O, 2.0 equiv), CHCl₃/MeOH (3:2), 0 °C, 1 h; (f) PMB-TCAI (O-(4-methoxybenzyl)-trichloroacetimidate, 3.0 equiv), CSA (10 mol%), CH₂Cl₂, 0 °C - 23 °C, 24 h; (g) LiSiPhMe₂ (2.6 equiv), Cul (1.3 equiv), THF, -78 °C, 15 min; (h) DBU (2.0 equiv), toluene, 23 °C, 12 h; (i) N₂H₄•H₂O, neat, 120 °C, 8 h; (j) I₂ (1.1 equiv), TMG (1,1,3,3-tetramethylguanidine, 6.0 equiv), Et₂O, 23 °C, 10 h, then 45 °C, 2 h, then neat, 3 h.

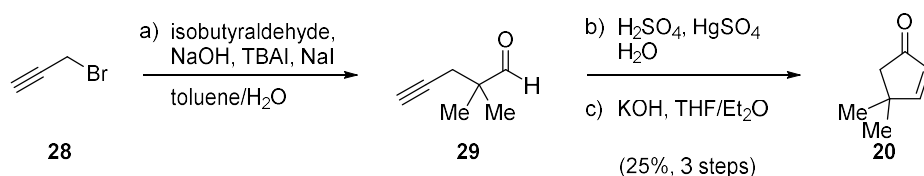
Starting from **24**, we introduced the hydroxyl group by the Baylis–Hillman reaction with formalin.^[12] The free hydroxyl group should be protected before following silyl Michael addition, but all silyl ether protection groups were proved to be incompatible with the strongly basic lithium reagent, leading to decomposition in the next step. After screening we found that PMB was a good protection group in this stage, which could be installed easily under acidic conditions with PMB-

TCAI^[13]. The silyl Michael addition with CuI^[14] worked well on the PMB protected intermediate **27** to form a pair of inseparable diastereomers, which could be epimerized to the desired diastereomer **23** by DBU treatment. Ketone **23** could be transferred to iodide **24** in 2 steps by Prof. Barton's methodology in moderate yield.^[15] (Scheme 3-3)

3.2.3 To Pd-catalyzed cascade

To synthesize the precursor of the key step in the Pd-catalyzed cyclization cascade, we started from commercially available enone **20**. However as **20** is expensive (\$ 540/g from Sigma-Aldrich), we synthesized **20** on a decagram scale in our lab. Many known procedures^[16] turned out to be difficult to handle or have low yields in large scale, so we developed a sequence that required only one purification and no column. Propargyl aldehyde **29** could be synthesized via phase transfer catalysis.^[17] The neutralized crude product of **29**, without further purification, could be hydrated to form a keto-aldehyde,^[18] followed by intramolecular aldol condensation and distillation to provide desired enone **20**.^[16a] (Scheme 3-4)

Scheme 3-4: Synthesis of enone **20**^a

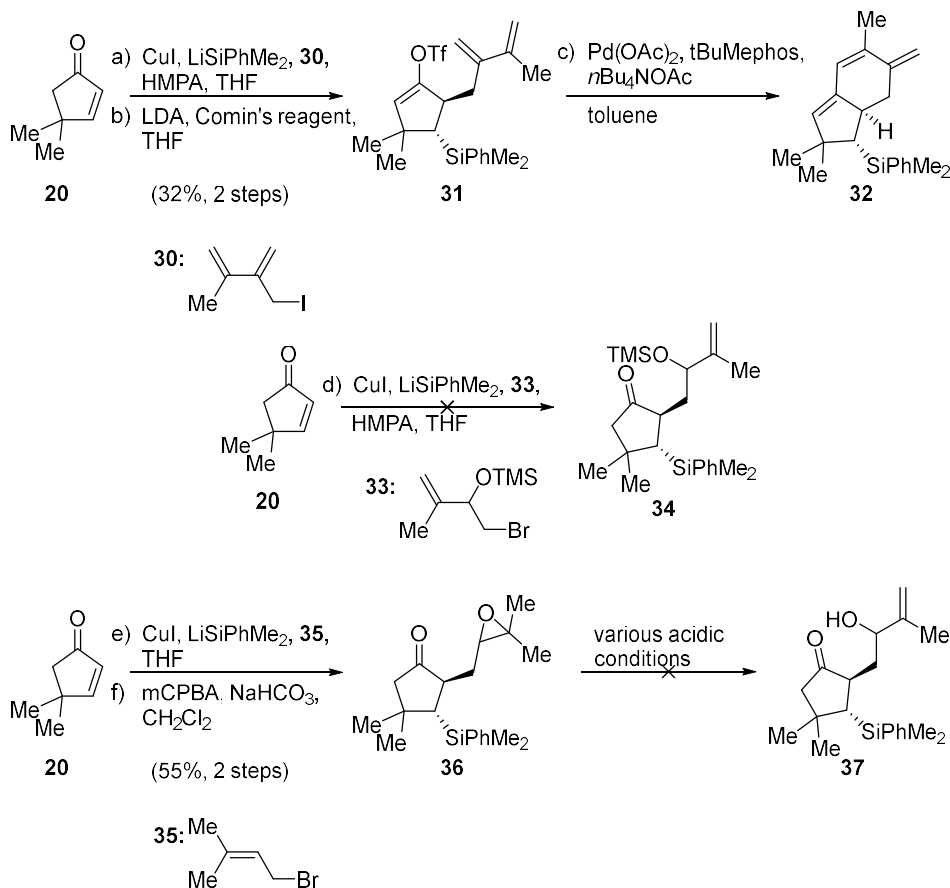


a. reaction conditions: (a) isobutyraldehyde (2.0 equiv), NaOH (2.0 equiv), NaI (10 mol%), TBAI (2 mol%), toluene/H₂O (2:1), 50 °C, 12 h; (b) H₂SO₄ (ex.), HgSO₄ (2 mol%), H₂O, 23 °C, 12 h; (c) KOH (5% in H₂O), THF/Et₂O (1:4), 45 °C, 12 h.

With abundant supply of **20**, we went on to explore an optimized route for the Pd-catalyzed cascade. The first trial started with conjugated diene side chain. After a one-pot Michael addition and allylation of **20**, using LDA as a base, triflate **31** could be formed with desired selectivity (KHMDS provided the undesired regioisomer). When **31** was treated with Pd-catalyzed cascade

condition, only a triene, tentatively assigned as **32**, was observed. We proposed that **32** was the product of β -H elimination of the allyl Pd(II) intermediate. (Scheme 3-5)

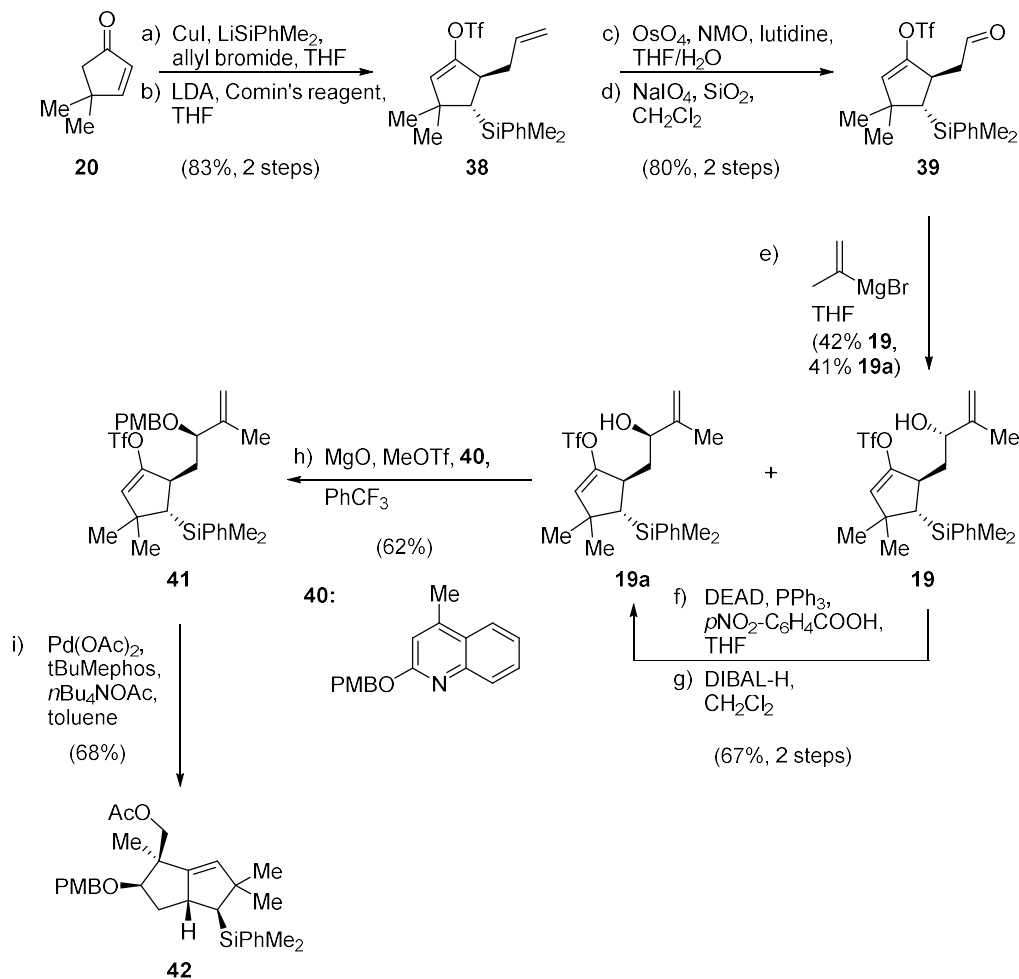
Scheme 3-5: Failed attempts on Pd-catalyzed cascade^a



a. reaction conditions: (a) CuI (1.0 equiv), LiSiPhMe₂ (2.0 equiv), THF, -78 °C, 20 min, then **30** (1.5 equiv), HMPA (2.5 equiv), -78 °C - 23 °C, 12 h; (b) LDA (4.0 equiv), Comin's reagent (3.0 equiv), THF, -78 °C, 3 h; (c) Pd(OAc)₂ (10 mol%), tBuMephos (15 mol%), *n*Bu₄NOAc (3.0 equiv), toluene, 90 °C, 12 h; (d) CuI (1.0 equiv), LiSiPhMe₂ (2.0 equiv), THF, -78 °C, 20 min, then **33** (1.5 equiv), HMPA (2.5 equiv), -78 °C - 23 °C, 12 h; (e) CuI (1.0 equiv), LiSiPhMe₂ (2.0 equiv), THF, -78 °C, 20 min, then **35** (1.5 equiv), -78 °C - 23 °C, 12 h; (f) mCPBA (1.2 equiv), NaHCO₃ (5.0 equiv), CH₂Cl₂, 0 °C.

As the conjugated diene side chain was proved to be incompatible, we proposed the allyl alcohol side chain instead. However, alkylation with bromide **33** failed to yield any desired product. We returned to allylation. Michael addition and allylation with bromide **35** worked smoothly. Epoxidation with *m*CPBA provided **36**, on which we tried many acidic conditions for rearrangement to the desired compound **37**, with all proposed conditions failing. (Scheme 3-5)

Scheme 3-6: Roue to Pd-catalyzed cascade precursor^a



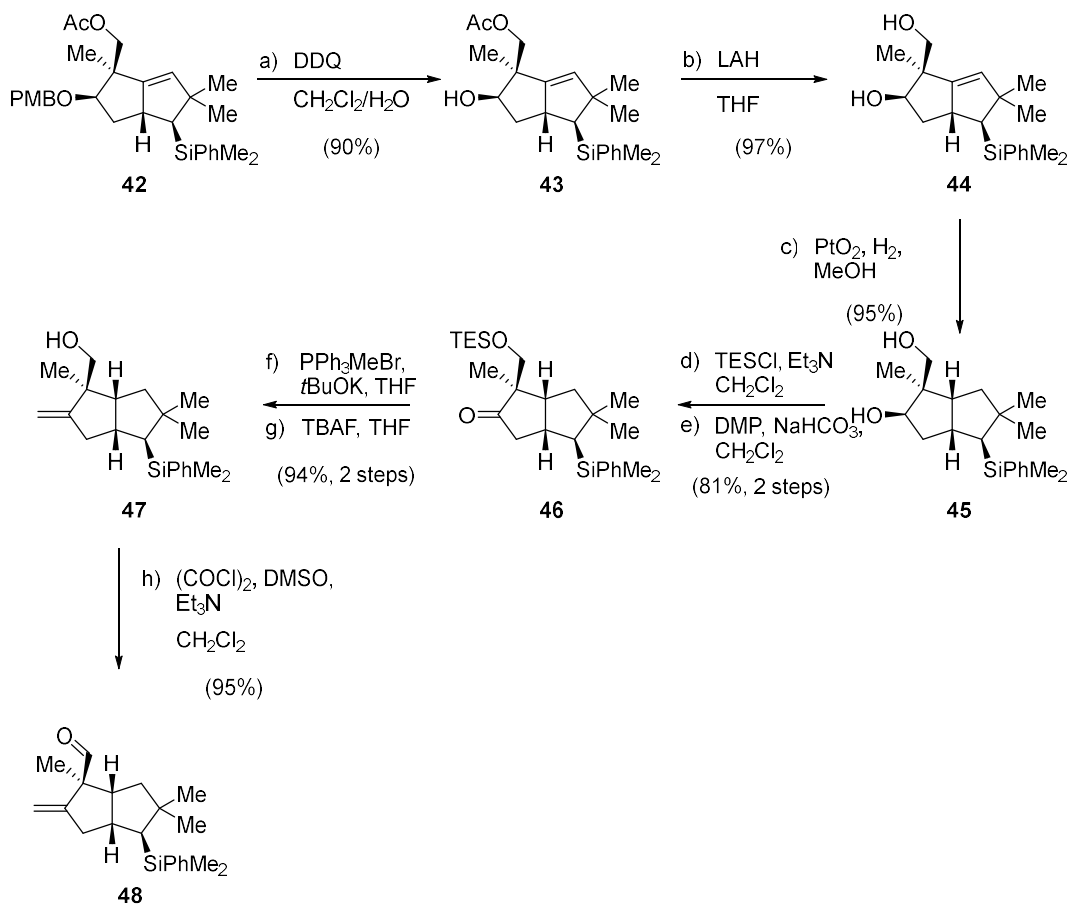
a. reaction conditions: (a) CuI (1.0 equiv), LiSiPhMe₂ (2.0 equiv), THF, -78 °C, 20 min, then allyl bromide (1.5 equiv), -78 °C - 23 °C, 12 h; (b) LDA (4.0 equiv), Comin's reagent (3.0 equiv), THF, -78 °C - 23 °C, 12 h; (c) OsO₄ (2 mol%), NMO (1.5 equiv), 2,6-lutidine (2.0 equiv), THF/H₂O (4:1), 0 °C - 23 °C, 12 h; (d) NaIO₄ (on SiO₂), CH₂Cl₂, 23 °C, 2 h; (e) isopropenylmagnesium bromide (1.5 equiv), THF, -78 °C, 2 h; (f) DEAD (2.0 equiv), PPh₃ (2.0 equiv), 4-nitrobenzoic acid (2.0 equiv), THF, 23 °C, 12 h; (g) DIBAL-H (3.5 equiv), CH₂Cl₂, 0 °C, 3 h; (h) MgO (3.0 equiv), MeOTf (3.0 equiv), **40** (3.0 equiv), PhCF₃, 0 °C - 23 °C, 12 h; (i) Pd(OAc)₂ (10 mol%), tBuMephos (15 mol%), *n*Bu₄NOAc (3.0 equiv), toluene, 90 °C, 12 h.

We then turned to build the allyl alcohol side chain with a nucleophilic attack. Triflate **38** could be synthesized via a similar route as **31** with allyl bromide. As the vinyl triflate was electron deficient, the double bond on side chain was selectively cleaved to form aldehyde **39**. The two-step sequence with NaIO₄ supported on silica gel^[19] provided better yield than the traditional one-pot procedure which provided a 60% yield. A Grignard attack to **39** resulted in a pair of separable

diastereomers, **19** and **19a**, with 1:1 ratio (the stereochemistry was tentatively assigned). We tested the cascade reaction with **19** and **19a**. Trace amounts of the desired cyclized product was observed only with **19**, and **19a** led to total decomposition. We proposed that the free alcohol was not good in palladium chemistry possibly due to protonation of organometallic intermediates. Silyl protection groups were put on first, but no improvement was observed. PMB protection was tested next. As the triflate **19a** was sensitive to both acidic and basic conditions, we found that PMB groups could be installed via neutral conditions^[20] and protected **41** worked well with Pd catalysis to form our desired bicycle **42**. However, the corresponding diastereomer of **41** from **19** resulted in low yields (<30%) in the cascade reaction. Thus, we inverted stereochemistry of **19** to **19a** via Mitsunobu reaction to push to **41**. (Scheme 3-6)

3.2.4 Synthesis of east fragment

Scheme 3-7: Synthesis of east fragment^a



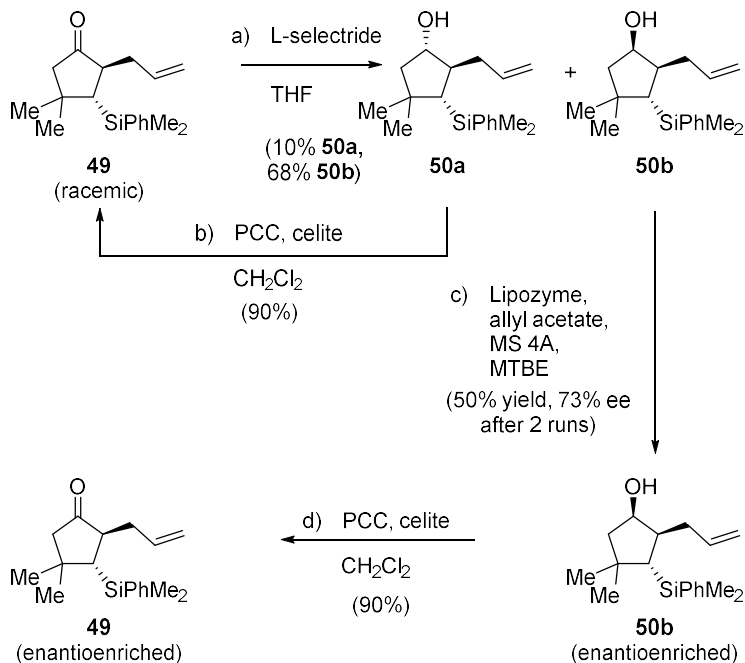
a. reaction conditions: (a) DDQ (1.5 equiv), $\text{CH}_2\text{Cl}_2/\text{H}_2\text{O}$ (10:1), 23 °C, 1 h; (b) LAH (1.2 equiv), THF, 0 °C, 1 h; (c) PtO_2 (10 mol%), H_2 (5 atm), MeOH, 23 °C, 12 h; (d) TESCl (1.05 equiv), Et_3N (3.0 equiv), CH_2Cl_2 , 0 °C - 23 °C, 12 h; (e) DMP (1.5 equiv), NaHCO_3 , CH_2Cl_2 , 0 °C - 23 °C, 3 h; (f) PPh_3MeBr (5.0 equiv), $t\text{BuOK}$ (4.0 equiv), THF, 23 °C, 6 h; (g) TBAF (1.1 equiv), THF, 0 °C, 2 h; (h) $(\text{COCl})_2$ (10 equiv), DMSO (20 equiv), CH_2Cl_2 , -78 °C, 3 h, then Et_3N (30 equiv), 2 h.

With the success on the Pd-catalyzed cascade key step, we continued to the desired east fragment. The alkene in **42** was extremely hindered such that both PMB and acetate groups had to be removed to get diol **44** before hydrogenation of the double bond to provide reasonable conversion. The hydrogenation of **44** only worked under elevated pressure, 5 atm, and with PtO_2 as catalyst, with 95% yield. To selectively protect the primary alcohol, we treated **45** with small excess amount of TESCl, and followed with a DMP oxidation to provide **46**. Then via a Wittig

reaction, deprotection followed by Swern oxidation with large excess amount of reagents, we could synthesize the east fragment **48**.

3.2.5 Route to enantioenriched intermediates

Scheme 3-8: Resolution of ketone **49**^a



a. reaction conditions: (a) L-selectride (1.5 equiv), THF, -78 °C - 0 °C, 2 h; (b) PCC (1.5 equiv), celite, CH₂Cl₂, 23 °C, 1.5 h; (c) Lipozyme, MS 4A, allyl acetate (5.0 equiv), MTBE, 50 °C, 18 h. (d) PCC (1.5 equiv), celite, CH₂Cl₂, 23 °C, 1.5 h.

To achieve asymmetric synthesis, with the enantioenriched west fragment, we needed to synthesize the east fragment asymmetrically. Prof. Hoveyda reported a methodology of asymmetrical silyl addition on **20** to provide **49**,^[21] which we succeeded on a 0.1 mmol scale (60% yield, 92% e.e.), but failed to reproduce on larger scales. For a better supply of material, we chose to do resolution on the racemic **49**. After extensive screening including CBS kinetic resolution,^[22] chiral sulfonamide resolution,^[23] we finally found that **49** could be selectively reduced by L-selectride to corresponding alcohol **50b**, while **50a** could be oxidized to recover ketone **49**. For

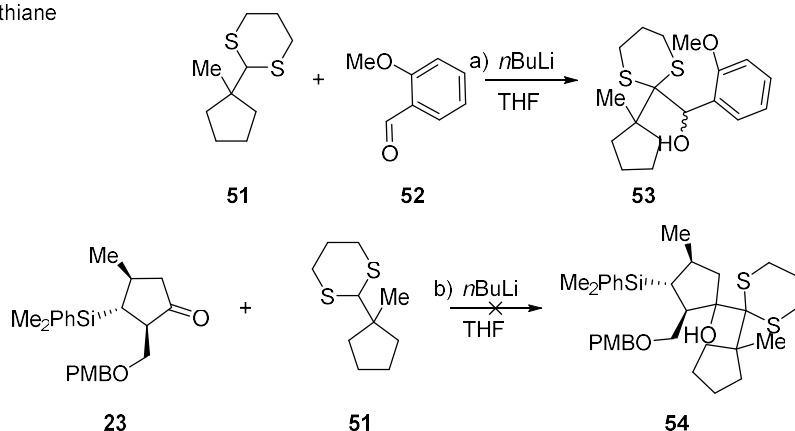
resolution of **50b**, many enzymes including Lipase and Novozyme[®] 435 were tested. Finally, Lipozyme[®] was found to be the good enzyme to resolve **50b**^[24] to provide desired enantiomer with 73% e.e. after 2 runs, and **49** by following PCC oxidation. By this process we could go on to synthesize enantioenriched **48** for further study on final B ring closure. (Scheme 3-8)

3.2.6 Model study on B ring formation

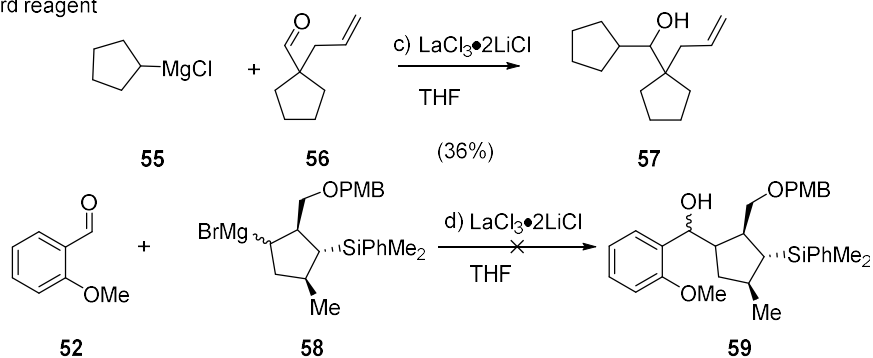
To construct the B ring, the first step would be a nucleophilic attack to couple the east and west fragments. As the reaction would be challenging due to hindrance, we tested a series of conditions with model compounds to explore possible solutions. The first attempt was the umpolung Corey-Seebach reaction.^[25] Model dithiane **51** could attack aldehyde **52** with *n*BuLi, but on the real hindered west fragment **23**, the dithiane attack did not result in any of the desired adduct. As the formation of continuous quaternary centers in one step was difficult, we turned to use the west fragment in the nucleophilic to attack on the aldehyde of east fragment. For model aldehyde **56**, simple Grignard reaction with **55** resulted in only reduction. The Grignard reaction catalyzed by La(III) salt, however, provided adduct **57** in moderate yield.^[26] But with real west fragment **58**, again no desired product was observed, only decomposition. Vinyl lithium reagents led to comparable results, working well with simple compound **60** but not real fragment **22**. We separated reduced product **63**, indicating that Li species did form but no nucleophilic attack happened. Finally, we tried the Nozaki–Hiyama–Kishi reaction.^[27] To our delight, we separated the desired adduct **64** with real fragment **22** and benzaldehyde. (Scheme 3-9)

Scheme 3-9: Model study on nucleophilic attack^a

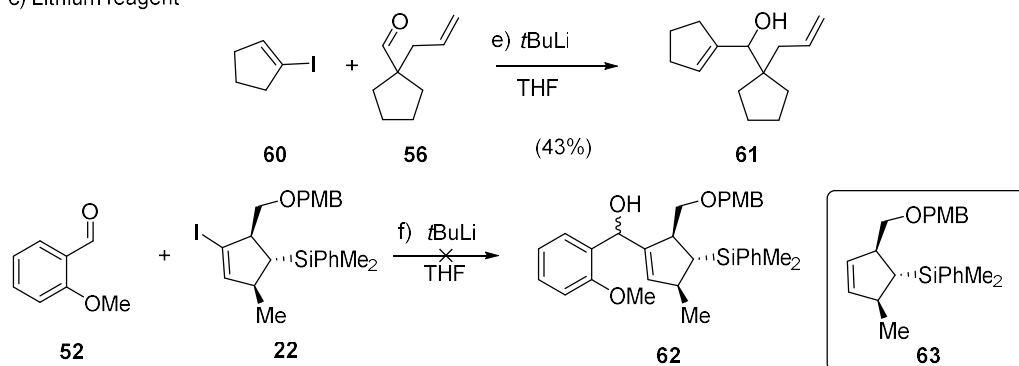
a) Dithiane



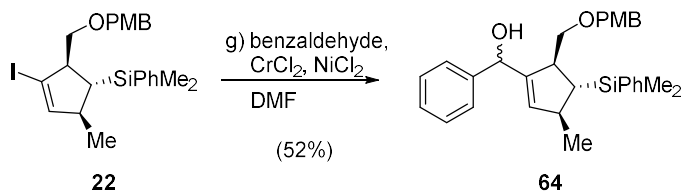
b) Grignard reagent



c) Lithium reagent



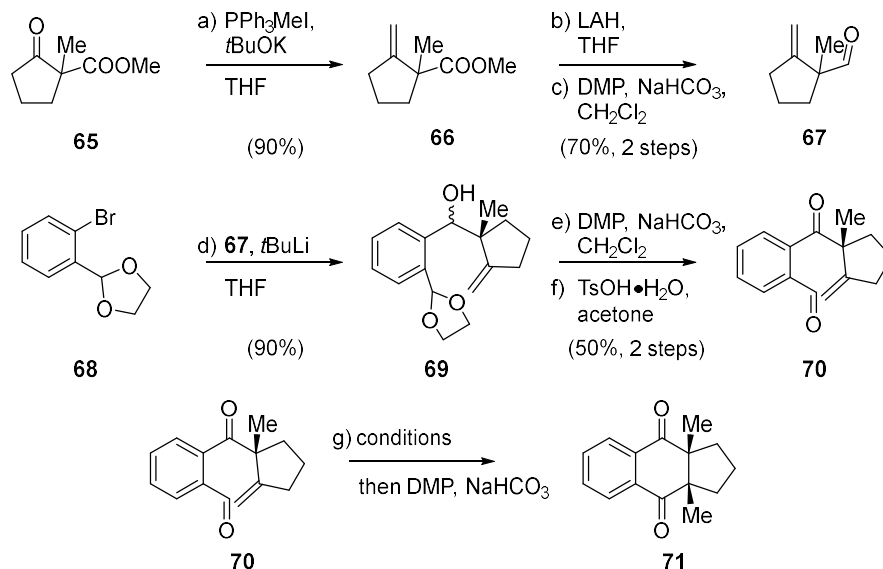
d) NHK reaction



a. reaction conditions: (a) **51** (1.0 equiv), **52** (1.2 equiv), *n*BuLi (2.0 equiv), THF, 0 °C - 23 °C, 12 h; (b) **51** (1.0 equiv), **23** (1.2 equiv), *n*BuLi (2.0 equiv), THF, 0 °C - 23 °C, 12 h; (c) **56** (1.0 equiv), **55** (1.2 equiv), LaCl₃·2LiCl (1.0 equiv), THF, 0 °C - 23 °C, 12 h; (d) **52** (1.0 equiv), **58** (2.0 equiv), LaCl₃·2LiCl (1.0 equiv), THF, 0 °C - 23 °C, 12 h; (e) **56** (1.0 equiv), **60** (1.2 equiv), *t*BuLi (2.5 equiv), THF, -78 °C, 1 h; (f) **22** (1.0 equiv), **52** (2.0 equiv), *t*BuLi (2.0 equiv), THF, -78 °C - 0 °C, 1 h; (g) **22** (1.3 equiv), benzaldehyde (1.0 equiv), CrCl₂ (7.0 equiv), NiCl₂ (5 mol%), DMF, 23 °C, 12 h.

We continued to explore on the radical cyclization for B ring closure. For a model compound with hindrance, we synthesized aldehyde **67** with quaternary center in 3 steps from keto-ester **65**. Then **67** was coupled with aryl bromide **68**^[28] to form model compound **70** after oxidation and deprotection. (Scheme 3-10)

Scheme 3-10: Benzaldehyde model study of radical cyclization^a



entry	conditions	yield ^b
1	SmI ₂ (2.5 equiv), <i>t</i> BuOH (2.5 equiv), THF, -78 °C	<10%
2	Fe(acac) ₃ (1.0 equiv), PhSiH ₃ (2.5 equiv), <i>i</i> PrOH (0.25M), 60 °C	32%
3	Fe(acac) ₃ (1.0 equiv), PhSiH ₂ (<i>O</i> / <i>Pr</i>) (2.5 equiv), <i>i</i> PrOH (0.25M), 60 °C	58%
4	Mn(dpm) ₃ (1.0 equiv), PhSiH ₂ (<i>O</i> / <i>Pr</i>) (2.5 equiv), <i>i</i> PrOH (0.25M), 23 °C	62%
5	Mn(dpm)₃ (1.0 equiv), PhSiH₂(<i>O</i>/<i>Pr</i>) (2.5 equiv), <i>i</i>PrOH (0.1M), 23 °C	66%
6	Co(dpm) ₂ (1.0 equiv), PhSiH ₂ (<i>O</i> / <i>Pr</i>) (2.5 equiv), TBHP (1.5 equiv), <i>i</i> PrOH (0.25M), 23 °C	31% ^c

a. reaction conditions: (a) PPh₃MeI (1.3 equiv), *t*BuOK (1.2 equiv), THF, 70 °C, 2 h; (b) LAH (1.0 equiv), THF, 23 °C, 90 min; (c) DMP (1.3 equiv), NaHCO₃, CH₂Cl₂, 0 °C - 23 °C, 1 h; (d) **67** (1.0 equiv), **68** (1.5 equiv), *t*BuLi (3.0 equiv), THF, -78 °C - 23 °C, 12 h; (e) DMP (1.3 equiv), NaHCO₃, CH₂Cl₂, 23 °C, 1 h; (f) TsOH·H₂O (10 mol%), acetone, 23 °C, 18 h.

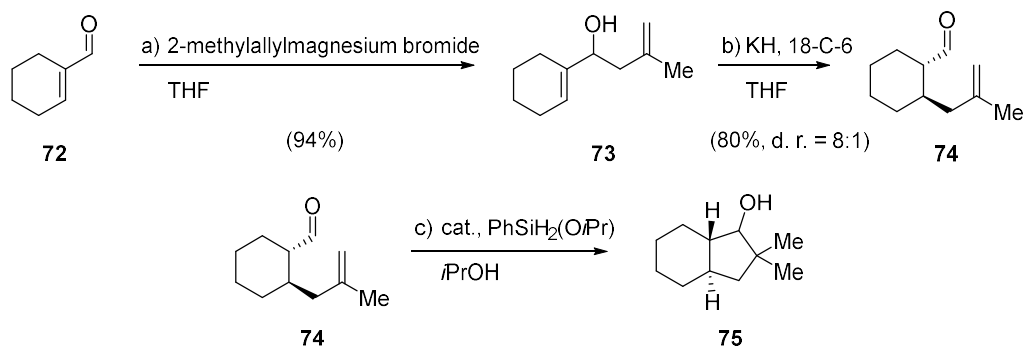
b. isolated yield

c. provide **71** without DMP oxidation, NMR yield

We tried a series of conditions on **70** for cyclization. To eliminate diastereomers for a better NMR to compare yields, the cyclized product was oxidized by DMP. SmI₂ reductive coupling provided only trace amounts of product (entry 1). We then turned to Mukaiyama radical conditions

reported by Prof. Bonjoch and Prof. Bradshaw et al.,^[29] which provided 32% yield (entry 2). Using Shenvi's silane^[30] instead of phenyl silane, a much better yield 58% could be achieved (entry 3). Further screening showed that Mn(dpm)₃ was a better catalyst (entry 4), and the yield was slightly better in lower concentration (entry 5). The cobalt catalyst directly resulted in diketone **71** without further oxidation, possibly because of Co(IV) as an oxidant, but the yield was lower (entry 6). (Scheme 3-10)

Scheme 3-11: Alkyl aldehyde model of radical cyclization^a



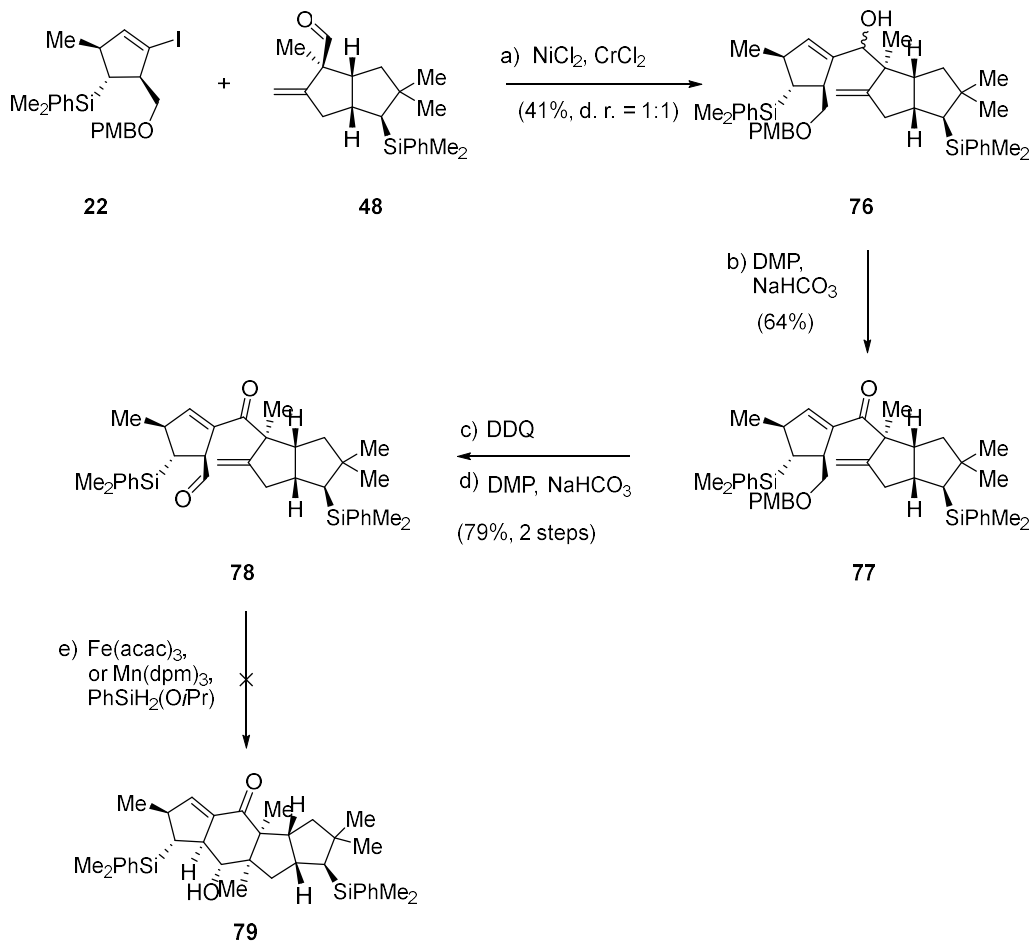
entry	catalyst	yield ^b
1	Fe(acac) ₃ (1.0 equiv)	60%
2	Co(dpm) ₂ (1.0 equiv), TBHP (1.5 equiv)	trace
3	Co(dpm) ₂ (10 mol%), TBHP (1.0 equiv)	18%
4	Mn(dpm) ₃ (1.0 equiv)	45%

a. reaction conditions: (a) 2-methylallylmagnesium bromide (1.5 equiv), THF, 0 °C, 1 h; (b) KH (4.0 equiv), 18-C-6 (4.0 equiv), THF, 23 °C, 8 h; (c) cat., PhSiH₂(O*i*Pr) (2.5 equiv), *i*PrOH, 23 °C, 12 h
b. isolated yield

As the real system would be alkyl aldehyde, we also tested the Mukaiyama reaction on model **74**, which could be synthesized from **72** by Grignard addition and oxy-Cope rearrangement. Different conditions of Mukaiyama radical reaction were tested on aldehyde **74**. A single diastereomer **75** was formed and Fe(acac)₃ was found to be the best catalyst (entry 1). (Scheme 3-11)

3.2.7 Trial of B ring formation

Scheme 3-12: Trial of B ring formation^a



a. reaction conditions: a) **48** (1.0 equiv), **22** (3.0 equiv), NiCl_2 (1 mol%), CrCl_2 (7.0 equiv), DMF, 40 °C, 48 h; (b) DMP (2.5 equiv), NaHCO_3 , CH_2Cl_2 , 23 °C, 3 h; (c) DDQ (1.5 equiv), $\text{CH}_2\text{Cl}_2/\text{H}_2\text{O}$, 23 °C, 1 h; (d) DMP (1.5 equiv), NaHCO_3 , CH_2Cl_2 , 23 °C, 1 h; (e) $\text{Fe}(\text{acac})_3$ (1.0 equiv) or $\text{Mn}(\text{dpm})_3$ (1.0 equiv), $\text{PhSiH}_2(\text{O}i\text{Pr})$ (2.5 equiv), $i\text{PrOH}$, 23 °C, 3 h.

With the success of model study, we turned to our real fragments **22** and **48**. The described Nozaki–Hiyama–Kishi reaction condition for **64** worked on them, however the conversion was very low. After screening, we found the optimized condition. With only 1 mol% NiCl_2 and higher concentration, we could separate 41% yield of the desired coupling product **76** as a pair of diastereomers (d. r. = 1:1). Then the two diastereomers was oxidized to one ketone **77**. Further

deprotection and oxidation provided aldehyde **78**. Unfortunately, trial of Mukaiyama radical cyclization on **78** resulted in only decomposition. It might be because that such HAT conditions could also react with enone^[31] and undesired transformation could happen.

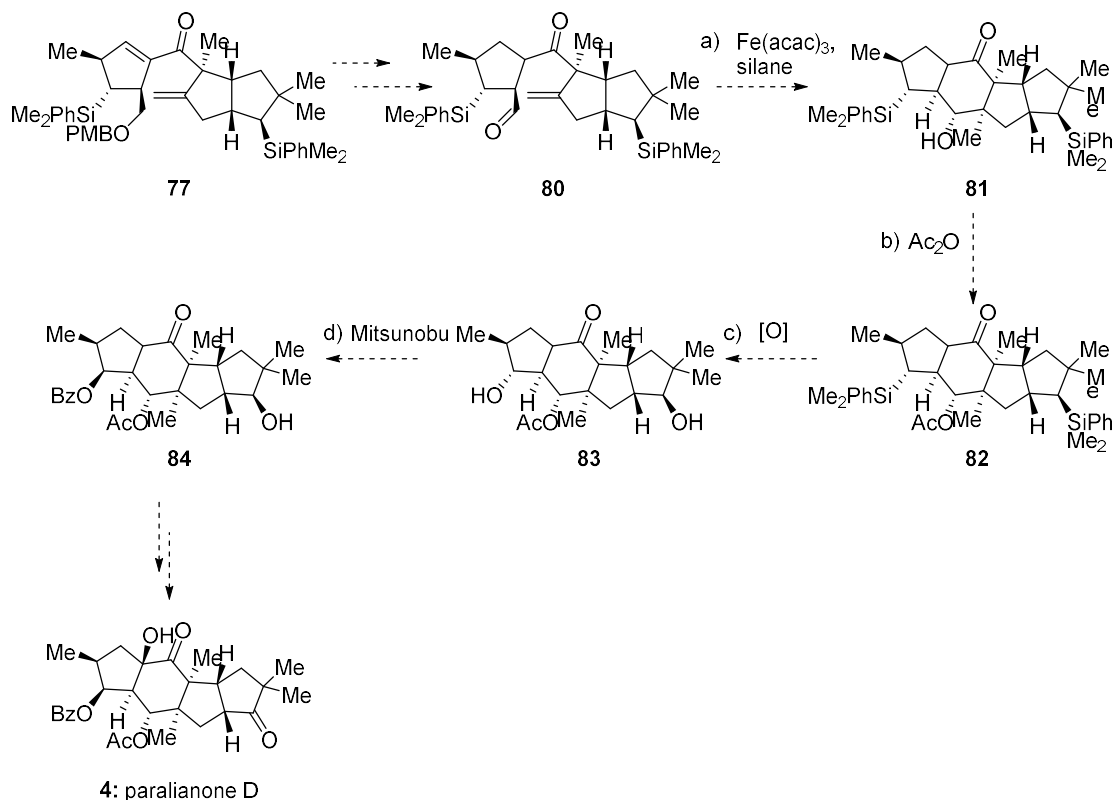
3.3 Summary and outlook

Based on our cascade methodology, we designed a synthetic route towards natural product *paralianone D*. We synthesized the west fragment in 8 steps from commercial chiral compound **26**. The east fragment was synthesized with our methodology in 18 steps from commercial enone **20**. To achieve enantioenriched east fragment, we used enzymatic resolution.

In model study, we succeeded in the Nozaki–Hiyama–Kishi coupling of two fragments and discovered an unprecedented Mukaiyama type radical cyclization of alkene and aldehyde. The Nozaki–Hiyama–Kishi reaction combined two fragments in the real system. However, the radical cyclization was not successful so far. We plan to try on other substrate without enone moiety for the cyclization, such as a 1,4-reduction to synthesis intermediate **80** prior to cyclization. Other reductive coupling like SmI₂ can also be considered. After the cyclization, we will build the whole skeleton of *paralianone D* (**4**), and a series of oxidation reactions and Mitsunobu reaction will lead to the target natural product. (Scheme 3-13)

The final steps of west fragment preparation also need optimization, due to very low yields and low purity. Prof. Reisman's vinyl iodide synthesis^[32] may be a possible choice.

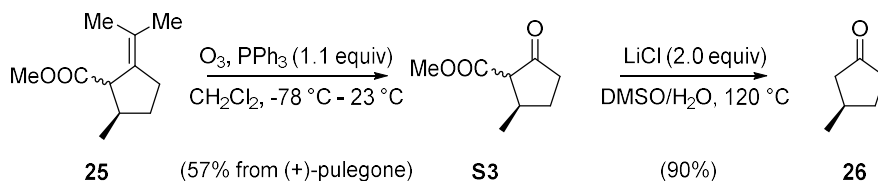
Scheme 3-13 Future plan



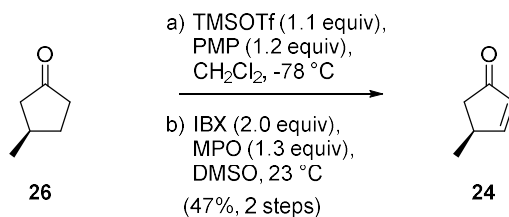
3.4 Experimental section

General Procedures. All reactions were conducted under an argon atmosphere with dry solvents under anhydrous conditions, unless otherwise noted. Dry tetrahydrofuran (THF), toluene, dimethylformamide (DMF), diethyl ether (Et_2O) and dichloromethane (CH_2Cl_2) were obtained by passing commercially available pre-dried, oxygen-free formulations through activated alumina columns. Yields refer to chromatographically and spectroscopically (^1H and ^{13}C NMR) homogeneous materials, unless otherwise stated. Reagents were purchased at the highest commercial quality and used without further purification, unless otherwise stated. Reactions were magnetically stirred and monitored by thin-layer chromatography (TLC) carried out on 0.25 mm E. SiliCycle silica gel plates silica gel plates (60F-254) using UV light as visualizing agent, and an ethanolic solution of phosphomolybdic acid and cerium sulfate, and heat as developing agents.

residue was purified by column chromatography (hexanes: EtOAc = 20:1→10:1) to provide ester **25** (24 g). All data were identical with literature report.^[8]

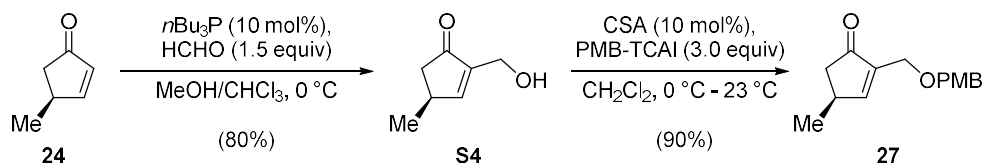


Ketone 26. To a solution of **25** (29.1 g, 0.16 mol, 1.0 equiv) in CH_2Cl_2 (200 mL) at -78°C was bubbled O_3 for 2 h until the solution turned blue, then PPh_3 (47 g, 0.18 mol, 1.1 equiv) was added. The solution was slowly warmed to 23°C and stirred for 12 h. Then it was concentrated and diluted by hexane. The resulted mixture was filtered through a pad of silica gel (hexanes: EtOAc = 3:1). The filtrate was concentrated, and the resultant residue was purified by column chromatography (hexanes: EtOAc = 30:1→20:1→3:1→2:1) to provide keto-ester **S3** (21.95 g, 57% from **S1**). All data were identical with literature report.^[9] To a solution of **S3** (21.8 g, 0.14 mol, 1.0 equiv) in DMSO (200 mL) and H_2O (10 mL) was added LiCl (12 g, 0.28 mol, 2.0 equiv). The mixture was heated to 120°C and stirred at that temperature for 12 h. Upon completion, the reaction contents were cooled to 23°C , diluted with water and extracted with Et_2O ($2 \times 200 \text{ mL}$). The combined organic layers were dried (Na_2SO_4), filtered and concentrated (40°C , 500 mm Hg). The resultant residue was purified by column chromatography (pentanes: Et_2O = 1:1) to provide ketone **26** (12.3 g, 90%). All data were identical with commercial sample from Sigma-Aldrich.



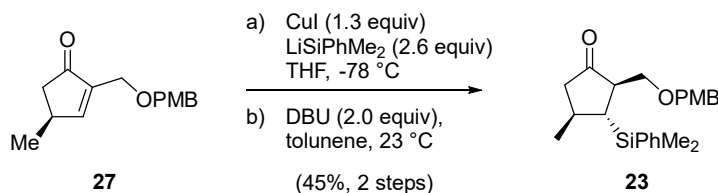
Enone 24. To a solution of **26** (3.92 g, 40 mmol, 1.0 equiv) in CH_2Cl_2 (120 ml) at -78°C was added PMP (7.44g, 48 mmol, 1.2 equiv) then TMSOTf (7.94 mL, 44 mmol, 1.1 equiv). The

resulted mixture was stirred at that temperature for 8 h. Upon completion, the reaction was quenched by addition of NaHCO₃ solution. The mixture was poured into a separatory funnel and the layers were separated. The organic layer was concentrated (40 °C, 500 mmHg). The residue was used in the next step without further purification. The IBX oxidation was conducted via reported procedure.^[11] The resulted mixture was purified by column chromatography (pentanes: Et₂O = 3:1→2:1) to provide enone **24** (1.8 g, 47% over 2 steps) as a pale-yellow oil. **24**: R_f = 0.31 (silica gel, hexanes:Et₂O, 2:1); ¹H NMR (500 MHz, CDCl₃) δ 7.58 (dd, *J* = 5.6 Hz, 2.5 Hz, 1 H), 6.12 (dd, *J* = 5.6 Hz, 2.1 Hz, 1 H), 3.06–2.99 (m, 1H), 2.59 (dd, *J* = 18.8 Hz, 6.4 Hz, 1 H), 1.94 (dd, *J* = 18.8 Hz, 2.2 Hz, 1 H), 1.04 (s, 3 H).



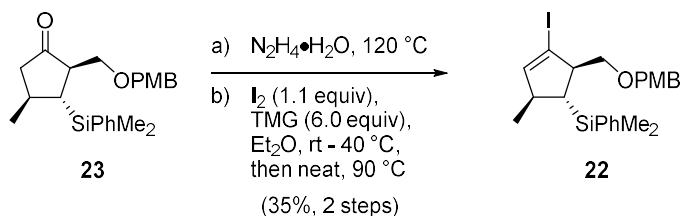
PMB ether 27. To a solution of **24** (2.97 g, 31 mmol, 1.0 equiv) and HCHO (37% in H₂O, 4.6 mL, 62 mmol, 2.0 equiv) in MeOH (60 ml) and CHCl₃ (60 ml) at 0 °C was added *n*Bu₃P (0.77 mL, 3.1 mmol, 10 mol%) in CHCl₃ (30 ml) dropwise over the course of 15 min. The mixture was stirred at that temperature for 1 h. Upon completion, the reaction was quenched by addition of water. The mixture was poured into a separatory funnel and the layers were separated. The aqueous layer was extracted by EtOAc (5× 100 mL). The combined organic layer was dried (Na₂SO₄), filtered and concentrated. The residue was purified by column chromatography (hexanes:EtOAc = 1:1) to provide **S4** (3.21 g, 80%) as a colorless oil. **S4**: R_f = 0.23 (silica gel, hexanes:EtOAc, 1:1); ¹H NMR (500 MHz, CDCl₃) δ 7.38 (dt, *J* = 2.5 Hz, 1.3 Hz, 1 H), 4.37–4.36 (m, 2 H), 2.98–2.95 (m, 1 H), 2.68 (dd, *J* = 19.0 Hz, 6.3 Hz, 1 H), 2.02 (dd, *J* = 19.0 Hz, 2.1 Hz, 1 H), 1.21 (d, *J* = 7.2 Hz, 3 H). To a solution of **S2** (3.15 g, 25 mmol, 1.0 equiv) and PMB-TCAI (21.1 g, 75 mmol, 3.0 equiv) in CH₂Cl₂ (100 mL) at 0 °C was added CSA (0.58 g, 2.5 mmol, 10 mol%). The mixture

was slowly warmed to 23 °C and stirred for 24 h. Upon completion, the reaction was quenched by addition of NaHCO₃ solution. The mixture was poured into a separatory funnel and the layers were separated. The aqueous layer was extracted by CH₂Cl₂ (2× 100 mL). The combined organic layer was dried (Na₂SO₄), filtered and concentrated. The residue was diluted by hexane (100 mL) and filtered to remove CCl₃CONH₂. The filtrate was concentrated and purified by column chromatography (hexanes:EtOAc = 10:1→5:1) to provide **27** (5.53 g, 90%) as a pale-yellow oil. **27**: R_f = 0.35 (silica gel, hexanes:EtOAc, 3:1); ¹H NMR (500 MHz, CDCl₃) δ 7.48–7.47 (m, 1 H), 7.28–7.26 (m, 2 H), 6.9–6.87 (m, 2 H), 4.50 (s, 2H), 4.16–4.15 (m, 2 H), 3.81 (s, 3 H), 2.95–2.93 (m, 1 H), 2.66 (dd, *J* = 18.9 Hz, 6.3 Hz, 1 H), 2.00 (dd, *J* = 18.9 Hz, 2.1 Hz, 1 H), 1.19 (d, *J* = 7.2 Hz, 3 H).



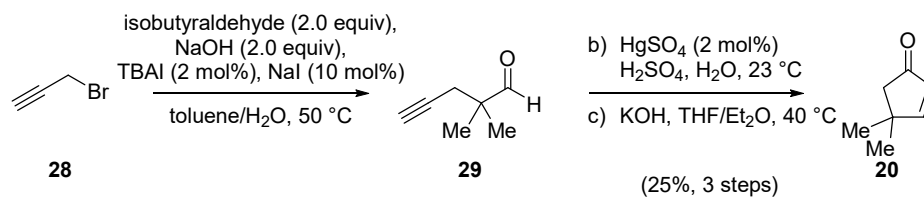
Ketone 23. To a suspension of CuI (2.72 g, 14.3 mmol, 1.3 equiv) in THF (50 mL) at 0 °C was added LiSiPhMe₂ (1.0 M in THF, 28.6 mL, 28.6 mmol, 2.6 equiv). The mixture was stirred at that temperature for 30 min. Then the mixture was cooled to -78 °C. A solution of **27** (2.71 g, 11 mmol, 1.0 equiv) in THF (30 mL) was added slowly over the course of 5 min. The mixture was stirred for additional 15 min. The reaction was quenched by addition of NH₄Cl solution. The mixture was warmed to 23 °C and poured into a separatory funnel and the layers were separated. The aqueous layer was extracted by EtOAc (2× 100 mL). The combined organic layer was dried (Na₂SO₄), filtered and concentrated. The residue was purified by column chromatography (hexanes:EtOAc = 10:1) to provide a mixture of diastereomers (2.05 g, 49%) with inseparable PhMe₂SiOH. The diastereomers were dissolved in toluene (40 mL) and DBU (1.64 mL, 11 mmol,

2.0 equiv) was added at 23 °C. The mixture was stirred at that temperature for 12 h. Upon completion, the reaction was quenched by addition of NH₄Cl solution. The mixture was poured into a separatory funnel and the layers were separated. The aqueous layer was extracted by EtOAc (2× 50 mL). The combined organic layer was dried (Na₂SO₄), filtered and concentrated. The residue was purified by column chromatography (hexanes:EtOAc = 10:1) to provide **23** (1.89 g, 45%) as a yellow oil with inseparable PhMe₂SiOH. **23**: R_f = 0.33 (silica gel, hexanes:EtOAc, 10:1); ¹H NMR (500 MHz, CDCl₃) δ 7.48–7.46 (m, 2 H), 7.36–7.32 (m, 3 H), 7.16 (d, *J* = 8.7 Hz, 2 H), 6.86 (d, *J* = 8.7 Hz, 2 H), 4.25 (dd, *J* = 39.8 Hz, 11.8 Hz, 2 H), 3.81 (s, 3 H), 3.62 (dd, *J* = 9.1 Hz, 2.9 Hz, 1 H), 3.05 (dd, *J* = 9.2 Hz, 3.4 Hz, 1 H), 2.37 (dd, *J* = 17.4 Hz, 6.4 Hz, 1 H), 2.11–2.02 (m, 2 H), 1.87 (dd, *J* = 17.5 Hz, 10.9 Hz, 1 H), 1.47 (d, *J* = 11.1 Hz, 1 H), 1.06 (d, *J* = 6.4 Hz, 3 H), 0.34 (s, 3 H), 0.32 (s, 3 H).

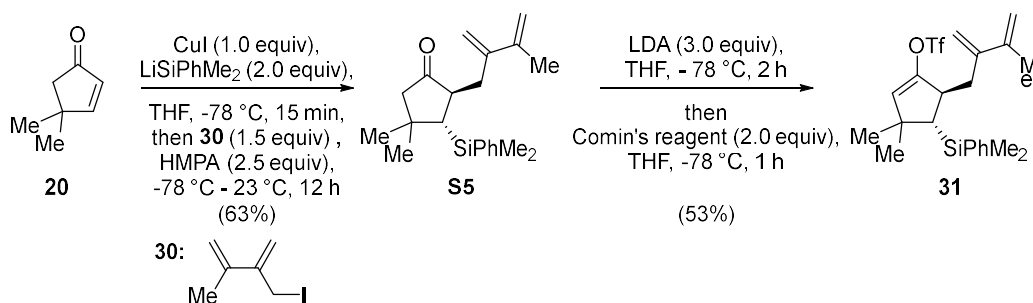


Iodide 22. **22** (0.96 g, 35%) was prepared via reported procedure^[15] from **23** (2.13 g, 5.6 mmol) **22**: R_f = 0.58 (silica gel, hexanes:EtOAc, 10:1); ¹H NMR (500 MHz, CDCl₃) δ 7.49–7.45 (m, 2 H), 7.36–7.33 (m, 3 H), 7.24–7.22 (m, 2 H), 6.88–6.86 (m, 2 H), 6.01 (t, *J* = 2.25 Hz, 1 H), 4.39 (dd, *J* = 31.1 Hz, 11.8 Hz, 2 H), 3.81 (s, 3 H), 3.40 (dd, *J* = 9.6 Hz, 4.5 Hz, 1 H), 3.17 (dd, *J* = 9.6 Hz, 3.6 Hz, 1 H), 2.77–2.74 (m, 1 H), 2.66–2.63 (m, 1 H), 1.28 (dd, *J* = 6.3 Hz, 5.7 Hz, 1 H), 0.96 (d, *J* = 6.8 Hz, 3 H), 0.25 (s, 3 H), 0.24 (s, 3 H).

East fragment

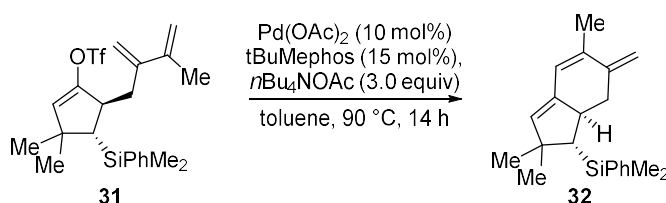


Enone 20. To a vigorously stirred solution of NaOH (96 g, 2.4 mol, 2.0 equiv), TBAI (8.8 g, 24 mmol, 2 mol%) and NaI (18 g, 0.12 mol, 10 mol%) in toluene (200 mL) and H₂O (150 mL) at 50 °C was added a solution of isobutyraldehyde (219 mL, 2.4 mol, 2.0 equiv) and **28** (132 mL, 1.2 mol, 1.0 equiv) in toluene (100 mL) slowly over the course of 5 h. The stirring was continued at that temperature for 18 h. Upon completion, the reaction was quenched and neutralized by HCl solution. The mixture was poured into a separatory funnel and the layers were separated. The organic layer was directly poured into a flask filled with H₂SO₄ (0.7 M in H₂O, 300 mL) and HgSO₄ (4.1 g, 14 mmol). And the mixture was stirred at 23 °C for 12 h. Upon completion, the reaction was quenched and neutralized by NaHCO₃ solution. The mixture was poured into a separatory funnel and the layers were separated. The aqueous layer was extracted by EtOAc (2 × 200 mL). The combined organic layer was concentrated. The residue was added to a flask with KOH (5 % in H₂O, 250 mL), THF (80 mL) and Et₂O (300 mL). The mixture was heated to 40 °C and refluxed at that temperature for 18 h. Upon completion, the reaction was quenched by addition of NH₄Cl solution. The mixture was poured into a separatory funnel and the layers were separated. The aqueous layer was extracted by Et₂O (200 mL). The mixture was distilled at 95 °C under atmosphere pressure to remove solvents. And following reduced pressure distillation (130 °C, 60-30 mmHg) provided **20** (33 g, 25% over 3 steps) as a yellow oil. All data of **20** were identical with commercial sample from Sigma-Aldrich.

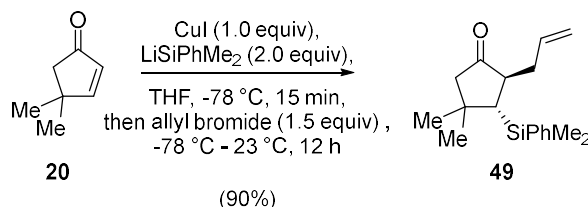


Triflate 31: To a suspension of CuI (190 mg, 1.0 mmol, 1.0 equiv) in THF (5 mL) was added LiPhSiMe₂ (1.0 M in THF, 2 mL, 2.0 equiv) and the mixture was stirred at that temperature for 30 min. Then it was cooled to -78 °C and a solution of **20** (110 mg, 1.0 mmol, 1.0 equiv) in THF (3 mL) was added over the course of 5 min. The mixture was stirred at that temperature for 15 min. HMPA (0.4 mL) and **30** (312 mg, 1.5 mmol, 1.5 equiv) was added and the mixture was slowly warmed to 23 °C. The stirring was continued for 12 h. Upon completion, the reaction was quenched by addition of NH₄Cl solution. The mixture was warmed to 23 °C and poured into a separatory funnel and the layers were separated. The aqueous layer was extracted by EtOAc (2 × 10 mL). The combined organic layer was dried (Na₂SO₄), filtered and concentrated. The residue was purified by column chromatography (hexanes:EtOAc = 40:1) to provide **S5** (205 mg, 63%) as a pale-yellow oil. **S5**: *R*_f = 0.41 (silica gel, hexanes:EtOAc, 10:1); ¹H NMR (500 MHz, CDCl₃) δ 7.53–7.51 (m, 2 H), 7.35–7.33 (m, 3 H), 5.07 (s, 1 H), 4.88 (s, 1 H), 4.84 (s, 1 H), 4.70 (s, 1 H), 2.55–2.51 (m, 1 H), 2.33 (ddd, *J* = 36.5 Hz, 14.7 Hz, 6.4 Hz, 2 H), 2.14 (d, *J* = 16.5 Hz, 1 H), 1.98 (dd, *J* = 16.7 Hz, 1.3 Hz, 1 H), 1.84 (s, 3 H), 1.45 (d, *J* = 9.7 Hz, 1 H), 1.11 (s, 3 H), 1.02 (s, 3 H), 0.38 (s, 3 H), 0.37 (s, 3 H). To a solution of *i*Pr₂NH (0.17 mL, 1.2 mmol) in THF (2 mL) at 0 °C was added *n*BuLi (2.5 M in THF, 0.4 mL, 1.0 mmol, 3.0 equiv). The mixture was stirred at that temperature for 15 min. Then the mixture was cooled to -78 °C and a solution of **S5** (108 mg, 0.33 mmol, 1.0 equiv) in THF (1 mL) was added. The mixture was stirred at that temperature for 2 h, and a solution of Comin's reagent (259 mg, 0.66 mmol, 2.0 equiv) in THF (1 mL) was added. The

stirring was continued at that temperature for 1 h. Upon completion, the reaction was quenched by addition of NaHCO₃ solution. The mixture was warmed to 23 °C and poured into a separatory funnel and the layers were separated. The aqueous layer was extracted by EtOAc (2 × 5 mL). The combined organic layer was dried (Na₂SO₄), filtered and concentrated. The residue was purified by column chromatography (Et₃N-buffered silica gel, hexanes) to provide **31** (86.1 mg, 53%) as a colorless oil. **31**: R_f = 0.69 (silica gel, hexanes:EtOAc, 10:1); ¹H NMR (500 MHz, CDCl₃) δ 7.52–7.50 (m, 2 H), 7.36–7.34 (m, 3 H), 5.40 (s, 1 H), 5.13 (s, 1 H), 4.96 (s, 1 H), 4.92 (s, 1 H), 4.73 (s, 1 H), 3.12–3.08 (m, 1 H), 2.50 (d, *J* = 14.0 Hz, 7.1 Hz, 1 H), 2.31 (d, *J* = 13.9 Hz, 6.9 Hz, 1 H), 1.84 (s, 3H), 1.49 (d, *J* = 5.2, 1 H), 1.18 (s, 3H), 1.07 (s, 3H), 0.40 (s, 3H), 0.30 (s, 3H).

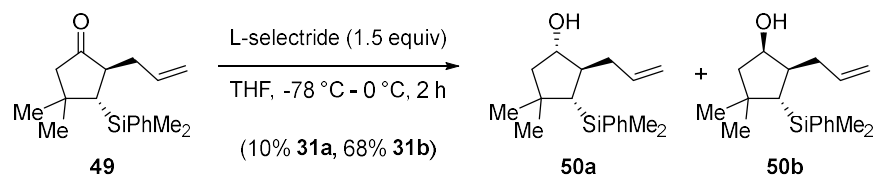


Triene 32. **31** (22.9 mg, 0.05 mmol, 1.0 equiv) was treated with standard procedure described in chapter 2. But only **32** was observed. **32**: ¹H NMR (500 MHz, CDCl₃) δ 7.56–7.55 (m, 2 H), 7.36–7.34 (m, 3 H), 5.94 (s, 1 H), 5.39 (d, *J* = 1.8 Hz, 1 H), 4.75 (s, 1 H), 4.59 (s, 1 H), 2.86–2.80 (m, 1 H), 2.22 (dd, *J* = 14.1 Hz, 5.1 Hz, 1 H), 1.85 (s, 3 H), 1.83 (tt, *J* = 13.7 Hz, 2.6 Hz, 1 H), 1.09 (d, *J* = 10.6 Hz, 1 H), 1.07 (s, 3 H), 1.01 (s, 3 H), 0.43 (s, 3 H), 0.41 (s, 3 H).

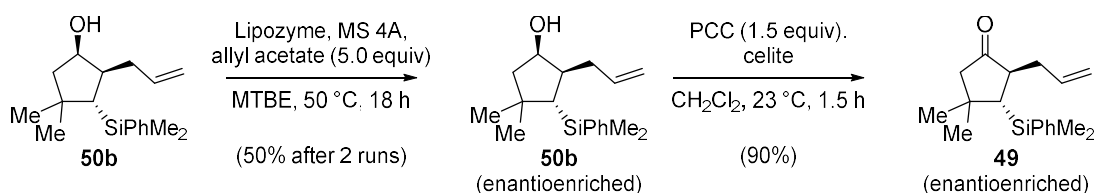


Ketone 49. **49** (53.9 g, 90%) was prepared by the procedure described for **S3** from **20** (23.1g, 0.21 mol, 1.0 equiv) as a yellow oil. **49**: R_f = 0.44 (silica gel, hexanes:EtOAc, 10:1); ¹H NMR (500 MHz, CDCl₃) δ 7.54–7.52 (m, 2 H), 7.37–7.35 (m, 3 H), 5.47–5.39 (m, 1 H), 4.90 (d,

$J = 10.2$ Hz, 1H), 4.62 (d, $J = 16.2$ Hz, 1 H), 2.45–2.37 (m, 2 H), 2.01 (dd, $J = 48.6$ Hz, 16.4 Hz, 2 H), 1.55 (s, 3 H), 1.41 (d, $J = 11.8$ Hz, 1 H), 1.13 (s, 3 H), 1.01 (s, 3 H), 0.43 (s, 6 H).

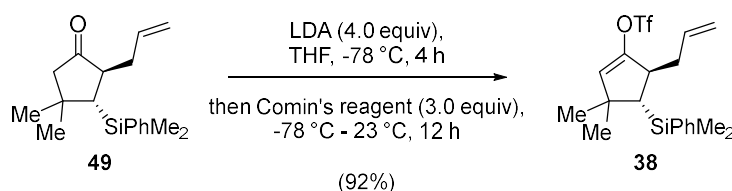


Alcohol 50a and 50b. To a solution of **49** (30.1 g, 105 mmol, 1.0 equiv) in THF (200 mL) at -78 °C was added L-selectride (1.0 M in THF, 160 mL, 160 mmol, 1.5 equiv). The mixture was stirred at that temperature for 1 h. Then it was warmed to 0 °C and stirred at that temperature for 1 h. Upon completion, the reaction was quenched by addition of NH_4Cl . The mixture was warmed to 23 °C and poured into a separatory funnel and the layers were separated. The aqueous layer was extracted by EtOAc (2×100 mL). The combined organic layer was dried (Na_2SO_4), filtered and concentrated. The residue was purified by careful column chromatography (hexanes:EtOAc = 100:1 \rightarrow 40:1 \rightarrow 30:1 \rightarrow 20:1) to provide **50b** (18.1 g, 60%) as a colorless oil, and a mixture of **50a** and **50b** (5.44 g, 18%, 5:4) was collected for recovery of **49**. **50a**: $R_f = 0.43$ (silica gel, hexanes:EtOAc, 5:1). **50b**: $R_f = 0.46$ (silica gel, hexanes:EtOAc, 5:1); $^1\text{H NMR}$ (500 MHz, CDCl_3) δ 7.54–7.51 (m, 2 H), 7.34–7.32 (m, 3 H), 5.86–5.75 (m, 1 H), 5.02–4.95 (m, 2 H), 4.25–4.21 (m, 1 H), 2.13–2.06 (m, 2 H), 1.99–1.88 (m, 1 H), 1.77 (dd, $J = 13.8$ Hz, 5.9 Hz, 1 H), 1.49 (d, $J = 13.8$ Hz, 3.9 Hz, 1 H), 1.23 (d, $J = 5.2$ Hz, 1 H), 1.05 (s, 3 H), 0.98 (s, 3 H), 0.37 (s, 3 H), 0.36 (s, 3 H).



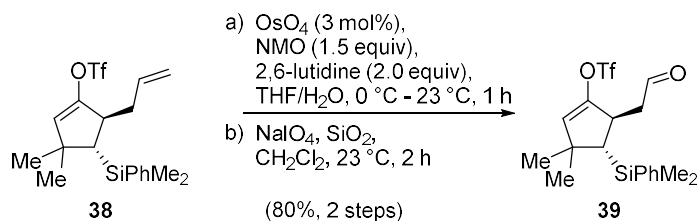
Enantioenriched 49. To a solution of **50b** (33.7 g, 117 mmol, 1.0 equiv) in *t*BuOMe (MTBE, 580 mL) at 50 °C was added Lipozyme[®] (11.6 g) and activated MS 4Å (104 g). Then

allyl acetate (54 mL, 0.58 mol, 5.0 equiv) was added. The mixture was refluxed at that temperature for 18 h. Then it was cooled to 23 °C and filtered. The filtrate was concentrated, and the residue was purified by column chromatography (hexanes:EtOAc = 40:1) to provide enantioenriched **50b** (24.3 g). The it was put in a second run with Lipozyme[®] (8.4 g), activated MS 4Å (75 g) and MTBE (420 mL) for 18 h. Then the mixture was cooled to 23 °C and filtered. The filtrate was concentrated, and the residue was purified by column chromatography (hexanes:EtOAc = 40:1) to provide enantioenriched **50b** (16.8 g, 50%, 70% e. e.) as a colorless oil. To a suspension of PCC (20.4 g, 95 mmol, 1.5 equiv) and celite (40 g) in CH₂Cl₂ (400 mL) at 23 °C was added enantioenriched **50b** (18.1 g, 63 mmol, 1.0 equiv). The mixture was stirred at that temperature for 90 min. Upon completion, the mixture was diluted by hexane (400 mL) and filtered through a pad of silica gel (hexanes:EtOAc = 5:1). The filtrate was concentrated, and the residue was purified by column chromatography (hexanes:EtOAc = 10:1) to provide enantioenriched **49** (17.2 g, 90%) as a pale-yellow oil. The mixture of racemic **50a** and **50b** was treated by the same condition to recover racemic **49** (4.87 g, 90%).



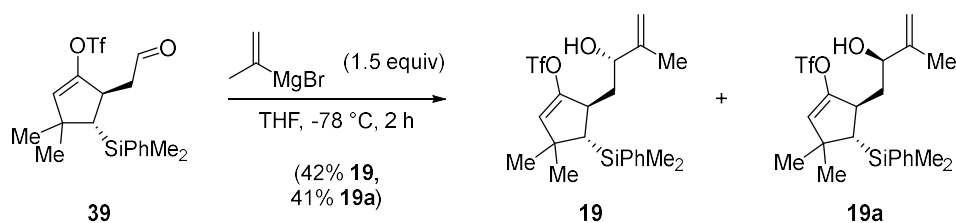
Triflate 38. To a solution of *i*Pr₂NH (19.6 mL, 0.14 mol) in THF (100 mL) at 0 °C was added *n*BuLi (2.5 M in THF, 48 mL, 0.12 mmol, 4.0 equiv). The mixture was stirred at that temperature for 15 min. Then the mixture was cooled to -78 °C and a solution of **49** (8.58 g, 30 mmol, 1.0 equiv) in THF (50 mL) was added. The mixture was stirred at that temperature for 4 h, and a solution of Comin's reagent (35.8 g, 90 mmol, 3.0 equiv) in THF (50 mL) was added. The mixture was slowly warmed to 23 °C and stirred at that temperature for 12 h. Upon completion,

the reaction was quenched by addition of NaHCO₃ solution. The mixture was poured into a separatory funnel and the layers were separated. The aqueous layer was extracted by EtOAc (2 × 200 mL). The combined organic layer was dried (Na₂SO₄), filtered and concentrated. The residue was filter through a pad of Et₃N-buffered silica gel (hexanes:EtOAc = 10:1). The filtrate was concentrated, and the residue was purified by column chromatography (Et₃N-buffered silica gel, hexanes) to provide **38** (22.9 g, 92%) as a colorless oil. **38**: *R*_f = 0.90 (silica gel, hexanes:EtOAc, 10:1); ¹H NMR (500 MHz, CDCl₃) δ 7.53–7.50 (m, 2 H), 7.37–7.35 (m, 3 H), 5.58–5.48(m, 1 H), 5.44 (d, *J* = 1.8 Hz, 1 H), 5.03–5.01 (m, 1 H), 4.81–4.76 (m, 1 H), 3.09–3.04 (m, 1 H), 2.24–2.17 (m, 1 H), 1.70 (ddd, *J* = 14.7 Hz, 8.8 Hz, 4.9 Hz, 1 H), 1.37 (d, *J* = 8.5 Hz, 1 H), 1.11 (s, 3H), 1.09 (s, 3H), 0.43 (s, 3H), 0.41 (s, 3H).



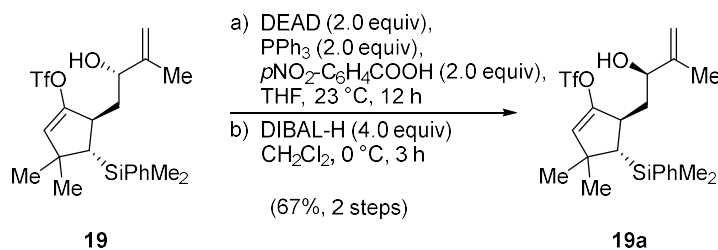
Aldehyde 39. To a solution of **38** (11.5 g, 27.5 mmol, 1.0 equiv) in THF (160 mL) at 0 °C was added OsO₄ (4% in H₂O, 0.8 mmol, 3 mol%). Then a solution NMO (4.83 g, 41 mmol, 1.5 equiv) in H₂O (40 mL) was added, followed by addition of 2,6-lutidine (7.1 ml, 55 mmol, 2.0 equiv). The mixture was slowly warmed to 23 °C and was stirred at that temperature for 1 h. Upon completion, the reaction was quenched by addition of Na₂S₂O₃ solution. The mixture was poured into a separatory funnel and the layers were separated. The aqueous layer was extracted by EtOAc (2 × 100 mL). The combined organic layer was dried (Na₂SO₄), filtered and concentrated. The residue was filter through a pad silica gel (EtOAc). The filtrated was concentrated and used in next step without further purification. NaIO₄ supported on SiO₂ was prepared by reported procedure^[19] from NaIO₄ (25.7 g, 120 mmol) and SiO₂ (100 g) in H₂O (50 mL). That supported NaIO₄ (110 g)

was suspended in CH₂Cl₂ (200 mL) and a solution of crude from the last step in CH₂Cl₂ (100 mL) was added. The mixture was stirred at 23 °C for 1 h. Upon completion, the reaction was filtered. The filtrate was concentrated, and the residue was purified by column chromatography (hexanes:EtOAc = 40:1→30:1) to provide **39** (18.5 g, 80%) as a colorless oil. **39**: R_f = 0.46 (silica gel, hexanes:EtOAc, 10:1); ¹H NMR (500 MHz, CDCl₃) δ 9.37 (m, 1 H), 7.50–7.48 (m, 2 H), 7.37–7.35 (m, 3 H), 5.51 (d, *J* = 1.9 Hz, 1 H), 3.49–3.44 (m, 1 H), 2.25 (ddd, *J* = 17.7 Hz, 7.0 Hz, 2.0 Hz, 1 H), 2.05 (dd, *J* = 17.5 Hz, 3.8 Hz, 1 H), 1.30 (d, *J* = 9.2 Hz, 1 H), 1.17 (s, 3H), 1.16 (s, 3H), 0.46 (s, 3H), 0.45 (s, 3H).

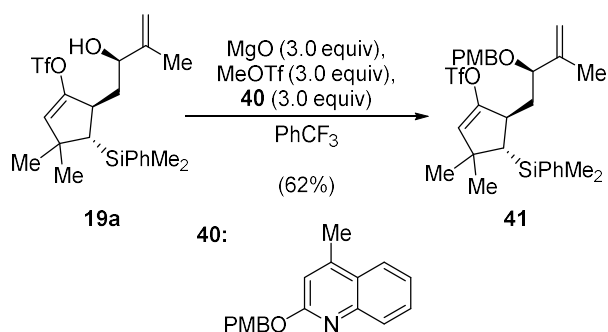


Alcohol 19 and 19a. To a solution of **39** (9.24 g, 22 mmol, 1.0 equiv) in THF (200 mL) at -78 °C was added isopropenylmagnesium bromide (0.5 M in THF, 66 mL, 33 mmol, 1.5 equiv). The mixture was stirred at that temperature for 2 h. Upon completion, the reaction was quenched by addition of NH₄Cl solution. The mixture was warmed to 23 °C and poured into a separatory funnel and the layers were separated. The aqueous layer was extracted by EtOAc (2 × 200 mL). The combined organic layer was dried (Na₂SO₄), filtered and concentrated. The residue was purified by careful column chromatography (hexanes:EtOAc = 100:1→50:1→10:1) to provide **19** (8.48 g, 42%) and **19a** (8.37 g, 41%) as colorless oil. **19**: R_f = 0.55 (silica gel, hexanes:EtOAc, 5:1); ¹H NMR (500 MHz, CDCl₃) δ 7.50–7.48 (m, 2 H), 7.37–7.35 (m, 3 H), 5.43 (d, *J* = 1.6 Hz, 1 H), 4.77 (s, 1 H), 4.69 (s, 1 H), 4.08–4.05 (m, 1 H), 3.25 (tt, *J* = 8.8 Hz, 2.3 Hz, 1 H), 1.50 (s, 3 H), 1.48–1.44 (m, 1 H), 1.36–1.30 (m, 1 H), 1.14 (s, 3 H), 1.11–1.10 (m, 4 H), 0.46 (s, 3 H), 0.46 (s, 3H). **19a**: R_f = 0.48 (silica gel, hexanes:EtOAc, 5:1); ¹H NMR (500 MHz, CDCl₃) δ 7.57–7.55

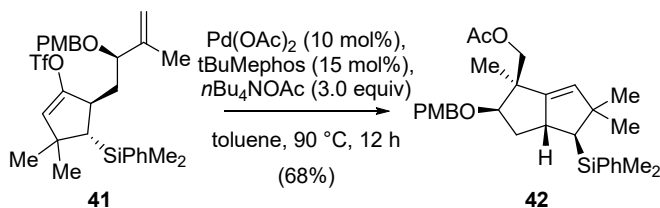
(m, 2 H), 7.38–7.37 (m, 3 H), 5.46 (d, $J = 1.4$ Hz, 1 H), 4.79 (s, 1 H), 4.74 (t, $J = 1.5$ Hz, 1 H), 3.79 (qui, $J = 4.1$ Hz, 1 H), 3.00–2.97 (m, 1 H), 1.61–1.57 (m, 2 H), 1.53 (s, 1 H), 1.51–1.50 (m, 1 H), 1.16 (s, 3 H), 1.14 (s, 3 H), 0.95 (d, $J = 4.1$ Hz, 1 H), 0.44 (s, 3 H), 0.42 (s, 3H).



Inversion. To a solution of **19** (8.48 g, 18.3 mmol, 1.0 equiv), PPh₃ (9.4 g, 36 mmol, 2.0 equiv) and 4-nitrobenzoic acid (6.0 g, 36 mmol, 2.0 equiv) in THF (100 mL) at 23 °C was added DEAD (40% in toluene, 16.4 mL, 36 mmol, 2.0 equiv) dropwise over the course of 10 min. The mixture was stirred at that temperature for 12 h. Upon completion, the reaction was quenched by addition of NaHCO₃ solution. The mixture was poured into a separatory funnel and the layers were separated. The aqueous layer was extracted by Et₂O (2 × 100 mL). The combined organic layer was dried (Na₂SO₄), filtered and concentrated. The residue was filtered through a pad of silica gel (hexanes:EtOAc = 10:1). The filtrate was concentrated and purified by column chromatography (hexanes:EtOAc = 100:1→50:1) to provide a yellow oil. The yellow oil was dissolved in CH₂Cl₂ (50 mL) and DIBAL-H (1.0 M in CH₂Cl₂, 50 mL, 50 mmol) was added at 0 °C. The mixture was stirred at that temperature for 3 h. Upon completion, the reaction was quenched by addition of Rochelle salt solution. The mixture was warmed to 23 °C and poured into a separatory funnel and the layers were separated. The aqueous layer was extracted by CH₂Cl₂ (2 × 100 mL). The combined organic layer was dried (Na₂SO₄), filtered and concentrated. The residue was purified by column chromatography (hexanes:EtOAc = 100:1→50:1) to provide **19a** (5.59 g, 67% over 2 steps).

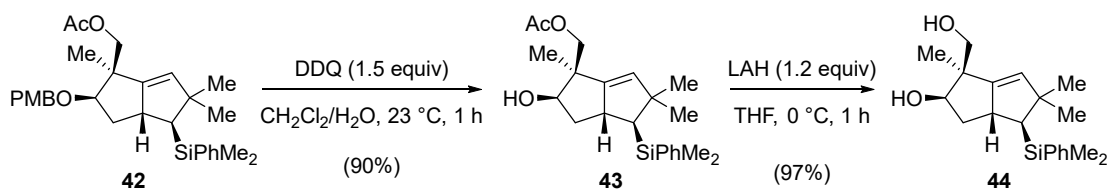


PMB ether 42. To **19a** (13.9 g, 30 mmol, 1.0 equiv), MgO (3.6 g, 90 mmol, 3.0 equiv) and **40**^[20] (25.1 g, 90 mmol, 3.0 equiv) in PhCF₃ (200 mL) was added MeOTf (9.84 mL, 90 mmol, 3.0 equiv) at 0 °C. The mixture was warmed to 23 °C and stirred at that temperature for 12 h. Upon completion, the mixture was filtered through celite (EtOAc). The filtrate was concentrated and filtered through a pad of Et₃N-buffered silica gel (hexanes:EtOAc = 50:1). The filtrate was concentrated, and the residue was purified by column chromatography (Et₃N-buffered silica gel, hexanes) to provide **41** as a pale-yellow oil (10.8 g, 62%). **41:** R_f = 0.63 (silica gel, hexanes:EtOAc, 10:1); ¹H NMR (500 MHz, CDCl₃) δ 7.48–7.43 (m, 2 H), 7.32–7.23 (m, 5 H), 6.89–6.87 (m, 2 H), 5.36 (d, *J* = 1.8 Hz, 1 H), 4.86 (d, *J* = 1.0 Hz, 2 H), 4.35 (d, *J* = 11.1 Hz, 1 H), 4.11 (d, *J* = 11.1 Hz, 1 H), 3.84–3.80 (m, 1 H), 3.81 (s, 3 H), 3.26 (tt, *J* = 8.9 Hz, 2.0 Hz, 1 H), 1.67 (ddd, *J* = 14.5 Hz, 10.8 Hz, 2.1 Hz, 1 H), 1.54 (s, 3 H), 1.39 (ddd, *J* = 14.7 Hz, 8.7 Hz, 2.6 Hz, 1 H), 1.22 (d, *J* = 9.1 Hz, 1 H), 1.06 (s, 3 H), 1.03 (s, 3H), 0.39 (s, 3 H), 0.38 (s, 3 H).



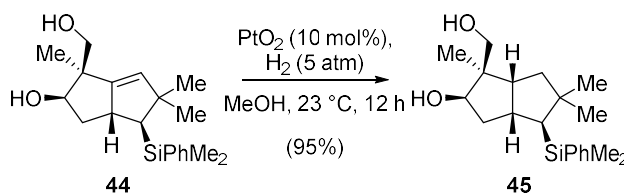
Acetate 42. To Pd(OAc)₂ (142 mg, 0.63 mmol, 10 mol%), tBuMephos (298 mg, 0.96 mmol, 15 mol%), *n*Bu₄OAc (5.74 g, 19 mmol, 3.0 equiv) in toluene (65 mL) was added **41** (3.70 g, 6.4 mmol, 1.0 equiv). The mixture in a sealed tube was heated to 90 °C and stirred at that temperature

for 12 h. Upon completion, the mixture was cooled to 23 °C and filtered through a pad of silica gel (EtOAc). The filtrate was concentrated, and the residue was purified by column chromatography (silica gel, hexanes:EtOAc = 30:1→20:1) to provide **42** (2.13 g, 68%) as a yellow oil. **42**: R_f = 0.60 (silica gel, hexanes:EtOAc, 5:1); ^1H NMR (500 MHz, CDCl_3) δ 7.52–7.50 (m, 2 H), 7.35–7.33 (m, 3 H), 7.22–7.20 (m, 2 H), 6.87–6.85 (m, 2 H), 5.12 (d, J = 2.6 Hz, 1 H), 3.89 (q, J = 11.2 Hz, 2 H), 3.85–3.81 (m, 1 H), 3.80 (s, 3 H), 2.77–2.71 (m, 1 H), 2.00 (s, 3 H), 1.96–1.92 (m, 1 H), 1.08 (d, J = 10.1 Hz, 1 H), 1.06 (s, 3 H), 1.00–0.94 (m, 1 H), 0.98 (s, 3 H), 0.96 (s, 3H), 0.36 (s, 3 H), 0.35 (s, 3 H).

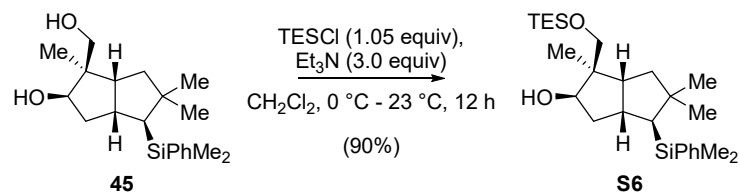


Diol 44. To a solution of **42** (3.34 g, 6.8 mmol, 1.0 equiv) in CH_2Cl_2 (60 mL) and H_2O (6 mL) was added DDQ (2.31 g, 10.2 mmol, 1.5 equiv). The mixture was stirred at 23 °C for 1 h. Upon completion, the reaction was quenched by addition of NaHCO_3 solution. The mixture was poured into a separatory funnel and the layers were separated. The aqueous layer was extracted by CH_2Cl_2 (2 × 50 mL). The combined organic layer was dried (Na_2SO_4), filtered and concentrated. The residue was purified by column chromatography (hexanes:EtOAc = 5:1) to provide **43** (2.27 g, 90%) as a pale-yellow oil. **43**: R_f = 0.18 (silica gel, hexanes:EtOAc, 5:1); ^1H NMR (500 MHz, CDCl_3) δ 7.52–7.50 (m, 2 H), 7.34–7.33 (m, 3 H), 5.17 (d, J = 2.6 Hz, 1 H), 4.12–4.09 (m, 1 H), 3.92 (q, J = 10.8 Hz, 2 H), 2.81 (tdd, J = 11.1 Hz, 6.7 Hz, 2.6 Hz, 1 H), 2.04 (s, 3 H), 1.92 (dt, J = 11.8 Hz, 6.3 Hz, 1 H), 1.07 Hz (d, J = 10.8 Hz, 1H), 1.01 (s, 3 H), 1.01 (s, 3 H), 0.98 (s, 3 H), 0.95–0.88 (m, 1H), 0.37 (s, 3 H), 0.37 (s, 3H). To a solution of **43** (2.23 g, 6.0 mmol, 1.0 equiv) in THF (50 mL) was added LAH (1.0 M in THF, 7.2 mL, 7.2 mmol, 1.2 equiv) at 0 °C. The mixture

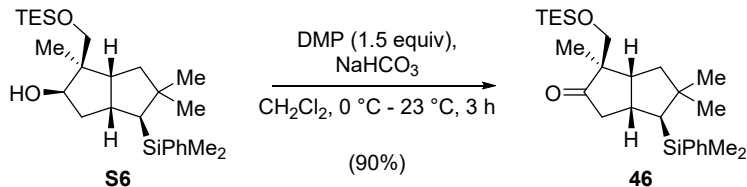
was stirred at that temperature for 1 h. Upon completion, the reaction was quenched by addition of Rochelle salt solution. The mixture was warmed to 23 °C and poured into a separatory funnel, and the layers were separated. The aqueous layer was extracted by EtOAc (2 × 50 mL). The combined organic layer was dried (Na₂SO₄), filtered and concentrated. The residue was purified by column chromatography (hexanes:EtOAc = 2:1) to provide **44** (1.92 g, 97%) as a white solid. **44**: *R_f* = 0.40 (silica gel, hexanes:EtOAc, 1:1); ¹H NMR (500 MHz, CDCl₃) δ 7.52–7.50 (m, 2 H), 7.34–7.33 (m, 3 H), 5.19 (d, *J* = 2.6 Hz, 1 H), 4.15–4.11 (m, 1 H), 3.43 (s, 2 H), 2.80–2.76 (tdd, *J* = 10.8 Hz, 6.6 Hz, 2.6 Hz, 1 H), 1.91 (dt, *J* = 11.6 Hz, 6.2 Hz, 1 H), 1.09 (d, *J* = 10.3 Hz, 1 H), 1.02 (s, 3 H), 1.00 (s, 3 H), 0.99 (s, 3 H), 0.96–0.91 (m, 1 H), 0.37 (s, 3 H), 0.37 (s, 3 H).



Diol 45. A Q-tube was equipped with PtO₂ (45 mg, 0.2 mmol, 10 mol%) and a solution of **44** (0.66 g, 2.0 mmol, 1.0 equiv) in MeOH (20 mL) was added. The Q-tube was purged with H₂ (5 atm), and the mixture was stirred at 23 °C for 12 h. Upon completion, the mixture was filtered through a pad of silica gel (EtOAc). The filtrate was concentrated to provide **45** (0.63 g, 95%) as a white solid. **45**: *R_f* = 0.40 (silica gel, hexanes:EtOAc, 1:1); ¹H NMR (500 MHz, CDCl₃) δ 7.50–7.49 (m, 2 H), 7.33–7.32 (m, 3 H), 3.96–3.92 (m, 1 H), 3.48–3.41 (m, 2 H), 2.47–2.40 (m, 1 H), 2.13–2.07 (m, 1 H), 1.91–1.86 (m, 1 H), 1.84 (d, *J* = 4.4 Hz, 1 H), 1.65 (t, *J* = 5.2 Hz, 1 H), 1.47 (t, *J* = 11.4 Hz, 1 H), 1.27 (dd, *J* = 12.2 Hz, 8.4 Hz, 1 H), 1.14–1.06 (m, 1 H), 0.94 (s, 3 H), 0.91 (s, 3 H), 0.86 (s, 3 H), 0.76 (d, *J* = 11.0 Hz, 1 H), 0.34 (s, 6 H).

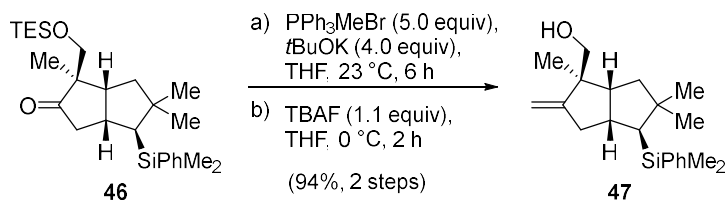


TES ether S6. To a solution of **45** (1.98 g, 6.0 mmol, 1.0 equiv) in CH₂Cl₂ (50 mL) was added TESCl (0.93 g, 6.18 mmol, 1.05 equiv) and Et₃N (2.46 mL, 17.7 mmol, 3.0 equiv) at 0 °C. The mixture was slowly warmed to 23 °C and stirred at that temperature for 12 h. Upon completion, the reaction was quenched by addition of NH₄Cl solution. The mixture was poured into a separatory funnel and the layers were separated. The aqueous layer was extracted by CH₂Cl₂ (2 × 50 mL). The combined organic layer was dried (Na₂SO₄), filtered and concentrated. The residue was purified by column chromatography (hexanes:EtOAc = 40:1→20:1→10:1) to provide **S6** (2.40 g, 90%) as a colorless oil. **S6**: *R*_f = 0.50 (silica gel, hexanes:EtOAc, 10:1); ¹H NMR (500 MHz, CDCl₃) δ 7.52–7.49 (m, 2 H), 7.33–7.31 (m, 3 H), 3.94 (ddd, *J* = 10.4 Hz, 6.1 Hz, 2.1 Hz, 1 H), 3.41 (dd, *J* = 33.9 Hz, 9.1 Hz, 2 H), 2.46–2.38 (m, 1 H), 2.38 (d, *J* = 2.1 Hz, 1 H), 2.02 (td, *J* = 10.6 Hz, 8.3 Hz, 1 H), 1.88 (ddd, *J* = 14.1 Hz, 7.9 Hz, 6.2 Hz, 1 H), 1.45 (t, *J* = 11.5 Hz, 1 H), 1.23 (dd, *J* = 12.2 Hz, 8.2 Hz, 1 H), 1.16–1.09 (m, 1 H), 0.94 (t, *J* = 8.0 Hz, 9 H), 0.92 (s, 3H), 0.88 (s, 3H), 0.84 (s, 3H), 0.74 (d, *J* = 11.0 Hz, 1 H), 0.58 (q, *J* = 8.0 Hz, 6 H), 0.33 (s, 3H), 0.33 (s, 3H).



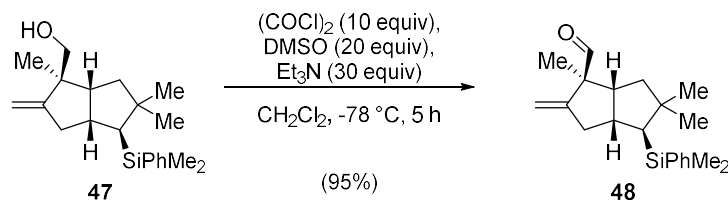
Ketone 46. DMP (1.84 g, 4.3 mmol, 1.5 equiv) and NaHCO₃ (1.8 g) was stirred in CH₂Cl₂ (10 mL) at 23 °C for 15 min. Then the mixture was cooled to 0 °C and a solution of **S6** (1.38 g, 2.9 mmol, 1.0 equiv) in CH₂Cl₂ (20 mL) was added. The mixture was slowly warmed to 23 °C and

stirred at that temperature for 3 h. Upon completion, the reaction was quenched by $\text{Na}_2\text{S}_2\text{O}_3/\text{NaHCO}_3$ solution. The mixture was poured into a separatory funnel and the layers were separated. The aqueous layer was extracted by CH_2Cl_2 (2×20 mL). The combined organic layer was dried (Na_2SO_4), filtered and concentrated. The residue was purified by column chromatography (hexanes:EtOAc = 10:1) to provide **46** (1.14 g, 90%) as a pale-yellow oil. **46**: R_f = 0.50 (silica gel, hexanes:EtOAc, 15:1); $^1\text{H NMR}$ (500 MHz, CDCl_3) δ 7.51–7.48 (m, 2 H), 7.34–7.31 (m, 3 H), 3.51 (d, J = 9.0 Hz, 1 H), 3.37 (d, J = 9.0 Hz, 1 H), 2.81–2.75 (m, 1 H), 2.71–2.66 (m, 1 H), 2.34 (dd, J = 19.4 Hz, 10.5 Hz, 1 H), 1.55 (dd, J = 19.6 Hz, 6.1 Hz, 1 H), 1.42 (dd, J = 12.0 Hz, 6.9 Hz, 1 H), 1.17 (t, J = 12.4 Hz, 1 H), 0.97 (s, 3 H), 0.97 (s, 3 H), 0.94 (t, J = 8.0 Hz, 9 H), 0.84 (s, 3 H), 0.75 (d, J = 8.1 Hz, 1 H), 0.56 (q, J = 8.0 Hz, 6 H), 0.38 (s, 3 H), 0.35 (s, 3 H).



Alkene 47. PPh_3MeBr (4.64 g, 13.0 mmol, 5.0 equiv) and $t\text{BuOK}$ (1.16 g, 10.3 mmol, 4.0 equiv) was stirred in THF (15 mL) at 23 °C for 15 min. Then a solution of **46** (1.15 g, 2.6 mmol, 1.0 equiv) in THF (10 mL) was added. The mixture was stirred at that temperature for 6 h. Upon completion, the reaction was quenched by addition of NH_4Cl solution. The mixture was poured into a separatory funnel and the layers were separated. The aqueous layer was extracted by Et_2O (2×20 mL). The combined organic layer was washed by H_2O and brine, dried (Na_2SO_4), filtered and concentrated. The residue was filtered through a pad of silica gel (hexanes:EtOAc = 40:1). The filtrate was concentrated, and the residue was used in the next step. To a solution of the residue in THF (25 mL) was added TBAF (1.0 M in THF, 2.8 mL, 2.8 mmol, 1.1 equiv) at 0 °C. The mixture was stirred at that temperature for 2 h. Upon completion, the reaction was quenched by

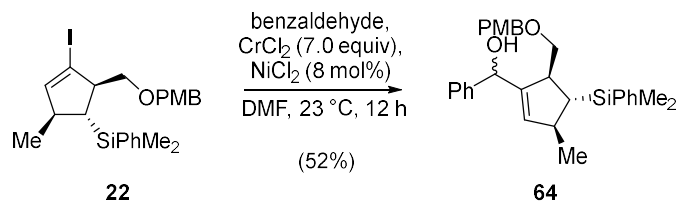
addition of NH_4Cl solution. The mixture was poured into a separatory funnel and the layers were separated. The aqueous layer was extracted by EtOAc (2×20 mL). The combined organic layer was dried (Na_2SO_4), filtered and concentrated. The residue was purified by column chromatography (hexanes:EtOAc = 20:1 \rightarrow 10:1) to provide **47** (802 mg, 94% over 2 steps) as a colorless oil. **47**: R_f = 0.28 (silica gel, hexanes:EtOAc, 10:1); ^1H NMR (500 MHz, CDCl_3) δ 7.52–7.50 (m, 2 H), 7.35–7.32 (m, 3 H), 4.90 (s, 1 H), 4.76–4.75 (m, 1 H), 3.30 (dd, J = 10.4 Hz, 2.8 Hz, 1 H), 3.19 (t, J = 10.2 Hz, 1 H), 2.48–2.26 (m, 2 H), 2.22–2.17 (m, 1H), 1.58 (d, J = 15.4 Hz, 1 H), 1.30 (dd, J = 11.8 Hz, 7.0 Hz, 1 H), 1.25 (dd, J = 10.0 Hz, 3.0 Hz, 1 H), 1.05 (q, J = 10.6 Hz, 1 H), 0.98 (s, 3 H), 0.89 (s, 3 H), 0.87 (s, 3 H), 0.65 (d, J = 8.5 Hz, 1 H), 0.37 (s, 3 H), 0.33 (s, 3 H).



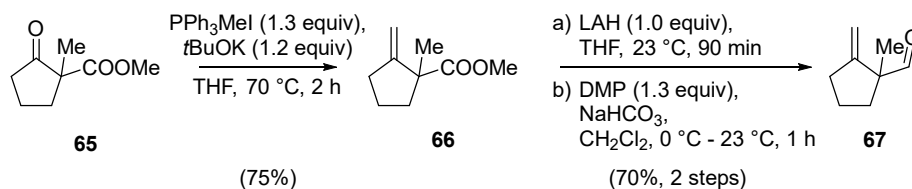
Aldehyde 48. To a solution of $(\text{COCl})_2$ (86 μL , 1.0 mmol, 10 equiv) in CH_2Cl_2 (1 mL) was added DMSO (0.14 mL, 2.0 mmol, 20 equiv) at -78 °C. The mixture was stirred at that temperature for 15 min. A solution of **47** (32.8 mg, 0.1 mmol, 1.0 equiv) in CH_2Cl_2 (1 mL) was added. The mixture was stirred at -78 °C for 3 h. Then Et_3N (0.42 mL, 3.0 mmol, 30 equiv) was added, and the stirring was continued at -78 °C for 2 h. Upon completion, the reaction was quenched by addition of H_2O . The mixture was warmed to 23 °C and poured into a separatory funnel, and the layers were separated. The aqueous layer was extracted by CH_2Cl_2 (2×2 mL). The combined organic layer was dried (Na_2SO_4), filtered and concentrated. The residue was purified by column chromatography (hexanes:EtOAc = 10:1) to provide **48** (30.9 mg, 95%) as a pale-yellow oil. **48**: R_f = 0.52 (silica gel, hexanes:EtOAc, 10:1); ^1H NMR (400 MHz, CDCl_3) δ 9.22 (s, 1 H), 7.52–

7.48 (m, 2 H), 7.35–7.32 (m, 3 H), 5.00 (s, 1 H), 4.87 (d, $J = 2.2$ Hz, 1 H), 2.85 (ddd, $J = 8.8$ Hz, 7.3 Hz, 12.1 Hz, 1 H), 2.63 (qd, $J = 9.5$ Hz, 2.9 Hz, 1 H), 2.27 (ddt, $J = 16.6$ Hz, 10.2 Hz, 2.8 Hz, 1 H), 1.58–1.53 (m, 1 H), 1.30 (dd, $J = 11.8$ Hz, 7.2 Hz, 1 H), 1.10 (s, 3 H), 1.08–1.02 (m, 1 H), 0.92 (s, 3 H), 0.90 (s, 3 H), 0.63 (d, $J = 9.1$ Hz, 1 H), 0.38 (s, 3 H), 0.34 (s, 3 H).

Model Study

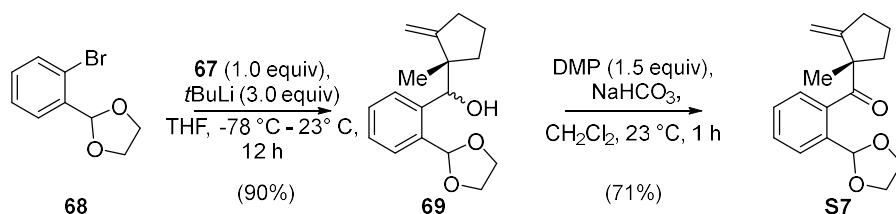


64. In glove box, to a solution of CrCl_2 (25 mg, 0.2 mmol, 7.0 equiv) in degassed DMF (0.5 mL) was added benzaldehyde (3.2 mg, 0.03 mmol, 1.0 equiv) and **22** (22 mg, 0.05 mmol, 1.3 equiv). Then NiCl_2 (0.3 mg, 8 mol%) was added. The mixture was stirred in glove box at 23 °C for 12 h. Upon completion, the reaction was diluted by Et_2O and quenched by addition of H_2O . The mixture was poured into a separatory funnel, and the layers were separated. The aqueous layer was extracted by Et_2O (2×2 mL). The combined organic layer was dried (Na_2SO_4), filtered and concentrated. The residue was purified by column chromatography (hexanes: $\text{EtOAc} = 10:1$) to provide **64** (7.3 mg, 52%, mixture of diastereomers) as a yellow oil. **64**(minor diastereomer): $R_f = 0.62$ (silica gel, hexanes: EtOAc , 5:1). **64**(major diastereomer): $R_f = 0.47$ (silica gel, hexanes: EtOAc , 5:1); $^1\text{H NMR}$ (400 MHz, CDCl_3) δ 7.39–7.20 (m, 10 H), 7.15 (d, $J = 8.6$ Hz, 2 H), 6.86 (d, $J = 8.6$ Hz, 2 H), 5.48 (s, 1 H), 4.75 (d, $J = 8.2$ Hz, 1 H), 4.28 (dd, $J = 25.4$ Hz, 10.4 Hz, 2 H), 3.81 (s, 3 H), 3.29 (t, $J = 9.2$ Hz, 1 H), 3.16 (dd, $J = 9.2$ Hz, 3.9 Hz, 1 H), 2.75–2.70 (m, 1 H), 2.65–2.61 (m, 1 H), 2.35 (t, $J = 7.5$ Hz, 1 H), 0.96 (d, $J = 6.8$ Hz, 3 H), 0.62 (t, $J = 6.0$ Hz, 1 H), 0.16 (s, 3 H), 0.14 (s, 3 H).



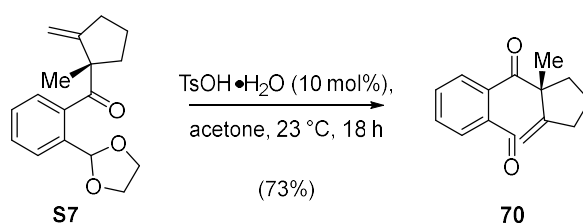
Aldehyde 67. To a solution of PPh_3MeI (9.05 g, 22 mmol, 1.3 equiv) in THF (50 mL) was added $t\text{BuOK}$ (2.32 g, 20 mmol, 1.2 equiv). The mixture was heated to 70 °C and refluxed at that temperature for 1 h. Then a solution of **65** (2.65 g, 17 mmol, 1.0 equiv) in THF (30 mL) was added. Reflux was continued at that temperature for 1 h. Upon completion, the reaction was quenched by addition of NH_4Cl solution. The mixture was cooled to 23 °C and poured into a separatory funnel, and the layers were separated. The aqueous layer was extracted by Et_2O (2 × 30 mL). The combined organic layer was dried (Na_2SO_4), filtered and concentrated (40 °C, 500 mmHg). The residue was purified by column chromatography (petanes: Et_2O = 10:1) to provide **66** (1.98 g, 75%) as a pale-yellow oil. **66**: R_f = 0.77 (silica gel, hexanes: EtOAc , 5:1); $^1\text{H NMR}$ (500 MHz, CDCl_3) δ 4.98 (t, J = 2.0 Hz, 1 H), 4.95 (t, J = 2.3 Hz, 1 H), 3.67 (s, 3 H), 2.51–2.38 (m, 2 H), 2.34 (dt, J = 12.5 Hz, 6.9 Hz, 1 H), 1.83–1.78 (m, 1 H), 1.69–1.65 (m, 1 H), 1.61–1.59 (m, 1 H), 1.33 (s, 3 H). To suspension of LAH (684 mg, 18 mmol, 1.0 equiv) in THF (20 mL) was added a solution of **66** (2.77 g, 18 mmol, 1.0 equiv) in THF (30 mL). The mixture was stirred at 23 °C for 90 min. Upon completion, the reaction was quenched by addition of Rochelle salt solution. The mixture was poured into a separatory funnel, and the layers were separated. The aqueous layer was extracted by Et_2O (2 × 30 mL). The combined organic layer was dried (Na_2SO_4), filtered and concentrated. The residue was filtered through a pad of silica gel (Et_2O). The filtrate was concentrated (40 °C, 500 mmHg) and used in next step. DMP (5.5 g, 13 mmol, 1.3 equiv) and NaHCO_3 (12 g) was stirred in CH_2Cl_2 (20 mL) at 23 °C for 15 min. Then a solution of the residue (10 mmol, 1.0 equiv) in CH_2Cl_2 (30 mL) was added at 0 °C. The mixture was slowly warmed to

23 °C and stirred at that temperature for 1 h. Upon completion, the reaction was quenched by Na₂S₂O₃/NaHCO₃ solution. The mixture was poured into a separatory funnel and the layers were separated. The aqueous layer was extracted by CH₂Cl₂ (2 × 20 mL). The combined organic layer was dried (Na₂SO₄), filtered and concentrated (40 °C, 550 mmHg). The residue was purified by column chromatography (petanes:Et₂O = 10:1) to provide **67** (0.97 g, 70% over 2 steps) as a pale-yellow oil. **67**: *R*_f = 0.64 (silica gel, hexanes:EtOAc, 10:1); ¹H NMR (500 MHz, CDCl₃) δ 9.31 (s, 1 H), 5.14 (t, *J* = 2.3 Hz, 1 H), 4.83 (t, *J* = 2.3 Hz, 1 H), 2.50–2.35 (m, 2 H), 2.20 (dt, *J* = 12.8 Hz, 6.8 Hz, 1 H), 1.76–1.70 (m, 2 H), 1.54 (qui, *J* = 6.6 Hz, 1 H), 1.21 (s, 3 H).

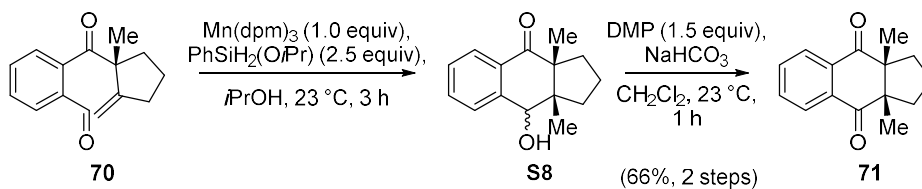


S7. To a solution of **68**^[28] (1.37 g, 6.0 mmol, 1.5 equiv) in THF (20 mL) was added *t*BuLi (1.6 M in THF, 7.5 mL, 12 mmol, 3.0 equiv) at -78 °C. The mixture was stirred at that temperature for 1 h. Then a solution of **67** (0.50 g, 4.0 mmol, 1.0 equiv) in THF (10 mL) was added. The mixture was slowly warmed to 23 °C and stirred at that temperature for 12 h. Upon completion, the reaction was quenched by addition of NH₄Cl solution. The mixture was poured into a separatory funnel and the layers were separated. The aqueous layer was extracted by EtOAc (2 × 20 mL). The combined organic layer was dried (Na₂SO₄), filtered and concentrated. The residue was purified by column chromatography (hexanes:EtOAc = 15:1→5:1) to provide **69** (0.99 g, 90%) as a mixture of diastereomers. DMP (193 mg, 0.45 mmol, 1.5 equiv) and NaHCO₃ (400 mg) was stirred in CH₂Cl₂ (2 mL) at 23 °C for 15 min. Then a solution of **69** (83 mg, 0.3 mmol, 1.0 equiv) in CH₂Cl₂ (1 mL) was added. The mixture was stirred at 23 °C for 1 h. Upon completion, the reaction was quenched by Na₂S₂O₃/NaHCO₃ solution. The mixture was poured into a separatory

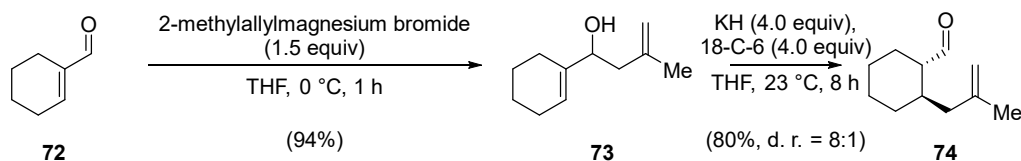
funnel and the layers were separated. The aqueous layer was extracted by CH_2Cl_2 (2×3 mL). The combined organic layer was dried (Na_2SO_4), filtered and concentrated. The residue was purified by column chromatography (hexanes:EtOAc = 10:1) to provide **S7** (59 mg, 71%) as a yellow oil. **S7**: R_f = 0.75 (silica gel, hexanes:EtOAc, 3:1); ^1H NMR (400 MHz, CDCl_3) δ 7.54–7.52 (m, 1 H), 7.38–7.34 (m, 1 H), 7.30–7.27 (m, 2 H), 5.94 (s, 1 H), 5.12 (t, J = 1.9 Hz, 1 H), 5.04 (t, J = 2.0 Hz, 1 H), 4.00–3.93 (m, 4 H), 2.53–2.48 (m, 1 H), 2.42–2.34 (m, 1 H), 2.13–2.04 (m, 1 H), 1.72–1.62 (m, 3 H), 1.38 (s, 3 H).



Model 70. To a solution of **S7** (59 mg, 0.22 mmol, 1.0 equiv) in acetone (2.5 mL) was added $\text{TsOH}\cdot\text{H}_2\text{O}$ (4 mg, 0.02 mmol, 10 mol%). The mixture was stirred at 23 °C for 18 h. Upon completion, the reaction was diluted by EtOAc and quenched by addition of NH_4Cl solution. The mixture was poured into a separatory funnel and the layers were separated. The aqueous layer was extracted by EtOAc (2×2 mL). The combined organic layer was dried (Na_2SO_4), filtered and concentrated. The residue was purified by column chromatography (hexanes:EtOAc = 8:1) to provide **70** (36 mg, 73%) as a pale-yellow oil. **70**: R_f = 0.46 (silica gel, hexanes:EtOAc, 5:1); ^1H NMR (500 MHz, CDCl_3) δ 9.91 (s, 1 H), 7.86–7.84 (m, 1 H), 7.59–7.52 (m, 2 H), 7.34–7.33 (m, 1 H), 5.13 (t, J = 2.0 Hz, 1 H), 5.00 (t, J = 2.1 Hz, 1 H), 2.52–2.47 (m, 1 H), 2.33–2.26 (m, 1 H), 2.10–2.04 (m, 1 H), 1.76–1.72 (m, 1 H), 1.67–1.61 (m, 2 H), 1.46 (s, 2 H).

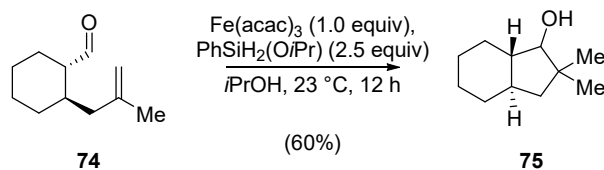


Diketone 71. To a flask equipped with $\text{Mn}(\text{dpm})_3$ (30.2 mg, 0.5 mmol, 1.0 equiv) was added a solution of **70** (11.4 mg, 0.5 mmol, 1.0 equiv) in *i*PrOH (0.5 mL). Then Shenvi's silane (22 μL , 1.25 mmol, 2.5 equiv) was added. The mixture was stirred at 23 °C for 3 h. Upon completion, the reaction was diluted with EtOAc and quenched by addition of H_2O . The mixture was poured into a separatory funnel and the layers were separated. The aqueous layer was extracted by EtOAc (2×2 mL). The combined organic layer was concentrated. The residue was filtered through a pad of silica gel (hexanes:EtOAc = 5:1) to provide a mixture of **S8** (2 diastereomers) and **71**. This mixture was then treated with DMP and NaHCO_3 via described procedure. Then column chromatography (hexanes:EtOAc = 10:1) provide **71** (7.5 mg, 66%) as a yellow oil. **71**: R_f = 0.64 (silica gel, hexanes:EtOAc, 5:1); ^1H NMR (500 MHz, CDCl_3) δ 8.08–8.06 (m, 2 H), 7.74–7.72 (m, 2 H), 2.39–2.34 (m, 2 H), 1.83–1.77 (m, 2 H), 1.71–1.66 (m, 2 H), 1.26 (s, 6 H).



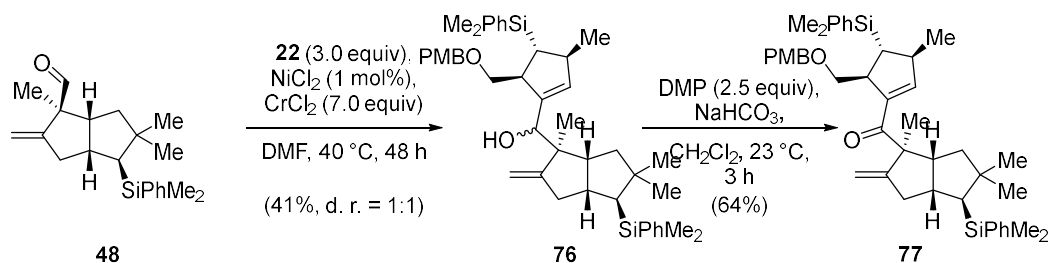
Model 74. To a solution of **72** (550 mg, 5.0 mmol, 1.0 equiv) in THF (30 mL) was added 2-methylallylmagnesium bromide (0.5 M in THF, 15 mL, 7.5 mmol, 1.5 equiv) at 0 °C. The mixture was stirred at that temperature for 1 h. Upon completion, the reaction was quenched by addition of NH_4Cl solution. The mixture was warmed to 23 °C and poured into a separatory funnel, and the layers were separated. The aqueous layer was extracted by EtOAc (2×30 mL). The combined organic layer was dried (Na_2SO_4), filtered and concentrated. The residue was purified by column chromatography (hexanes:EtOAc = 1:1) to provide **73** (777 mg, 94%) as a colorless oil. **73**: R_f = 0.43 (silica gel, hexanes:EtOAc, 5:1); ^1H NMR (500 MHz, CDCl_3) δ 5.72–5.70 (m, 1 H), 4.88–4.87 (m, 1 H), 4.82–4.81 (m, 1 H), 4.10–4.08 (m, 1 H), 2.26–2.24 (m, 2 H), 2.10–2.02 (m, 2 H), 1.97–1.92 (m, 1 H), 1.70–1.53 (m, 4 H), 1.67 (d, J = 2.8 Hz, 1 H). To a suspension of KH (80

mg, 2.0 mmol, 4.0 equiv) in THF (4 mL) was added a solution of **73** (83 mg, 0.5 mmol, 1.0 equiv) in THF (3 mL) at 0 °C. Then a solution of 18-C-6 (528 mg, 2.0 mmol, 4.0 equiv) in THF (3 mL) was added at 0 °C. The mixture was slowly warmed to 23 °C and stirred at that temperature for 8 h. Upon completion, the reaction was quenched by addition of NH₄Cl solution. The mixture was poured into a separatory funnel and the layers were separated. The aqueous layer was extracted by Et₂O (2 × 10 mL). The combined organic layer was dried (Na₂SO₄), filtered and concentrated (40 °C, 550 mmHg). The residue was purified by column chromatography (petanes:Et₂O = 30:1→20:1) to provide **74** (66 mg, 80%, d. r. = 8:1) as a pale-yellow oil. **74**: R_f = 0.66 (silica gel, hexanes:EtOAc, 5:1); ¹H NMR (500 MHz, CDCl₃) δ 9.56 (d, *J* = 3.6 Hz, 1 H), 4.76 (s, 1 H), 4.68 (s, 1 H), 2.13–2.10 (m, 1 H), 2.04–2.00 (m, 1 H), 1.86–1.81 (m, 3 H), 1.77–1.71 (m, 3 H), 1.69 (s, 3 H), 1.41–1.36 (m, 1 H), 1.34–1.25 (m, 3 H), 0.97–0.92 (m, 1 H).



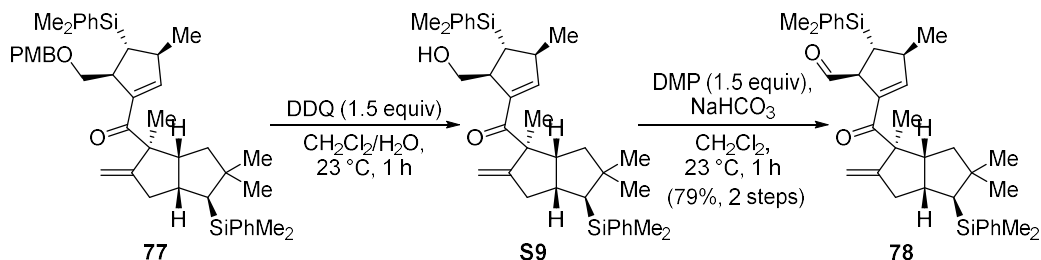
Alcohol 75. To a flask equipped with Fe(acac)₃ (17.6 mg, 0.05 mmol, 1.0 equiv) was added a solution of **74** (8.3 mg, 0.05 mmol, 1.0 equiv) in *i*PrOH (0.5 mL). Then Shenvi's silane (22 μL, 0.12 mmol, 2.5 equiv) was added. The mixture was stirred 23 °C for 12 h. Upon completion, the mixture was concentrated, and the residue was purified by column chromatography (hexanes:EtOAc = 10:1) to provide **75** (5.0 mg, 60%, single diastereomer) as a colorless oil. **75**: R_f = 0.30 (silica gel, hexanes:EtOAc, 10:1); ¹H NMR (500 MHz, CDCl₃) δ 3.24 (dd, *J* = 8.6 Hz, 5.5 Hz, 1 H), 1.98–1.96 (m, 1 H), 1.78–1.75 (m, 2 H), 1.73–1.71 (m, 1 H), 1.59–1.58 (m, 1 H), 1.26–1.25 (m, 1 H), 1.18–1.08 (m, 6 H), 1.04 (s, 3 H), 0.93 (s, 3H).

Ring B formation



Ketone 77. To a vial equipped with CrCl₂ (35 mg, 0.3 mmol, 7.0 equiv) in glove box was added a solution of **48** (13.7 mg, 42 μmol, 1.0 equiv) in degassed DMF (0.15 mL). Then **22** (62 mg, 0.13 mmol, 3.0 equiv) and NiCl₂ (0.8 mg/mL in DMF, 65 μL, 0.42 μmol, 1 mol%) was added. The mixture was stirred at 40 °C for 48 h. Then the mixture was moved out from glove box and cooled to 23 °C. H₂O (0.5 mL) and Et₂O (0.5 mL) was added. The mixture was poured into a separatory funnel and the layers were separated. The aqueous layer was extracted by Et₂O (2 × 1 mL). The combined organic layer was dried (Na₂SO₄), filtered and concentrated. The residue was purified by column chromatography (hexanes:EtOAc = 20:1→5:1) to provide **76** (12.0 mg, 41%, d. r. = 1:1) as a pale-yellow oil. And 3.0 mg of **48** and 13 mg of **22** was recovered. **76**: R_f = 0.32 and 0.41 (silica gel, hexanes:EtOAc, 10:1). A suspension of DMP (34 mg, 80 μmol, 2.5 equiv) and NaHCO₃ (60 mg) in CH₂Cl₂ (0.2 mL) was stirred for 15 min at 23 °C and a solution of **76** (22 mg, 32 μmol, 1.0 equiv) in CH₂Cl₂ was added. The mixture was stirred at that temperature for 3 h. Upon completion, the reaction was quenched by addition of Na₂S₂O₃/NaHCO₃ solution. The mixture was poured into a separatory funnel and the layers were separated. The aqueous layer was extracted by CH₂Cl₂ (2 × 1 mL). The combined organic layer was dried (Na₂SO₄), filtered and concentrated. The residue was purified by column chromatography (hexanes:EtOAc = 10:1→5:1) to provide **78** (14.1 mg, 64%) as a colorless oil. **78**: R_f = 0.60 (silica gel, hexanes:EtOAc, 10:1); ¹H NMR (500 MHz, CDCl₃) δ 7.49–7.43 (m, 4 H), 7.33–7.26 (m, 6 H), 7.18–7.17 (m, 2 H), 6.87–6.84 (m, 2 H), 6.61 (dd, *J* = 2.6 Hz, 1.6 Hz, 1 H), 4.89 (s, 1 H), 4.85 (s, 1 H), 4.31 (dd, *J* = 28.0 Hz,

11.7 Hz, 2 H), 3.81 (s, 3 H), 3.40 (dd, $J = 9.2$ Hz, 5.7 Hz, 1 H), 3.31 (dd, $J = 9.2$ Hz, 3.7 Hz, 1 H), 3.20–3.19 (m, 1 H), 3.16–3.12 (m, 1 H), 2.72–2.69 (m, 1 H), 2.45–2.40 (m, 1 H), 2.22–2.17 (m, 1 H), 1.36–1.33 (m, 1 H), 1.23–1.19 (m, 1 H), 1.11 (s, 3 H), 0.97 (d, $J = 7.0$ Hz, 3 H), 0.92–0.87 (m, 2 H), 0.88 (s, 3 H), 0.85 (s, 3 H), 0.56 (d, $J = 9.2$ Hz, 1 H), 0.32 (s, 3 H), 0.27 (s, 3 H), 0.23 (s, 3 H), 0.22 (s, 3 H).



Aldehyde 78. To a solution of **77** (14 mg, 20 μ mol, 1.0 equiv) in CH_2Cl_2 (0.4 mL) and H_2O (40 μ L) was added DDQ (6.8 mg, 30 μ mol, 1.5 equiv). The mixture was stirred at 23 $^\circ\text{C}$ for 1 h. Upon completion, the reaction was quenched by addition of NaHCO_3 solution. The mixture was poured into a separatory funnel and the layers were separated. The aqueous layer was extracted by CH_2Cl_2 (2×1 mL). The combined organic layer was dried (Na_2SO_4), filtered and concentrated. The residue was purified by column chromatography (hexanes:EtOAc = 20:1 \rightarrow 10:1) to provide **S9** (inseparable with 4-methoxybenzaldehyde, 86% NMR yield) as a pale-yellow oil. **S9**: $R_f = 0.35$ (silica gel, hexanes:EtOAc, 10:1). The mixture was used in the next step. A suspension of DMP (10 mg, 24 μ mol, 1.5 equiv) and NaHCO_3 (20 mg) in CH_2Cl_2 (0.1 mL) was stirred at 23 $^\circ\text{C}$ for 15 min. Then a solution of **S9** (16 μ mol) in CH_2Cl_2 (0.1 mL) was added. The mixture was stirred at that temperature for 1 h. Upon completion, the reaction was quenched by addition of $\text{Na}_2\text{S}_2\text{O}_3/\text{NaHCO}_3$ solution. The mixture was poured into a separatory funnel and the layers were separated. The aqueous layer was extracted by CH_2Cl_2 (2×0.5 mL). The combined organic layer was dried (Na_2SO_4), filtered and concentrated. The residue was purified by column

chromatography (hexanes:EtOAc = 10:1) to provide **78** (8.5 mg, 79 % in 2 steps) as a pale-yellow oil. **78**: R_f = 0.50 (silica gel, hexanes:EtOAc, 10:1); $^1\text{H NMR}$ (500 MHz, CDCl_3) δ 9.53 (d, J = 3.0 Hz, 1 H), 7.49–7.46 (m, 4 H), 7.36–7.31 (m, 6 H), 6.88 (dd, J = 2.5 Hz, 1.9 Hz, 1 H), 4.94–4.93 (m, 2 H), 3.63–3.61 (m, 1 H), 3.14–3.09 (m, 1 H), 2.89–2.86 (m, 1 H), 2.49–2.44 (m, 1 H), 2.07–2.02 (m, 1 H), 1.33 (m, 1 H), 1.21–1.18 (m, 1 H), 1.10 (s, 3 H), 0.97 (d, J = 7.0 Hz, 3 H), 0.89 (s, 3 H), 0.88–0.84 (m, 2 H), 0.85 (s, 3 H), 0.47 (d, J = 9.3 Hz, 1 H), 0.35 (s, 3 H), 0.30 (s, 3 H), 0.29 (s, 3 H), 0.29 (s, 3 H).

3.5 References

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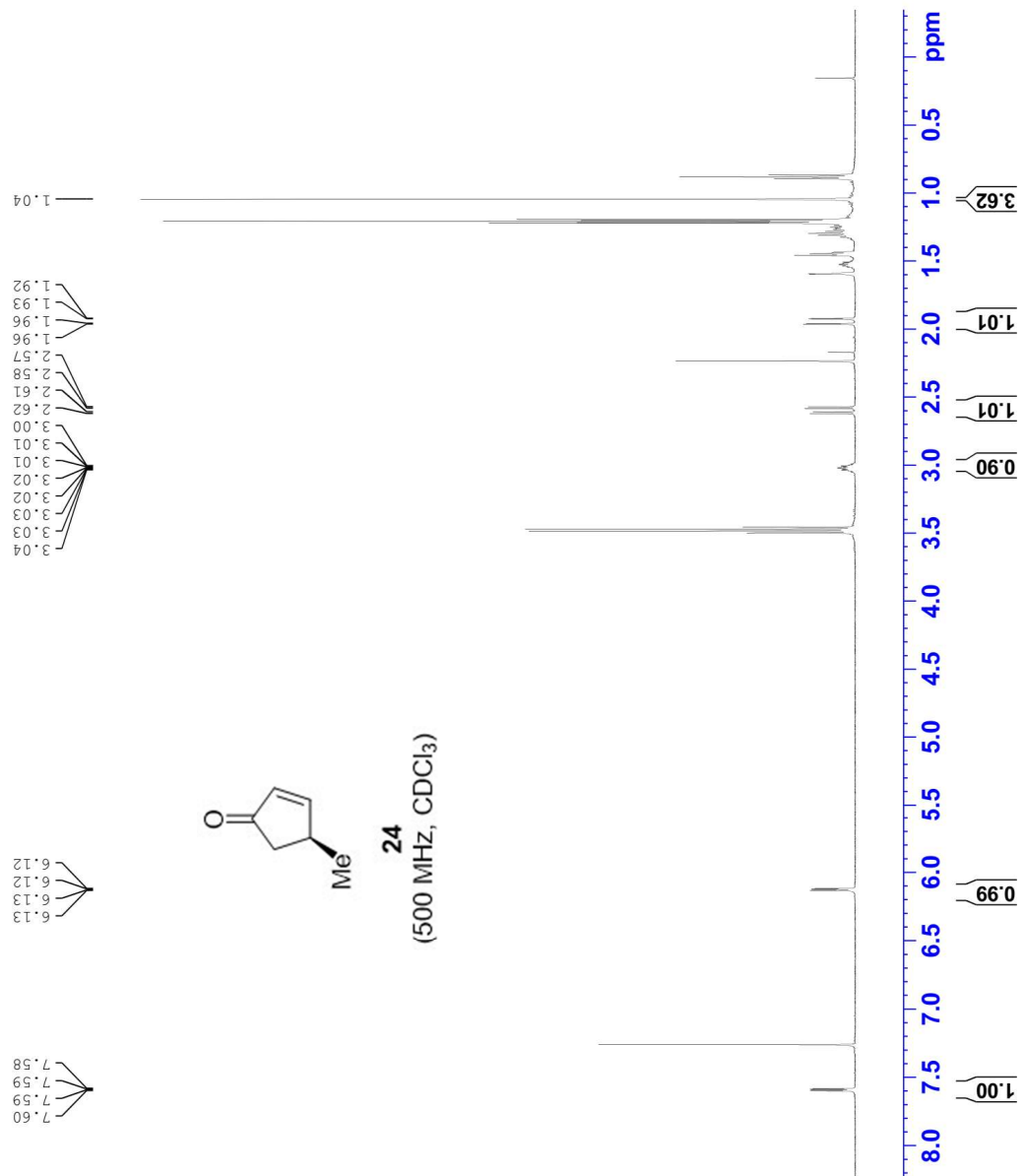
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3.6 NMR data of selected intermediates

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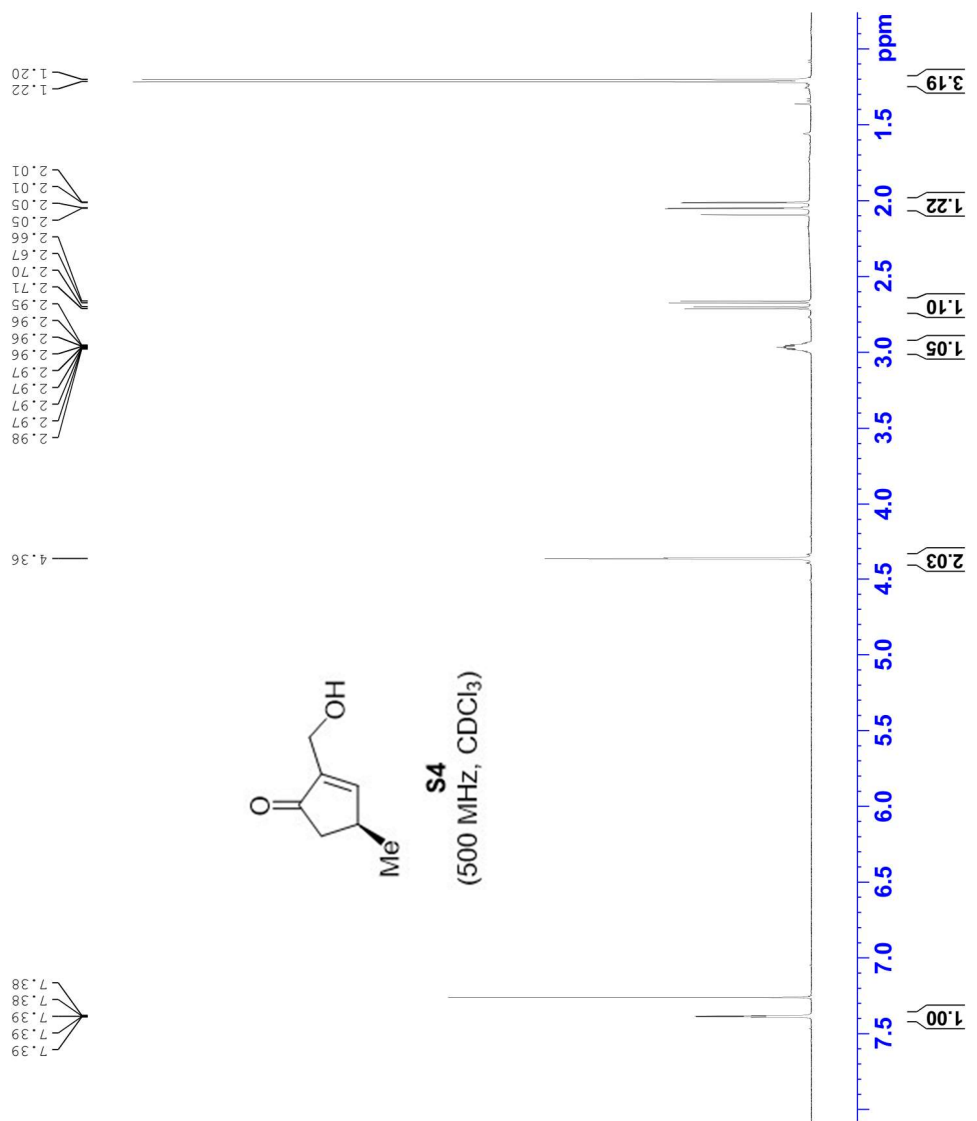
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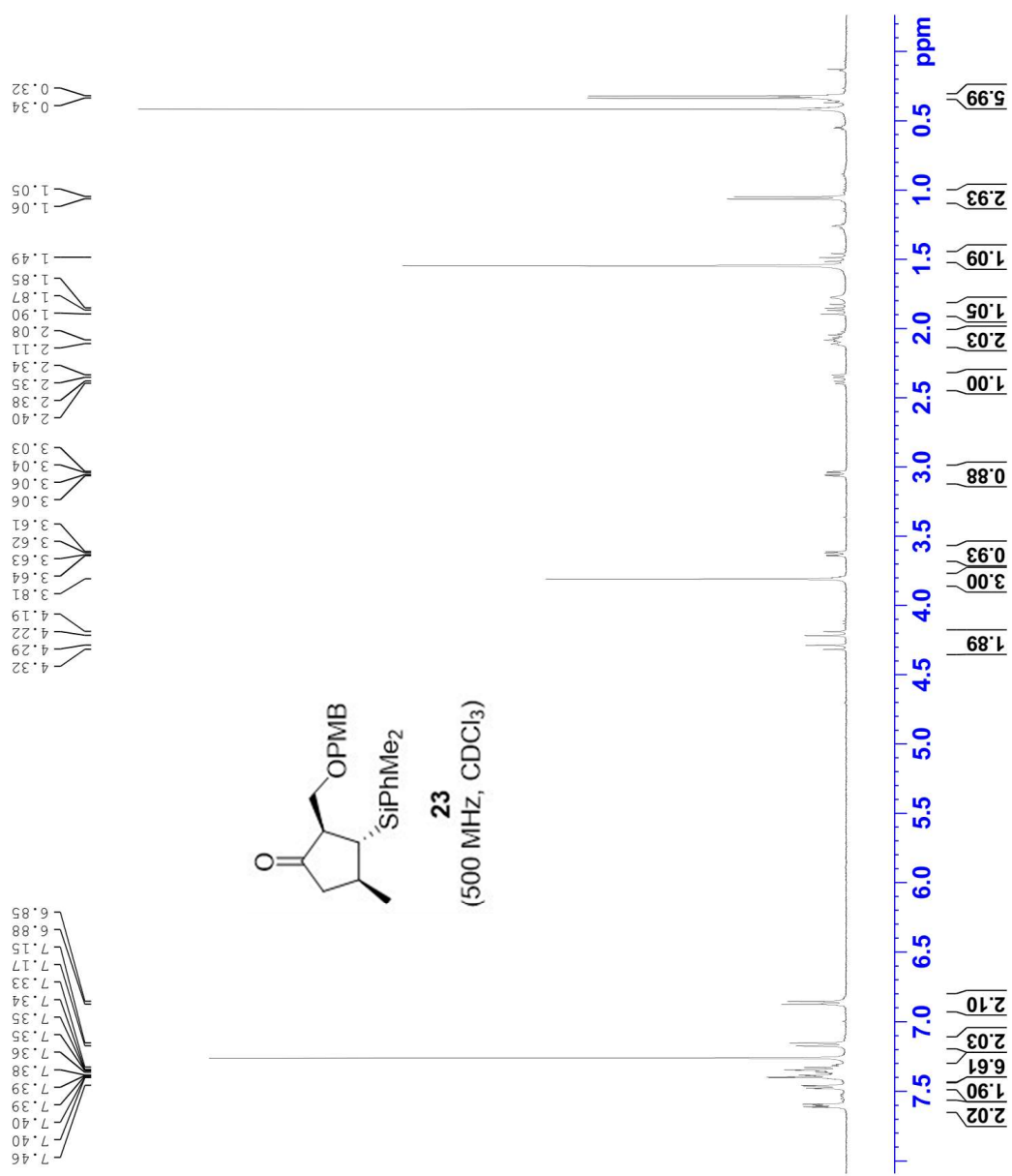
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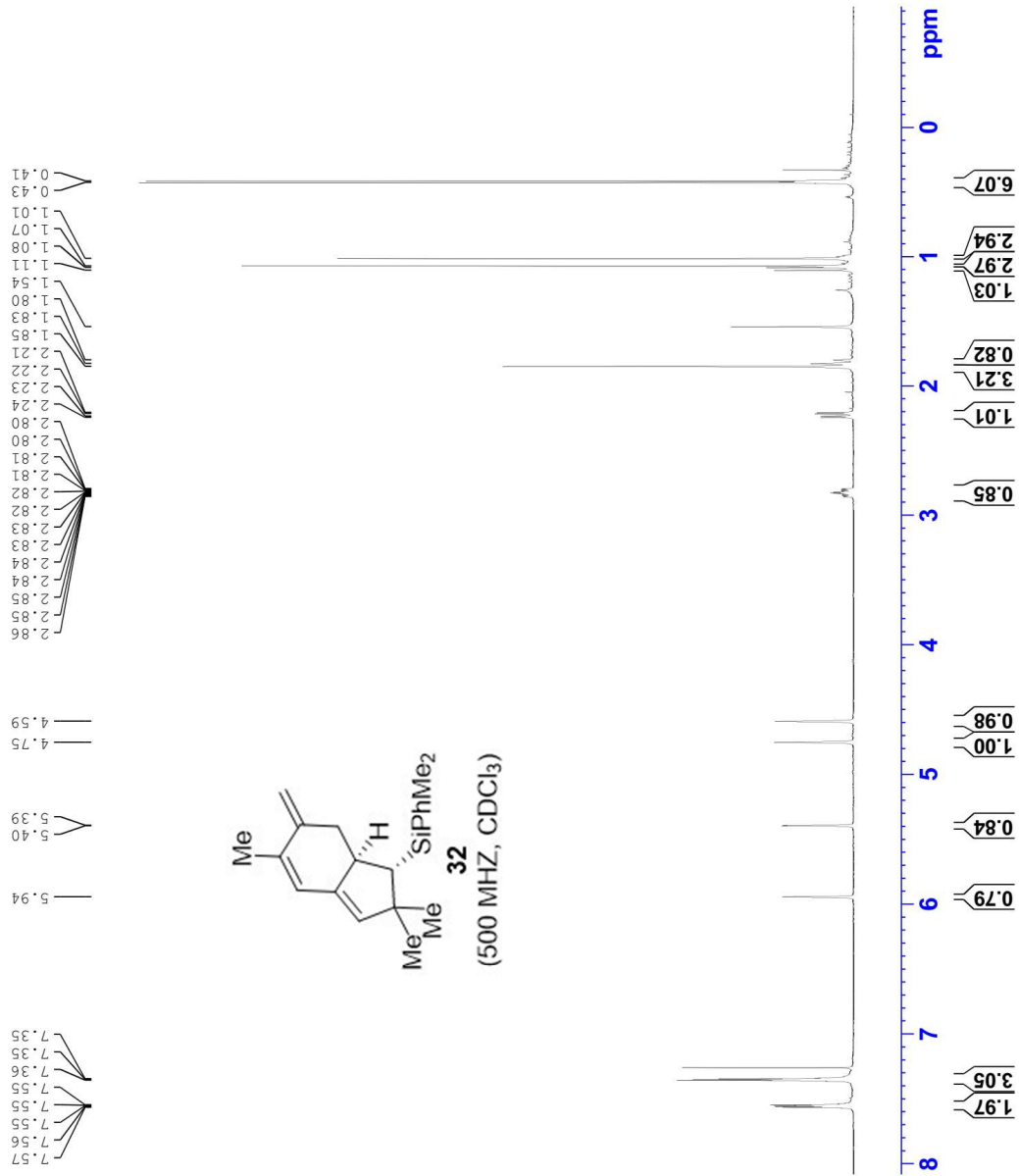


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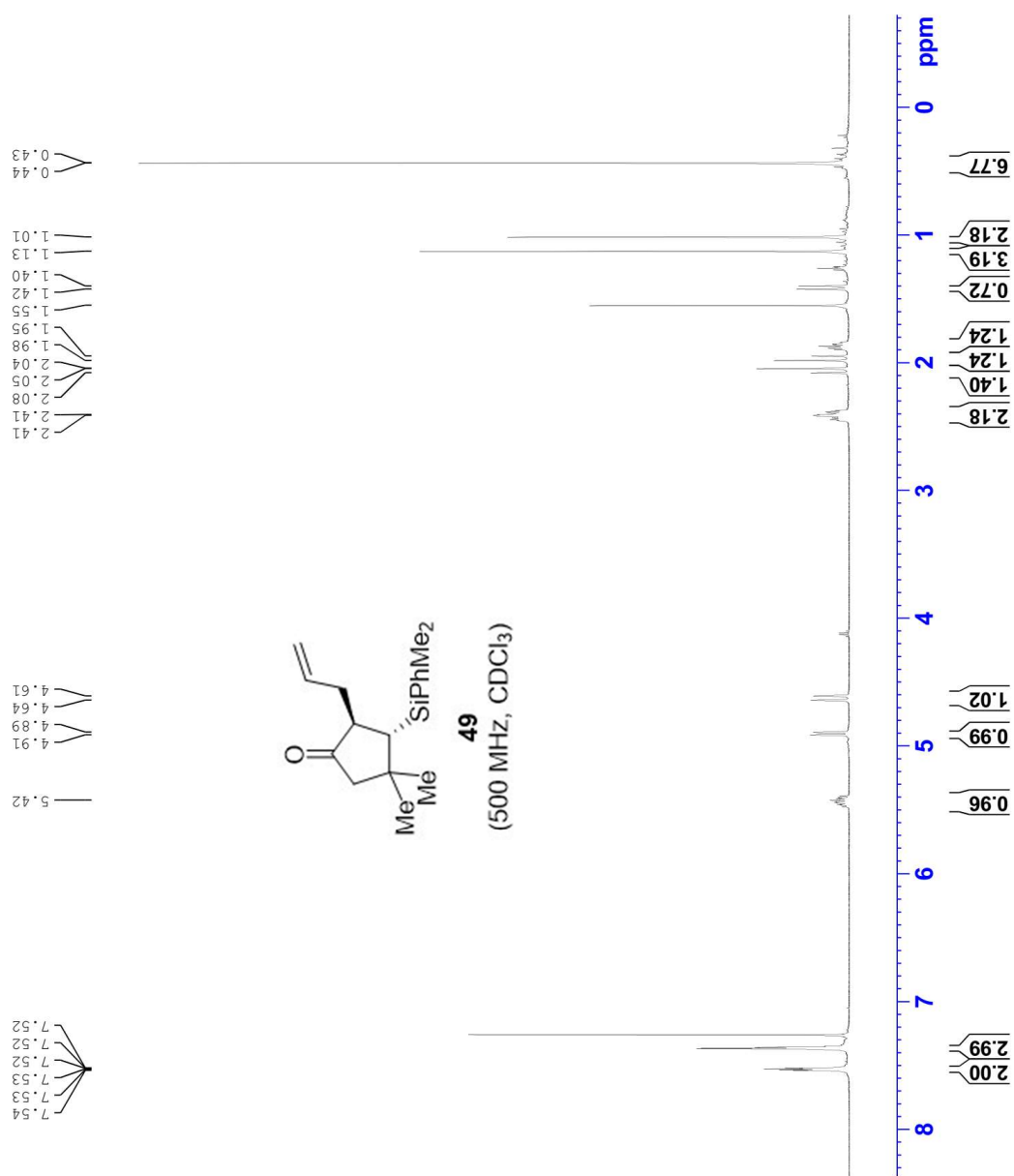


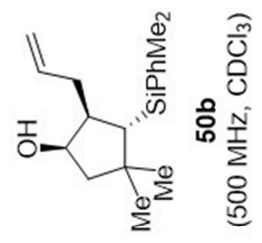
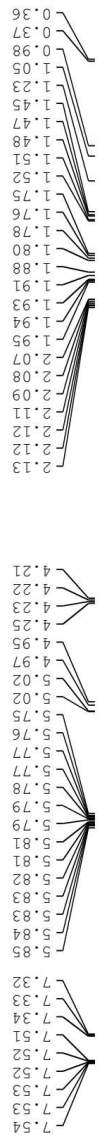
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 RG 196.79
 DW 50.000 usec
 DE 10.00 usec
 TE 294.3 K
 D1 2.00000000 sec
 TD0 1

==== CHANNEL f1 =====
 SF01 500.1330885 MHz
 NUC1 1H
 P1 9.90 usec
 PLW1 12.19999981 W

F2 - Processing parameters
 SI 65536
 SF 500.1300134 MHz
 EM EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00





Current Data Parameters
 NAME yh-8-62-1
 EXPNO 10
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20200704
 Time_ 17.23 h
 INSTRUM spect
 PROBHD z140678_0010 ()
 PULPROG zg30
 TD 38460
 SOLVENT CDCl3

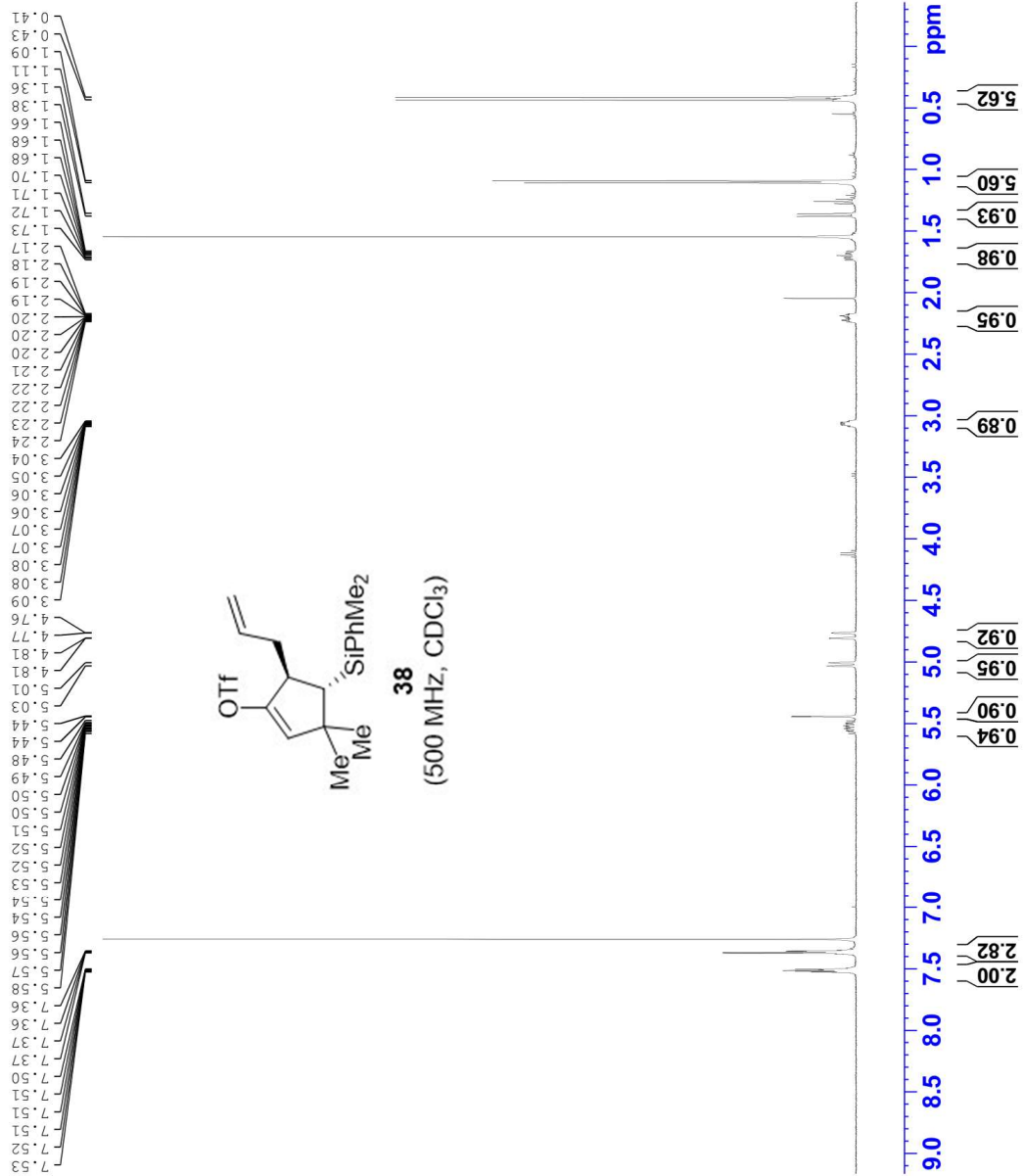
NS 8
 DS 0
 SWH 6410.256 Hz
 FIDRES 0.333347 Hz
 AQ 2.9998801 sec
 RG 211.9
 DW 78.000 usec
 DE 6.50 usec
 TE 297.1 K
 D1 1.5000000 sec
 TD0 1
 SF01 400.1528010 MHz
 NUC1 1H
 P0 2.28 usec
 P1 6.85 usec
 PLW1 31.98900032 W

F2 - Processing parameters
 SI 131072
 SF 400.1500096 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00

Current Data Parameters
 NAME yh-8-119
 EXPNO 10
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20200810
 Time_ 20.29 h
 INSTRUM spect
 PROBHD z140678_0010 (zg30
 PULPROG 38460
 TD 38460
 SOLVENT CDCl3
 NS 8
 DS 0
 SWH 6410.256 Hz
 FIDRES 0.333347 Hz
 AQ 2.9998801 sec
 RG 211.9
 DW 78.000 usec
 DE 6.50 usec
 TE 297.2 K
 D1 1.50000000 sec
 TD0 1
 SF01 400.1528010 MHz
 NUC1 1H
 P0 2.28 usec
 P1 6.85 usec
 PLW1 31.98900032 W

F2 - Processing parameters
 SI 131072
 SF 400.1500098 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00

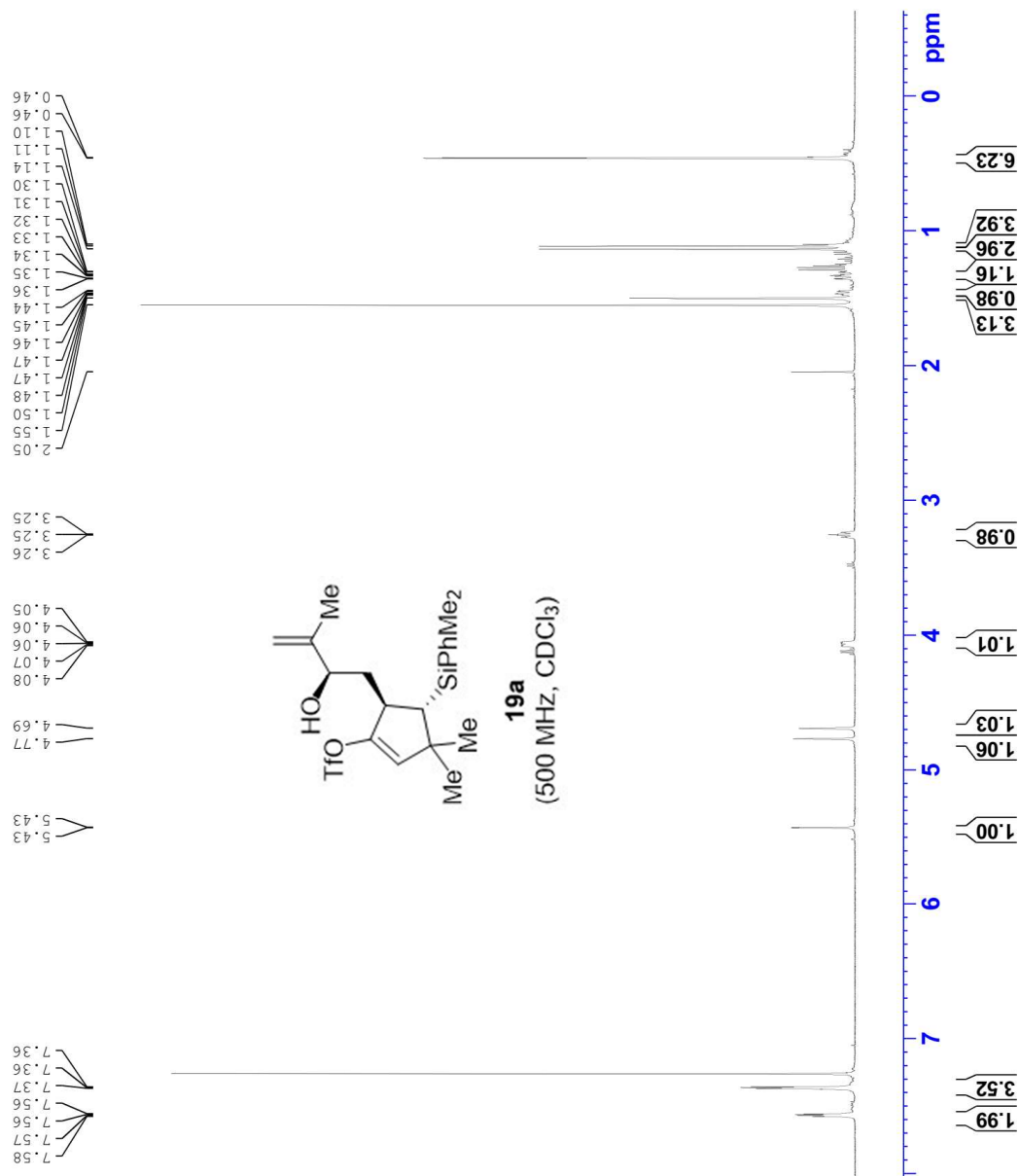


Current Data Parameters
 NAME Yh-7-94-1
 EXPNO 1
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20191124
 Time_ 19.54
 INSTRUM spect
 PROBHD 5 mm PATXI 1H/
 PULPROG zg
 TD 5998
 SOLVENT CDCl3
 NS 6
 DS 0
 SWH 10000.000 Hz
 FIDRES 0.166672 Hz
 AQ 2.9999001 sec
 RG 196.79
 DW 50.000 usec
 DE 10.00 usec
 TE 294.6 K
 DI 4.0000000 sec
 TD0 1

==== CHANNEL f1 =====
 SFO1 500.1330885 MHz
 NUC1 1H
 P1 3.30 usec
 PLW1 12.19999981 W

F2 - Processing parameters
 SI 65536
 SF 500.1300134 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00

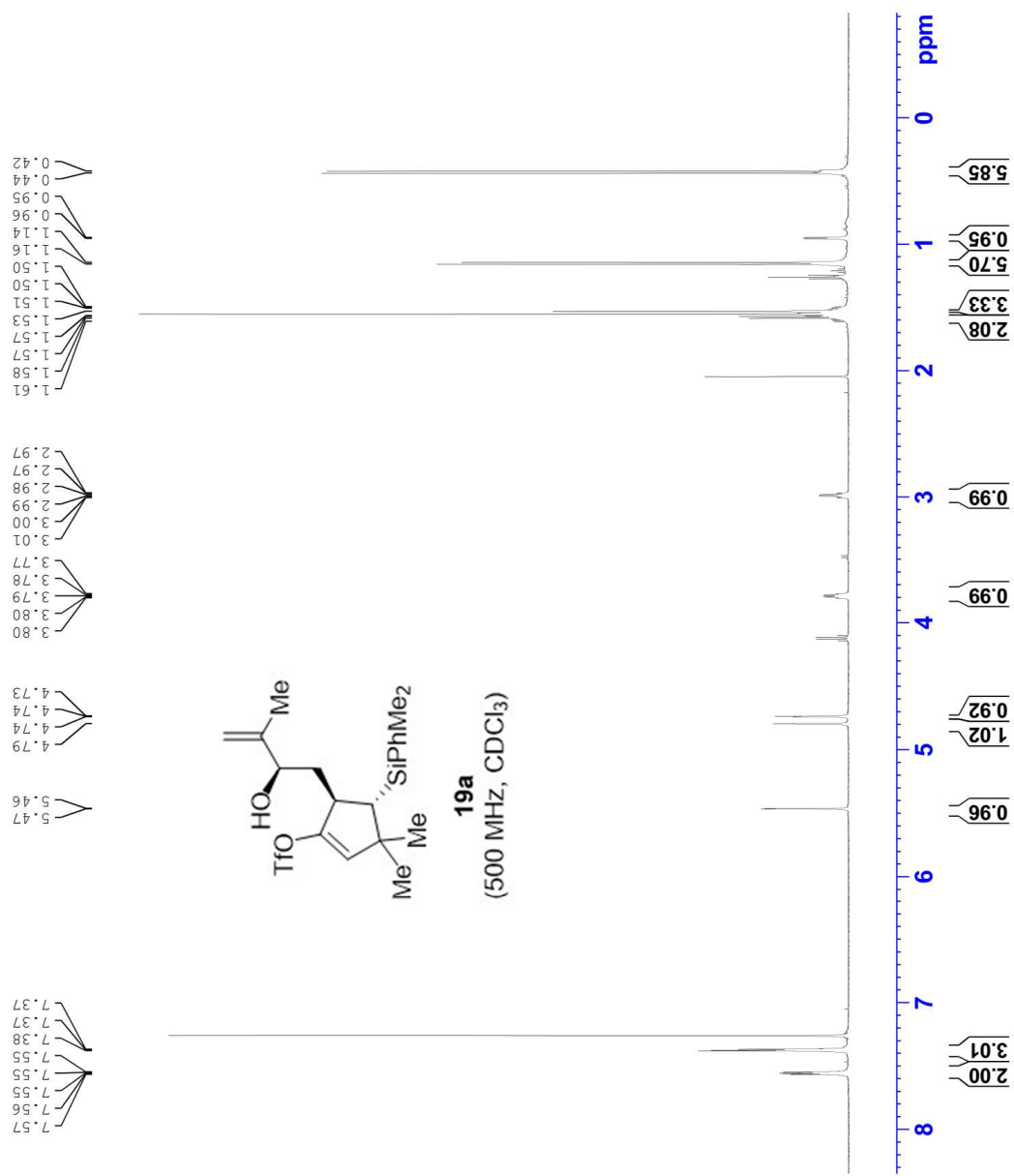


Current Data Parameters
 NAME yh-7-94-2
 EXPNO 1
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20191124
 Time 19.58
 INSTRUM spect
 PROBHD 5 mm PATXI 1H/
 PULPROG zg
 TD 59998
 SOLVENT CDCl3
 NS 6
 DS 0
 SWH 10000.000 Hz
 FIDRES 0.166672 Hz
 AQ 2.9999001 sec
 RG 196.79
 DW 50.000 usec
 DE 10.000 usec
 TE 294.7 K
 D1 4.0000000 sec
 TDO 1

=====
 CHANNEL f1
 SF01 500.1330885 MHz
 NUC1 1H
 P1 3.30 usec
 PLW1 12.19999981 W

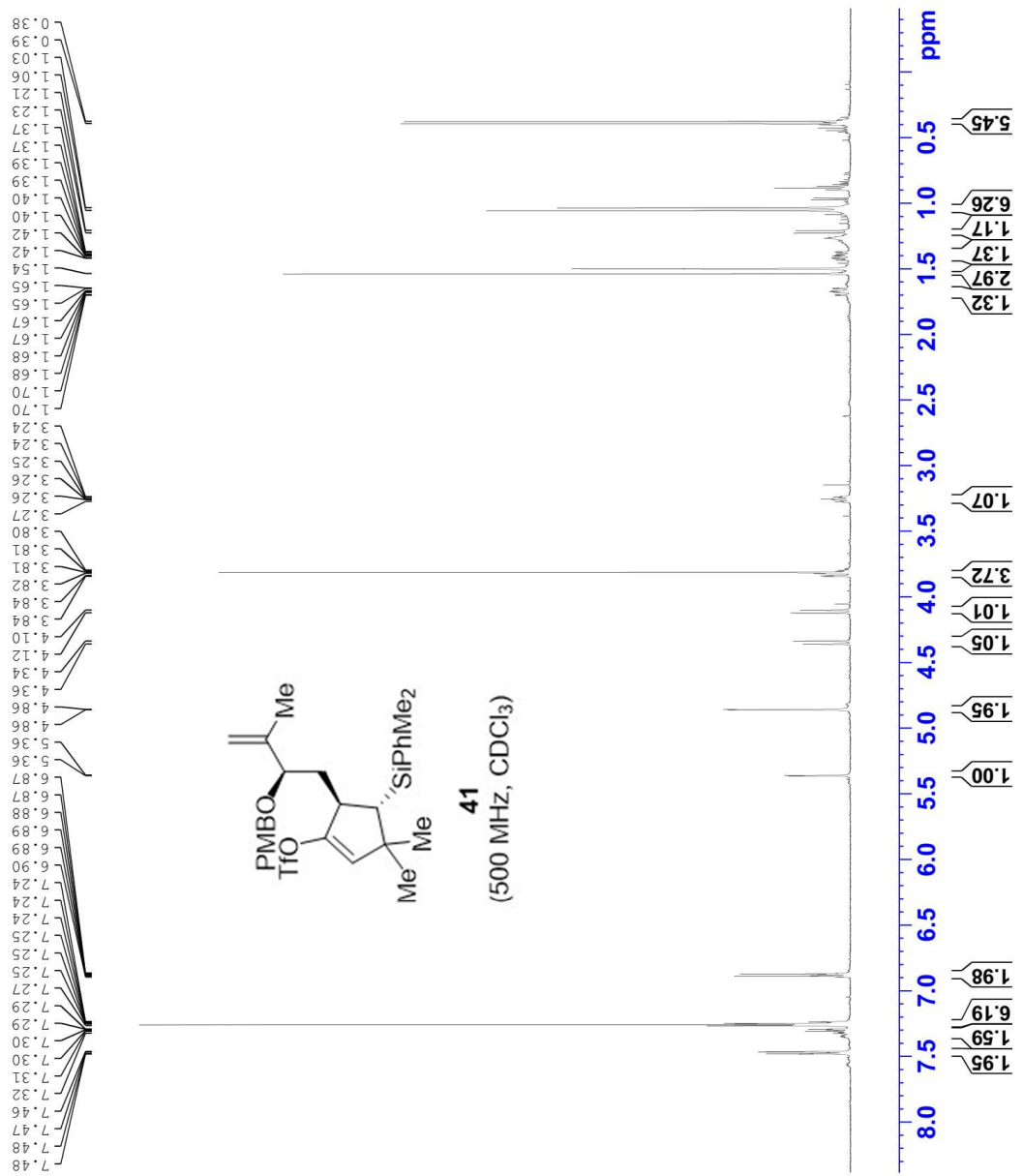
F2 - Processing parameters
 SI 65536
 SF 500.1300135 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00



Current Data Parameters
 NAME yh-10-106
 EXPNO 1
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20210806
 Time 17.17 h
 INSTRUM spect
 PROBHD Z800701_0094 (z30)
 PULPROG zg30
 TD 48076
 SOLVENT CDC13
 NS 8
 DS 0
 SWH 8012.820 Hz
 FIDRES 0.333340 Hz
 AQ 2.9999423 sec
 RG 196.79
 DW 62.400 usec
 DE 6.50 usec
 TE 298.2 K
 D1 1.50000000 sec
 TD0 1
 SFO1 500.1335009 MHz
 NUC1 1H
 P0 3.37 usec
 P1 10.10 usec
 PLW1 25.00300026 W

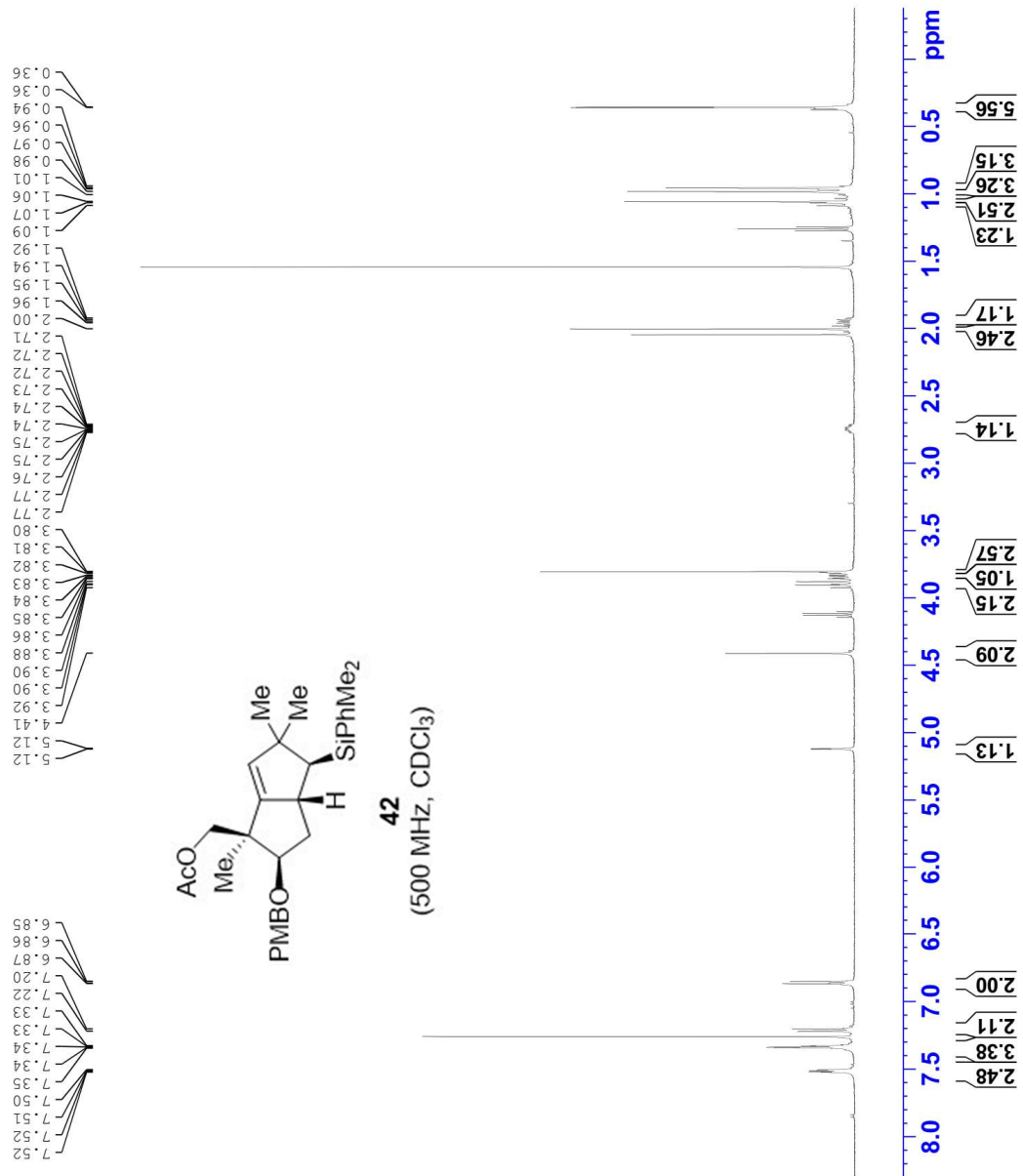
F2 - Processing parameters
 SI 131072
 SF 500.1300130 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00



Current Data Parameters
 NAME yh-10-116
 EXPNO 1
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20210824
 Time 18.04 h
 INSTRUM spect
 PROBHD z113652_0187 ()
 PULPROG zg30
 TD 48076
 SOLVENT CDC13
 NS 8
 DS 0
 SWH 8012.820 Hz
 FIDRES 0.333340 Hz
 AQ 2.9999423 sec
 RG 105.66
 DW 62.400 usec
 DE 6.50 usec
 TE 298.1 K
 D1 1.5000000 sec
 TD0 1
 SF01 499.8734991 MHz
 NUC1 1H
 P0 3.20 usec
 F1 9.60 usec
 PLW1 20.98900032 W

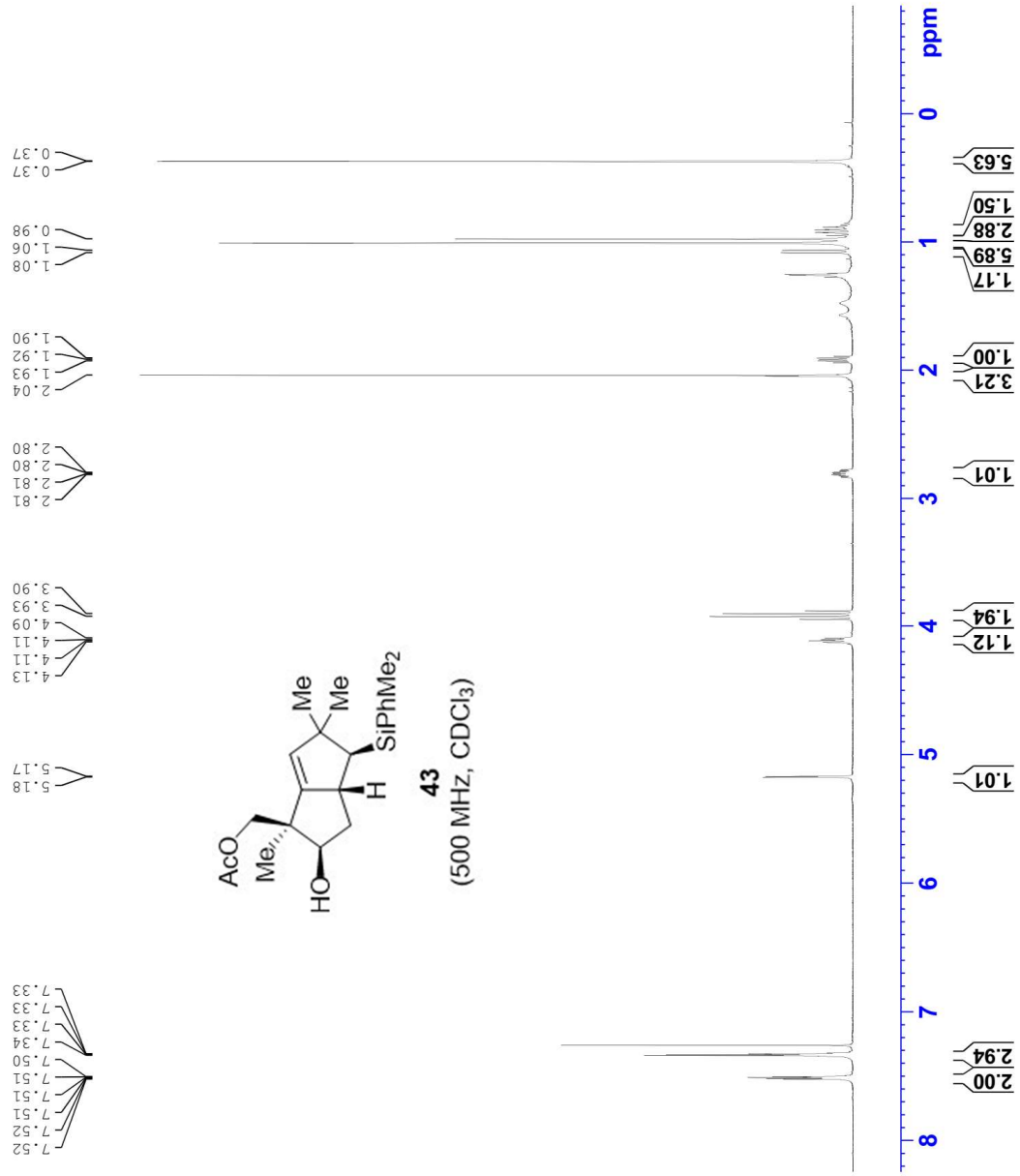
F2 - Processing parameters
 SI 131072
 SF 499.8700124 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00



Current Data Parameters
 NAME Yh-7-110
 EXPNO 1
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20191205
 Time_ 17.30 h
 INSTRUM spect
 PROBHD Z113652_0187 (zg)
 PULPROG 59998
 TD CDC13
 NS 6
 DS 0
 SWH 10000.000 Hz
 FIDRES 0.333344 Hz
 AQ 2.9999001 sec
 RG 86.16
 DW 50.000 usec
 DE 6.50 usec
 TE 295.0 K
 D1 4.0000000 sec
 TD0 1
 SF01 499.8730869 MHz
 NUC1 1H
 P1 3.58 usec
 PLW1 18.2500000 W

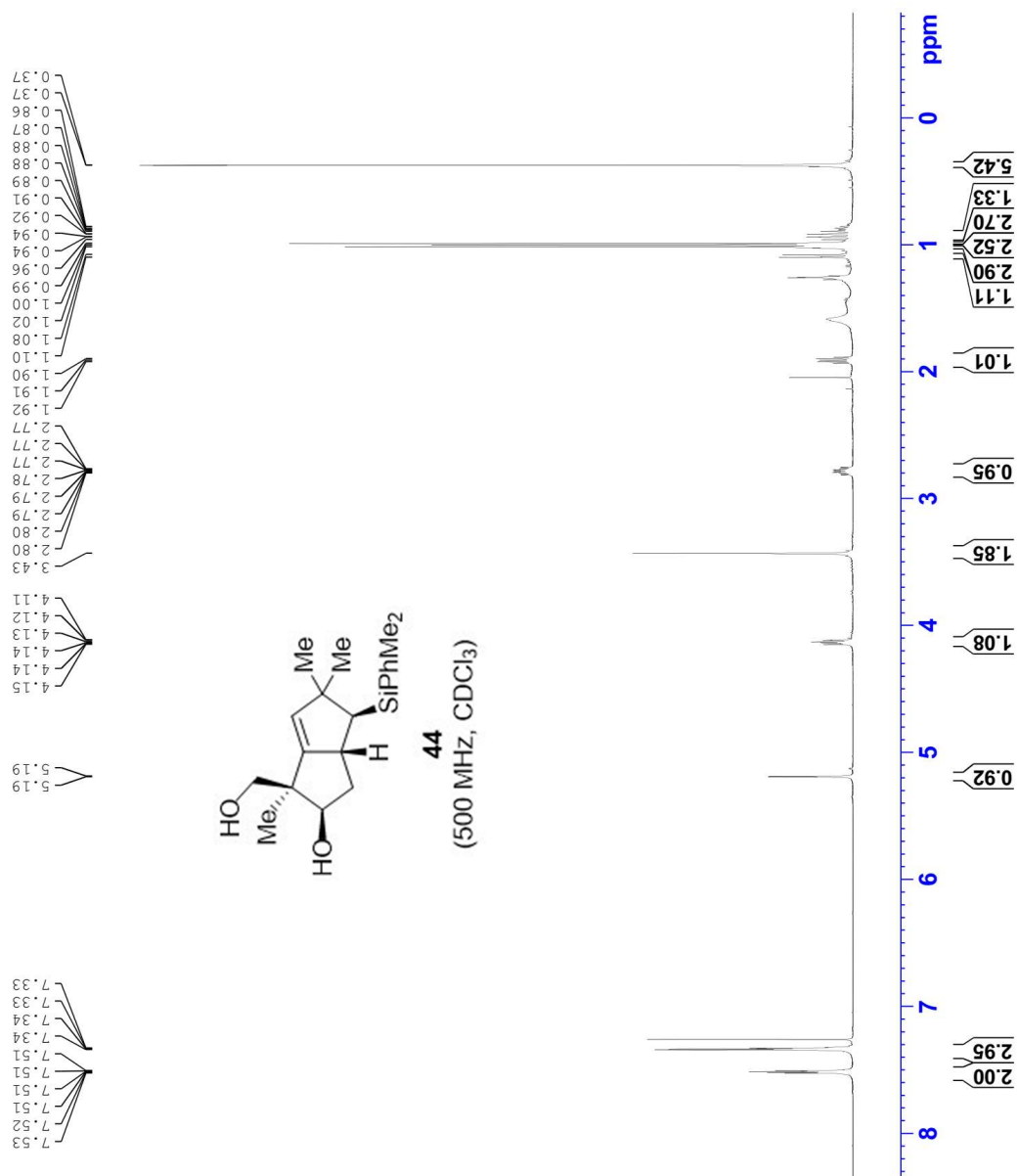
F2 - Processing parameters
 SI 65536
 SF 499.8700124 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00



Current Data Parameters
 NAME Yh-7-148-1
 EXPNO 1
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20200122
 Time_ 21.20 h
 INSTRUM spect
 PROBHD z113652_0187 (ZG
 PULPROG 59998
 TD CDC13
 NS 8
 DS 0
 SWH 10000.000 Hz
 FIDRES 0.333344 Hz
 AQ 2.9999001 sec
 RG 105.66
 DW 50.000 usec
 DE 6.50 usec
 TE 297.0 K
 D1 4.0000000 sec
 TD0 1
 SF01 499.8730869 MHz
 NUC1 1H
 P1 3.58 usec
 PLW1 18.25000000 W

F2 - Processing parameters
 SI 65536
 SF 499.8700125 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00



Current Data Parameters
 NAME yt-10-122-1-crude-2
 EXPNO 1
 PROCNO 1

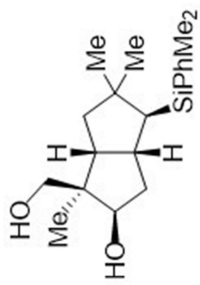
F2 - Acquisition Parameters

Date 20210831
 Time 11.10 h
 INSTRUM spect
 PROBHD z800701_0094 (z930)
 PULPROG zg30
 TD 48076
 SOLVENT CDCl3
 NS 8
 DS 0
 SWH 8012.820 Hz
 FIDRES 0.333340 Hz
 AQ 2.9999423 sec
 RG 196.79
 DW 62.400 usec
 DE 6.50 usec
 TE 298.1 K
 D1 1.50000000 sec
 TD0 1
 SF01 500.1335009 MHz
 NUC1 1H
 P0 3.37 usec
 P1 10.10 usec
 PLW1 25.00300026 W

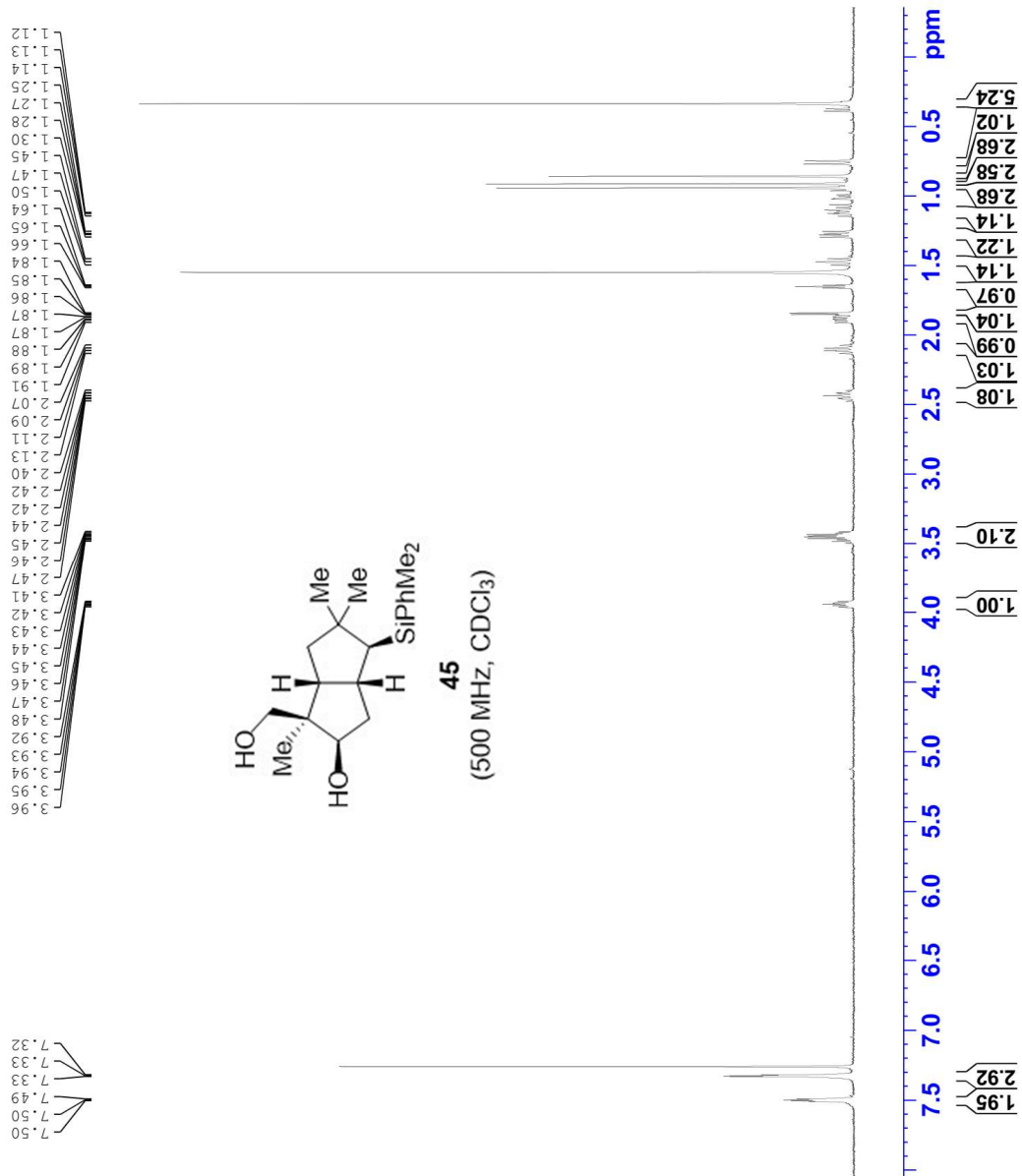
F2 - Processing parameters

SI 131072
 SF 500.1300128 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00

7.50
7.49
7.49
7.33
7.33
7.33
3.96
3.95
3.94
3.92
3.48
3.47
3.46
3.45
3.44
3.43
3.42
3.41
2.47
2.46
2.45
2.44
2.42
2.42
2.40
2.13
2.11
2.09
1.91
1.89
1.88
1.87
1.87
1.86
1.85
1.84
1.66
1.65
1.64
1.50
1.47
1.45
1.30
1.28
1.27
1.25
1.14
1.13



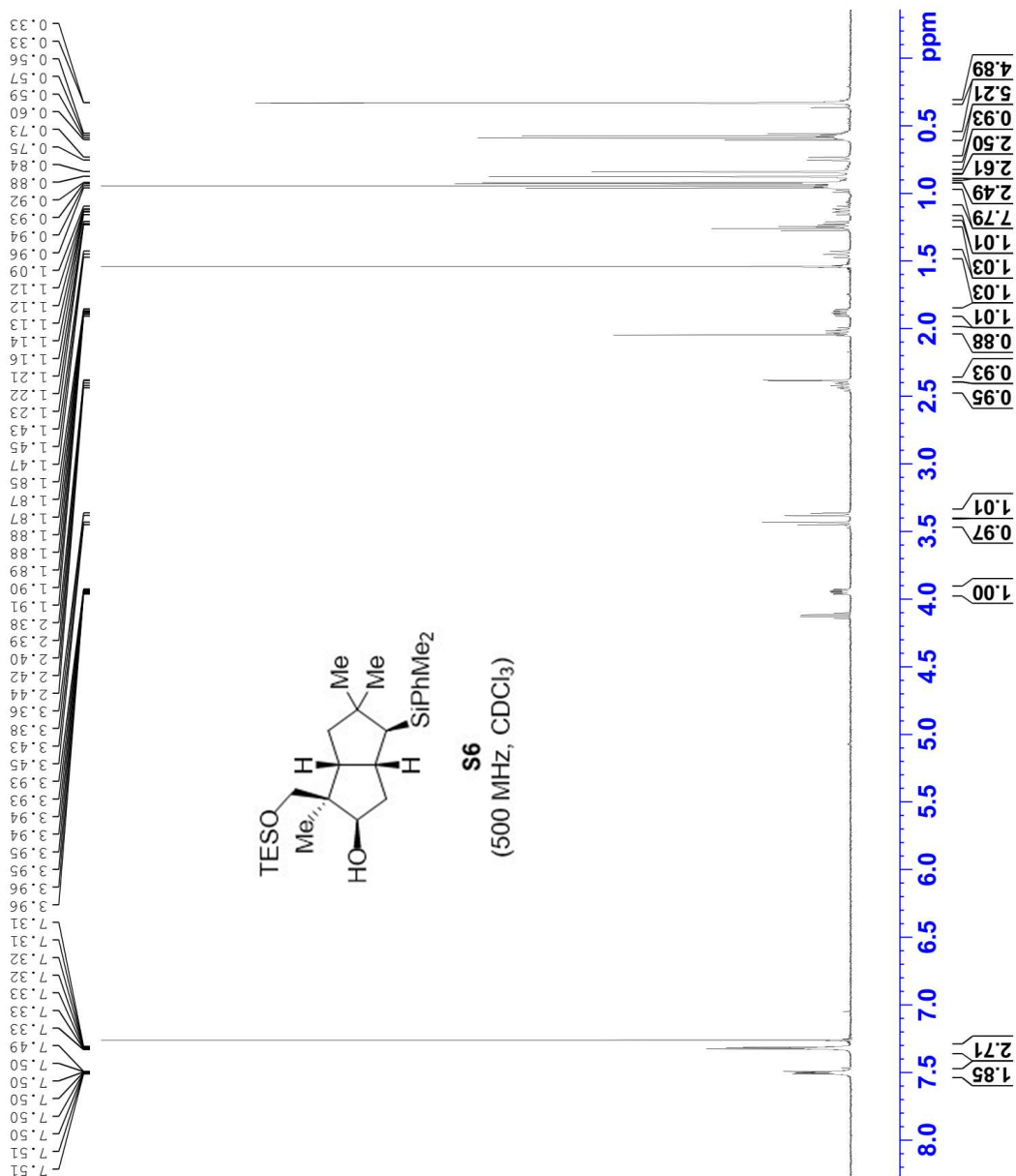
308



Current Data Parameters
 NAME yh-10-123-2
 EXPNO 1
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20210901
 Time_ 16.29 h
 INSTRUM spect
 PROBHD z800701_0094 (zg30)
 PULPROG 48076
 SOLVENT CDCl3
 NS 8
 DS 0
 SWH 8012.820 Hz
 FIDRES 0.333340 Hz
 AQ 2.9999423 sec
 RG 196.79
 DM 62.400 usec
 DE 6.50 usec
 TE 296.2 K
 D1 1.5000000 sec
 TD0 1
 SF01 500.1335009 MHz
 NUC1 1H
 P0 3.37 usec
 P1 10.10 usec
 PLW1 25.00300026 W

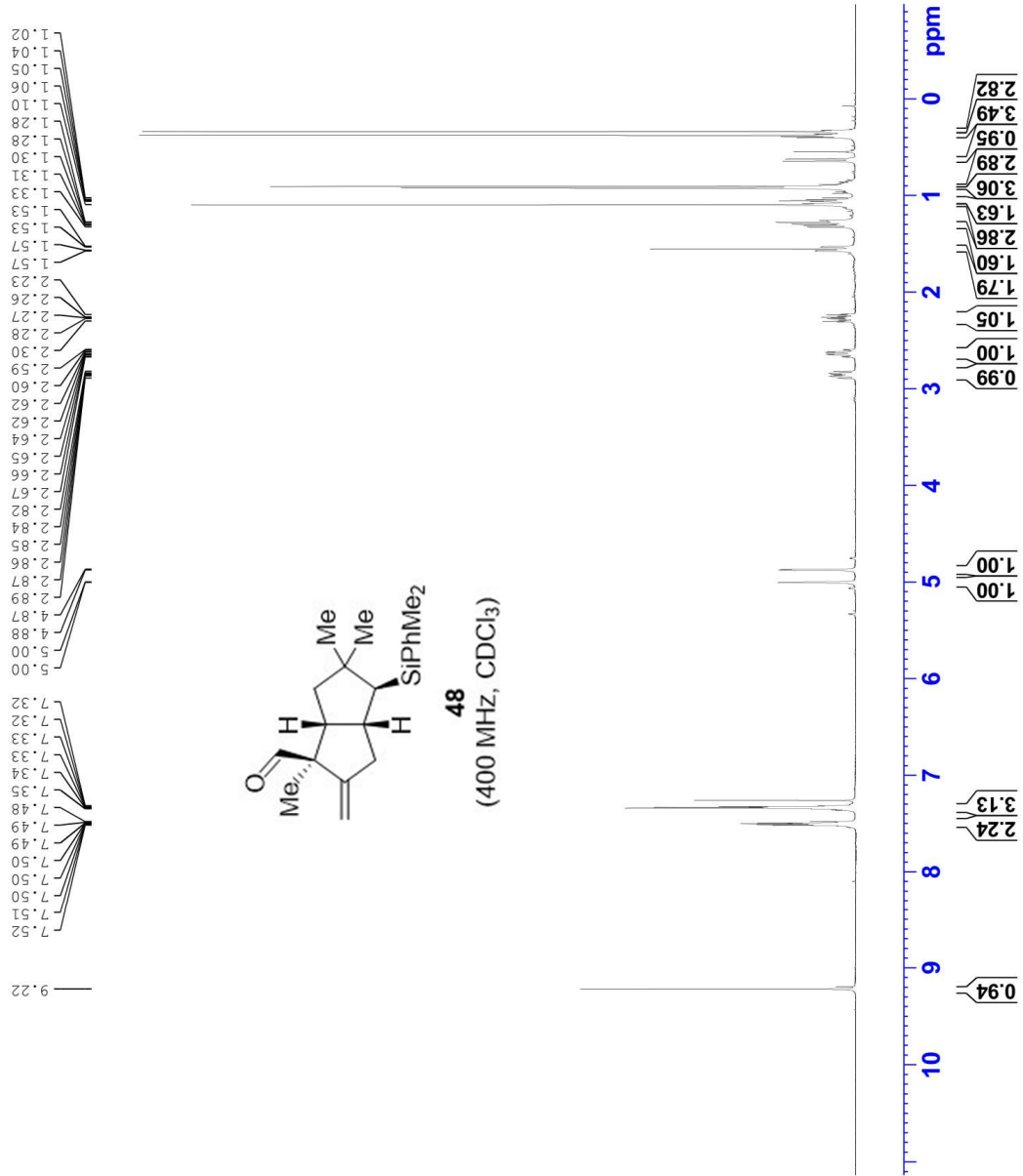
F2 - Processing Parameters
 SI 131072
 SF 500.1300126 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00



Current Data Parameters
 NAME Yh-9-47
 EXPNO 12
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20201022
 Time_ 1.41 h
 INSTRUM spect
 PROBHD Z140678_0010 (z930
 PULPROG 38460
 TD 8
 SOLVENT CDCl3
 NS 8
 DS 0
 SWH 6410.256 Hz
 FIDRES 0.333347 Hz
 AQ 2.9998801 sec
 RG 103.18
 DW 78.000 usec
 DE 6.50 usec
 TE 297.2 K
 D1 1.50000000 sec
 TD0 1
 SFO1 400.1528010 MHz
 NUC1 1H
 P0 2.28 usec
 P1 6.85 usec
 PLW1 31.98900032 W

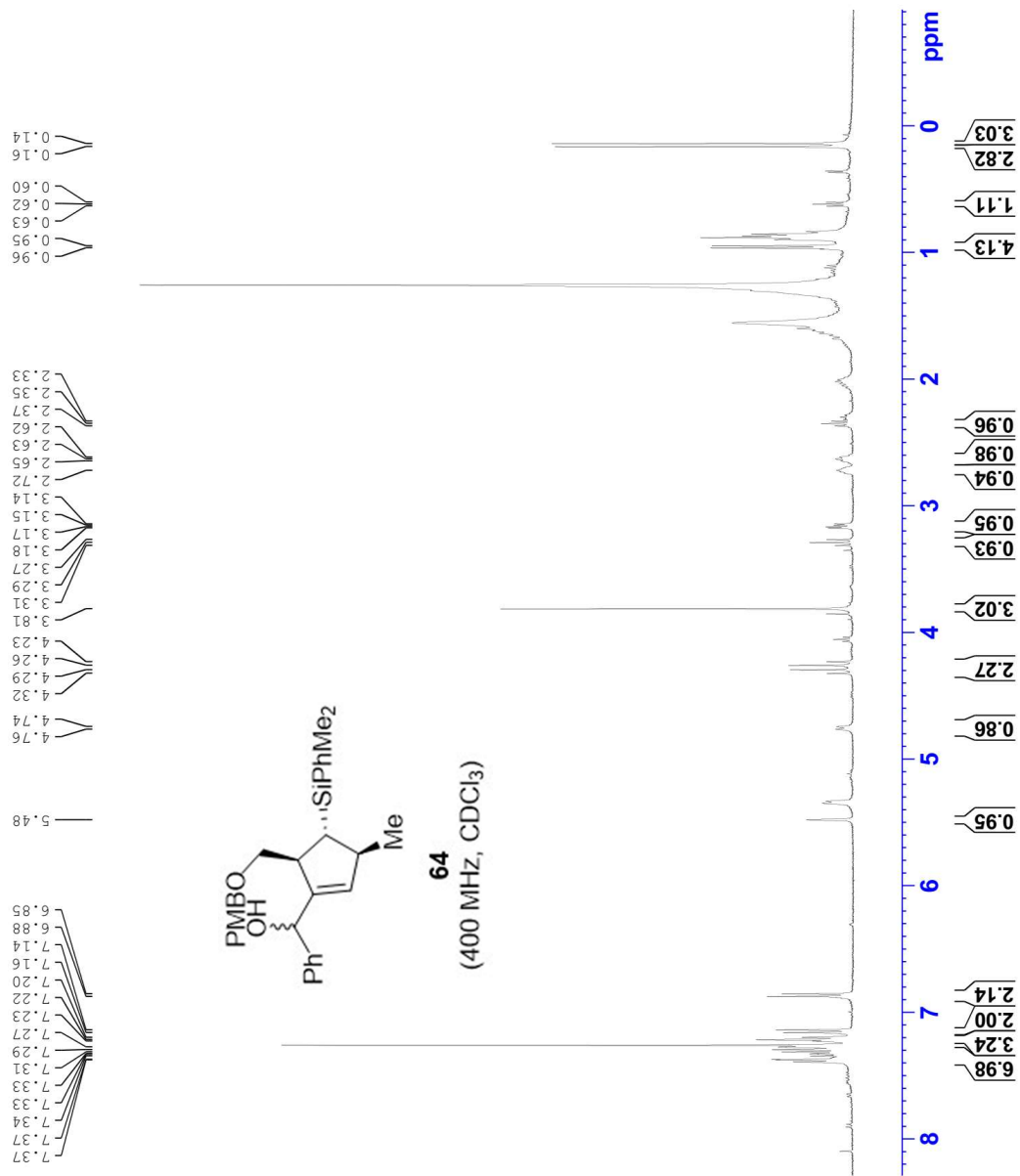
F2 - Processing parameters
 SI 131072
 SF 400.1500096 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00



Current Data Parameters
 NAME yh-10-5-a
 EXPNO 10
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20211020
 Time_ 18.51 h
 INSTRUM spect
 PROBHD Z140678_0010 ()
 PULPROG zg30
 TD 38460
 SOLVENT CDCl3
 NS 8
 DS 0
 SWH 6410.256 Hz
 FIDRES 0.333347 Hz
 AQ 2.9998801 sec
 RG 165.16
 DW 78.000 usec
 DE 6.50 usec
 TE 298.2 K
 D1 1.50000000 sec
 TD0 1
 SFO1 400.1528010 MHz
 NUC1 1H
 P0 2.28 usec
 P1 6.85 usec
 PLW1 31.98900032 W

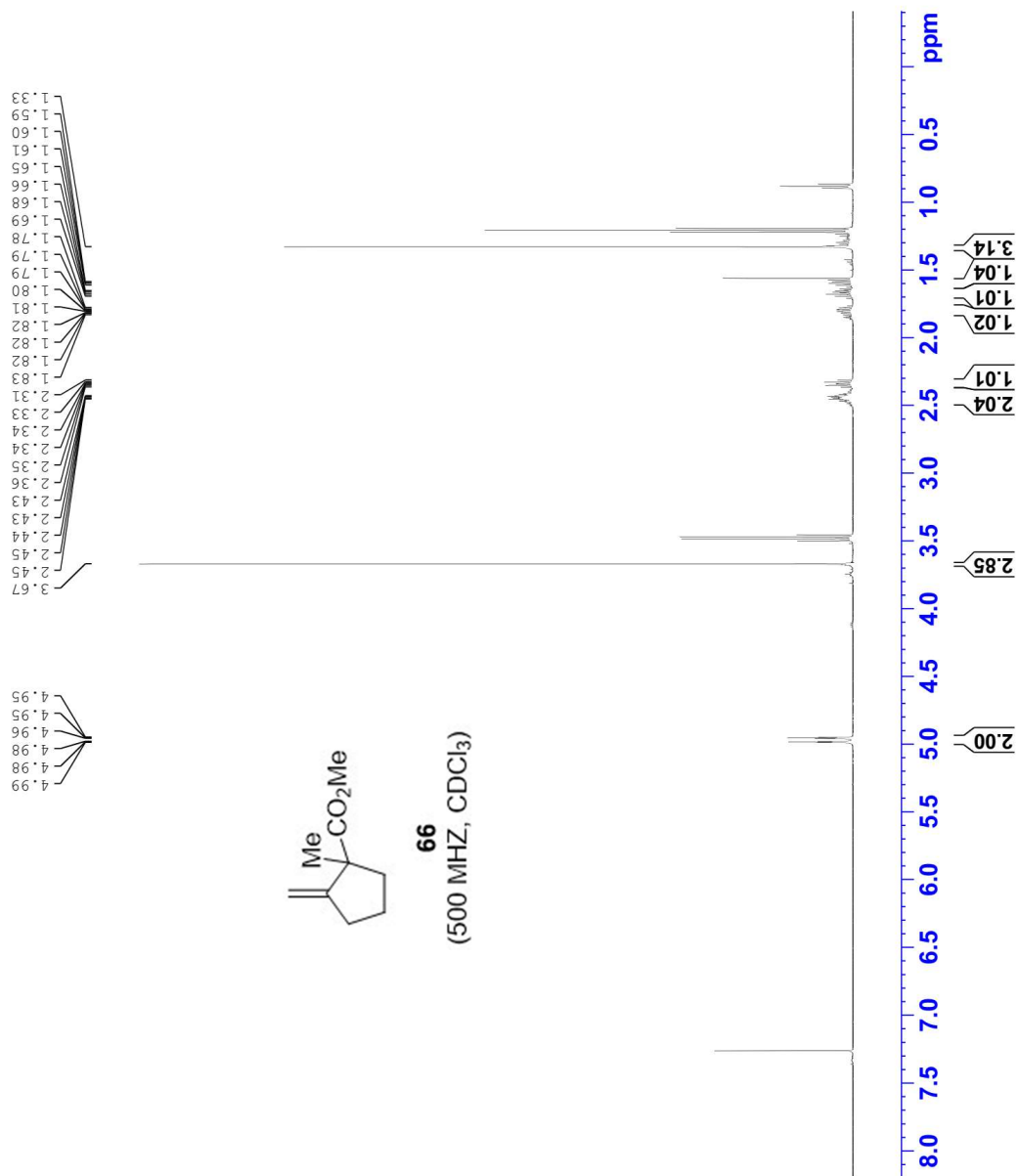
F2 - Processing parameters
 SI 131072
 SF 400.1500099 MHz
 EM EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00



Current Data Parameters
 NAME Yh-10-41
 EXPNO 1
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20210512
 Time_ 14.19 h
 INSTRUM spect
 PROBHD Z800701_0094 (
 PULPROG zg30
 TD 48076
 SOLVENT CDCl3
 NS 8
 DS 0
 SWH 8012.820 Hz
 FIDRES 0.333340 Hz
 AQ 2.9999423 sec
 RG 196.79
 DW 62.400 usec
 DE 6.50 usec
 TE 298.2 K
 D1 1.50000000 sec
 TD0 1
 SFO1 500.1335009 MHz
 NUC1 1H
 P0 3.37 usec
 P1 10.10 usec
 PLW1 25.00300026 W

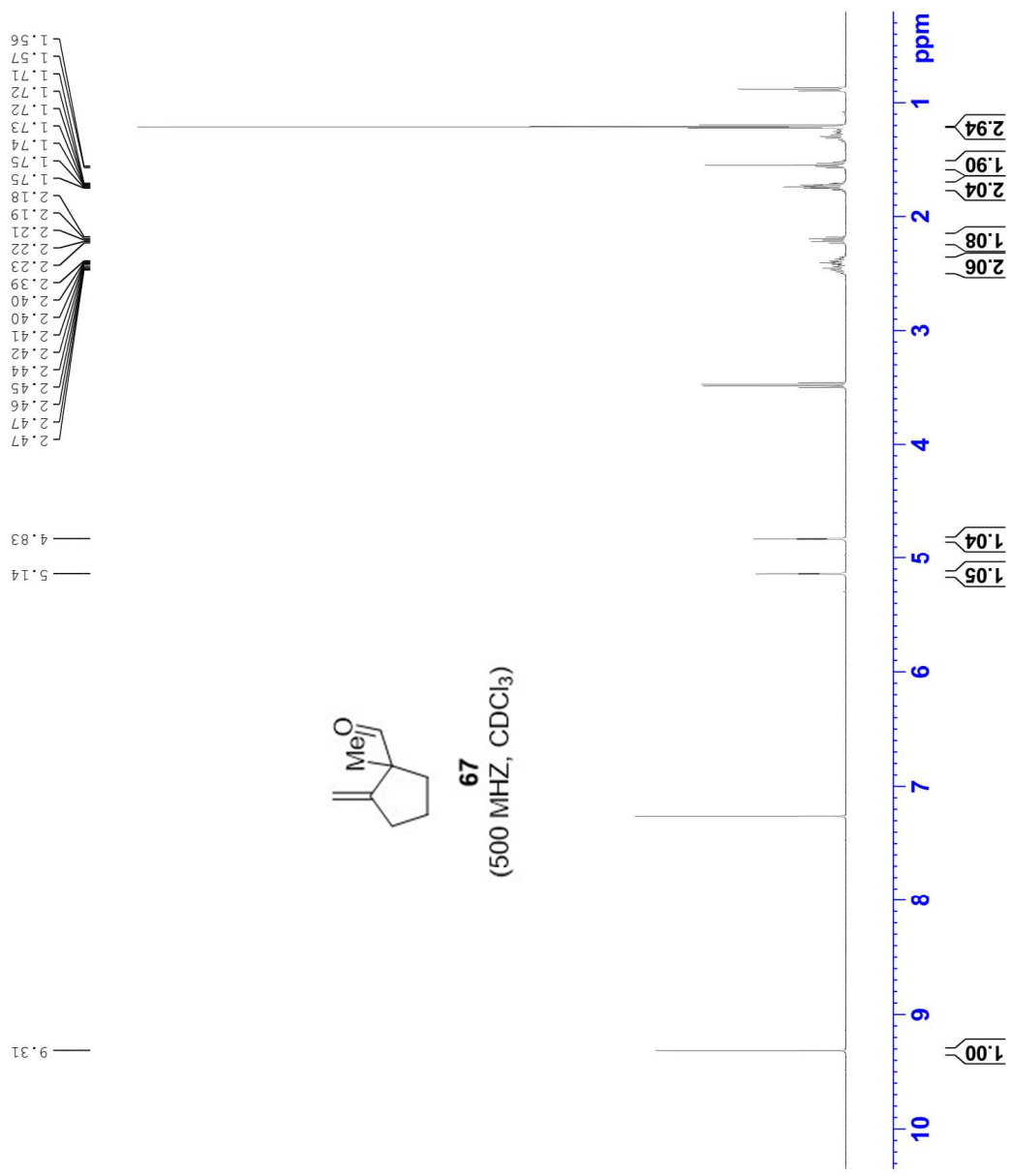
F2 - Processing parameters
 SI 131072
 SF 500.1300125 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00



Current Data Parameters
 NAME yh-10-44
 EXPNO 1
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20210514
 Time_ 12.56 h
 INSTRUM spect
 PROBHD z800701_0094 (z930)
 PULPROG zg30
 TD 48076
 SOLVENT CDCl3
 NS 8
 DS 0
 SWH 8012.820 Hz
 FIDRES 0.333340 Hz
 AQ 2.9999423 sec
 RG 196.79
 DW 62.400 usec
 DE 6.50 usec
 TE 298.1 K
 D1 1.50000000 sec
 TD0 1
 SF01 500.1335009 MHz
 NUC1 1H
 P0 3.37 usec
 P1 10.10 usec
 PLW1 25.00300026 W

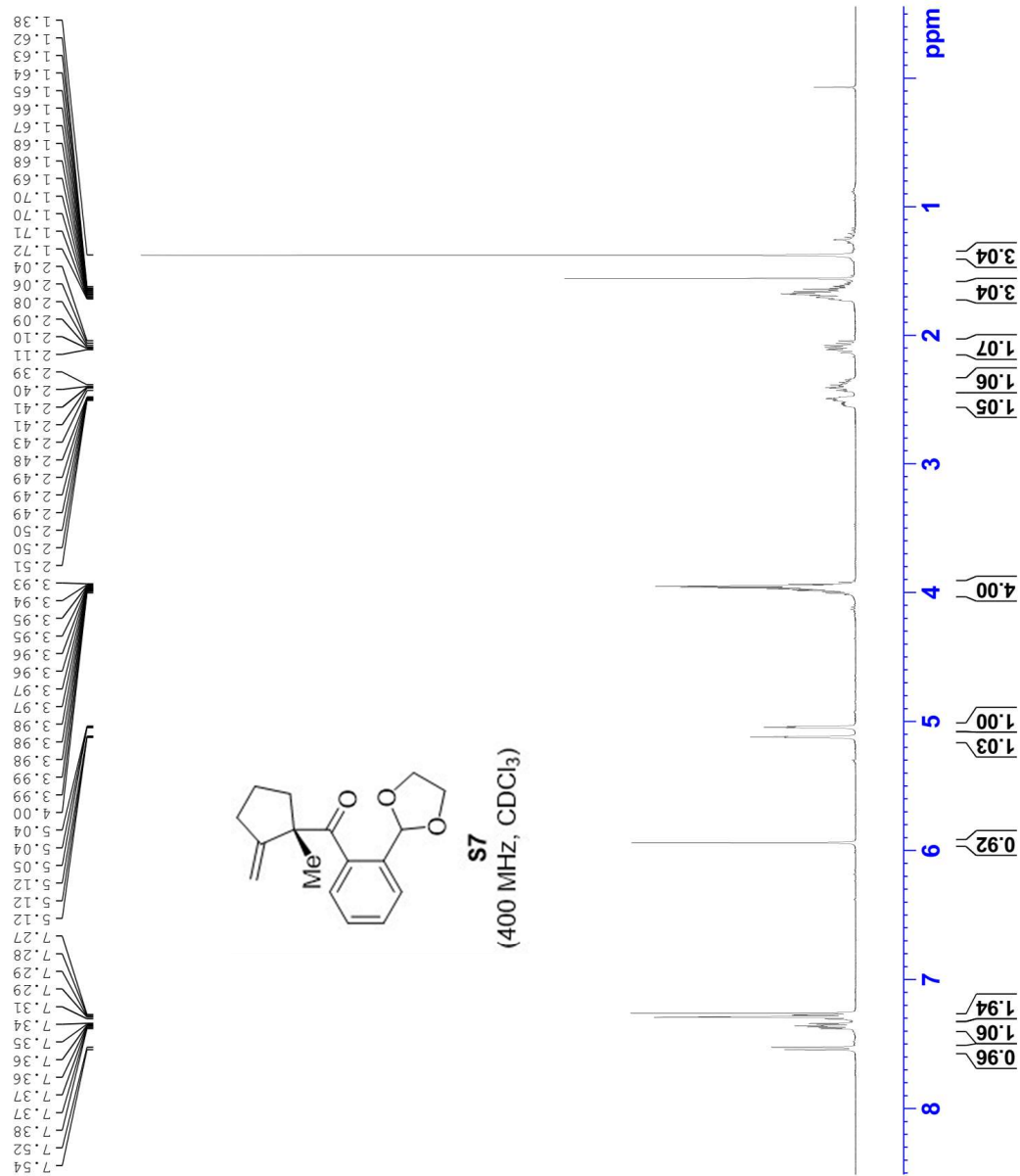
F2 - Processing Parameters
 SI 131072
 SF 500.1300125 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00



Current Data Parameters
 NAME Yh-10-47
 EXPNO 10
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20210518
 Time_ 14.35 h
 INSTRUM spect
 PROBHD Z140678_0010 (Z930
 PULPROG 38460
 TD 38460
 SOLVENT CDCl3
 NS 8
 DS 0
 SWH 6410.256 Hz
 FIDRES 0.333347 Hz
 AQ 2.9998801 sec
 RG 143.42
 DW 78.000 usec
 DE 6.50 usec
 TE 298.2 K
 D1 1.50000000 sec
 TD0 1
 SFO1 400.1528010 MHz
 NUC1 1H
 P0 2.28 usec
 P1 6.85 usec
 PLW1 31.98900032 W

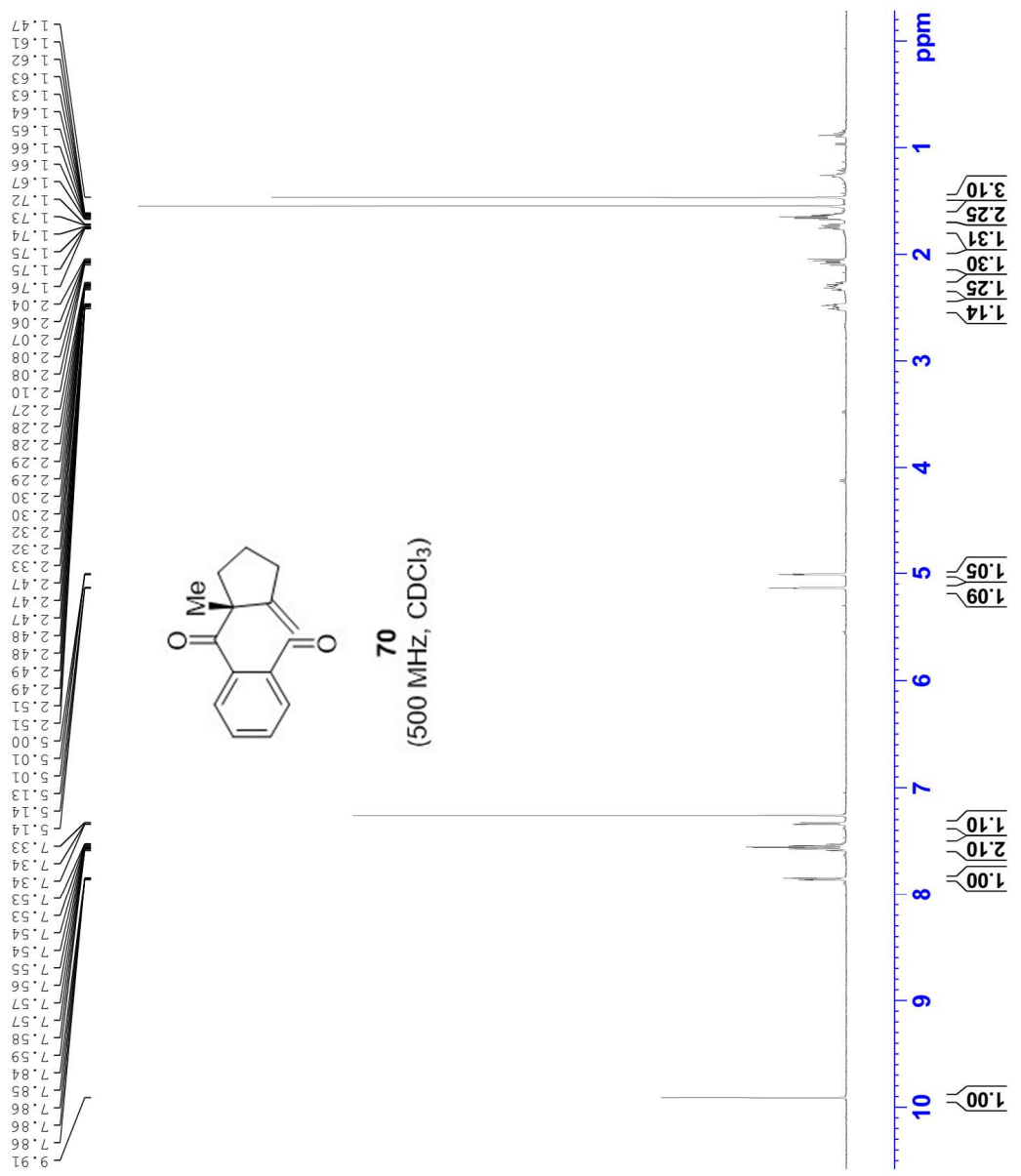
F2 - Processing parameters
 SI 131072
 SF 400.1500097 MHz
 WDM EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00



Current Data Parameters
 NAME Yh-10-49
 EXPNO 1
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20210520
 Time_ 11.26 h
 INSTRUM spect
 PROBHD Z800701_0094 (z930)
 PULPROG zg30
 TD 48076
 SOLVENT CDC13
 NS 8
 DS 0
 SWH 8012.820 Hz
 FIDRES 0.333340 Hz
 AQ 2.9999423 sec
 RG 196.79
 DM 62.400 usec
 DE 6.50 usec
 TE 298.2 K
 D1 1.5000000 sec
 TDO 1
 SFO1 500.1335009 MHz
 NUC1 1H
 P0 3.37 usec
 P1 10.10 usec
 PLW1 25.00300026 W

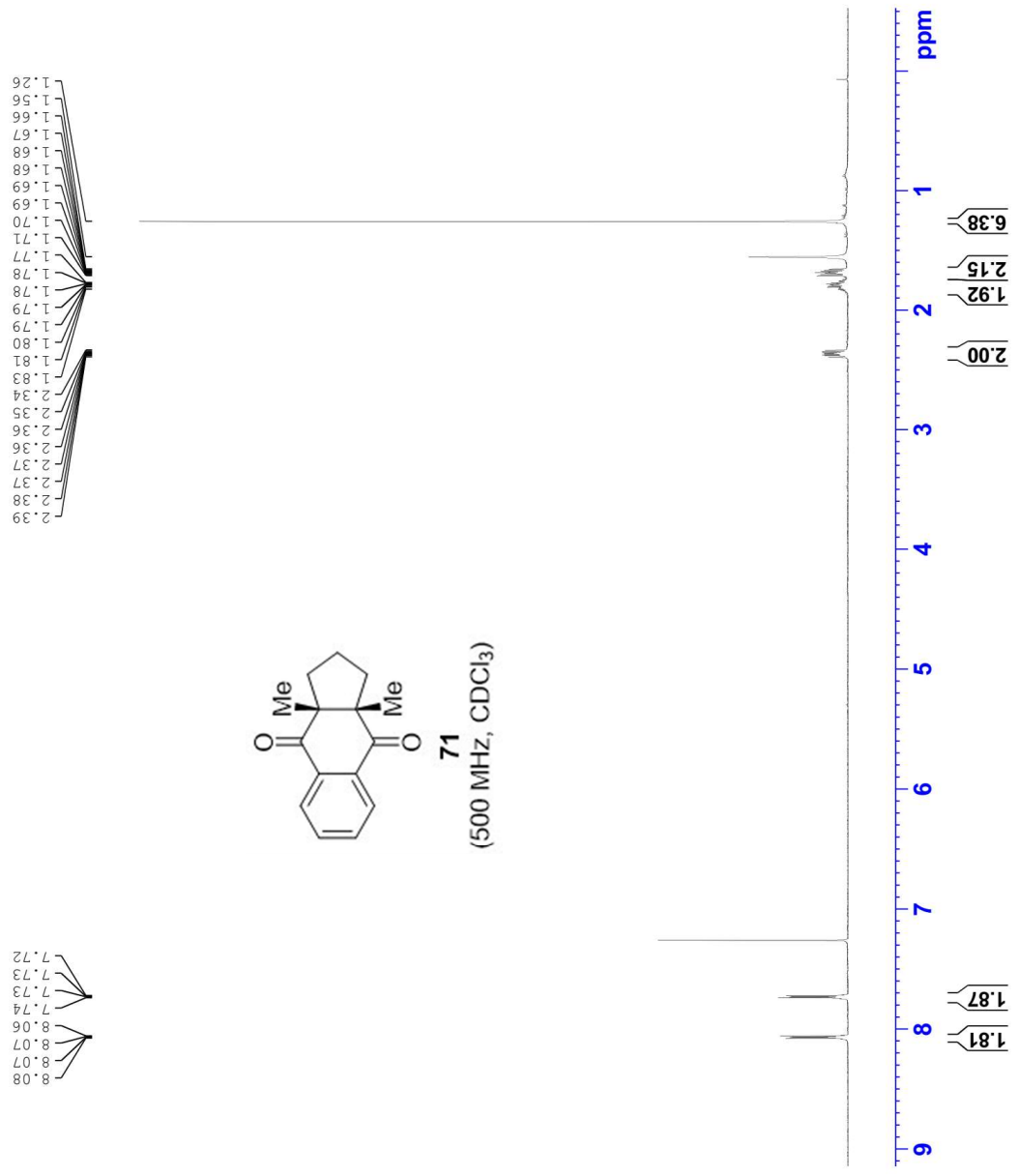
F2 - Processing Parameters
 SI 131072
 SF 500.1300126 MHz
 WDM EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00



Current Data Parameters
 NAME Yh-10-52-1
 EXPNO 1
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20210523
 Time_ 12.45 h
 INSTRUM spect
 PROBHD z800701_0094 (zg30
 PULPROG 48076
 TD 8076
 SOLVENT CDCl3
 NS 8
 DS 0
 SWH 8012.820 Hz
 FIDRES 0.333340 Hz
 AQ 2.9999423 sec
 RG 196.79
 DW 62.400 usec
 DE 6.50 usec
 TE 298.1 K
 D1 1.50000000 sec
 TD0 1
 SFO1 500.1335009 MHz
 NUC1 1H
 P0 3.37 usec
 P1 10.10 usec
 PLW1 25.00300026 W

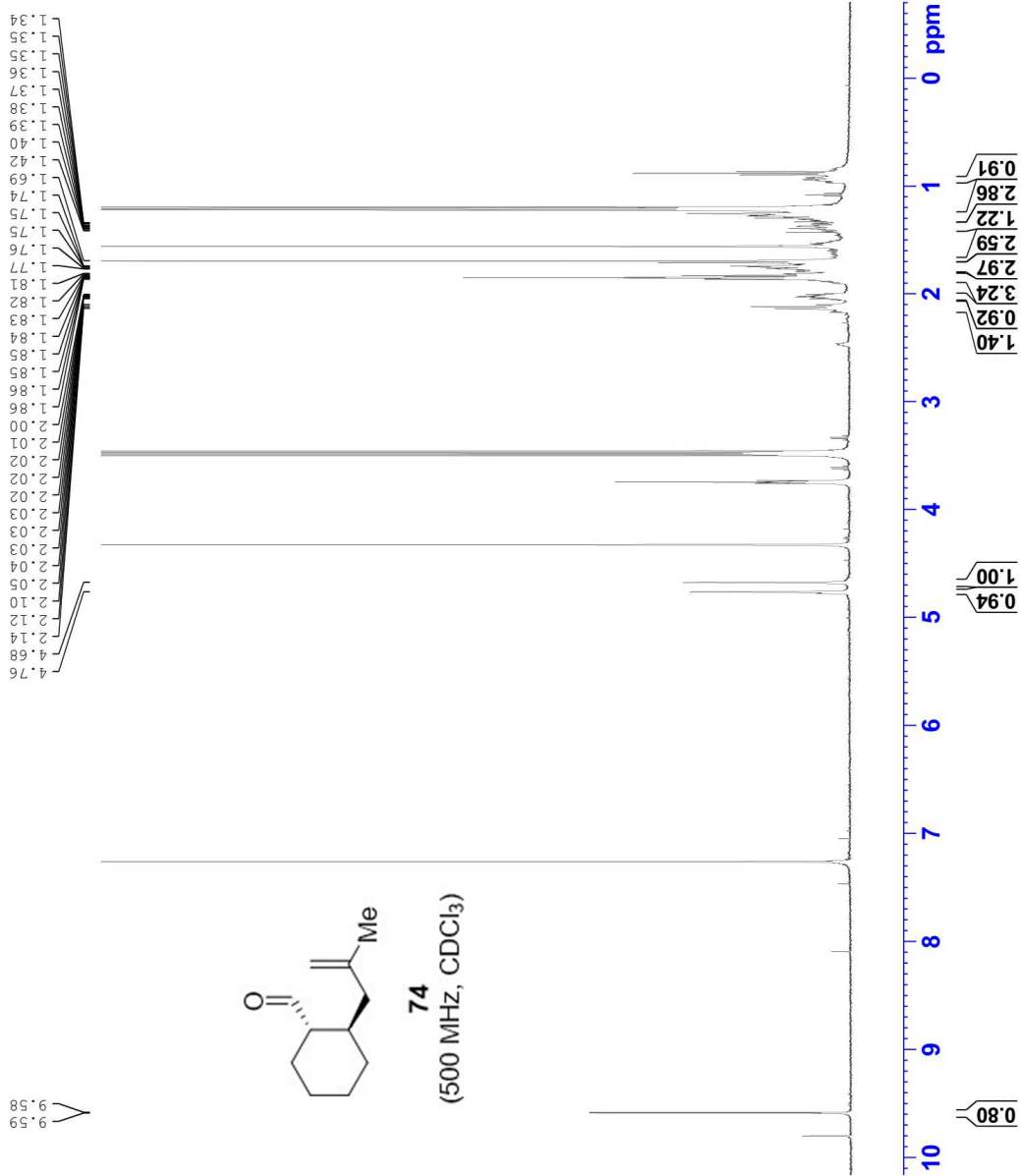
F2 - Processing parameters
 SI 131072
 SF 500.1300126 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00



Current Data Parameters
 NAME yh-10-152-sm
 EXPNO 1
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20210930
 Time_ 17.05 h
 INSTRUM spect
 PROBHD Z113652_0187 (
 PULPROG zg30
 TD 48076
 SOLVENT CDCl3
 NS 8
 DS 0
 SWH 8012.820 Hz
 FIDRES 0.333340 Hz
 AQ 2.9999423 sec
 RG 189.89
 DW 62.400 usec
 DE 6.50 usec
 TE 298.1 K
 D1 1.50000000 sec
 TDO 1
 SFO1 499.8734991 MHz
 NUC1 1H
 P0 3.20 usec
 P1 9.60 usec
 PLW1 20.98900032 W

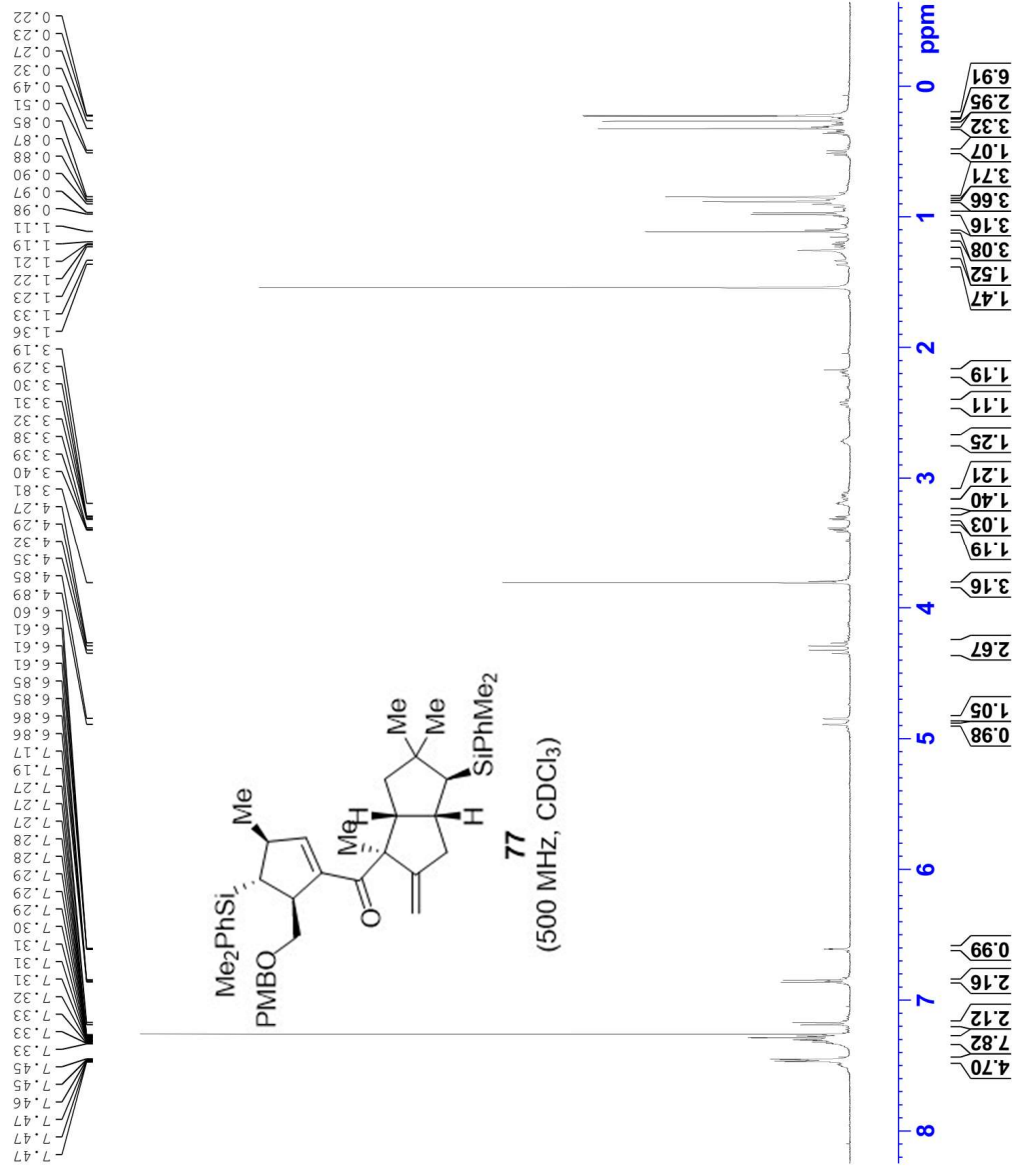
F2 - Processing parameters
 SI 131072
 SF 499.8700123 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00



Current Data Parameters
 NAME yh-11-1
 EXPNO 1
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20211023
 Time 17.03 h
 INSTRUM spect
 PROBHD Z800701_0094 (
 PULPROG zg30
 TD 48076
 SOLVENT CDCl3
 NS 8
 DS 0
 SWH 8012.820 Hz
 FIDRES 0.333340 Hz
 AQ 2.9999423 sec
 RG 196.79
 DW 62.400 usec
 DE 6.50 usec
 TE 298.1 K
 D1 1.5000000 sec
 TD0 1
 SFO1 500.1335009 MHz
 NUC1 1H
 P0 3.37 usec
 F1 10.10 usec
 PLW1 25.00300026 W

F2 - Processing parameters
 SI 131072
 SF 500.1300121 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00



Current Data Parameters
 NAME yh-11-3
 EXPNO 1
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20211025
 Time 19.22 h
 INSTRUM spect
 PROBHD z8c0701_0094 (zg30
 PULPROG 48076
 TD 48076
 SOLVENT CDCl3
 NS 32
 DS 0
 SWH 8012.820 Hz
 FIDRES 0.333340 Hz
 AQ 2.9999423 sec
 RG 196.79
 DW 62.400 usec
 DE 6.50 usec
 TE 298.2 K
 DL 1.5000000 sec
 TD0 1
 SF01 500.1335009 MHz
 NUC1 1H
 P0 3.37 usec
 P1 10.10 usec
 PLW1 25.00300026 W

F2 - Processing parameters
 SI 131072
 SF 500.1300118 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00

